S2 Table. Primers used in this study

pPROPET co-expression plasmids	MCS1	MCS2
pPROPET/G$^{1-132}$U$^{199-359}$_Psp	*hrpG*$^{1-132}$ (template pPROEX/*hrpG*) Forward: 5' – GACCAATCATGAGCTCGATGGATTTC-3' (ligated to compatible vector's NcoI site) Reverse: 5' – AGGGTCGTGCGGCCGCATCACGCGG- 3'	*hrcU*$^{199-359}$ (template pET16b/*hrcU*$^{199-359}$) Forward: 5' - CTGGTGCAAGTCATATGAGTGAAAAAACCGA-3' Reverse: 5'-CCTGTTACGCCTGGATCCCAGCTCC-3'
pPROPET/VJ_Psp	*hrpV* (template pET16b/*hrpV*) Forward: 5' – AGGATTAGTCATGATTGAGGTAAAG-3' (ligated to compatible vector's NcoI site) Reverse: 5' – CGTCTAGAAGCTTGCGTCC-3'	*hrpJ* (template pPROEX/*hrpJ*) Forward: 5'-GCTCAGAACCGTCATATGAAAATCG-3' Reverse: 5'-CGACAGGGCAAGCTTGTTCAGAAAG-3' (ligated blunt to vector's SacI site after Klenow treatment)
pPROPET/GJ_Eamy	*hrpG* (template: genomic DNA *Eamy*) Forward: 5' – GCCATCATTCATGAGATCAACTGAATTGCAG C – 3' Reverse: 5' - GCAACTCTCGAGTCTCAACCATGGTTTCC -3'	*hrpJ* (template: genomic DNA *Eamy*) Forward: 5' – GAAGGTTCGCCCATATGAAAATTGC – 3' Reverse: 5' - GCCAGACAAACAGAGCTCTCATTTA - 3'
pPROPET/GV_Eamy	*hrpG* (as in 3)	*hrpV* (template: genomic DNA *Eamy*) Forward: 5'-GTATCGGCATATGAATAATCCTTGC-3' Reverse: 5'-CCGGC GAATTCAGAAAAAATAACC-3'
pPROPET/GVJ_Eamy	*hrpG* (as in 3)	*hrpV/hrpJ* *hrpV* as in 4. *hrpJ* was first subcloned to pET26b using primers in 3, digested from there XbaI(blunt)/SacI and ligated to pPROpET/*hrpG*/*hrpV* in sites EcoRI(blunt)/SacI)
pPROPET/VJ_Eamy	*hrpV*(template genomic DNA *Eamy*) Forward: 5' – GGTATCGGTCCATGGATAATCCTTG – 3' Reverse: 5' - GAAAGGACGCAAGCTTGGATCAATC -3'	*hrpJ* (template genomic DNA *Eamy*) (as in 3)
pPROPET/GU$^{199-360}$_Eamy	*hrpG* (as in 3)	*hrcU*$^{199-360}$ (template: genomic DNA *Eamy*) Forward: 5' - GCTGGTACTGCATATGCTGGACTTCGGCC -3' Reverse: 5'- GGTAATAGTTGAGCTCATTCGGCTTCCAGTTC -3'

Site directed mutagenesis plasmid 1 derivative	**Forward primer**	**Reverse primer**
pPROPET/$G^{1-132}U^{199-359}$_APTH_Psp	5' – GACATGCTGCTGGTGGCGCCGACGCACTACGCGG- 3'	5'-CCGCGTAGTGCGTCGGCGCCACCAGCAGCATGTC-3'
***Psp hrpJ* knock-out allele generation**		
A1	5'-CTGGCGATACATTTTGCGG-3'	
A2	5'-CCCTATAGTGAGTCGAATTCAGGACGGTTCTGAGCCTG-3'	
B1	5'-CAGGTTGGCGCGCAGATC-3'	
B2	5'-GAATTCGACTCACTATAGGGATCAAGTCATCAACTTTC-3'	
RT-qPCR analysis	**Forward primer**	**Reverse primer**
hrpL (159 bp amplicon)	5' –AGCCGCAGACCTGGTTGTG- 3'	5' -ATTGCCTGTGCCCGTCTACC3-'
16S rDNA *(157 bp amplicon)*	5'-GGAATCTGCCTGGTAGTGGGG- 3'	5'-GGCTCACCAAGGCGACGAT 3-'