

Roles of Hsf1, Sfl1 and Skn7 in mediating heat-shock protein genes and sustaining asexual cycle, stress tolerance and virulence in *Beauveria bassiana*

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Table S1. Paired primers used for the manipulation of *hsf1*, *sfl1* and *skn7* in *B. bassiana*.

Primer	Paired sequences (5'–3')*	Purpose
Hsf1up-F/R	AAAAAGAATTGCTGCGAGAGTCAAGAGAG / AAAAAAGCTTGTGTCGGTTGTGGGGTTAT	Cloning <i>hsf1</i> 5'-end (1045 bp)
Hsf1dn-F/R	AAAAACTCGAGAGGAGGAGAGCGTGGAGAAG / AAAAACTAGTGGGGTTATGAACACCTCCAG	Cloning <i>hsf1</i> 3'-end (1072 bp)
Sfl1up-F/R	AAAAAGAATTGCTAGATTGTAGCACAGACGGG / AAAAAGAGCTCACGAGGGTGAAGTGAATAAC	Cloning <i>sfl1</i> 5'-end (1583 bp)
Sfl1dn-F/R	AAAAATCTAGAGCTCGGTCCCAAAGAAAACA / AAAAAAGATCTCGAATCTGGCTCTCAGGTGTAT	Cloning <i>sfl1</i> 3'-end (1432 bp)
Skn7up-F/R	AAAAAGAATTGCTGCGACTGTGTTGTTGGC / AAAAAACCGGGGTTGACGACGGGGTGAAAAG	Cloning <i>skn7</i> 5'-end (2113 bp)
Skn7dn-F/R	AAAAAAGATCTATGCCGTAAGTTGCGTCGTC / AAAAACTAGTCGTCGTTAGGCTGTCTGCTGT	Cloning <i>skn7</i> 3'-end (2298 bp)
Hsf1fl-F/R	<u>GGGGACCACTTTGTACAAGAAAGCTGGGTGACCAGCGTCACGGCAAGAAT</u> / <u>GGGGACAAGTTTGTACAAAAAAGCAGGCTGGACTGAATGCGACAGGCGTT</u>	Cloning full-length <i>hsf1</i> (5390 bp)
Sfl1fl-F/R	<u>GGGGACCACTTTGTACAAGAAAGCTGGGTGGGGAGAGAACAGATTGGAC</u> / <u>GGGGACAAGTTTGTACAAAAAAGCAGGCTTCGTTGGACTACTCCTTTTC</u>	Cloning full-length <i>sfl1</i> (4336 bp)
Skn7fl-F/R	<u>GGGGACCACTTTGTACAAGAAAGCTGGGTCTTAGCGATGTGAAGCAGGTT</u> / <u>GGGGACAAGTTTGTACAAAAAAGCAGGCTCAGAGCACCAACAAACAACAT</u>	Cloning full-length <i>skn7</i> (5811 bp)
pHsf1-F/R	CAAGCACCTCACAACAACAAG / CTCACGCTTGGCTTTGGCT	PCR detecting <i>hsf1</i>
pSfl1-F/R	CACTACACTCGTCTTTACCGCCAG / TCGGGAGGCAGTTGAACAGGT	PCR detecting <i>sfl1</i>
pSkn7-F/R	TCGAACTCTTCTTCCCGCG / TGGTCTCTCCACAGCATT	PCR detecting <i>skn7</i>
sbHsf1-F/R	TGTGACTCAAGGTGAAAAAAG / GAATATATTGTGGCAGTGGGT	Southern probe for <i>hsf1</i>
sbSfl1-F/R	GTAGGTAGGTATCACTTGCC / GCACCTGGAGAGAGAAAAAT	Southern probe for <i>sfl1</i>
sbSkn7-F/R	TTCTCCTCTCATTCCGTCTT / GACACCGAAGGAGGTTGACAG	Southern probe for <i>skn7</i>
qHsf1-F/R	GTCCGATGAACCAAGTCTT/AAAGGAGTCCAGGGATCT	qRT-PCR detecting <i>hsf1</i>
qSfl1-F/R	ATCCAGCATCTAATATCA/TAAAGTATTGGGAGAGAA	qRT-PCR detecting <i>sfl1</i>
qSkn7-F/R	GAACAATGCCAGTGAATT / CGACAACAACAAGGTAT	qRT-PCR detecting <i>skn7</i>
qActin-F/R	TCTCTACGGCAACATTGTC / TGCTGGAAGGTGGATAGG	qRT-PCR detecting γ -actin gene

