

Supplementary material

Development of selective agents targeting 5HT_{1A} receptor with subnanomolar activities based on coumarin core

K. Ostrowska,^{*a} D. Grzeszczuk,^a M. Głuch-Litwin,^b A. Gryboś,^b A. Siwek,^b Ł. Dobrzycki^c and B. Trzaskowski^d

^a Department of Organic Chemistry, Faculty of Pharmacy, Medical University of Warsaw, 1 Banacha Str., 02 097 Warsaw, Poland. E-mail: kostrowska@wum.edu.pl

^b Department of Pharmacobiology, Faculty of Pharmacy, Jagiellonian University Collegium Medicum, Kraków, Poland

^c Crystallochemistry Laboratory, Faculty of Chemistry, University of Warsaw, 1 Pasteura Str., 02 093 Warsaw, Poland

^d Centre of New Technologies, University of Warsaw, 2C Banacha Str., 02-097 Warsaw, Poland

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Single crystal X-ray diffraction data

The X-ray measurements of **3b** and **6d** were performed at 100(2) K on a Bruker D8 Venture Photon100 diffractometer equipped with a TRIUMPH monochromator and a MoK α fine focus sealed tube ($\lambda=0.71073$ Å). The frames were collected with Bruker APEX2 program^{S1} and integrated with the Bruker SAINT software package^{S2} using a narrow-frame algorithm. The deposition number CCDC 1541420 for **3b** and CCDC 1541465 for **6d** contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk or by contacting The Cambridge Crystallographic Data Centre, 12, Union Road, Cambridge CB2 1EZ, UK; fax 44 1223 336033

Crystals of 6-acetyl-4,7-dimethyl-5-[3-(morpholin-4-yl)propoxycoumarin **6d** are twinned by pseudomerohedry with every third layer of reflections: $0kl$, $3kl$, $\bar{3}kl$, $6kl$, $\bar{6}kl$, ... almost ideally overlapped (see Figure S1). The twin lattice has pseudo orthorhombic symmetry with base-centered unit cell and twin obliquity calculated for (100) plane and [610] lattice vector equal to 0.38°. The twin domains are related by the (100) mirror reflection and compared with the single component real lattice. At the twin boundary the morpholine groups are meshed together similarly as in the single lattice of **6d**. In the crystals of 6-acetyl-4,7-dimethyl-5-[3-(morpholin-4-yl)propoxycoumarin **6d** two methyl groups (C9A/C9B and C12A/C12B) have disordered hydrogen atoms over two positions. These positions however are not equally occupied by the H atoms. This observation is confirmed by H-NMR measurement of the crystalline powder of **6d** where the signal splitting for C9 and C12 methyl groups is visible. The structure of **6d**, due to the lack of typical hydrogen bond donors, is dominated by weak interactions. The molecule of 4,7-dimethyl-5-{3-[4-(2-fluorophenyl)piperazin-1-yl]propoxy}coumarin (**3b**) is shorter C₃H₆ linker analogue of the 4,7-dimethyl-5-{4-[4-(1-fluorobenzyl)-piperazin-1-yl]-butoxy} coumarin^{S3} which has four carbon atoms aliphatic chain between chromene and piperazine fragments. Due to different conformation of these molecules their packing in the crystal lattices is different. Similarly as in the **6d** structure in the **3b** there are also only weak intermolecular interactions. The shortest contacts are observed between two neighboring coumarin fragments with O...H distance yielding 2.52 Å.

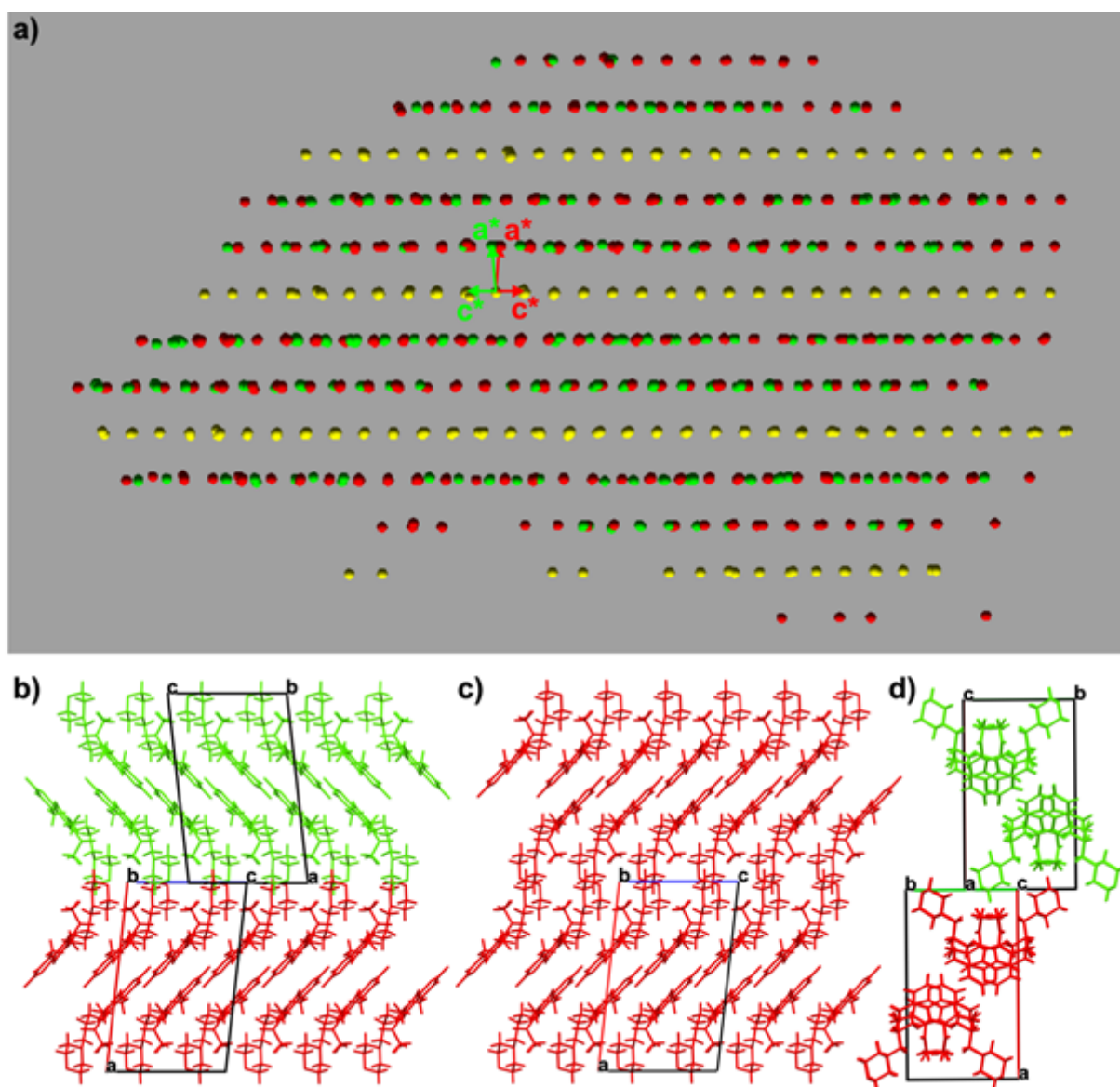


Figure S1 Twinning of the crystals of 6-acetyl-4,7-dimethyl-5-[3-(morpholin-4-yl)propoxycoumarin **6b** – visualization of reciprocal lattice – composite reflections are coloured in yellow a); possible relation between twin real lattices b), d); visualization of single component real lattice c).

The crystal of **6d** is twinned with part of reflection separated and the others overlapped, thus the further processing of the data was based on two twin domains. The integration of the diffraction spots using a monoclinic unit cell yielded a total of 27947 reflections to a maximum θ angle of 25.43° (0.83 \AA resolution), of which 3346 were independent (average redundancy 8.352, completeness = 99.6%, $R_{int}=4.98\%$, $R_{sig}=4.62\%$) and 2502 (74.78%) were greater than $2\sigma(F^2)$. The final cell constants of $a=17.1277(16) \text{ \AA}$, $b=9.9821(9) \text{ \AA}$,

$c=10.6929(9)$ Å, $\beta=96.349(2)^\circ$, $V=1817.0(3)$ Å³, are based upon the refinement of the XYZ-centroids of 9147 reflections above $20 \sigma(I)$ with $5.922^\circ < 2\theta < 50.87^\circ$ belonging to two twin components. Data were corrected for absorption effects using the multi-scan method (TWINABS).^{S4} The ratio of minimum to maximum apparent transmission was 0.830. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.9580 and 0.9940. The structure was solved and refined using SHELXTL software package^{S5} using the space group $P2_1/c$, with $Z=4$ for the formula unit, $C_{20}H_{25}NO_5$, with HKLF5 reflection format based on two components of the twin. The refinement gave twin fractions equal to 0.805(1) and 0.195(1). After the refinement converged the reflection file was converted to HKLF4 format with help of WinGX software.^{S6} The final anisotropic full-matrix least-squares refinement on F^2 with 242 variables converged at $R1=4.40\%$, for the observed data and $wR2=12.38\%$ for all data. The goodness-of-fit was 1.065. The largest peak in the final difference electron density synthesis was $0.295 e^-/\text{Å}^3$ and the largest hole was $-0.240 e^-/\text{Å}^3$ with an RMS deviation of $0.049 e^-/\text{Å}^3$. On the basis of the final model, the calculated density was 1.314 g/cm^3 and $F(000)$, 768 e^- . In the crystal lattice of DG405 two methyl groups are disordered with hydrogen atoms occupying alternative positions. Relative orientation of H atoms of each CH_3 group was refined together with the occupancy ratio yielding $0.74(2):0.26(2)$ and $0.17(3):0.83(3)$ for C9A/C9B and C12A/C12B methyl group respectively.

The structure of **6b**, due to the lack of typical hydrogen bond donors, is dominated by weak interactions. These interactions are visualized in Figure 2S with the help of Hirshfeld surface analysis approach^{S7} performed in CrystalExplorer software.^{S8} The closest interatomic contacts can be observed between carbonyl O atom of the coumarin fragment and one of the H atom of the ordered methyl group as sharp spikes in the fingerprint plot^{S6} displayed in Figure S2b), with the O...H distance equal to 2.44 Å. In the structure there are also some intermolecular contacts between carbon coumarin fragment atoms corresponding to π - π interactions (see Figure S2). The average distance between RMS planes^{S9} fitted to chromene skeleton of neighboring molecules is equal to 3.53 Å. The rings have a large parallel separation, thus the overlay of the chromene skeletons is limited to C3 and C4 atoms of the ring. The other part of this molecule interacts with another chromene moiety which is, however, slightly tilted but with intermolecular distance from C2 atom to the average plane equal to 3.78 Å.

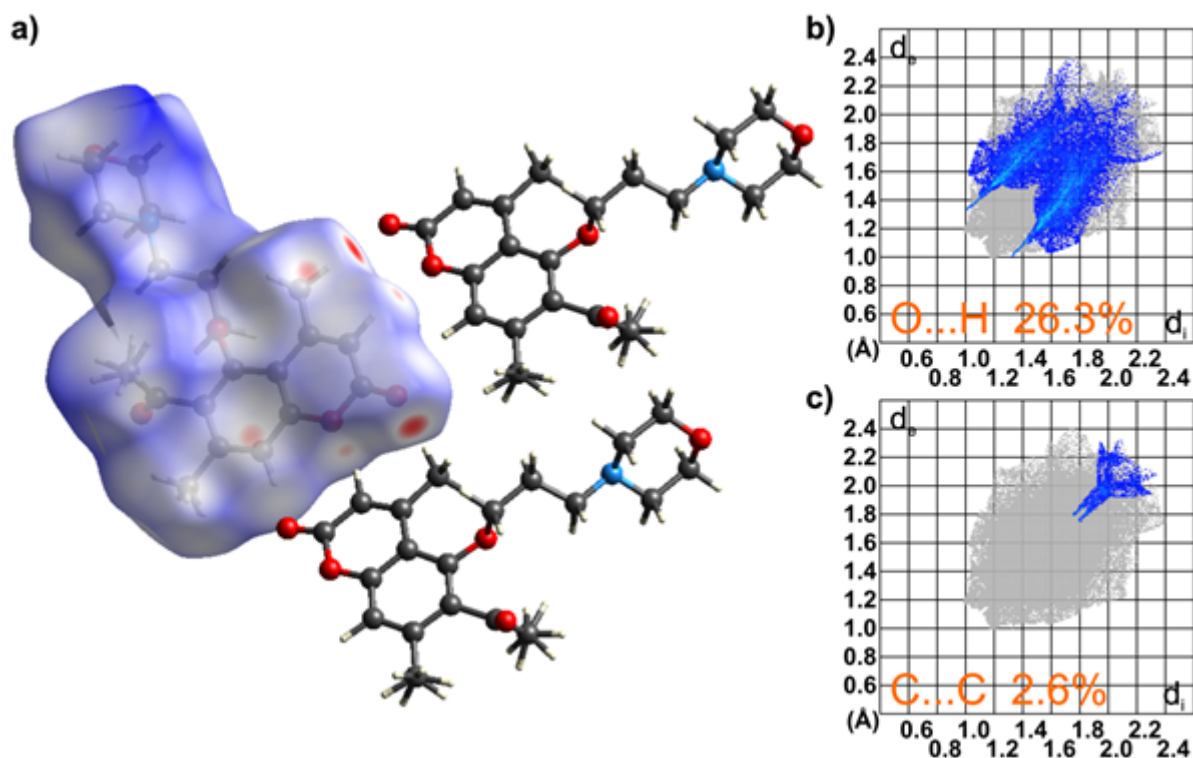


Figure S2 Hirshfeld surface generated for **6b** a) with selected 2D fingerprint plots presenting d_e and d_i distances for O...H a) and C...C b) atomic pairs in the crystal lattice.

In the case of **3b** the frames were integrated with the Bruker SAINT software package^{S4} using a narrow-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 22213 reflections to a maximum θ angle of 25.05° (0.84 \AA resolution), of which 3690 were independent (average redundancy 6.020, completeness = 99.8%, $R_{int}=2.26\%$, $R_{sig}=1.32\%$) and 3303 (89.51%) were greater than $2\sigma(F^2)$. The final cell constants of $a=7.7995(6) \text{ \AA}$, $b=9.8117(8) \text{ \AA}$, $c=14.8527(12) \text{ \AA}$, $\alpha=78.6250(19)^\circ$, $\beta=82.4050(18)^\circ$, $\gamma=69.9762(18)^\circ$, $V=1044.36(14) \text{ \AA}^3$, are based upon the refinement of the XYZ-centroids of 9962 reflections above $20 \sigma(I)$ with $6.037^\circ < 2\theta < 50.75^\circ$. Data were corrected for absorption effects using the multi-scan method (SADABS).^{S10} The ratio of minimum to maximum apparent transmission was 0.930. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.9390 and 0.9880. The structure was solved and refined using SHELXTL Software Package^{S5} using the space group $P\bar{1}$, with $Z=2$ for the formula unit, $C_{24}H_{27}FN_2O_3$. The final anisotropic full-matrix least-squares refinement on F^2 with 274 variables converged at $R1=3.32\%$, for the observed data and $wR2=8.99\%$ for all data. The goodness-of-fit was 1.042. The largest peak in the final difference electron density synthesis was $0.199 \text{ e}/\text{\AA}^3$ and the

largest hole was $-0.198 \text{ e}^-/\text{\AA}^3$ with an RMS deviation of $0.037 \text{ e}^-/\text{\AA}^3$. On the basis of the final model, the calculated density was 1.305 g/cm^3 and $F(000)$, 436 e^- .

Similarly as in the **6d** structure in the **3b** there are also only weak intermolecular interactions. The shortest contacts are observed between two neighboring coumarin fragments with O...H distance yielding 2.52 Å. This is visualized in Figure S3a, which shows also the Hirshfeld surface^{S7,S8} generated for one molecule. The fingerprint plot^{S9} for O...H contacts are shown in Figure S3b with wide spikes corresponding to O...H interaction between two coumarin fragments.

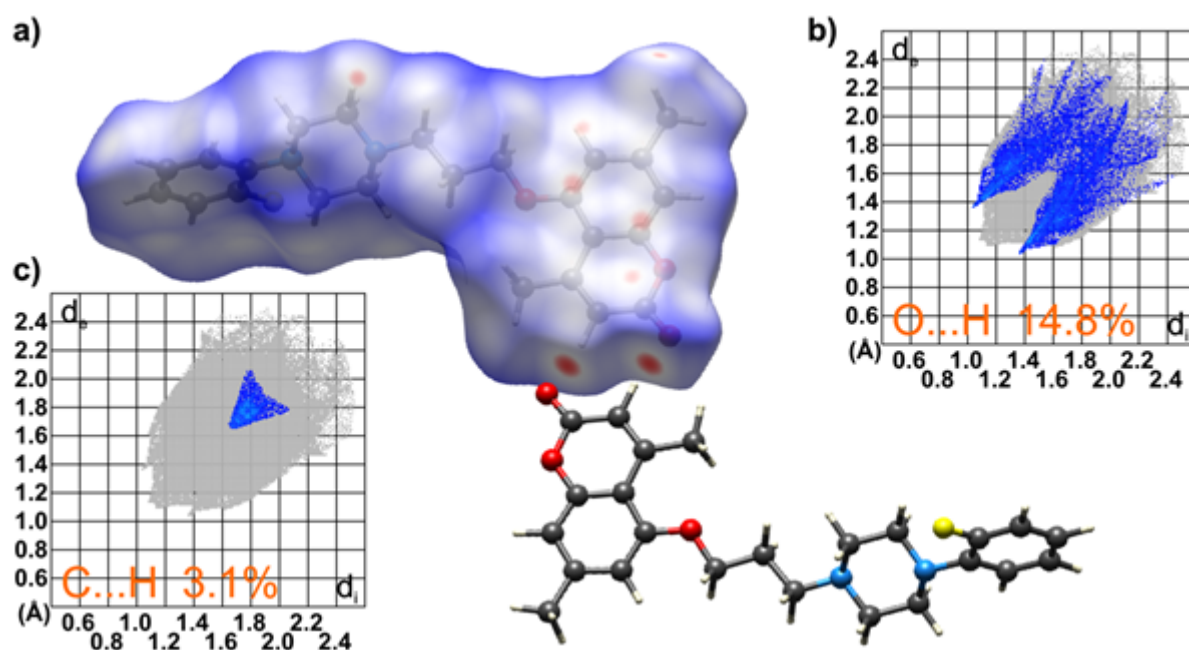


Figure S3 Hirshfeld surface generated for **3b** **a)** with selected 2D fingerprint plots presenting d_e and d_i distances for O...H **a)** and C...C **b)** atomic pairs in the crystal lattice.