*Underlined regions denote the sites of restriction enzyme for the deletion of each target gene (*EcoRI/HindIII* and *XhoI/SpeI* for *hsf1*, *EcoRI/SacI* and *XbaI/BglII* for *sfl1*, and *EcoRI/XmaI* and *BglII/SpeI* for *skn7*) via homogenous recombination of 5' and 3' fragments separated by *bar* marker or the gateway exchange fragments for complementation of each target gene.

Table S2. Paired primers used for assessing transcript levels of 46 genes in the mutants of *hsf1*, *sfl1* and *skn7* versus *B. bassiana* wild-type via qRT-PCR.

Gene ID	Gene	Annotation	Sequences (5'-3') of paired primers
Involved in growth and asexual development			
19887954	<i>fluG</i>	developmental protein	CCTCCCTAGTTTGGTCGCTTTCTC / CGCTGTCGGTAATCTGCTCCTC
19885980	<i>flbA</i>	developmental regulator	CCAATCCACTCGCCGCTCTC / CGGAGGAAAAGAGAATCGGTAGAGG
19890000	<i>flbB</i>	bZIP transcription factor	GCACTGACACGCCGACAAGAGC / CCGCCGCCGAAGCCTGTTG
19886193	<i>flbC</i>	C ₂ H ₂ conidiation transcription factor	TCCATCTCCAACCTGCTGGGTCTC / GGCGGCGTAGGCGGAAGG
19890271	<i>flbD</i>	MYB conidiophore development protein	CGGCAAGCGATGGGCAGAGATTG / ACGAGCAAGGTGACGGTAGAGGTG
19889611	<i>hyd1</i>	hydrophobin 1	TTCTCAGCGATCTTGATCTT / GCACTTGTTCGATTGG
19886027	<i>hyd2</i>	hydrophobin 2	CATGGTGAAAGGATCTG / ATCTTGGTCGTCTTCTCG
Involved in Na⁺ efflux			
19889202	<i>Ena1</i>	P-type Na ⁺ /K ⁺ ATPase 1	TTCTACCGACGATAACT / TATTCTCAGCCAGGACAT
19892062	<i>Ena2</i>	P-type Na ⁺ /K ⁺ ATPase 2	TAATACCAGGCAAGAAGT / TTAGGACGACAATGAGAG
19886191	<i>Ena3</i>	P-type Na ⁺ /K ⁺ ATPase 3	GAATCTCAGCAGACTCAT / AATCACGAATCTTCTCT
19887969	<i>Ena4</i>	P-type Na ⁺ /K ⁺ ATPase 4	CGATGATGACAGCAAGTT / CGATAGAGCACCACAGAT
19890673	<i>Ena5</i>	P-type Na ⁺ /K ⁺ ATPase 5	CAAGGATACAAGCGGTAA / CGACGAAGTAATGAGGTT
HSP-coding genes			
19890898	<i>hsp20</i>	heat shock protein 20	TCAGGAGATGAGAAGAAC / GCTGTGATGATGTTTGT
19891700	<i>hsp30a</i>	heat shock protein 30	TCTCCCTTGGTCGTGAT/GTATTGGTCGCTGCTCTC
19885069	<i>hsp30b</i>	heat shock protein 30	ATGGCTTCTTCTTCC/AGTATGTGTCAAACGTC
19886748	<i>hsp40a</i>	heat shock protein 40	GAGATTGACCTCTATGAG / CTCCTGAATTTGACCTC
19891452	<i>hsp40b</i>	heat shock protein 40	AGAAGACTACTACAAGATCC / TTCCTGGCAGTATCATC
19889942	<i>hsp40c</i>	heat shock protein 40	ATTTACACGACGAACAAG / TCGGAGTCCATAGTAAGC
19888317	<i>hsp40d</i>	heat shock protein 40	CAGGTACTTTCCGACAAG / CGAAAATAGACGAGAAGAAC
19885590	<i>hsp40e</i>	heat shock protein 40	GACAAGAACAAGGACGAT / ACCTCTCAGCAGAAAATTC
19883188	<i>hsp40f</i>	heat shock protein 40	AATCGGTGCTCTAAGTT / TGTCGAAAGAATTCGTAAG
19888479	<i>hsp60a</i>	heat shock protein 60	TTTACATCTTCACTACGA/CCTTCTAATCAGTTCT
19892463	<i>hsp70a</i>	heat shock protein 70	GTGGTAAAGGCATGATTT / GAGTCAGAAGTTGGAAG
19889799	<i>hsp70b</i>	heat shock protein 70	GGACCTATGGCTCAACAG / AATGCGACACCAGAAAAG
19891640	<i>hsp70c</i>	heat shock protein 70	TTGGCTCAGTTGAATGTG / GCAAGTCTGAAAGCAATA
19892721	<i>hsp70d</i>	heat shock protein 70	TCTTGTAGCCGTTGATT / GGTATTATCTCCTCTTAC
19884009	<i>hsp70e</i>	heat shock protein 70	AGCAGAAGCAATGTCAAT/TGGCTGATGAAGAAGTTG
19886620	<i>hsp70f</i>	heat shock protein 70	AGCCGTCTTCTTCTCT/GCCTTGATGTACTCTGTG
19883269	<i>hsp70g</i>	heat shock protein 70	CATGGTTCTCCTCAAGAT / CTGGTTGCTGTTGAAGTA
19888598	<i>hsp70h</i>	heat shock protein 70	TGCTGGTGGTATGATGAC / TCGGAGAAGGTAGAGAAGA
19886917	<i>hsp70i</i>	heat shock protein 70	ATTATGGCACTGTCAATG / TGATACGGTTTCTTGAT
19885360	<i>hsp70j</i>	heat shock protein 70	ACATCAAGGTCACTAAGC / CTTCTCTCGTCTTCGTC
19889859	<i>hsp70k</i>	heat shock protein 70	TCAACATTCACGGCATTTC / CTTCTGTGCAAGTCCAT
19884520	<i>hsp70l</i>	heat shock protein 70	TCTGTCCTTAAACGATACC / TCACGAAGAGCAGTAATC
19883953	<i>hsp70m</i>	heat shock protein 70	ACTTACTTCAATATGGC / TAGGTACTTCTCTCAAG
19886284	<i>hsp70n</i>	heat shock protein 70	CATTGTTGTCTATTGACT / TCTCCTGATCTCCACTA
19883808	<i>hsp70o</i>	heat shock protein 70	TACAATCTGACTGAAGAACATATC / ATCGTTGAGCCTGACATT
19889528	<i>hsp90</i>	heat shock protein 90	TTCTCTCCCTCATCATCAAC/CTTGTAGCGGACCTTGTC
19885295	<i>hsp104</i>	heat shock protein 104	CTTTCGCCACAATTTAC/ACACCGATGTAGGTATCTT
19884587	<i>hsp78</i>	heat shock protein 78	ATACAGACCTGCCGCTAT/TATGTTGCTGTGACCATTTG