In the structure of 4,7-dimethyl-5-{3-[4-(2-fluorophenyl)piperazin-1-yl]propoxy}coumarin **3b** the π - π interaction between two neighboring coumarin fragments is more pronounced than in the case of **6d**. Indeed C...C distances for **3b** are shorter than in **6d** (see Figures S2c and S3c). In **3b** coumarin fragments are forming dimmers with average distance between RMS planes^{S10} fitted to the chromene skeleton of 3.30 Å what is presented in Figure S4.

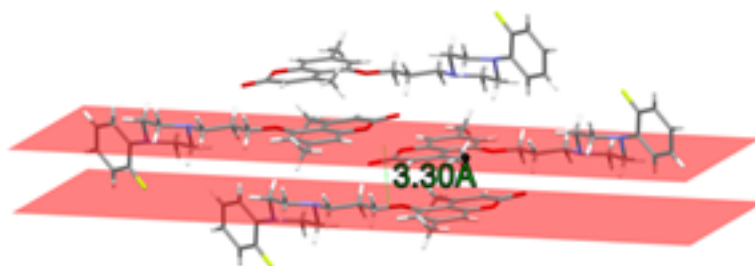


Figure S4 Stacking of molecules in the structure of 4,7-dimethyl-5-{3-[4-(2-fluorophenyl)piperazin-1-yl]propoxy}coumarin **3b**.

In the case of both structures all non-hydrogen atoms were refined anisotropically. All hydrogen atoms were placed in calculated positions and refined within the riding model. All CH₃ groups, including disordered ones in **6d**, were free to rotate along C-C bonds. The temperature factors of hydrogen atoms were not refined and were set to be equal to either 1.2 or 1.5 times larger than U_{eq} of the corresponding heavy atom. The atomic scattering factors were taken from the International Tables^{S12} molecular graphics was prepared using Diamond 3.2.^{S13} Crystal data and refinement parameters for **3b** and **6d** are collected in Table S1.

Table S1

Identification code	6b	3a
Chemical formula	C ₂₀ H ₂₅ NO ₅	C ₂₄ H ₂₇ FN ₂ O ₃
<i>M</i>	359.41	410.47
<i>T</i> / K	100(2)	100(2)
λ / Å	0.71073	0.71073
Crystal size/ mm	0.068×0.327×0.456	0.134×0.292×0.694
Crystal system	monoclinic	triclinic
Space group	<i>P</i> 2 ₁ / <i>c</i>	<i>P</i> $\bar{1}$
Unit cell dimensions	<i>a</i> =17.1277(16)Å <i>b</i> =9.9821(9)Å <i>c</i> =10.6929(9)Å β =96.349(2)°	<i>a</i> =7.7995(6)Å <i>b</i> =9.8117(8)Å <i>c</i> =14.8527(12)Å α =78.6250(19)° β =96.349(2)° γ =69.9762(18)°
<i>V</i> / Å ³	1817.0(3)Å ³	1044.36(14)Å ³
<i>Z</i> , <i>D_x</i> / g·cm ⁻³	4, 1.314	2, 1.305
μ / mm ⁻¹	0.094	0.092
<i>F</i> (000)	768	436
θ_{min} , θ_{max}	2.96°, 25.43°	2.95°, 25.05°

Reflections collected/ independent	27947/ 3346 [$R_{int}=0.0498$]*	22213/ 3690 [$R_{int}=0.0226$]
Completeness	99.6%	99.8%
Absorption correction	multi-scan	multi-scan
T_{max} T_{min}	0.994, 0.958	0.998, 0.939
Data / restraints / parameters	3346 / 0 / 242	3690 / 0 / 274
Goof F^2	1.065	1.042
Final R indices	2502 data; $I > 2\sigma(I)$ $R_I=0.0440$, $wR_2=0.1122$ all data $R_I=0.0553$, $wR_2=0.1238$	3303 data; $I > 2\sigma(I)$ $R_I=0.0332$, $wR_2=0.0859$ all data $R_I=0.0377$, $wR_2=0.0899$
ρ_{max} ρ_{min}	0.295 eÅ ⁻³ , -0.240 eÅ ⁻³	0.199 eÅ ⁻³ , -0.198 eÅ ⁻³

* Data scaling based on two twin components with HKLF5 refinement giving twin fractions equal to 0.805(1) and 0.195(1). Final refinement performed on merged reflections in HKLF4 format with twinning effect included.

- S1 APEX2. Bruker AXS Inc., Madison, Wisconsin, USA, 2013
- S2 SAINT. Bruker AXS Inc., Madison, Wisconsin, USA, 2013.
- S3 K. Ostrowska, D. Grzeszczuk, D. Maciejewska, I. Młynarczuk-Biały, A. Czajkowska, A. Sztokfisz, Ł. Dobrzycki and H. Kruszewska *Monatsh. Chem.*, 2016, **147**, 1615.
- S4 TWINABS. Bruker AXS Inc., Madison, Wisconsin, USA, 2012.
- S5 G. M. Sheldrick, *Acta Crystallogr.*, 1990, **A46**, 467.
- S6 L. J. Farrugia *J. Appl. Cryst.*, 2012, **45**, 849.
- S7 M. A. Spackman and D. Jayatilaka, *Cryst. Eng. Comm.*, 2009, **11**, 19.
- S8 S. K. Wolff, D. J. Grimwood, J. J. McKinnon, M. J. Turner, D. Jayatilaka and M. A. Spackman, University of Western Australia, CrystalExplorer, 2012.
- S9 M. A. Spackman, and J. McKinnon J. Fingerprinting intermolecular interactions in molecular crystals. *Cryst. Eng. Comm.*, **2002**, **4**, 378.
- S10 C. F. Macrae, I. J. Bruno, J. A. Chisholm, P. R. Edgington, P. McCabe, E. Pidcock, L. Rodriguez-Monge, R. Taylor, J. van de Streek and P. A. Wood, *J. Appl. Cryst.*, **2008**, **41**, 466.
- S11 SADABS, Bruker AXS Inc., Madison, Wisconsin, USA, 2012.

S12 A. J. C. Wilson. *International Tables for Crystallography*. Kluwer: Dordrecht, 1992.

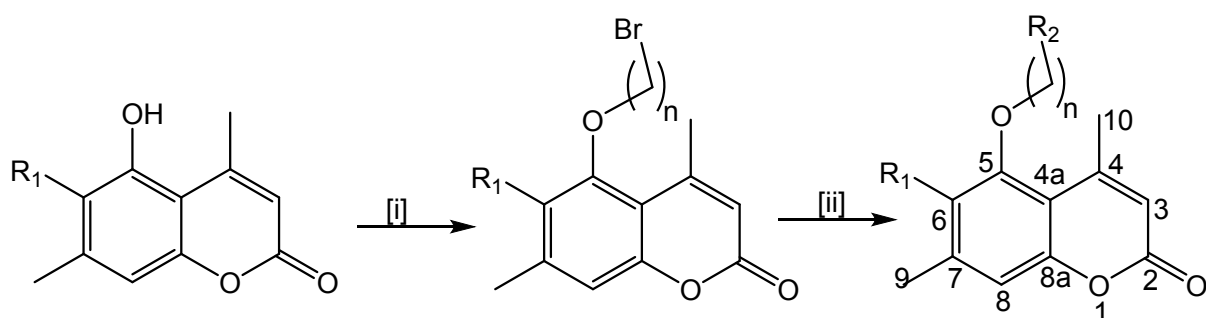
S13 Diamond – Crystal and Molecular Structure Visualization Crystal Impact – K. Brandenburg & H. Putz GbR, Rathausgasse 30, D-53111 Bonn, 2012.

ADME analysis

molecule	problems ^a	MW ^b	dipole ^c	SASA ^d	volume ^e	donorHB ^f	accptHB ^g	glob ^h	logP ⁱ	Ro5 ^j	Ro3 ^k
3a	0	424.51	8.70	768.98	1379.41	0	6.25	0.78	4.62	0	0
3b	0	410.49	9.71	732.31	1316.40	0	6.25	0.79	4.22	0	0
3c	0	424.51	8.70	768.98	1379.41	0	6.25	0.78	4.62	0	0
3d	0	452.52	5.73	772.93	1408.08	0	8.25	0.79	3.579	0	0
4a	0	408.50	7.12	769.84	1362.50	0	8.25	0.77	3.137	0	1
4b	0	394.47	7.88	727.88	1291.23	0	8.25	0.79	2.676	0	1
4c	0	450.54	5.10	790.37	1440.06	0	10.25	0.78	2.467	0	1
4d	0	436.51	5.03	756.48	1378.15	0	10.25	0.79	2.07	0	1
5a	0	436.55	7.39	796.07	1444.61	0	7	0.78	4.545	0	0
5b	0	422.52	7.18	755.04	1369.73	0	7	0.79	4.024	0	0
5c	0	478.59	7.34	810.50	1518.20	0	9	0.79	3.792	0	0
5d	0	464.56	5.45	808.09	1473.45	0	9	0.77	3.487	0	0
6a	0	331.41	7.59	636.53	1120.21	0	6.95	0.82	2.302	0	0
6b	0	317.38	7.68	598.65	1055.54	0	6.95	0.84	1.867	0	0
6c	0	373.45	4.67	663.95	1200.96	0	8.95	0.82	1.652	0	0
6d	0	359.42	6.46	620.30	1132.24	0	8.95	0.85	1.142	0	0
7a	0	451.52	7.82	794.94	1436.93	0	7.25	0.77	3.686	0	0
7b	0	437.49	6.25	768.72	1380.31	0	7.25	0.78	3.284	0	0
7c	0	493.56	9.39	831.20	1527.27	0	9.25	0.77	3.099	0	0
7d	0	479.53	8.34	791.08	1451.43	0	9.25	0.78	2.498	0	0
8a	0	440.97	7.45	780.25	1407.92	0	6.25	0.78	4.943	0	1
8b	0	426.94	9.91	741.96	1339.08	0	6.25	0.79	4.474	0	0
8c	0	483.01	6.68	802.56	1482.52	0	8.25	0.78	4.166	0	0
8d	0	468.98	3.80	795.04	1442.77	0	8.25	0.78	3.935	0	0
9a	0	475.41	6.02	803.59	1447.53	0	6.25	0.77	5.343	1	1
9b	0	461.39	9.75	767.97	1385.81	0	6.25	0.78	4.927	0	1
9c	0	517.45	7.41	825.85	1528.87	0	8.25	0.78	4.669	1	0
9d	0	503.42	2.51	792.57	1463.55	0	8.25	0.79	4.23	1	0
10a	0	431.53	7.56	796.83	1429.93	0	7.75	0.77	3.682	0	1
10b	0	417.51	12.39	766.70	1369.61	0	7.75	0.78	3.276	0	0
10d	0	459.54	7.36	805.52	1465.82	0	9.75	0.77	2.729	0	0
11a	0	407.51	7.44	760.96	1355.93	0	7.75	0.78	3.403	0	0
11b	0	393.49	9.80	710.01	1280.83	0	7.75	0.80	2.923	0	0
11c	0	449.55	3.16	800.64	1449.74	0	9.75	0.77	2.831	0	0
11d	0	435.52	6.20	758.67	1383.27	0	9.75	0.79	2.343	0	0

^aproblems – number of property or descriptor values that fall outside the 95% range of similar values for known drugs; ^bMW - molecular weight (Da); ^cdipole - dipole moment (D); ^dSASA - solvent accessible surface (Å²); ^evolume - total molecular volume (Å³); ^fdonorHB - estimated number of hydrogen bonds that would be donated by the solute to water molecules in an aqueous solution; ^gaccptHB - estimated number of hydrogen bonds that would be accepted by the solute from water molecules in an aqueous solution; ^hglob - globularity descriptor defined as $4\pi r^2/SASA$; ⁱlogP - octanol/water partition coefficient; ^jRo5 - number of violations of Lipinski's rule of five; ^kRo3 . number of violations of Jorgensen's rule of three.

NMR and HR-MS spectra analysis



1a $R_1=H$

1b $R_1=COCH_3$

2a $R_1=H, n=4$

2b $R_1=H, n=3$

2c $R_1=COCH_3, n=4$

2d $R_1=COCH_3, n=3$

3a-11a $R_1=H, n=4$

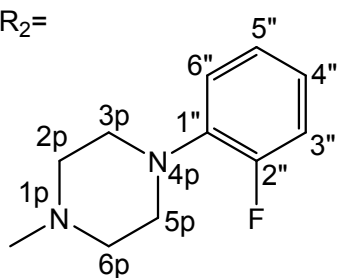
3b-11b $R_1=H, n=3$

3c-10c $R_1=COCH_3, n=4$

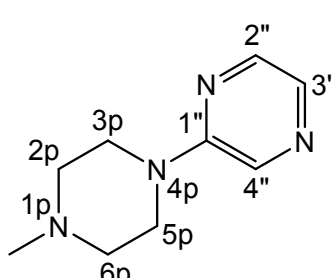
3d-10d $R_1=COCH_3, n=3$

p=piperazine

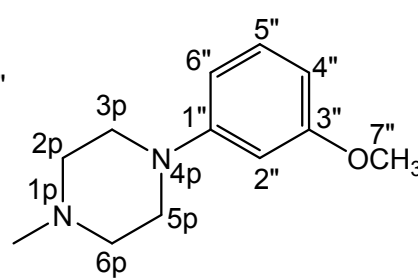
$R_2=$



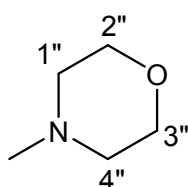
3a, 3b, 3c, 3d



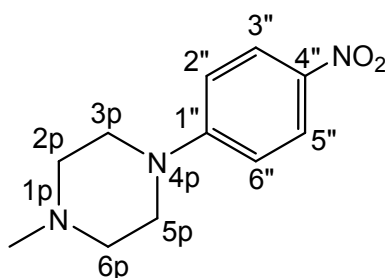
4a, 4b, 4c, 4d



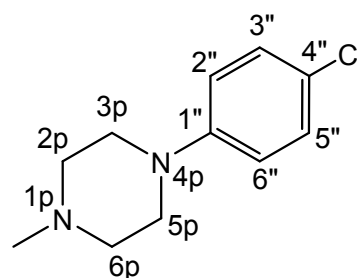
5a, 5b, 5c, 5d



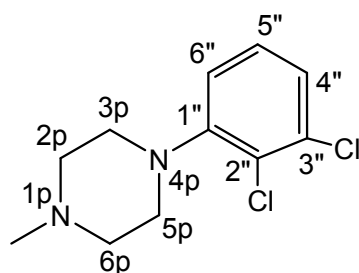
6a, 6b, 6c, 6d



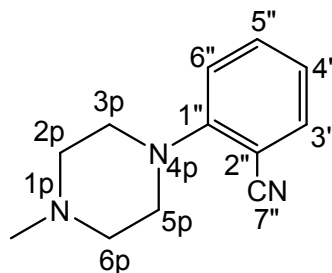
7a, 7b, 7c, 7d



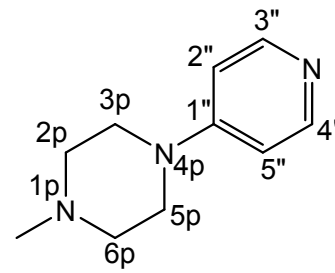
8a, 8b, 8c, 8d



9a, 9b, 9c, 9d



10a, 10b, 10d



11a, 11b, 11c, 11d

[i]: $Br(CH_2)_3Br$ or $Br(CH_2)_4Br$, KI, K_2CO_3 , ACN, MW

[ii]: HR_2 ; KI, K_2CO_3 , ACN, MW

4.2.1 5-(3-bromopropoxy)-4,7-dimethylcoumarin (**2b**)

White solid, MP: 135-137 °C, yield 99 %, Rf = 0.62, ¹H NMR (300 MHz, CDCl₃): δ = 6.77 (1H, m, H-8), 6.56 (1H, d, *J* = 0.9 Hz, H-6), 6.06 (1H, d, *J* = 1.2 Hz, H-3), 4.20 (2H, t, *J* = 5.8 Hz, H-1'), 3.60 (2H, t, *J* = 6.3 Hz, H-3'), 2.56 (3H, d, *J* = 1.5 Hz, H-9), 2.40 (5H, m, H-10, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 161.0 (C-2), 157.0 (C-8a), 155.6 (C-4), 153.8 (C-5), 143.3 (C-7), 113.9 (C-3), 110.8 (C-8), 108.4 (C-4a), 108.2 (C-6), 66.7 (C-1'), 32.3 (C-2'), 29.8 (C-3'), 24.7 (C-10), 22.2 (C-9); TOF MS ES+: [M+Na]⁺ calcd for C₁₄H₁₅O₃Na Br (333.1418) found 333.0100.