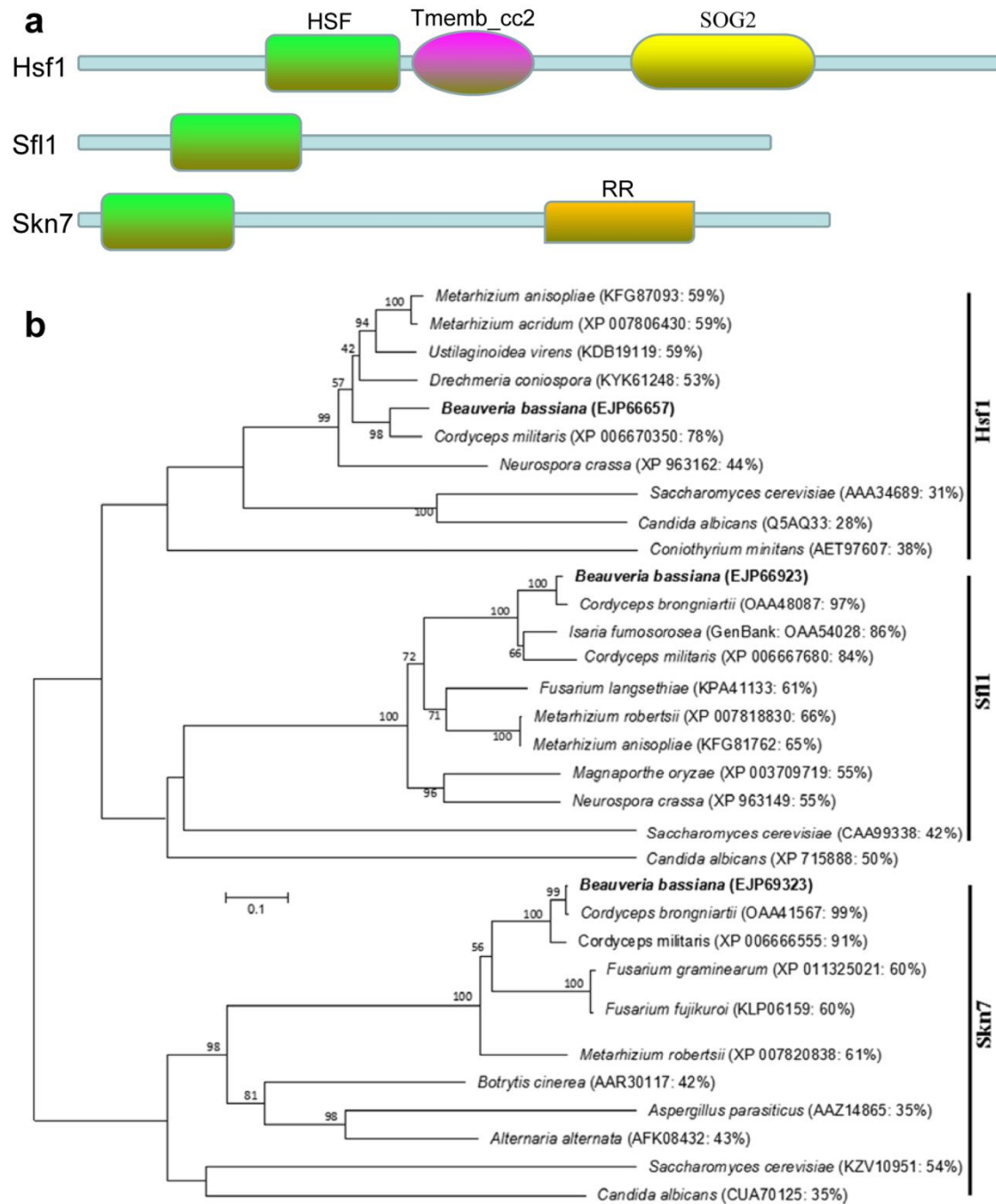


Fig. S1. Structural and phylogenetic features of three HSF-containing genes found in the genome database of *B. bassiana*. **(a)** Conserved domains predicted from three HSFs via sequence alignment and online blast analysis at <http://blast.ncbi.nlm.nih.gov/blast.cgi>. **(b)** Phylogenetic tree for three HSFs of *B. bassiana* and other fungi based on a neighbor-joining method in MEGA7 at <http://www.megasoftware.net>. The NCBI accession code of a given protein and its sequence identity to that of *B. bassiana* are given in the parentheses following each fungal name. Scale bar: branch length proportional to genetic distance.

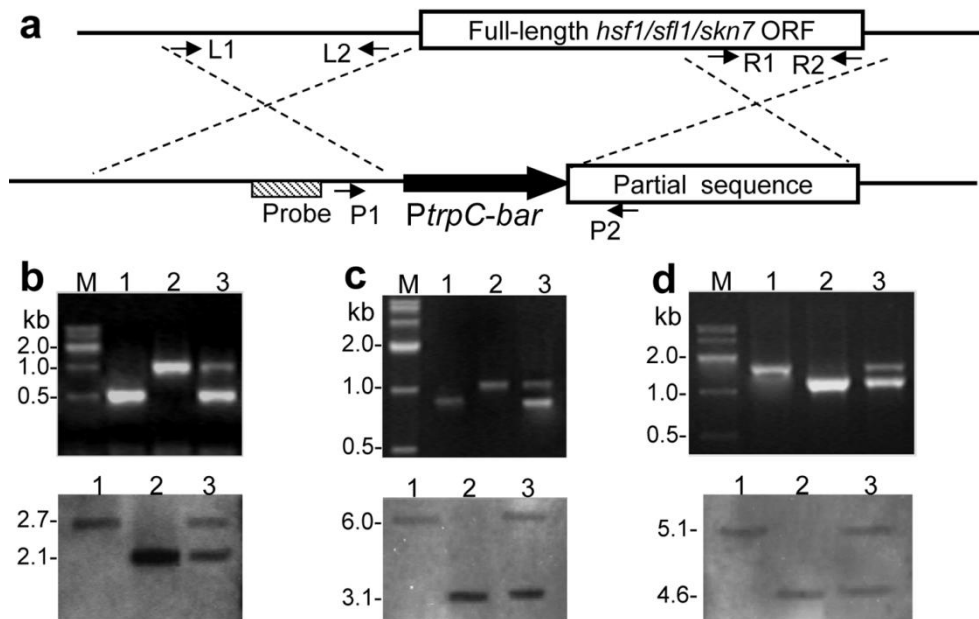


Fig. S2. Construction and identification of *B. bassiana* *hsf1*, *sfl1* and *skn7* mutants. **(a)** Schematic diagram for the deletion strategy of each target gene. **(b–d)** The *hsf1*, *sfl1* and *skn7* mutants identified via PCR (*upper panel*) and Southern blot hybridization (*lower panel*), respectively. *Lane 1*: wild-type strain. *Lane 2*: deletion mutant. *Lane 3*: complemented mutant. Genomic DNAs used for Southern blotting of *hsf1*, *sfl1* and *skn7* were digested with *XhoI/XhoI*, *XbaI/XbaI* and *BstXI/EcoRI*, respectively.

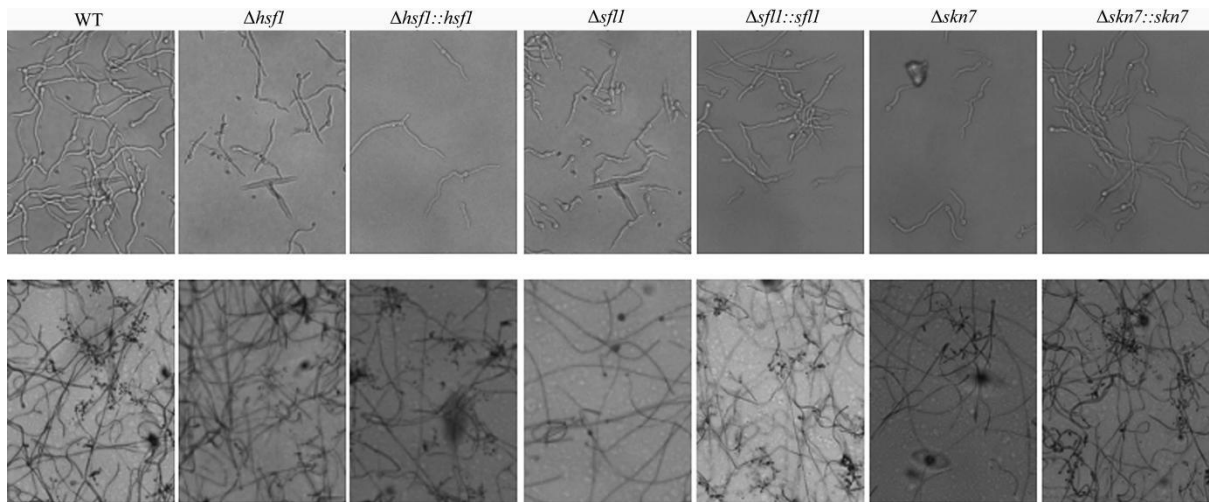


Fig. S3. Images of germ tubes and early hyphae after conidia were cultivated on GM plates for 16 h (up) and conidiophores and conidia after conidia were cultivated on SDAY plates for 3 days (down), visualized at 400× magnification.