4.2.2 6-acetyl-5-(3-bromopropoxy)-4,7-dimethylcoumarin (**2d**)

White solid, MP: 146-148 °C, yield 90 %, Rf = 0.54, ¹H NMR (300 MHz, CDCl₃): δ = 6.99 (1H, d, *J* = 0.9 Hz, H-8), 6.19 (1H, d, *J* = 1.5 Hz, H-3), 3.96 (2H, t, *J* = 6.1 Hz, H-1'), 3.52 (2H, t, *J* = 6.4 Hz, H-3'), 2.61 (3H, d, *J* = 1.5 Hz, H-12), 2.55 (3H, d, *J* = 1.2 Hz, H-9), 2.29 (m, 5H, H-10, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 204.4 (C-11), 160.1 (C-2), 154.8 (C-8a), 153.8 (C-4), 151.9 (C-5), 139.3 (C-7), 133.7 (C-6), 116.2 (C-8), 115.6 (C-3), 112.6 (C-4a), 76.0 (C-1'), 33.1 (C-12), 32.8 (C-2'), 28.9 (C-3'), 22.7 (C-10), 19.5 (C-9); TOF MS ES+: [M+Na]⁺ calcd for C₁₆H₁₇O₄Na Br (375.0192) found 375.0208.

4.3.1 4,7-dimethyl-5-{3-[4-(2-fluorophenyl)piperazin-1-yl]propoxy}coumarin (**3b**)

White solid, MP: 113 –115 °C; Rf=0.47; yield 60 %; ¹H NMR (300 MHz, CDCl₃): δ = 7.04 (4H, m, H-3'', H-4'', H-5'', H-6''), 6.75 (1H, q, *J* = 0.8 Hz, H-8), 6.55 (1H, d, *J* = 0.9 Hz, H-6), 6.05 (1H, d, *J* = 1.2 Hz, H-3), 4.12 (2H, t, *J* = 6.3 Hz, H-1'), 3.15 (4H, t, *J* = 4.6 Hz, H-3p, H-5p), 2.64 (6H, m, H-2p, H-6p, H-3'), 2.59 (3H, d, *J* = 1.5 Hz, H-9), 2.39 (3H, s, H-10), 2.10 (2H, m, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 161.1 (C-2), 157.6 (C-8a), 157.3 (C-2'', J_{F-C}=238 Hz), 155.5 (C-5), 154.3 (C-4), 154.2 (C-7), 143.3 (C-1''), 124.6 (C-5'', J_{F-C}=124.9 Hz), 122.8 (C-4'', J_{F-C}=8 Hz), 119.2 (C-6''), 116.4 (C-3'', J_{F-C}=20 Hz), 116.2 (C-3), 113.7 (C-8), 110.4 (C-4a), 108.4 (C-6), 108.1 (C-1'), 67.4 (C-3p, C-5p), 55.5 (C-2p), 53.6 (C-6p), 50.6 (C-3'), 26.6 (C-2'), 24.8 (C-1'), 22.2 (C-9); TOF MS ES+: [M+Na]⁺ calcd for C₂₄H₂₇O₃N₂FNa (433.4428) found 433.1901.

4.3.2 4,7-dimethyl-5-{3-[4-(pyrazin-2-yl)piperazin-1-yl]propoxy}coumarin (**4b**)

White solid, MP: 116 –118 °C; Rf = 0.11, yield 58%; ¹H NMR (300 MHz, CDCl₃): δ = 8.15 (1H, d, *J* = 1.5 Hz, H-2''), 8.06 (1H, m, H-4''), 7.86 (1H, d, *J* = 2.4 Hz, H-3''), 6.75 (1H, q, *J* =

1.0 Hz, H-8), 6.55 (1H, d, $J = 0.9$ Hz H-6), 6.05 (1H, d, $J = 1.2$ Hz, H-3), 4.13 (2H, t, $J = 6.3$ Hz, H-1'), 3.64 (4H, t, $J = 4.6$ Hz, H-3p, H-5p), 2.62 (m, 6H, H-2p, H-6p, H-3'), 2.58 (3H, s, H-9), 2.39 (3H, s, H-10), 2.15 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.1$ (C-2), 157.3 (C-8a), 155.5 (C-5), 154.1 (C-1''), 143.2 (C-4), 141.9 (C-7), 131.2 (C-2'', C-3''), 113.7 (C-4''), 110.4 (C-3, C-8a), 108.3 (C-4a), 108.1 (C-6), 67.2 (C-1'), 55.5 (C-3p, C-5p), 52.9 (C-2p, C-6p), 44.6 (C-3'), 24.8 (C-10, C-2'), 22.1 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{22}\text{H}_{26}\text{O}_3\text{N}_4\text{Na}$ (417.4301) found 417.1914.

4.3.3 4,7-dimethyl-5-{3-[4-(3-methoxyphenyl)piperazin-1-yl]propoxy}coumarin (**5b**)

White solid, MP: 148–149 °C; Rf = 0.29, yield 61%, ^1H NMR (300 MHz, CDCl_3): $\delta = 7.17$ (1H, t, $J = 8.1$ Hz, H-5''), 6.74 (1H, q, $J = 1$ Hz, H-8), 6.54 (2H, m, H-6'', H-6), 6.47 (1H, t, $J = 2.2$ Hz, H-2''), 6.43 (1H, dd, $J = 11.0, 3.6$ Hz, H-4''), 6.04 (1H, d, $J = 1.2$ Hz, H-3), 4.15 (2H, t, $J = 6.3$ Hz, H-1'), 3.79 (3H, s, H-7''), 3.22 (4H, t, $J = 5.1$ Hz, H-3p, H-5p), 2.64 (6H, m, H-2p, H-6p, H-3'), 2.58 (3H, s, H-9), 2.38 (3H, s, H-10), 2.09 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.1$ (C-2), 160.8 (C-3''), 157.3 (C-8a), 155.5 (C-5), 154.1 (C-4), 152.6 (C-1''), 143.2 (C-7), 130.0 (C-5''), 113.7 (C-3), 110.4 (C-8), 109.1 (C-2''), 108.4 (C-4a), 108.1 (C-6), 104.8 (C-4''), 102.9 (C-6''), 67.4 (C-1'), 55.5 (C-7''), 55.4 (C-3p, C-5p), 53.4 (C-2p, C-6p), 49.0 (C-2'), 26.6 (C-10), 24.8 (C-3'), 22.2 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{25}\text{H}_{30}\text{O}_4\text{N}_2\text{Na}$ (445.4771) found 445.2117.

4.3.4 4,7-dimethyl-5-[3-(morpholin-4-yl)propoxy]coumarin (**6b**)

White solid, MP: 118 – 120 °C; Rf = 0.16, yield 74 %; ^1H NMR (300 MHz, CDCl_3): $\delta = 6.73$ (1H, q, $J = 1.0$ Hz, H-8), 6.54 (1H, d, $J = 0.9$ Hz, H-6), 6.03 (1H, d, $J = 1.2$ Hz, H-3), 4.09 (2H, t, $J = 6.3$ Hz, H-1'), 3.73 (4H, t, $J = 4.6$ Hz, H-2'', H-3''), 2.57 (3H, s, H-9), 2.54 - 2.46 (6H, m, H-1'', H-4'', H-3'), 2.38 (3H, s, H-10), 2.05 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.1$ (C-2), 157.3 (C-8a), 155.5 (C-5), 154.1 (C-4), 143.2 (C-7), 113.6 (C-3), 110.3 (C-8), 108.3 (C-4a), 108.0 (C-6), 67.3 (C-1'), 67.0 (C-2'', C-3''), 55.8 (C-3'), 53.9 (C-1', C-4'), 26.4 (C-2', C-10), 22.1 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{18}\text{H}_{23}\text{O}_4\text{NNa}$ (340.3383) found 340.1540.

4.3.5 4,7-dimethyl-5-{3-[4-(4-nitrophenyl)piperazin-1-yl]propoxy}coumarin (**7b**)

White solid, MP: 195 – 198 °C; Rf = 0.15, yield 45%; ^1H NMR (300 MHz, CDCl_3): $\delta = 8.14$ (2H, d, $J = 9.3$ Hz, H-3'', H-5''), 6.84 (2H, m, H-2'', H-6''), 6.75 (1H, d, $J = 0.6$ Hz, H-8), 6.54 (1H, d, $J = 0.9$ Hz, H-6), 6.05 (1H, d, $J = 1.2$ Hz, H-3), 4.13 (2H, t, $J = 6.3$ Hz, H-1'), 3.46

(4H, m, H-3p, H-5p), 2.63 (6H, m, H-2p, H-6p, H-3'), 2.58 (3H, d, $J = 1.2$ Hz, H-9), 2.39 (3H, s, H-10), 2.10 (3H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.0$ (C-2), 157.2 (C-1''), 155.5 (C-4, C-5), 154.8 (C-8a), 153.9 (C-7), 143.2 (C-4''), 126.1 (C-3'', C-5''), 113.8 (C-4a), 113.0 (C-6), 110.5 (C-2'', C-6''), 108.4 (C-3), 108.1 (C-8), 67.2 (C-1'), 55.3 (C-3'), 52.9 (C-3p, C-5p), 47.1 (C-2p, C-6p), 26.64 (C-3'), 24.8 (C-10), 22.2 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{24}\text{H}_{27}\text{O}_5\text{N}_3\text{Na}$ (460.4496) found 460.1515

4.3.6 4,7-dimethyl-5-{3-[4-(4-chlorophenyl)piperazin-1-yl]propoxy}coumarin (**8b**)

White solid, MP: 153 – 155 °C, Rf = 0.05, yield 71%; ^1H NMR (300 MHz, CDCl_3): $\delta = 7.22$ (2H, m, H-3'', H-5''), 6.85 (2H, m, H-2'', H-6''), 6.75 (1H, q, $J = 1.0$ Hz, H-8), 6.54 (1H, d, $J = 0.9$ Hz, H-6), 6.05 (1H, d, $J = 1.2$ Hz, H-3), 4.12 (2H, t, $J = 6.3$ Hz, H-1'), 3.19 (4H, t, $J = 3$ Hz, H-3p, H-5p), 2.64 (6H, m, H-2p, H-6p, H-3'), 2.59 (3H, s, H-9), 2.39 (3H, s, H-10), 2.09 (3H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.1$ (C-2), 157.2 (C-8a), 155.1 (C-5), 154.0 (C-4, C-1''), 143.3 (C-7), 129.2 (C-3'', C-5''), 117.7 (C-2'', C-6''), 113.8 (C-4'', C-3), 110.6 (C-8), 108.4 (C-4a), 108.1 (C-6), 67.2 (C-1'), 55.5 (C-3p), 53.2 (C-5p), 49.0 (C-2p, C-6p), 26.3 (C-3'), 24.8 (C-10), 23.8 (C-2'), 22.2 (C-9); TOF MS ES+: $[\text{M}+\text{NaCl}]^+$ calcd for $\text{C}_{24}\text{H}_{27}\text{O}_3\text{N}_2\text{NaCl}$ (449.8974) found 449.1619.

4.3.7 4,7-dimethyl-5-{3-[4-(2,3-dichlorophenyl)piperazin-1-yl]propoxy}coumarin (**9b**)

White solid, MP: 170 – 173 °C; Rf = 0.17, yield 82%; ^1H NMR (300 MHz, CDCl_3): $\delta = 7.16$ (2H, m, H-4'', H-6''), 6.97 (1H, m, H-5''), 6.75 (1H, q, $J = 0.9$ Hz, H-8), 6.55 (1H, s, 1H, H-6), 6.05 (1H, d, $J = 1.2$ Hz, H-3), 4.12 (2H, t, $J = 6.3$ Hz, H-1'), 3.09 (4H, s, H-3p, H-5p), 2.67 (6H, m, H-2p, H-6p, H-3'), 2.59 (3H, s, H-9), 2.39 (3H, s, H-10), 2.09 (3H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 161.0$ (C-2), 155.6 (C-8a, C-5), 154.7 (C-4), 150.6 (C-1''), 143.3 (C-7), 134.4 (C-3'', C-5''), 127.8 (C-2'', C-6''), 119.0 (C-4''), 113.9 (C-3), 110.7 (C-8), 108.4 (C-4a), 108.2 (C-6), 67.2 (C-1'), 55.5 (C-3p, C-5p), 53.3 (C-2p, C-6p), 24.8 (C-3', C-10), 22.2 (C-2', C-9); TOF MS ES+: $[\text{M}+2\text{Cl}]^+$ calcd for $\text{C}_{24}\text{H}_{26}\text{O}_3\text{N}_2\text{Cl}_2$ (483.3425) found 483.1234.

4.3.8 4,7-dimethyl-5-{3-[4-(2-cyanophenyl)piperazin-1-yl] propoxy}coumarin (**10b**)

White solid, MP: 171 – 172 °C; Rf = 0.14, yield 83%; ^1H NMR (300 MHz, CDCl_3): $\delta = 7.58$ (1H, dd, $J = 7.9, 1.5$ Hz, H-3''), 7.49 (1H, m, H-4''), 7.01 (2H, m, H-5'', H-6''), 6.75 (1H, m, H-8), 6.55 (1H, d, $J = 0.9$ Hz, H-6), 6.05 (1H, d, $J = 1.2$ Hz, H-3), 4.12 (2H, t, $J = 6.3$ Hz, H-1'), 3.27 (4H, t, $J = 4.6$ Hz, H-3p, H-5p), 2.72 (6H, m, H-2p, H-6p, H-3'), 2.59 (3H, d, $J = 1.2$

Hz, H-9), 2.39 (3H, s, H-10), 2.10 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): δ = 161.1 (C-2), 157.3 (C-8a), 155.5 (C-1'', C-5), 154.1 (C-4), 143.2 (C-7), 134.5 (C-3''), 134.0 (C-5''), 118.9 (C-7''), 118.6 (C-4''), 113.7 (C-6''), 110.5 (C-2''), 108.3 (C-3), 108.1 (C-4a, C-8), 106.2 (C-6), 67.2 (C-1'), 58.6 (C-3p), 55.3 (C-5p), 53.3 (C-2p), 51.6 (C-6p), 24.8 (C-3'), 22.2 (C-10, C-2'), 22.1 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{25}\text{H}_{27}\text{O}_3\text{N}_3\text{Na}$ (440.4604) found 440.1962.

4.3.9 4,7-dimethyl-5-(3-(4-(pyridin-4-yl)piperazin-1-yl)propoxy)coumarin (**11b**)

White solid, MP: 157 – 159 °C; Rf = 0.06, yield 48 %; ^1H NMR (300 MHz, CDCl_3): δ = 8.28 (2H, m, H-4'', H-3''), 6.75 (1H, d, J = 0.6 Hz, H-8), 6.69 (2H, dd, J = 5.1, 1.5 Hz, H-2'', H-5''), 6.54 (1H, d, J = 7.0 Hz, H-6), 6.05 (1H, d, J = 1.2 Hz, H-3), 4.12 (2H, t, J = 6.3 Hz, H-1'), 3.38 (4H, t, J = 5.1 Hz, H-3p, H-5p), 2.61 (6H, m, H-2p, H-6p, H-3'), 2.58 (3H, s, H-9), 2.39 (3H, s, H-10), 2.08 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): δ = 160.0 (C-2), 157.3 (C-8a), 155.5 (C-5), 155.4 (C-18), 154.0 (C-4), 148.6 (C-3'', C-4''), 143.2 (C-7), 113.7 (C-3), 110.4 (C-8), 108.4 (C-2'', C-5''), 108.0 (C-4a, C-6), 67.2 (C-1'), 55.2 (C-3p, C-5p), 52.8 (C-2p, C-6p), 46.2 (C-3'), 26.7 (C-10), 24.7 (C-2'), 22.1 (C-9); TOF MS ES+: $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{23}\text{H}_{27}\text{O}_3\text{N}_3\text{Na}$ (394.4594) found 394.2139.

4.3.10 6-acetyl-4,7-dimethyl-5-{3-[4-(2-fluorophenyl)piperazin-1-yl]propoxy}coumarin (**3d**)

White solid, MP: 99 – 102 °C; Rf = 0.24, yield 56 %; ^1H NMR (300 MHz, CDCl_3): δ = 6.97 (5H, m, H-8, H-3'', H-4'', H-5'', H-6''), 6.18 (1H, d, J = 1.5 Hz, H-3), 3.90 (2H, t, J = 6.7 Hz, H-1'), 3.15 (4H, t, J = 4.5 Hz, H-3p, H-5p), 2.67 (4H, m, H-2p, H-6p), 2.62 (3H, d, J = 1.2 Hz, H-3'), 2.54 (3H, m, H-12), 2.55 (3H, s, H-9), 2.29 (3H, d, J = 0.6 Hz, H-10), 2.00 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): δ = 204.6 (C-11), 160.2 (C-2), 157.5 (C-2'', $J_{\text{F-C}}=245$ Hz), 154.9 (C-8a), 154.4 (C-4), 152.3 (C-5), 140.11 (C-1'', $J_{\text{F-C}}=9$ Hz), 139.4 (C-7), 124.7 (C-5'', $J_{\text{F-C}}=4$ Hz), 122.8 (C-4'', $J_{\text{F-C}}=8$ Hz), 119.2 (C-6'', $J_{\text{F-C}}=3$ Hz), 116.3 (C-3'', $J_{\text{F-C}}=20$ Hz), 116.2 (C-8), 115.4 (C-3), 112.7 (C-4a), 110.9 (C-6), 76.6 (C-1'), 54.6 (C-3'), 53.3 (C-3p, C-5p), 50.4 (C-2p, C-6p), 32.7 (C-12), 27.1 (C-2'), 22.7 (C-10), 19.5 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{26}\text{H}_{29}\text{O}_4\text{N}_2\text{FNa}$ (475.2009) found 475.2025.