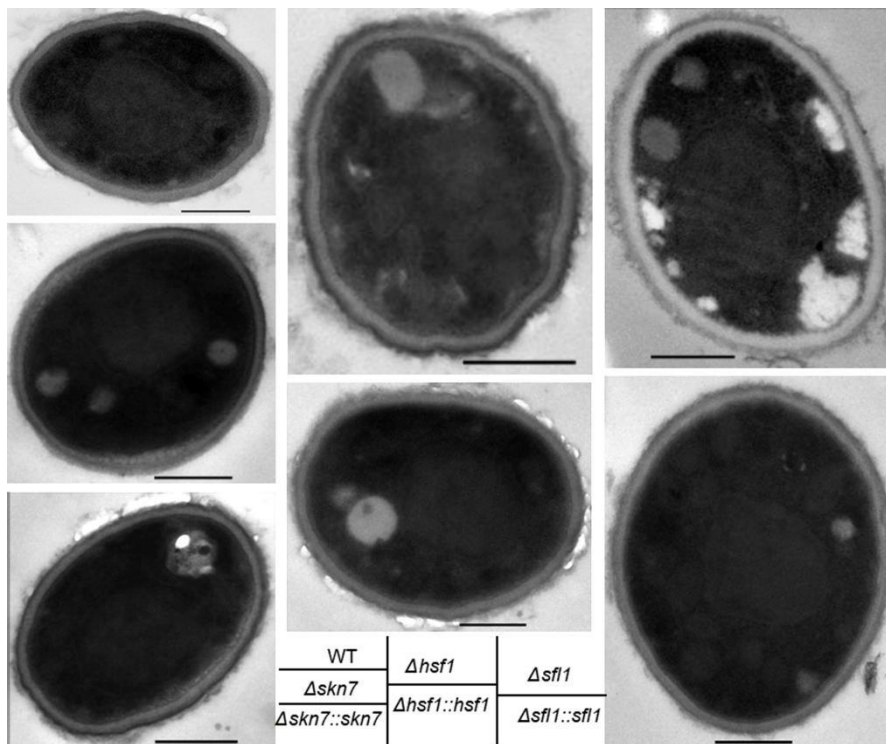


Fig. S4. Images for the ultrathin sections of conidia observed under transmission electron microscopy (TEM). Briefly, aerial conidia were washed three times in PBS (pH 7.0) and fixed overnight at 4 °C with 2% paraformaldehyde and 2.5% (v/v) glutaraldehyde in 0.1 M phosphate buffer (pH 7.2). After post-fixed in 1% OsO₄, conidia were dehydrated in a graded ethanol series, embedded in Spurr resin and stained with 2% uranyl acetate and Reynold's lead solution. The samples were examined under an H-7650 transmission electron microscope (Hitachi). Scale: 0.5 μm.

Table S3. Relative transcript levels of HSPs gene in the 3-d-old SDAY cultures of *hsf1*, *sfl1* and *skn7* mutants versus WT grown at 25°C or then shocked at 40°C for 1 h.