4.3.11 6-acetyl-4,7-dimethyl-5-{3-[4-(pyrazin-2-yl)piperazin-1-yl]propoxy}coumarin (**4d**)

White solid, MP: 118 – 120 °C; Rf = 0.11, yield 58 %; ^1H NMR (300 MHz, CDCl_3): δ = 8.15 (1H, d, J = 1.5 Hz, H-2''), 8.07 (1H, m, H-4''), 7.87 (1H, d, J = 2.4 Hz, H-3''), 6.98 (1H, d, J = 0.6 Hz, H-8), 6.18 (1H, d, J = 1.2 Hz, H-3), 3.91 (t, J = 6.6 Hz, 2H, H-1'), 3.67 (4H, m, H-3p,

H-5p), 2.60 (9H, m, H-2p, H-6p, H-3', H-12), 2.55 (3H, s, H-9), 2.29 (3H, d, $J = 0.6$ Hz, H-10), 2.03 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 204.55$ (C-11), 160.2 (C-2), 155.2 (C-1''), 154.9 (C-5), 154.4 (C-4), 152.2 (C-8a), 141.9 (C-2''), 139.4 (C-7), 133.7 (C-3''), 133.2 (C-4''), 131.2 (C-6), 116.1 (C-3), 115.4 (C-8), 112.7 (C-4a), 76.6 (C-1'), 54.6 (C-3'), 52.9 (C-3p, C-5p), 44.6 (C-2p, C-6p), 32.7 (C-12), 27.2 (C-2'), 22.7 (C-10), 19.5 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{24}\text{H}_{28}\text{O}_4\text{N}_4\text{Na}$ (459.2011) found 459.2008.

4.3.12 6-acetyl-4,7-dimethyl-5-{3-[4-(3-methoxyphenyl)piperazin-1-yl]propoxy}coumarin (**5d**)

White solid, MP: 93 – 95 °C; Rf = 0.22, yield 55 %; ^1H NMR (300 MHz, CDCl_3): $\delta = 7.17$ (1H, t, $J = 8.2$ Hz, H-5''), 6.97 (1H, d, $J = 0.6$ Hz, H-2''), 6.55 (1H, m, H-8), 6.47-6.40 (2H, m, H-4'', H-6''), 6.17 (1H, d, $J = 1.2$ Hz, H-3), 3.90 (2H, t, $J = 6.6$ Hz, H-1'), 3.79 (3H, s, H-7''), 3.27 (4H, t, $J = 4.8$ Hz, H-3p, H-5p), 2.64 (6H, m, H-2p, H-6p, H-3'), 2.61 (3H, d, $J = 1.5$ Hz, H-12), 2.55 (3H, s, H-9), 2.29 (2H, d, $J = 0.6$ Hz, H-10), 2.00 (2H, t, $J = 7$ Hz, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 204.6$ (C-11), 160.8 (C-3''), 160.1 (C-2), 154.8 (C-5), 152.6 (C-4), 152.2 (C-8a), 139.3 (C-1''), 133.7 (C-7), 130.0 (C-5''), 116.2 (C-6), 115.4 (C-3), 112.7 (C-8), 109.1 (C-4''), 105.8 (C-4a), 104.8 (C-6''), 102.8 (C-2''), 76.6 (C-1'), 55.4 (C-3'), 54.6 (C-3p, C-5p), 53.2 (C-7''), 49.0 (C-2p, C-6p), 32.7 (C-12), 27.0 (C-2'), 22.7 (C-10), 19.5 (C-9); TOF MS ES+: $[\text{M}+\text{H}]^+$ calcd for $\text{C}_{27}\text{H}_{33}\text{O}_5\text{N}_2$ (465.2374) found 465.2389.

4.3.13 6-acetyl-4,7-dimethyl-5-[3-(morpholin-4-yl)propoxycoumarin (**6d**)

White solid, MP: 128 – 131 °C; Rf = 0.27, yield 80 %; ^1H NMR (300 MHz, CDCl_3): $\delta = 6.97$ (1H, d, $J = 0.3$ Hz, H-8), 6.18 (1H, d, $J = 0.6$ Hz, H-3), 3.88 (2H, t, $J = 6.6$ Hz, H-1'), 3.74 (4H, t, $J = 5$ Hz, H-2'', H-3''), 2.61 (3H, d, $J = 1$ Hz, H-12), 2.54 (3H, s, H-9), 2.48 (6H, m, H-1'', H-4'', H-3'), 2.29 (3H, s, H-10), 1.96 (2H, d, $J = 6.7$ Hz, H-2'); NMR ^{13}C (75 MHz, CDCl_3): $\delta = 204.5$ (C-11), 160.1 (C-2), 154.8 (C-8a), 152.1 (C-5), 143.3 (C-4), 139.3 (C-7), 133.7 (C-6), 116.2 (C-8), 115.4 (C-3), 113.8 (C-4a), 112.7 (C-1'), 67.1 (C-2''), 76.4 (C-3''), 66.7 (C-1''), 55.0 (C-4''), 32.7 (C-12), 26.6 (C-2'), 24.7 (C-3'), 22.6 (C-10), 19.5 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{20}\text{H}_{25}\text{O}_5\text{NNa}$ (382.3826) found 382.1630.

4.3.14 6-acetyl-4,7-dimethyl-5-{3-[4-(4-nitrophenyl)piperazin-1-yl]propoxy}coumarin (**7d**)

White solid, MP: 172 – 174 °C; Rf = 0.27, yield 45 %; ^1H NMR (300 MHz, CDCl_3): $\delta = 8.14$ (2H, d, $J = 9.3$ Hz, H-3'', H-5''), 6.98 (1H, s, H-8), 6.85 (2H, d, $J = 9.3$ Hz, H-2'', H-6''), 6.18 (1H, s, H-3), 3.91 (2H, t, $J = 6.5$ Hz, H-1'), 3.47 (4H, m, H-3p, H-5p), 2.61 (3H, s, H-12),

2.58 (6H, m, H-2p, H-6p, H-3'), 2.55 (3H, s, H-9), 2.29 (3H, s, H-10), 2.12 (2H, m, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 204.6 (C-11), 160.6 (C-2), 154.8 (C-5, C-1''), 152.1 (C-4), 147.5 (C-8a), 139.2 (C-7), 133.7 (C-4''), 126.1 (C-5'', C-3''), 116.2 (C-6), 115.5 (C-3), 113.1 (C-8), 113.0 (C-2'', C-6''), 112.7 (C-4a), 78.2 (C-1'), 54.5 (C-3'), 52.6 (C-3p, C-5p), 47.0 (C-2p, C-6p), 32.8 (C-12), 24.8 (C-2'), 22.6 (C-10), 19.5 (C-9); TOF MS ES+: [M+Na]⁺ calcd for C₂₆H₂₉O₆N₃Na (502.4840) found 502.1955.

4.3.15 6-acetyl-4,7-dimethyl-5-{3-[4-(4-chlorophenyl)piperazin-1-yl]propoxy}coumarin (**8d**)

White solid, MP: 135 – 138 °C; R_f = 0.05, yield 86%; ¹H NMR (300 MHz, CDCl₃): δ = 7.20 (2H, m, H-3'', H-5''), 6.97 (1H, d, *J* = 0.9 Hz, H-8), 6.84 (2H, m, H-2'', H-6''), 6.17 (1H, d, *J* = 1.5 Hz, H-3), 3.90 (2H, t, *J* = 6.7 Hz, H-1'), 3.17 (4H, m, H-3p, H-5p), 2.58 (9H, m, H-12, H-2p, H-6p, H-3'), 2.55 (3H, s, H-9), 2.29 (3H, s, H-10), 1.98 (2H, m, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 204.6 (C-11), 160.2 (C-2), 154.8 (C-8a), 154.2 (C-5), 152.2 (C-4), 150.0 (C-1''), 139.4 (C-7), 133.7 (C-4''), 129.1 (C-5''), 117.4 (C-2'', C-6''), 116.1 (C-6), 115.4 (C-8), 113.7 (C-3), 112.7 (C-3''), 110.4 (C-4a), 76.6 (C-1'), 54.5 (C-3p, C-5p), 53.3 (C-2p, C-6p), 49.2 (C-12), 32.7 (C-2'), 27.2 (C-3'), 22.7 (C-10), 19.5 (C-9); TOF MS ES+: [M+NaCl]⁺ calcd for C₂₆H₂₉O₄N₂NaCl (491.9318) found 491.1702.

4.3.16 6-acetyl-4,7-dimethyl-5-{3-[4-(2,3-dichlorophenyl)piperazin-1-yl]propoxy}coumarin (**9d**)

White solid, MP: 142 – 145 °C; R_f = 0.24, yield 54 %; ¹H NMR (300 MHz, CDCl₃): δ = 7.15 (2H, m, H-4'', H-5''), 6.98 (2H, m, H-8, H-6''), 6.18 (1H, d, *J* = 1.2 Hz, H-8), 3.90 (2H, t, *J* = 6.7 Hz, H-1'), 3.08 (4H, s, H-3p, H-5p), 2.63 (9H, m, H-2p, H-6p, H-3', H-12), 2.56 (3H, s, H-9), 2.29 (3H, s, H-10), 1.99 (2H, m, H-2'); ¹³C NMR (75 MHz, CDCl₃): δ = 204.5 (C-11), 160.2 (C-2), 154.9 (C-8a), 154.4 (C-5), 152.3 (C-4), 151.3 (C-1''), 139.4 (C-7), 134.2 (C-5''), 133.7 (C-3''), 127.7 (C-6''), 124.9 (C-4''), 118.8 (C-2''), 116.1 (C-8), 115.4 (C-3, C-6), 112.7 (C-4a), 76.6 (C-1'), 54.6 (C-3p, C-5p), 53.4 (C-2p), 51.4 (C-6p), 32.7 (C-12), 27.2 (C-2'), 22.7 (C-3'), 22.8 (C-10), 19.5 (C-9); TOF MS ES+: [M+Na]⁺ calcd for C₂₆H₂₈O₄N₂Cl₂Na (525.3821) found 525.1320.

4.3.17 6-acetyl-4,7-dimethyl-5-{3-[4-(2-cyanophenyl)piperazin-1-yl]propoxy}coumarin (**10d**)

White solid, MP: 103 – 106 °C; R_f = 0.42, yield 87 %; ¹H NMR (300 MHz, CDCl₃): δ = 7.59 (1H, dd, *J* = 7.9, 1.5 Hz, H-5''), 7.51 (1H, m, H-3''), 7.03 (2H, m, H-4'', H-6''), 7.00 (1H, s, H-

8), 6.19 (1H, H-3), 3.92 (2H, t, $J = 6.6$ Hz, H-1'), 3.29 (4H, s, H-3p, H-5p), 2.72 (4H, s, H-2p, H-6p), 2.63 (3H, s, H-12), 2.61 (2H, m, H-3'), 2.57 (3H, s, H-9), 2.30 (3H, s, H-10), 2.09 (2H, t, $J = 6.1$ Hz, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 204.6$ (C-11), 160.2 (C-2), 155.7 (C-8a), 154.8 (C-5''), 154.3 (C-5), 152.2 (C-4), 143.2 (C-7), 139.4 (C-3''), 134.5 (C-2''), 134.0 (C-6''), 118.9 (C-4''), 116.2 (C-3), 115.4 (C-8), 112.7 (C-4a), 108.1 (C-6), 106.3 (C-1'), 76.5 (C-11), 55.4 (C-1'), 54.5 (C-3'), 53.2 (C-3p, C-5p), 51.4 (C-2p, C-6p), 32.8 (C-12), 26.9 (C-10), 22.7 (C-2'), 19.5 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{27}\text{H}_{29}\text{O}_4\text{N}_3\text{Na}$ (482.2736) found 482.2663.

4.3.18 6-acetyl-4,7-dimethyl-5-(3-(4-(pyridin-4-yl)piperazin-1-yl)propoxy)coumarin (**11d**)

White solid, MP: 113 –115 °C; Rf = 0.85, yield 75 %; ^1H NMR (300 MHz, CDCl_3): $\delta = 8.26$ (2H, m, H-3'', H-4''), 6.97 (1H, s, H-8), 6.66 (2H, m, H-2'', H-5''), 6.17 (1H, d, $J = 1.5$ Hz, H-3), 3.90 (2H, t, $J = 6.7$ Hz, H-1'), 3.33 (4H, m, H-3p, H-5p), 3.00 (4H, m, H-2p, H-6p), 2.66 (6H, m, H-12, H-9), 2.29 (5H, m, H-10, H-3'), 1.97 (2H, m, H-2'); ^{13}C NMR (75 MHz, CDCl_3): $\delta = 204.5$ (C-11), 160.2 (C-2), 155.4 (C-5), 155.1 (C-4), 154.8 (C-8a), 152.1 (C-1''), 150.6 (C-3''), 150.1 (C-4''), 150.0 (C-7), 116.1 (C-6), 115.3 (C-3), 112.6 (C-8), 108.5 (C-4a), 108.4 (C-2'', C-5''), 76.5 (C-1'), 54.4 (C-3'), 47.1 (C-3p, C-5p), 46.0 (C-2p, C-6p), 32.7 (C-12), 27.2 (C-2'), 22.6 (C-10), 19.4 (C-9); TOF MS ES+: $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{25}\text{H}_{29}\text{O}_4\text{N}_1\text{Na}$ (458.4756) found 458.2053.

NMR spectra

2b
DG013 w CDC13 + TMS

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Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

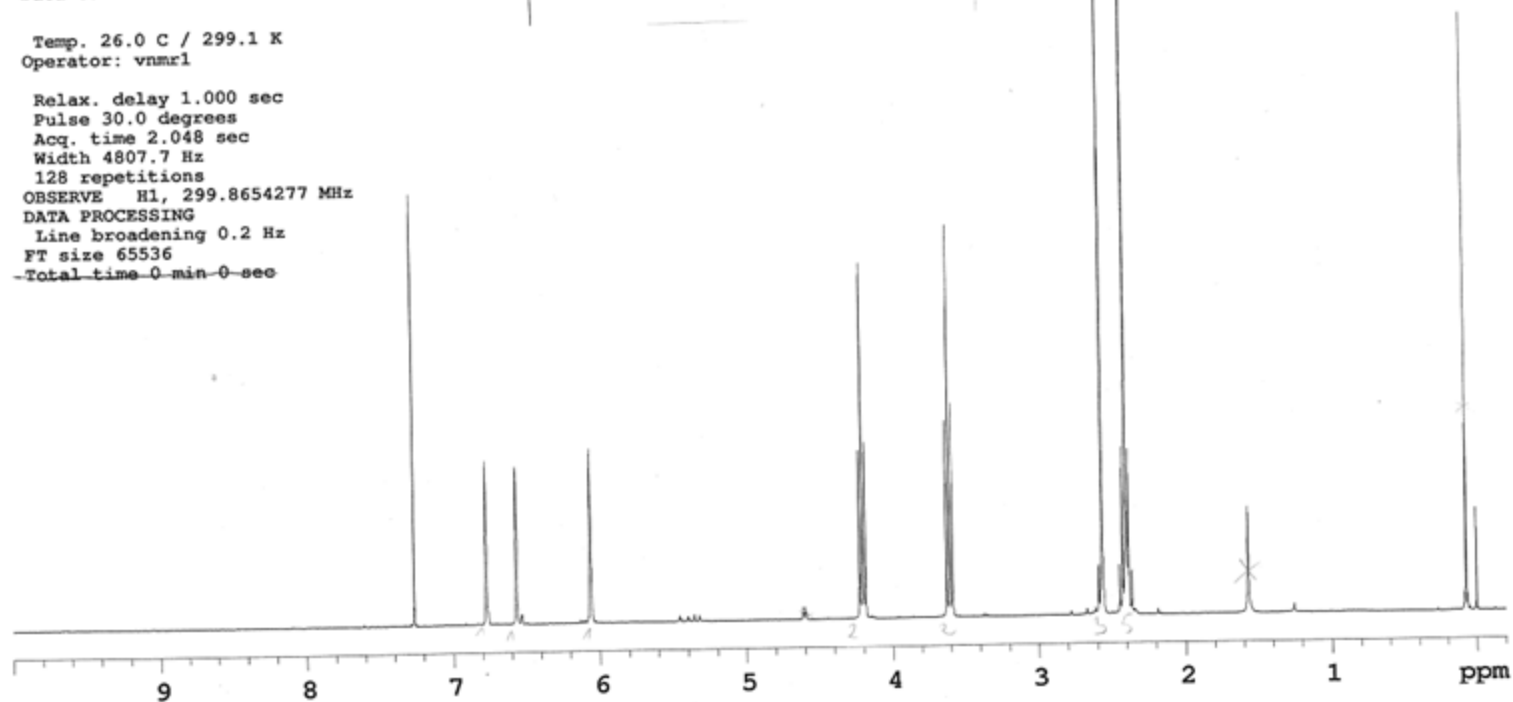
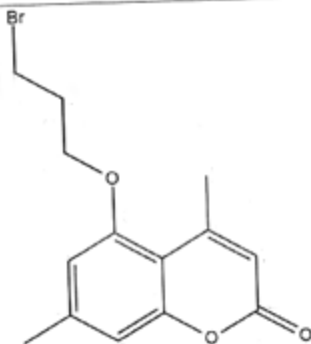
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Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Nov 12 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654277 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
-Total time 0 min 0 sec

2b



DG-413 w CDCl3 13C-NMR

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

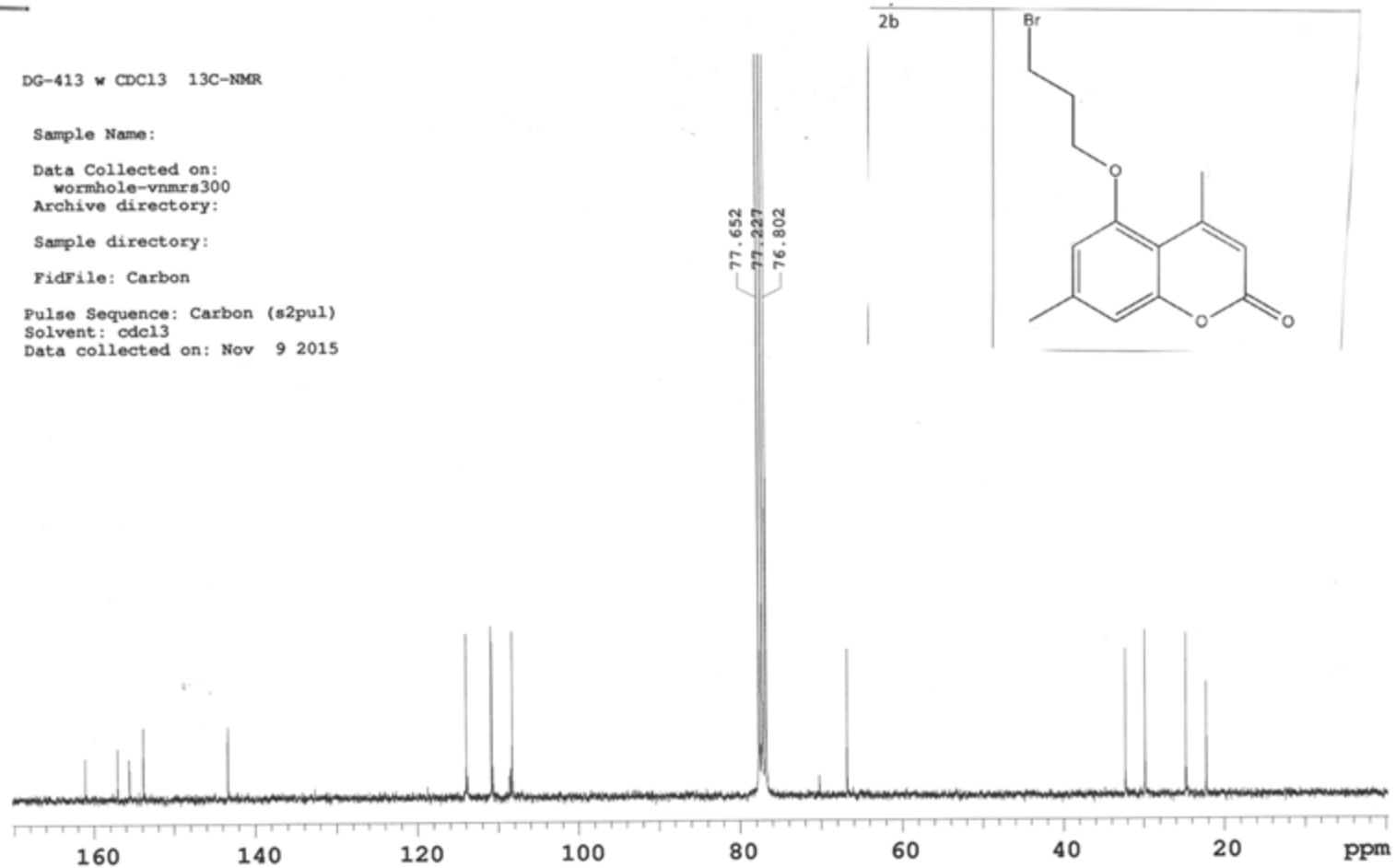
Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)

Solvent: cdcl3

Data collected on: Nov 9 2015



3b

DG414a w CDC13 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

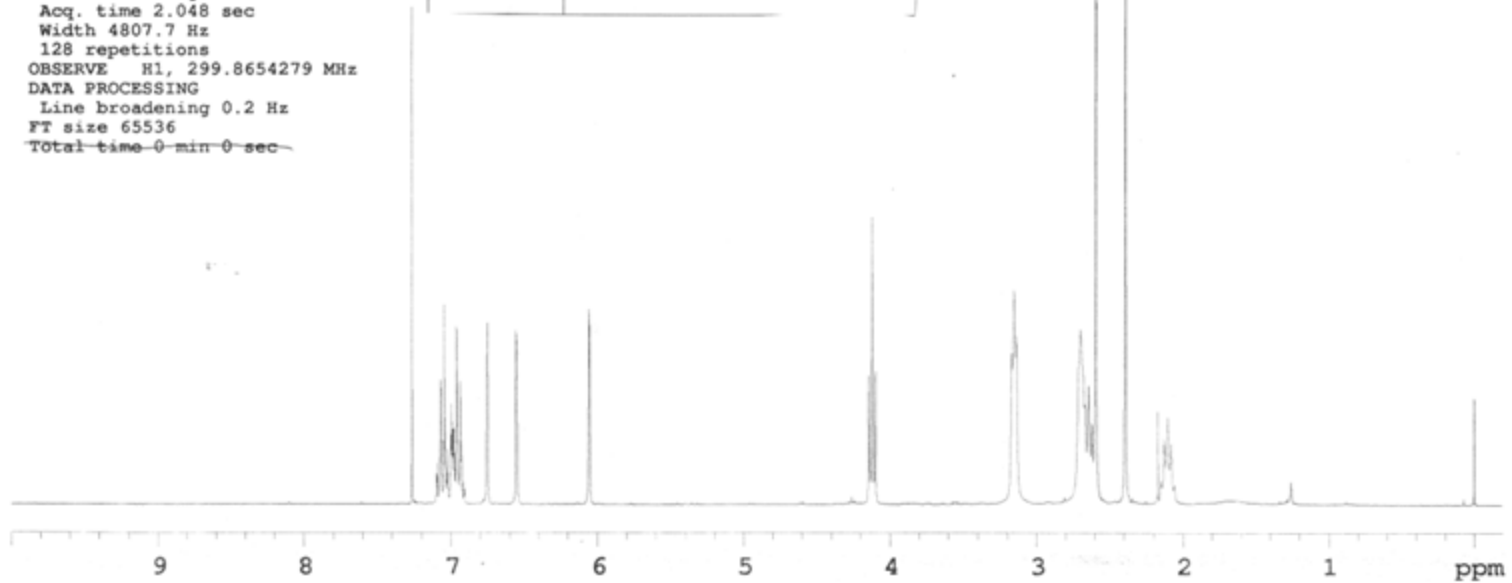
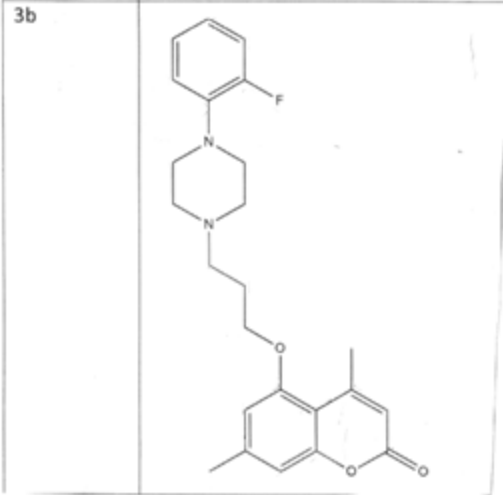
Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Sep 21 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654279 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec



3b

DG 414a 13C

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

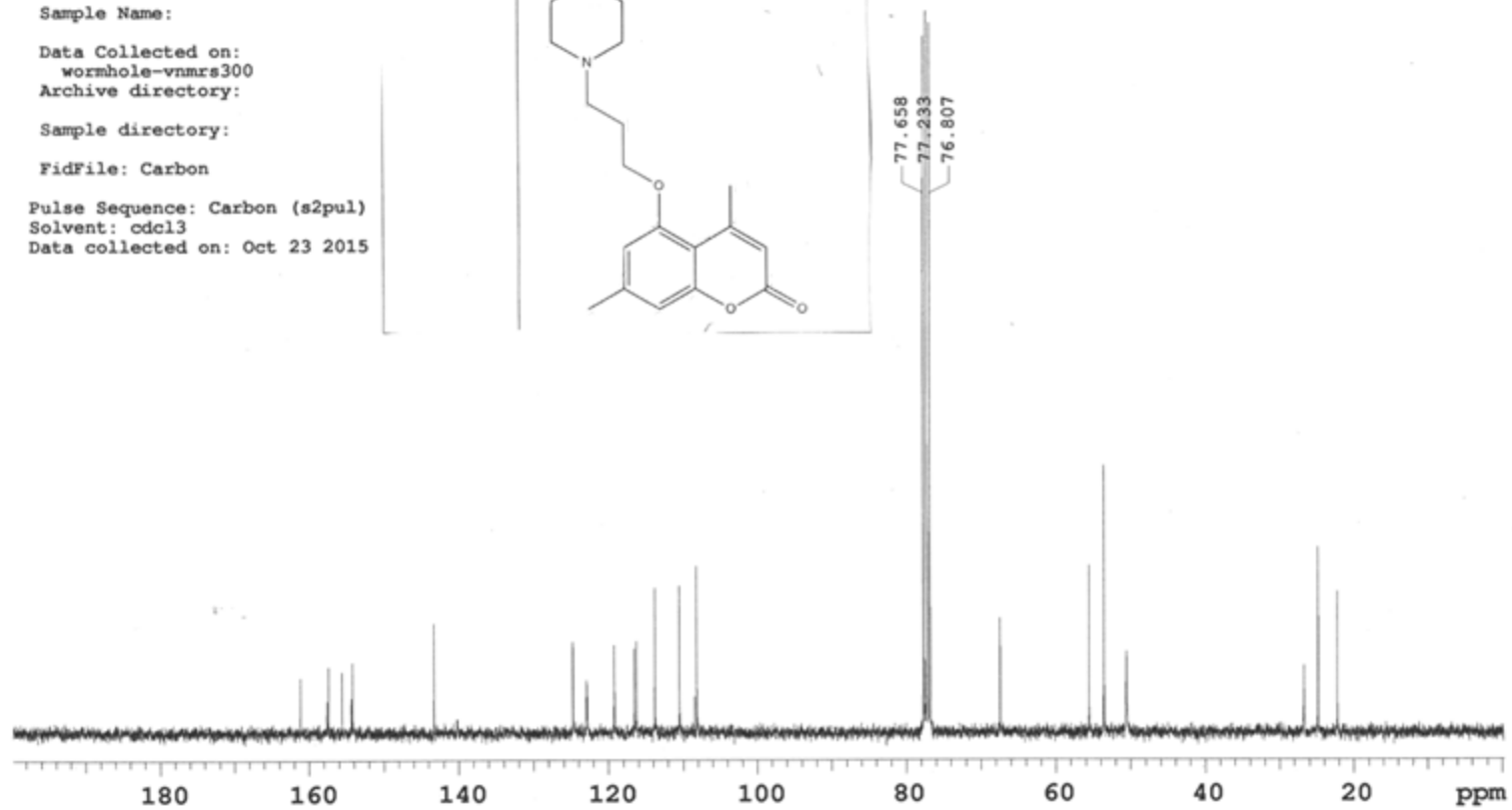
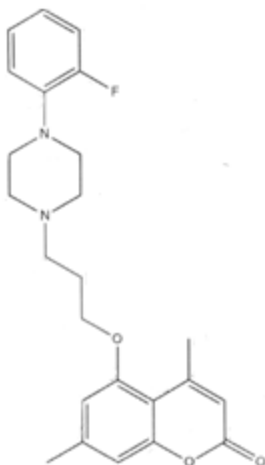
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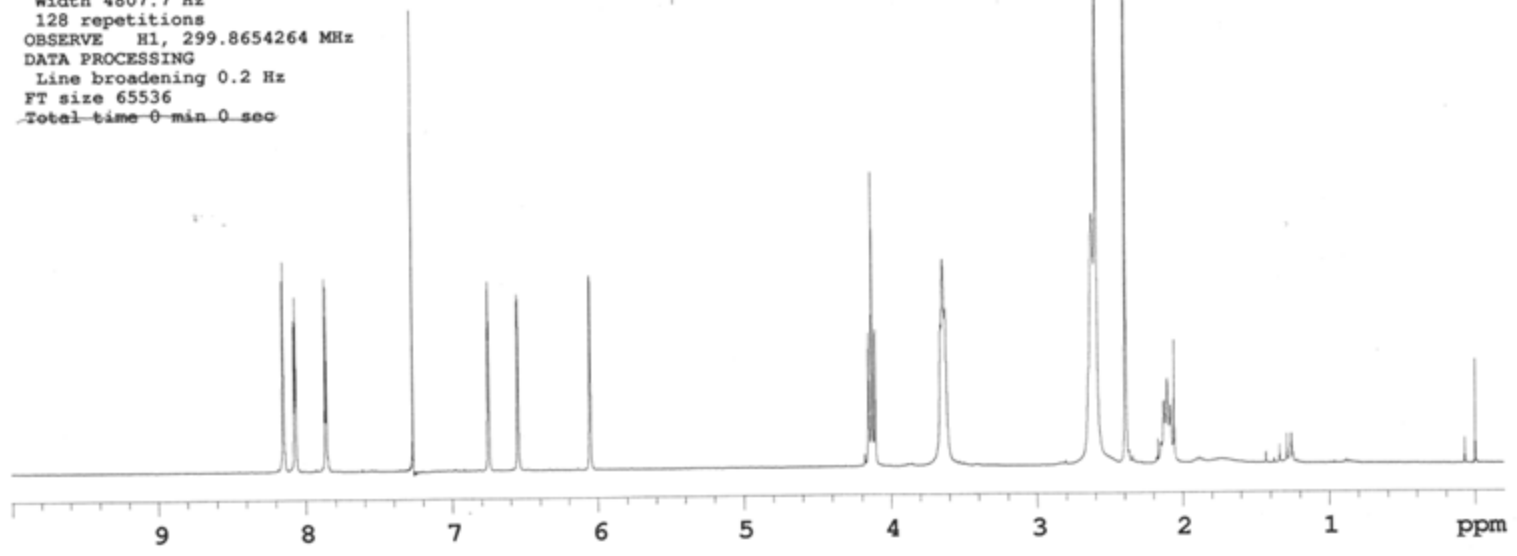
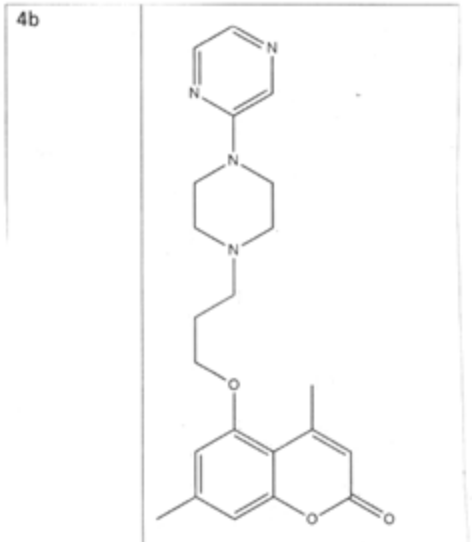
Pulse Sequence: Carbon (s2pul)

Solvent: cdcl3

Data collected on: Oct 23 2015



46
DG-415 w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Oct 21 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654264 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec



CHORG
DG-415 13C w CDCl3

161.10
157.28
155.49
154.09

143.24
141.93

131.24

113.71
110.45
108.34
108.08

77.53
77.22
76.90

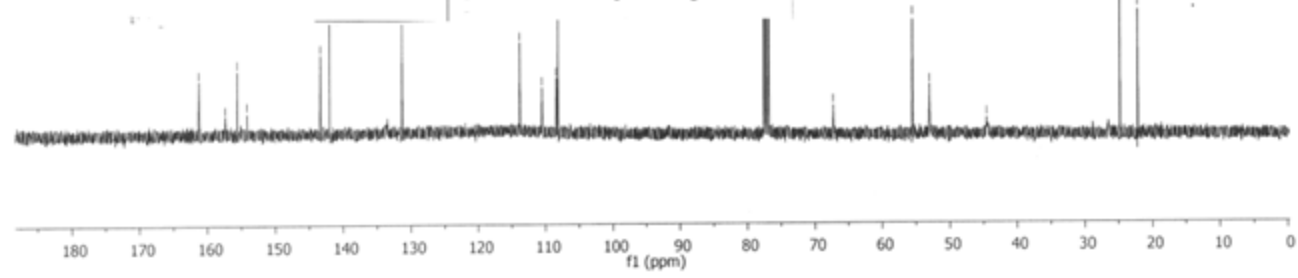
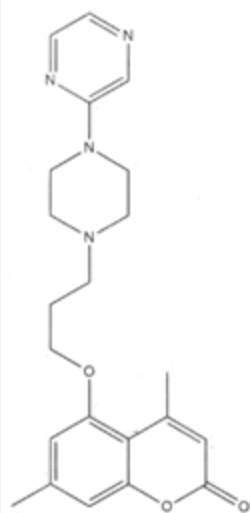
67.22