HSP genes	Relative transcript levels (mean ± SD) in normal cultures*							Relative transcript levels (mean ± SD) in heat-shocked cultures*						
	WT	$\Delta hsf1$	$\Delta hsf1::hsf1$	$\Delta sfl1$	$\Delta sfl1::sfl1$	$\Delta skn7$	$\Delta skn7::skn7$	WT	$\Delta hsf1$	$\Delta hsf1::hsf1$	$\Delta sfl1$	$\Delta sfl1::sfl1$	$\Delta skn7$	$\Delta skn7::skn7$
<i>hsp20</i>	1.00±0.09	13.5±0.97	1.71±0.35	1.38±0.11	1.02±0.08	0.24±0.01	0.97±0.02	1.00±0.07	2.14±0.03	1.00±0.02	0.11±0.02	0.95±0.04	0.47±0.03	1.00±0.02
<i>hsp30a</i>	1.00±0.09	1.25±0.14	0.97±0.13	0.83±0.02	1.00±0.02	0.58±0.01	1.00±0.01	1.00±0.03	0.10±0.01	0.95±0.03	0.78±0.02	0.97±0.04	0.54±0.02	0.95±0.03
<i>hsp30b</i>	1.00±0.11	1.50±0.10	1.02±0.06	0.23±0.02	0.96±0.02	0.25±0.01	0.96±0.03	1.03±0.31	0.43±0.04	1.00±0.03	0.09±0.01	0.93±0.06	0.94±0.02	1.00±0.02
<i>hsp40a</i>	1.00±0.06	0.17±0.01	1.00±0.01	1.03±0.06	1.02±0.06	0.62±0.03	0.93±0.04	1.04±0.39	0.54±0.03	1.00±0.02	0.79±0.04	1.00±0.03	3.34±0.33	1.35±0.39
<i>hsp40b</i>	1.00±0.10	0.10±0.00	0.95±0.04	1.33±0.02	1.01±0.02	0.45±0.02	0.96±0.04	1.01±0.14	1.21±0.04	1.00±0.03	0.17±0.04	0.95±0.03	1.14±0.07	1.00±0.05
<i>hsp40c</i>	1.00±0.06	6.30±0.31	1.63±0.35	1.09±0.02	0.99±0.02	0.24±0.01	0.99±0.01	1.00±0.02	0.30±0.01	1.00±0.01	0.61±0.05	1.00±0.03	0.14±0.01	0.93±0.10
<i>hsp40d</i>	1.01±0.13	7.45±0.67	1.71±0.47	1.03±0.07	1.02±0.06	0.19±0.01	0.95±0.04	1.00±0.12	1.07±0.02	1.00±0.01	0.14±0.01	0.99±0.01	0.07±0.00	0.87±0.01
<i>hsp40e</i>	1.01±0.14	8.99±0.90	1.74±0.18	1.57±0.17	1.11±0.02	0.20±0.02	0.95±0.04	1.00±0.04	0.84±0.03	1.00±0.02	0.21±0.00	1.00±0.00	0.12±0.00	0.84±0.03
<i>hsp40f</i>	1.00±0.12	5.53±0.12	1.67±0.50	0.95±0.02	1.00±0.02	0.28±0.00	0.97±0.03	1.00±0.08	0.49±0.04	0.97±0.03	0.24±0.01	1.00±0.01	0.18±0.00	1.00±0.00
<i>hsp60</i>	1.00±0.10	35.9±11.7	0.97±0.05	0.68±0.14	1.00±0.11	0.42±0.10	0.96±0.21	1.00±0.12	2.13±0.22	1.14±0.10	0.50±0.09	0.93±0.01	1.07±0.25	1.01±0.17
<i>hsp70a</i>	1.00±0.09	0.77±0.05	1.00±0.03	1.68±0.11	0.98±0.11	0.77±0.05	1.10±0.18	1.00±0.06	0.06±0.00	0.96±0.04	0.07±0.00	1.00±0.00	2.19±0.02	1.00±0.01
<i>hsp70b</i>	1.00±0.05	0.11±0.01	0.95±0.07	0.29±0.02	0.94±0.04	2.60±0.08	1.02±0.07	1.00±0.06	2.43±0.07	1.00±0.05	0.14±0.01	0.85±0.04	0.06±0.01	0.87±0.02
<i>hsp70c</i>	1.00±0.03	15.7±1.10	1.96±0.37	2.28±0.13	0.98±0.12	0.17±0.01	0.90±0.00	1.00±0.03	1.42±0.03	1.00±0.02	0.09±0.01	0.87±0.02	0.06±0.01	0.83±0.01