55.48
52.97

44.46

24.80
22.18

4b



5b

DG-416 w CDCl3 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

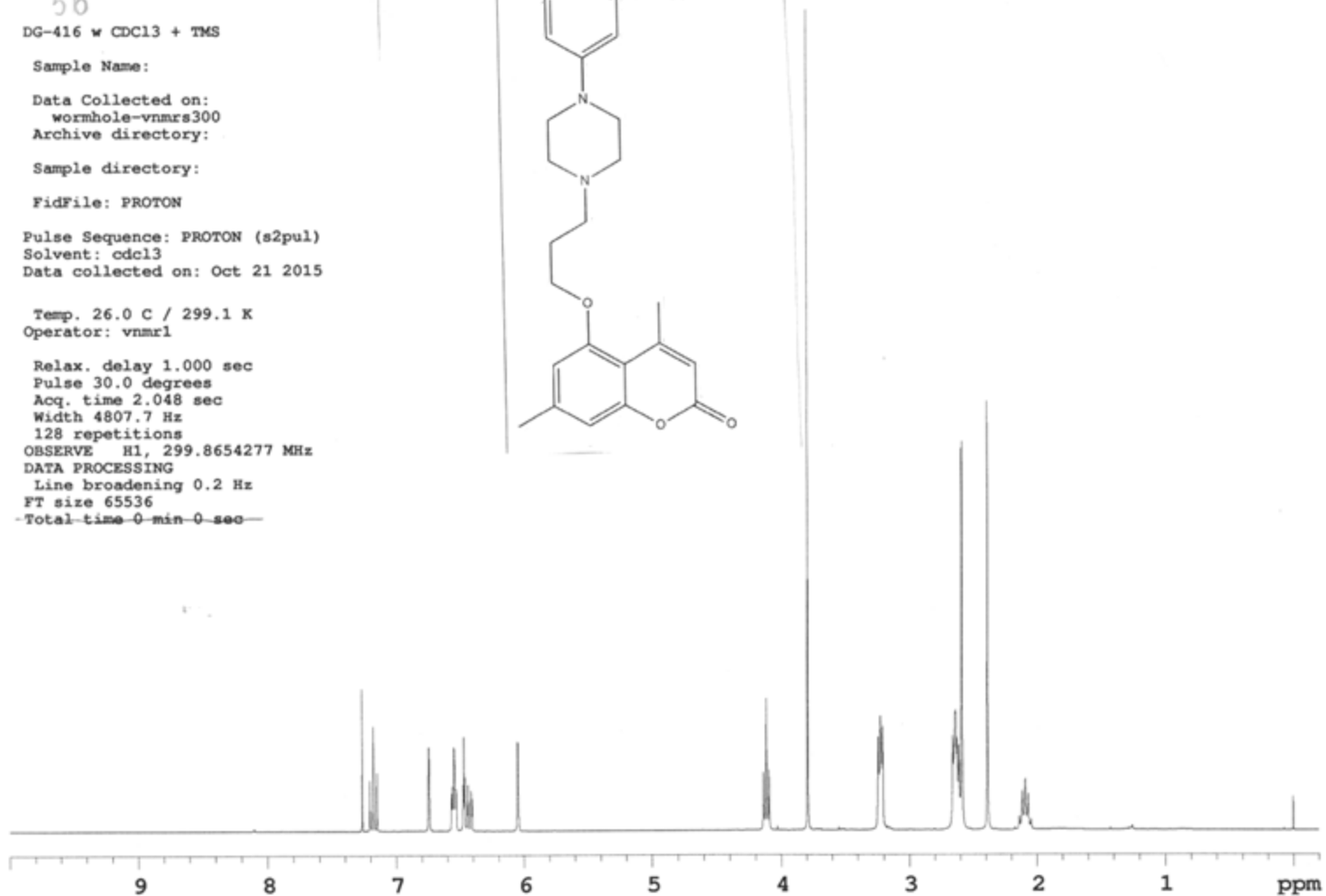
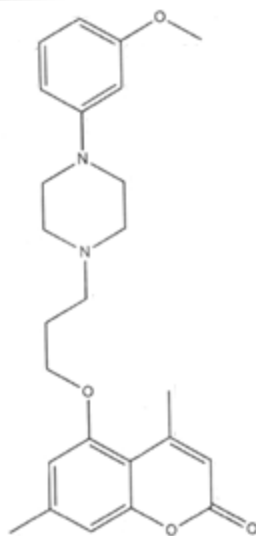
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Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Oct 21 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654277 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
~~Total time 0 min 0 sec~~

5b



DG 416 13C

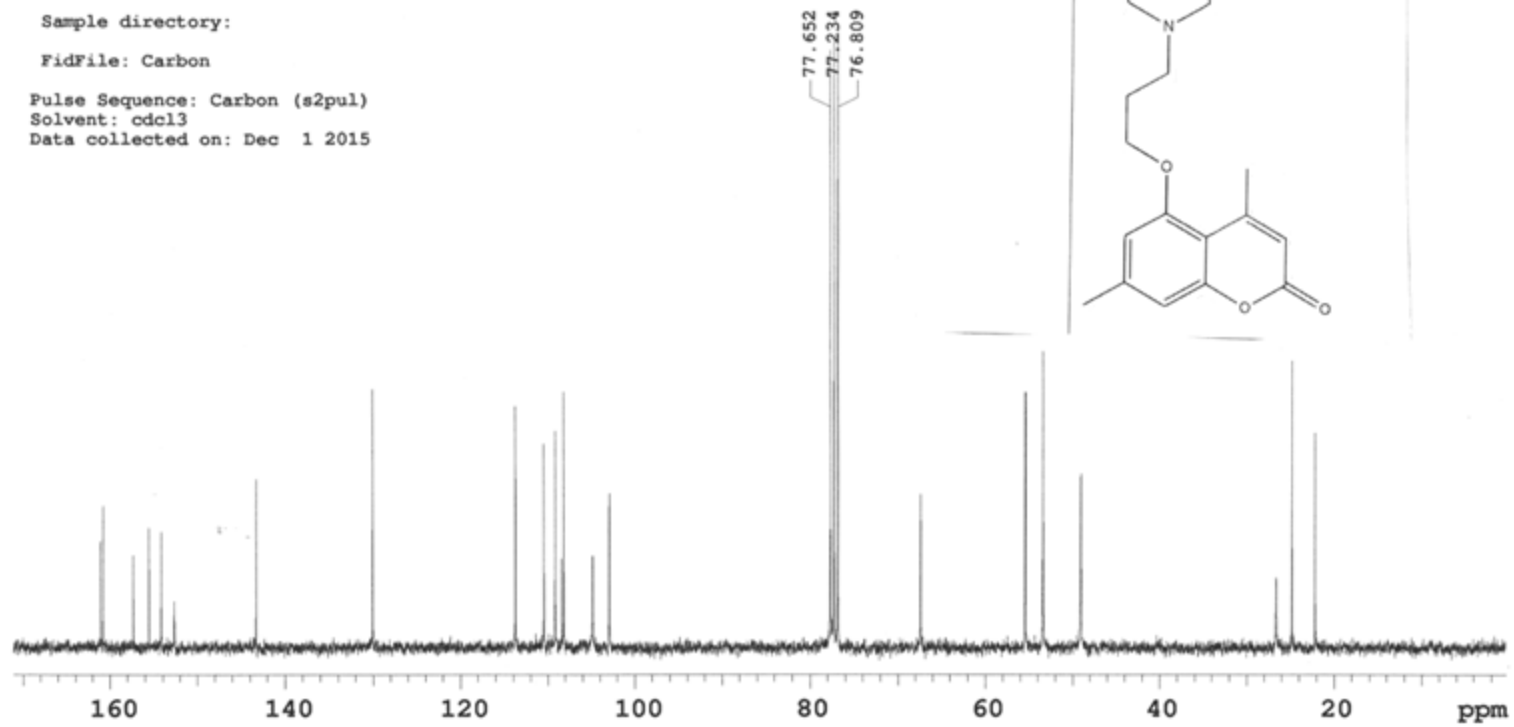
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Dec 1 2015



6b

DG-417 w CDCl3 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

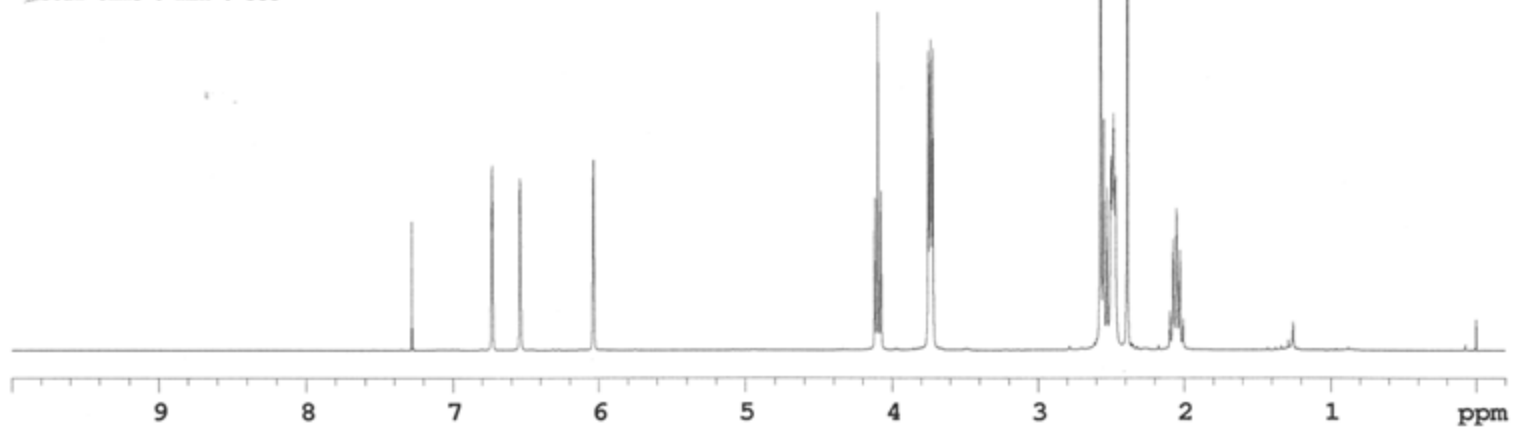
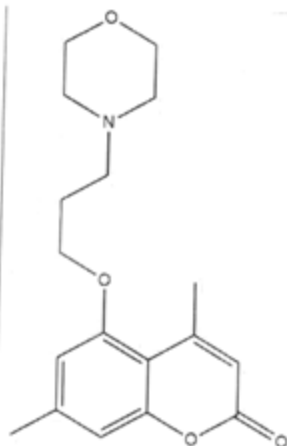
FidFile: PROTON

Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Oct 21 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654230 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

6b



DG 417 13C

Sample Name:

Data Collected on:
wormhole-vnmrs300

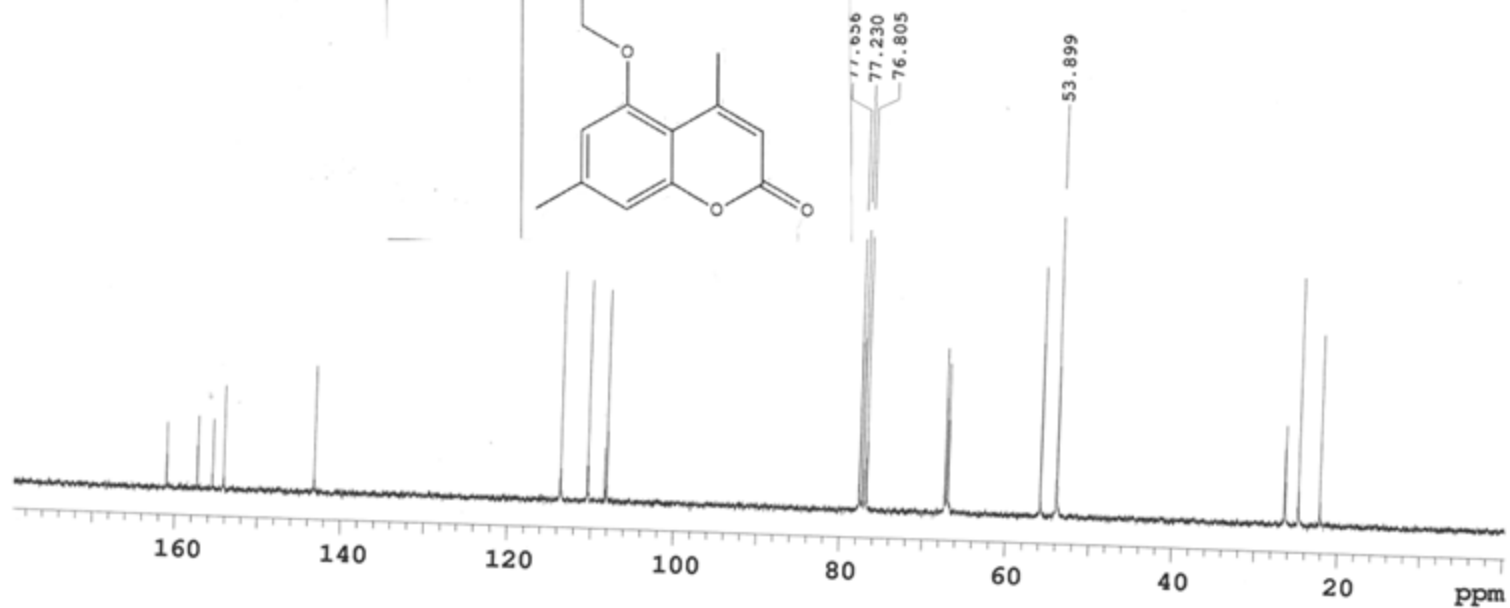
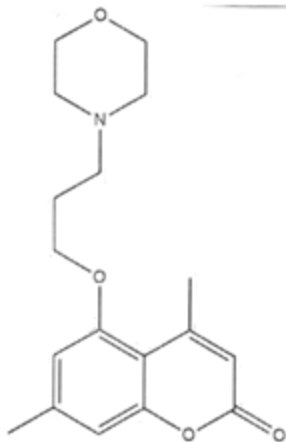
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Sample directory:

FidFile: Carbon

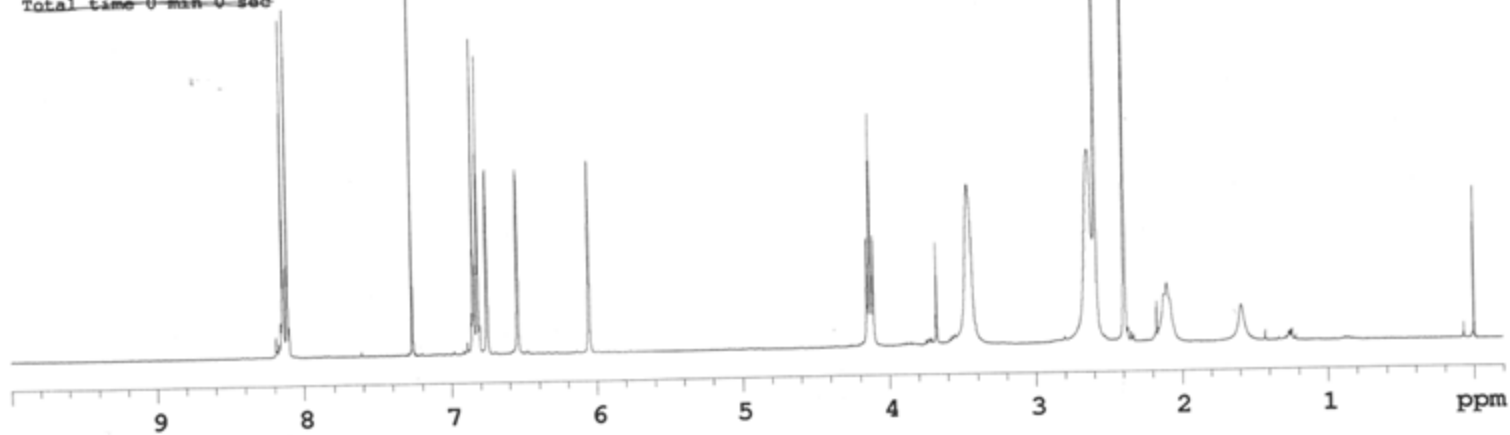
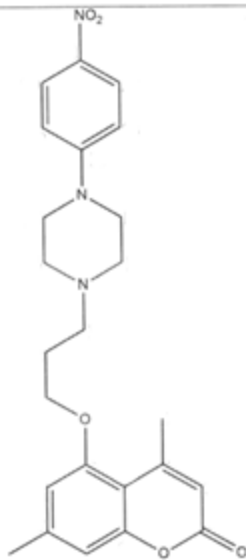
Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Dec 1 2015

6b



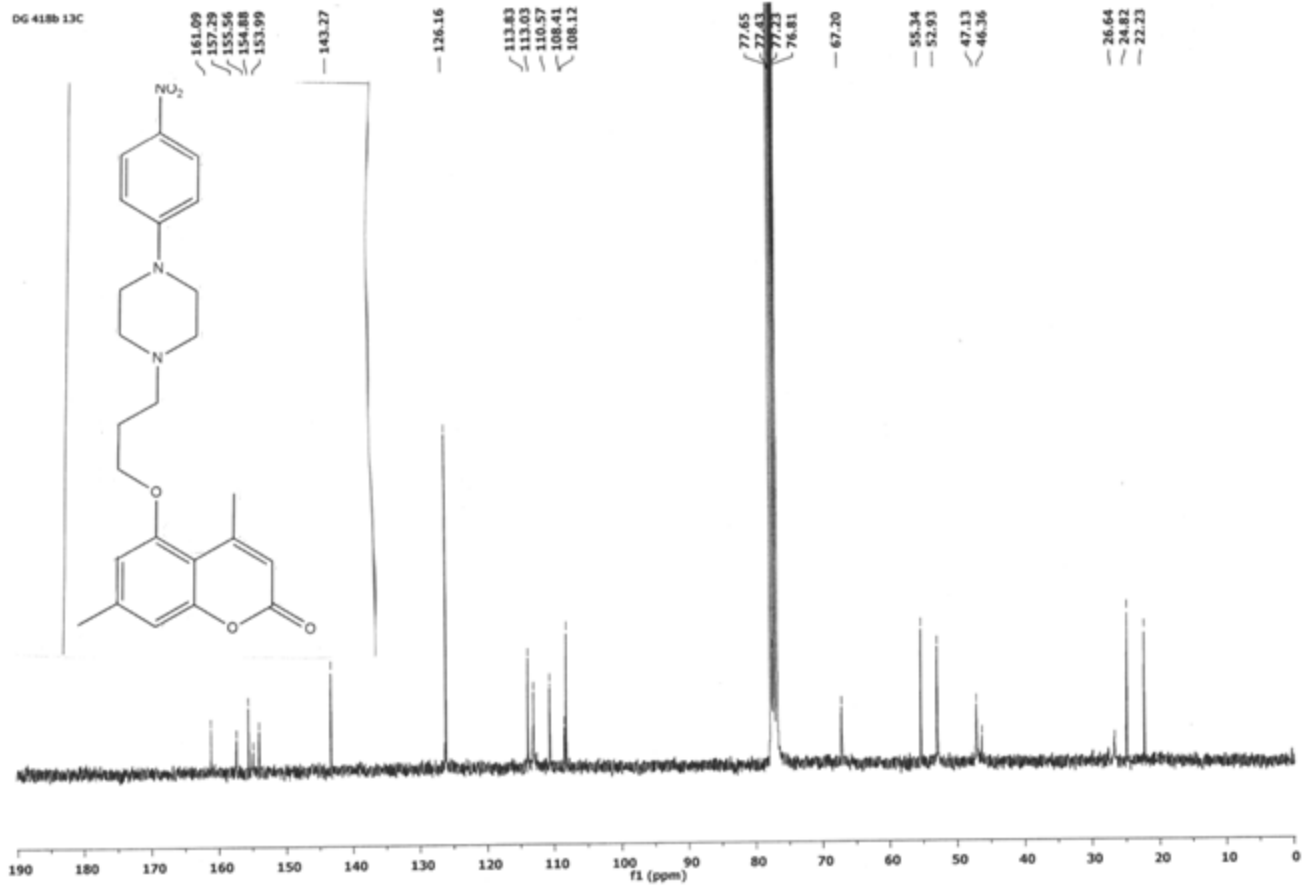
7b
DG-418b w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Oct 21 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654274 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

7b

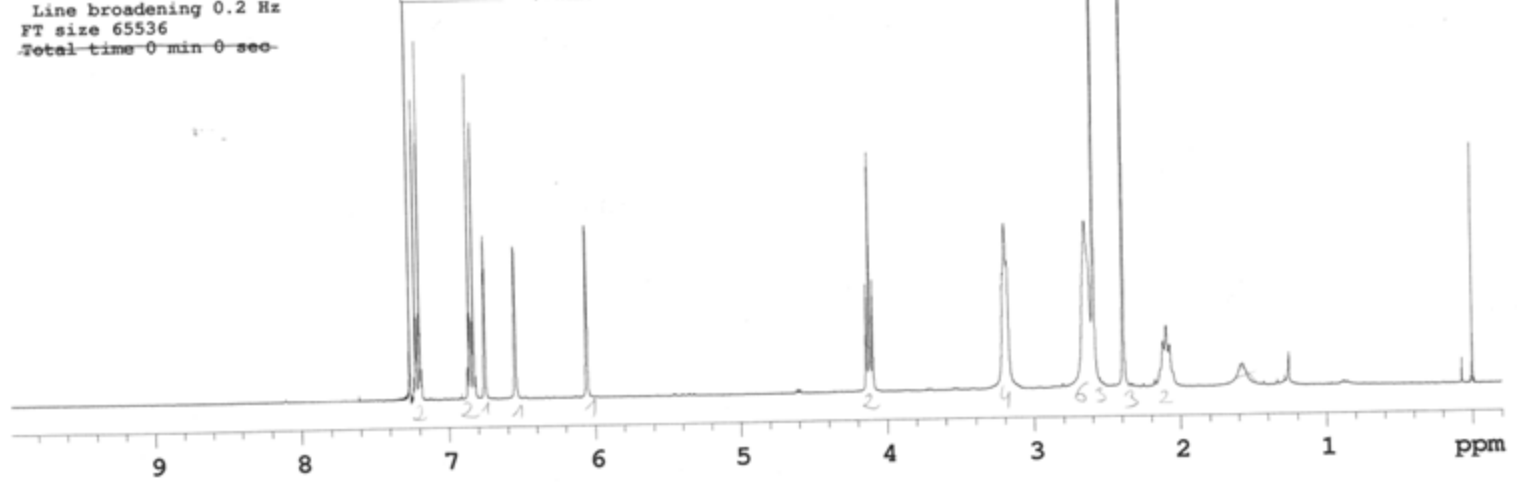
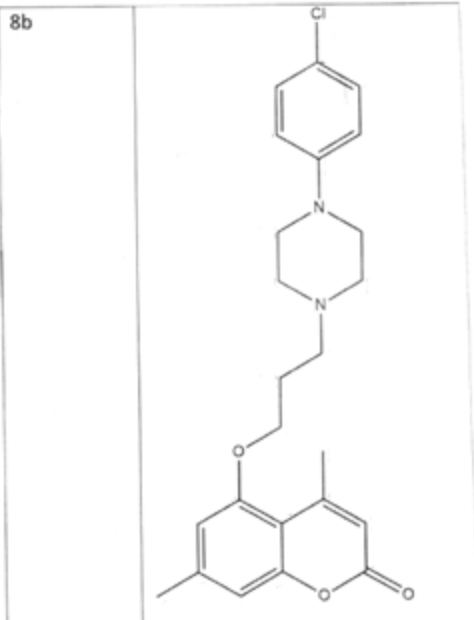


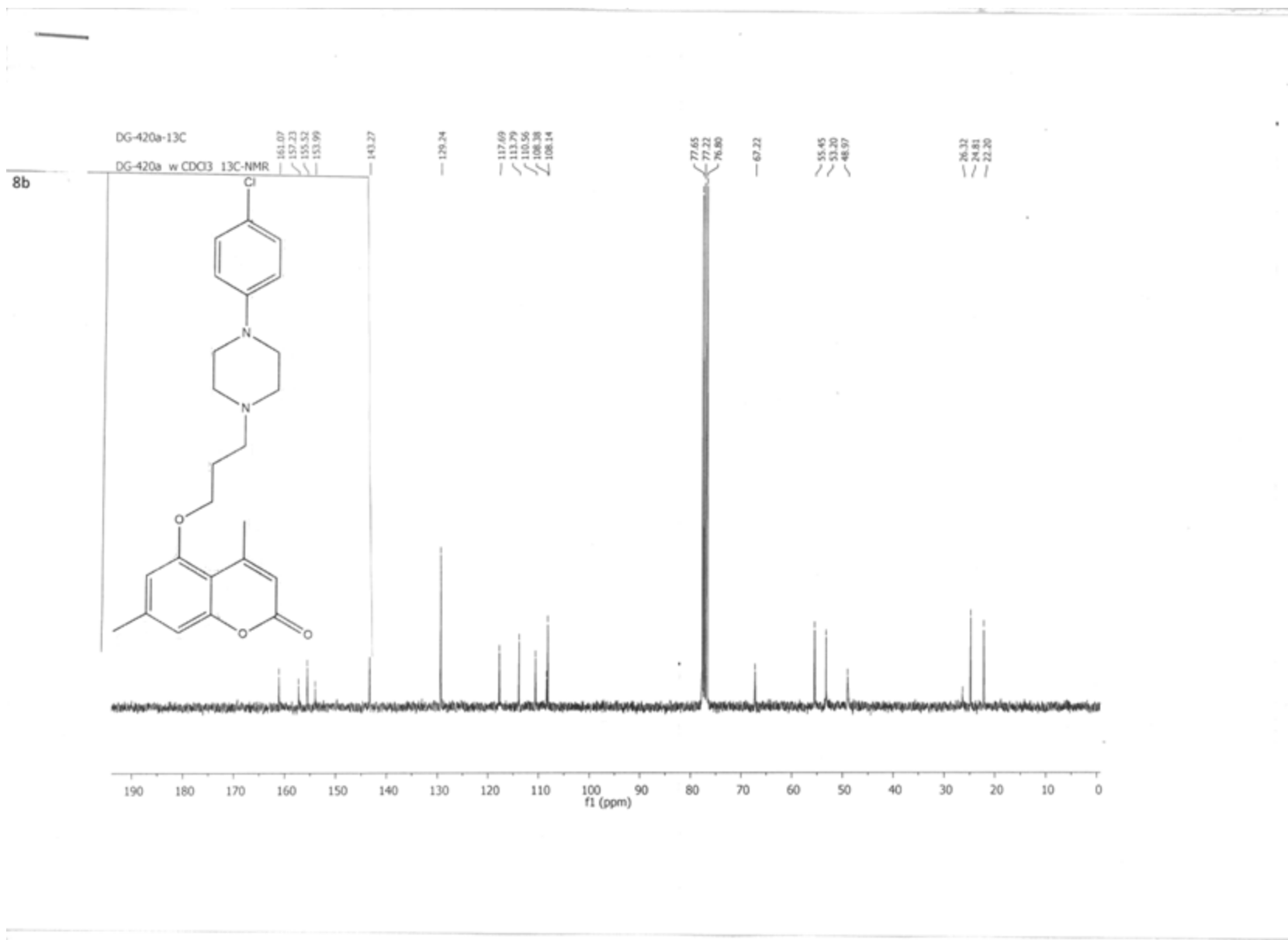
7b

06 418b 13C



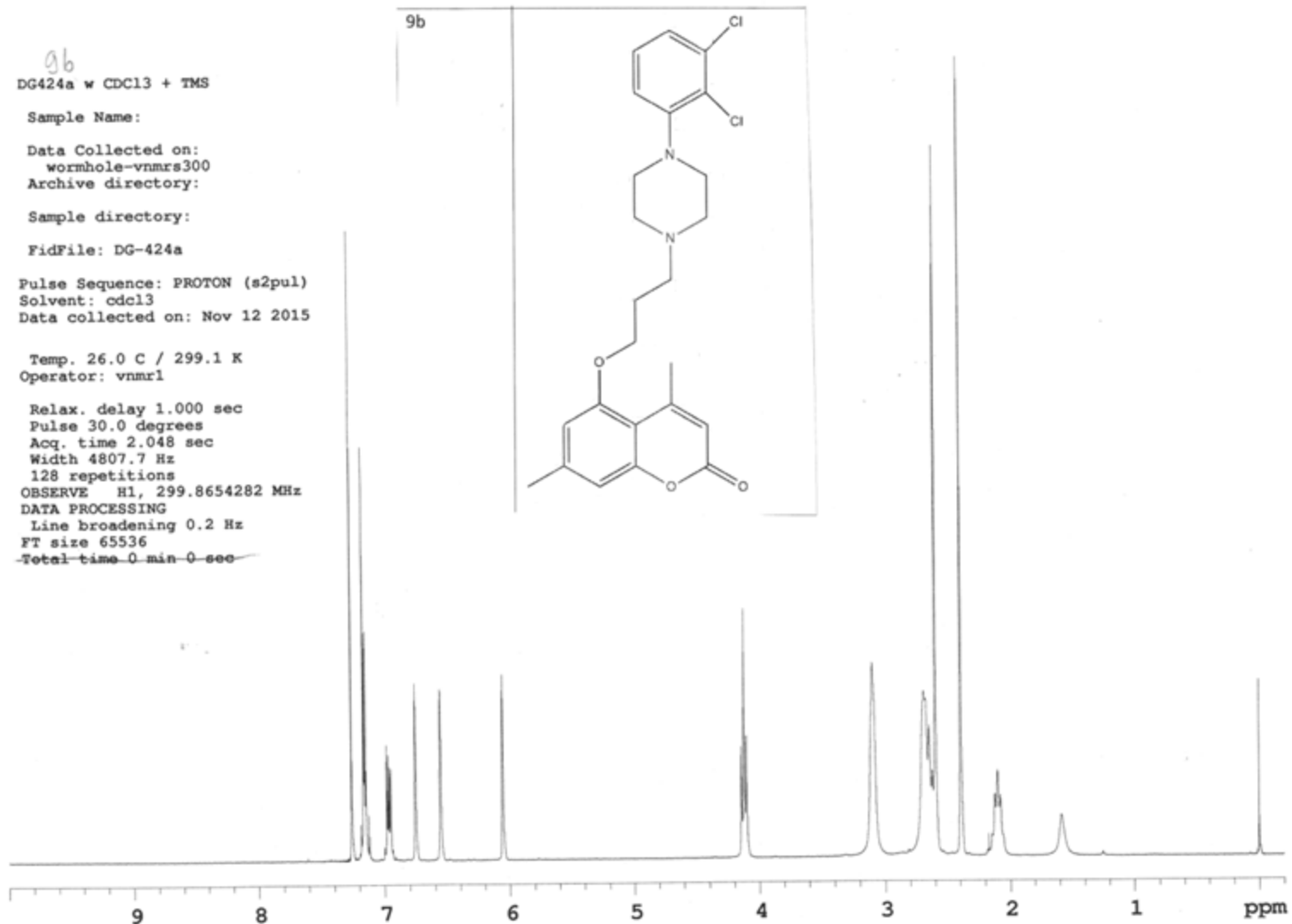
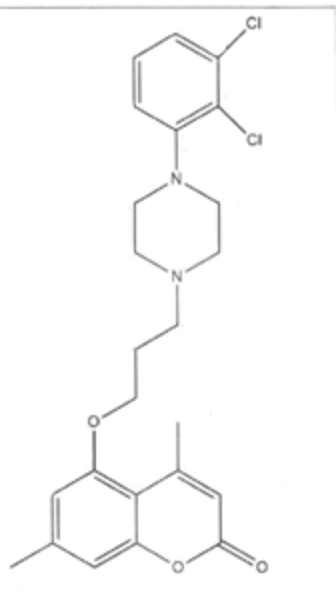
8b
 DG 420a w CDCl3 + TMS
 Sample Name:
 Data Collected on:
 wormhole-vnmrs300
 Archive directory:
 Sample directory:
 FidFile: DG-420a
 Pulse Sequence: PROTON (s2pul)
 Solvent: cdcl3
 Data collected on: Nov 2 2015
 Temp. 26.0 C / 299.1 K
 Operator: vnmr1
 Relax. delay 1.000 sec
 Pulse 30.0 degrees
 Acq. time 2.048 sec
 Width 4807.7 Hz
 128 repetitions
 OBSERVE H1, 299.8654279 MHz
 DATA PROCESSING
 Line broadening 0.2 Hz
 FT size 65536
 Total time 0 min 0 sec





9b
DG424a w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: DG-424a
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Nov 12 2015
Temp. 26.0 C / 299.1 K
Operator: vnmrl
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654282 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
~~Total time 0 min 0 sec~~

9b



DG 424a 13C

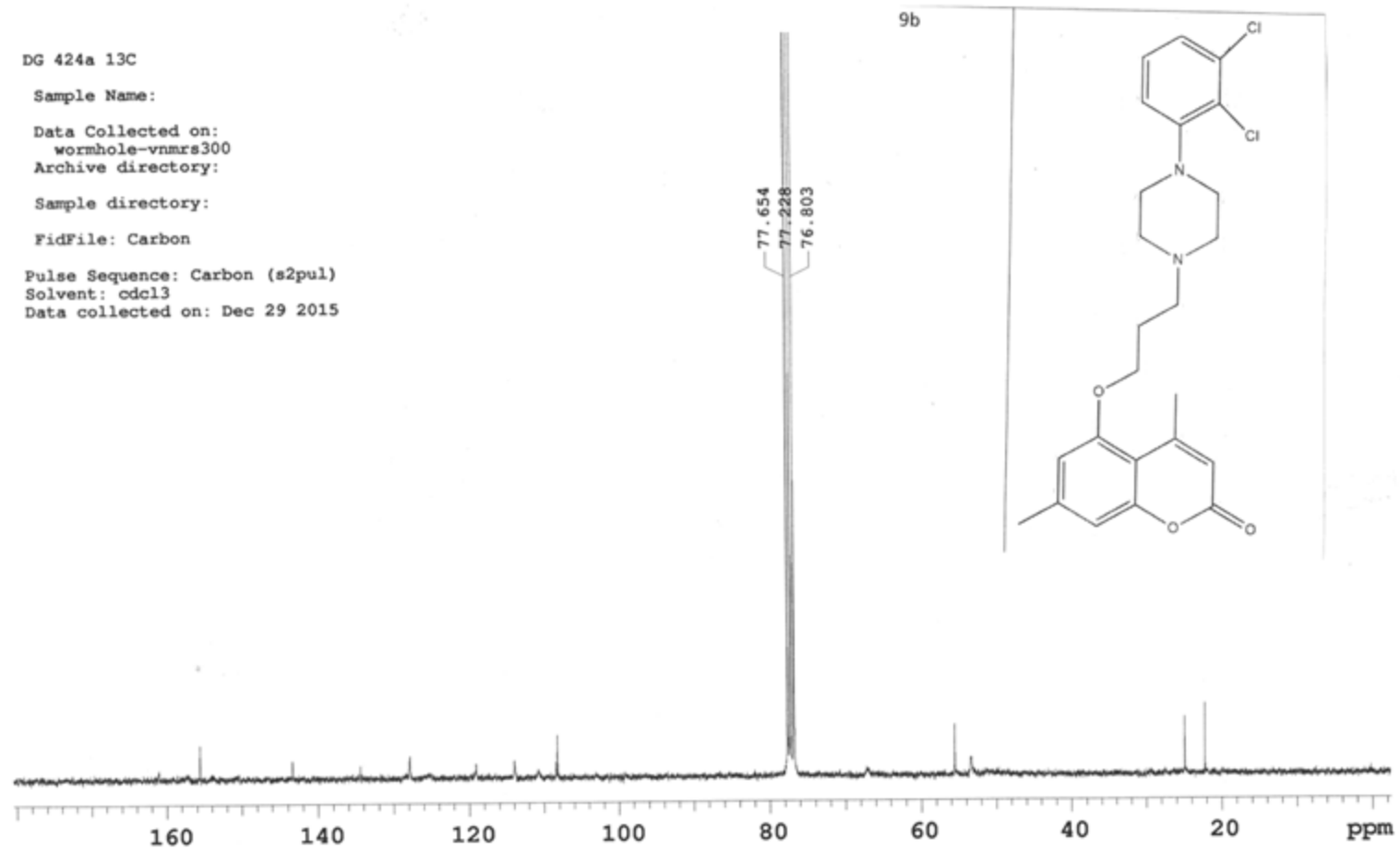
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Dec 29 2015