<i>hsp70d</i>	1.01±0.14	17.4±0.83	1.84±0.07	1.74±0.14	1.02±0.09	0.74±0.03	1.01±0.02	1.00±0.05	1.82±0.01	1.00±0.01	0.07±0.00	0.95±0.05	0.06±0.01	0.85±0.03
<i>hsp70e</i>	1.00±0.10	0.92±0.02	1.00±0.02	0.16±0.00	0.97±0.06	0.20±0.01	0.94±0.06	1.09±0.50	0.83±0.08	1.00±0.05	0.13±0.01	0.90±0.01	0.34±0.06	1.00±0.04
<i>hsp70f</i>	1.00±0.10	0.90±0.03	1.00±0.03	0.16±0.01	0.94±0.05	0.15±0.00	0.94±0.05	1.00±0.04	0.91±0.04	1.00±0.03	0.12±0.01	0.90±0.00	0.06±0.00	0.95±0.04
<i>hsp70g</i>	1.00±0.08	0.06±0.00	1.00±0.00	0.51±0.06	0.95±0.02	2.82±0.04	0.99±0.03	1.00±0.08	2.32±0.08	1.00±0.06	0.09±0.01	0.88±0.03	1.38±0.14	1.03±0.06
<i>hsp70h</i>	1.00±0.07	0.59±0.07	0.95±0.00	0.33±0.03	1.00±0.02	2.60±0.08	0.98±0.06	1.00±0.02	0.53±0.03	1.00±0.02	0.48±0.01	1.00±0.00	0.45±0.07	1.00±0.05
<i>hsp70i</i>	1.00±0.03	0.47±0.01	1.00±0.01	0.12±0.01	0.96±0.04	2.78±0.22	0.98±0.02	1.00±0.06	1.22±0.03	1.00±0.02	0.17±0.01	0.98±0.02	0.52±0.04	1.00±0.03
<i>hsp70j</i>	1.00±0.06	4.82±0.24	1.62±0.36	2.01±0.03	0.99±0.03	0.44±0.01	0.96±0.02	1.00±0.05	1.84±0.17	1.00±0.12	0.12±0.01	0.90±0.01	0.07±0.00	0.87±0.02
<i>hsp70k</i>	1.00±0.06	4.17±0.23	1.03±0.15	1.31±0.11	0.98±0.11	2.30±0.11	1.03±0.11	1.00±0.08	0.81±0.04	1.00±0.03	0.52±0.02	1.00±0.01	0.42±0.04	1.00±0.03
<i>hsp70l</i>	1.00±0.11	0.46±0.09	0.96±0.01	0.15±0.03	0.84±0.03	2.39±0.08	1.02±0.08	1.00±0.09	1.48±0.04	1.00±0.03	0.43±0.03	1.00±0.02	0.74±0.09	1.01±0.16
<i>hsp70m</i>	1.00±0.10	17.7±0.71	1.71±0.36	1.41±0.12	1.09±0.06	0.11±0.01	0.97±0.05	1.00±0.04	1.01±0.00	1.00±0.00	0.06±0.00	0.87±0.04	0.06±0.00	1.00±0.00
<i>hsp70n</i>	1.00±0.01	15.4±0.75	1.70±0.19	1.07±0.04	1.01±0.04	0.38±0.04	0.94±0.02	1.00±0.07	0.71±0.03	1.00±0.02	0.03±0.00	0.86±0.03	0.92±0.01	1.00±0.01
<i>hsp70o</i>	1.01±0.15	14.1±0.78	1.69±0.12	1.37±0.08	1.02±0.06	0.12±0.00	0.98±0.04	1.00±0.05	0.98±0.05	1.00±0.04	0.06±0.00	0.84±0.03	0.05±0.00	0.94±0.07
<i>hsp90</i>	1.00±0.02	0.75±0.02	1.01±0.02	0.33±0.01	0.99±0.02	0.54±0.01	1.00±0.00	1.00±0.07	0.23±0.02	1.00±0.01	0.48±0.04	0.97±0.03	1.06±0.04	1.00±0.02
<i>hsp104</i>	1.00±0.04	2.73±0.21	0.98±0.12	0.63±0.02	1.00±0.02	0.36±0.00	1.00±0.00	1.00±0.09	0.42±0.05	1.00±0.04	3.61±0.39	1.46±0.09	0.57±0.04	1.00±0.03
<i>hsp78</i>	1.00±0.00	0.93±0.00	1.00±0.00	0.15±0.00	0.90±0.00	0.20±0.01	0.80±0.01	1.00±0.08	0.66±0.01	1.00±0.01	0.63±0.03	0.97±0.03	3.71±0.86	1.44±0.13

* Red: HSP genes significantly upregulated by ≥ 2 -fold. Green: HSP genes significantly downregulated by $\geq 50\%$.