10b

DG-419 w CDC13 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)

Solvent: cdcl3

Data collected on: Oct 21 2015

Temp. 26.0 C / 299.1 K

Operator: vnmr1

Relax. delay 1.000 sec

Pulse 30.0 degrees

Acq. time 2.048 sec

Width 4807.7 Hz

128 repetitions

OBSERVE H1, 299.8654270 MHz

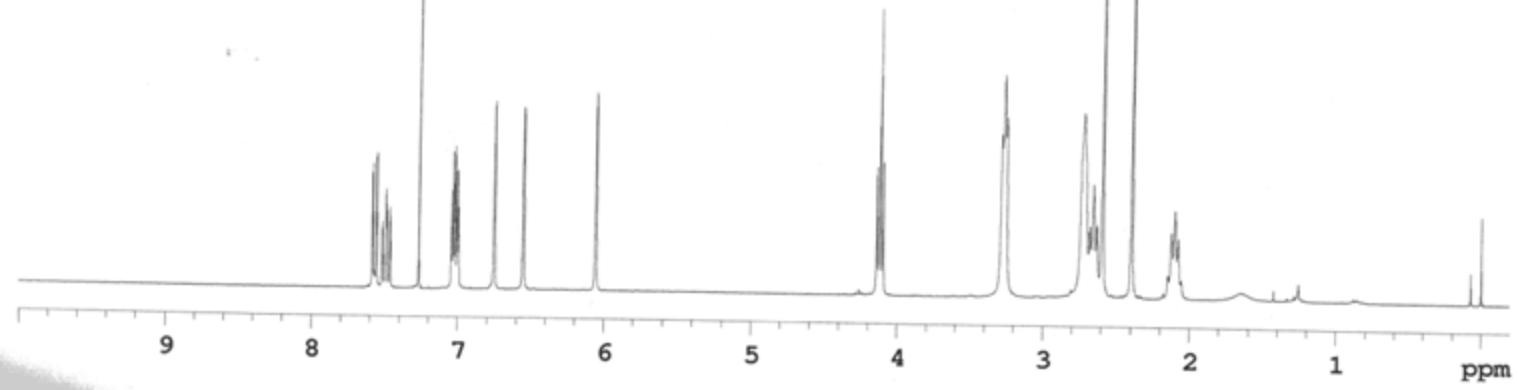
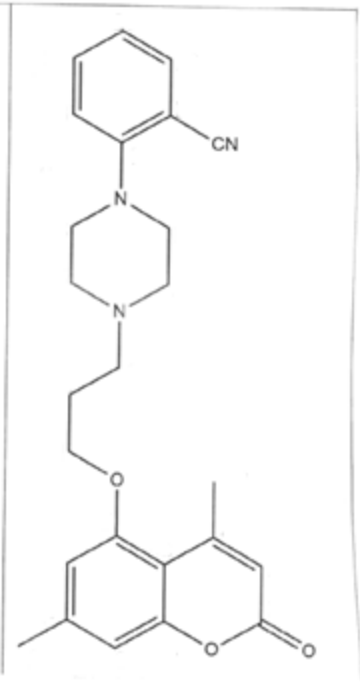
DATA PROCESSING

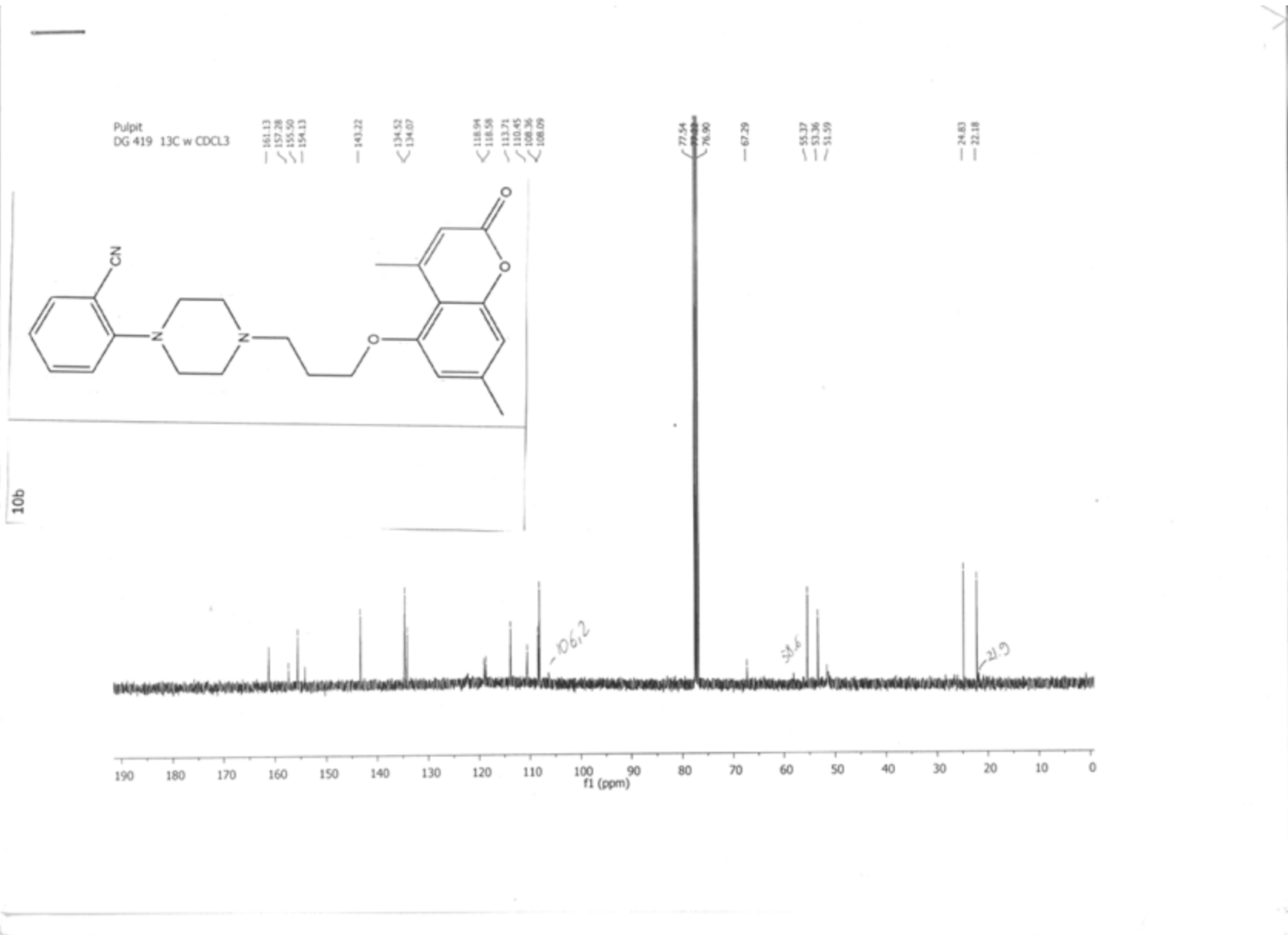
Line broadening 0.2 Hz

FT size 65536

Total time 0 min 0 sec

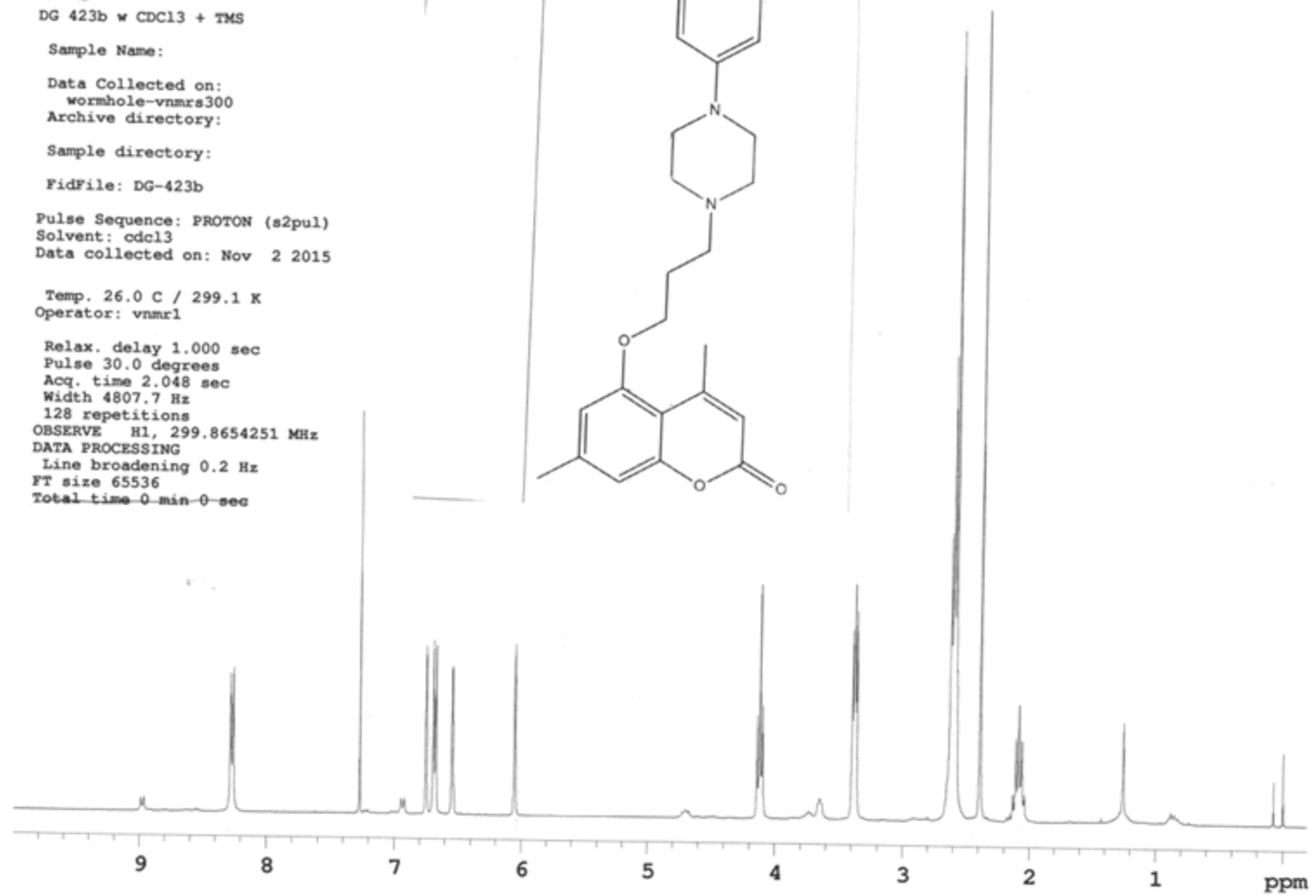
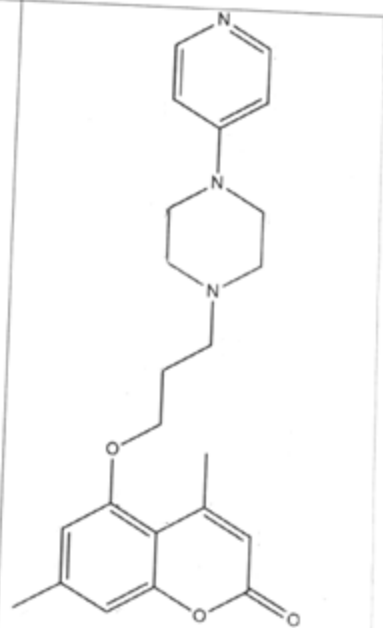
10b





11b
DG 423b w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: DG-423b
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Nov 2 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654251 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

11b



DG-423b-13C

DG-423b w CDCl3 13C-NMR

161.06
157.30
155.48
155.40
154.06
148.59
140.22

113.68
110.94
108.35
108.07

67.23

55.26

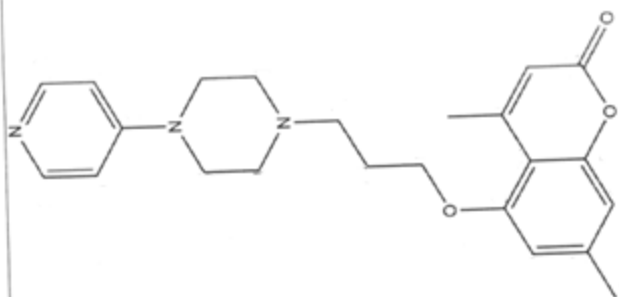
52.85

46.16

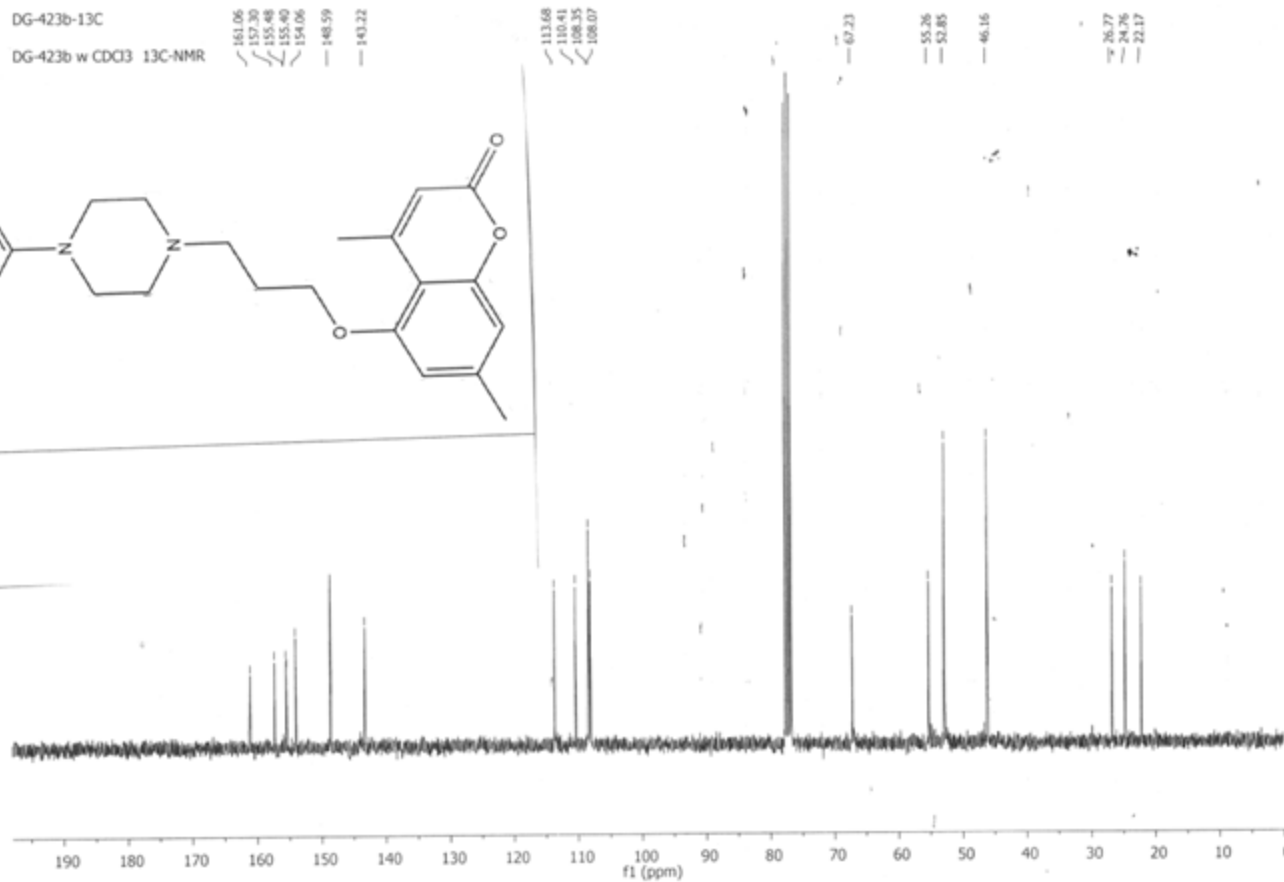
26.77

24.76

22.17



11b



2d

DG401a w CDCl3 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)

Solvent: cdcl3

Data collected on: Apr 13 2015

Temp. 26.0 C / 299.1 K

Operator: vnmr1

Relax. delay 1.000 sec

Pulse 30.0 degrees

Acq. time 2.048 sec

Width 4807.7 Hz

128 repetitions

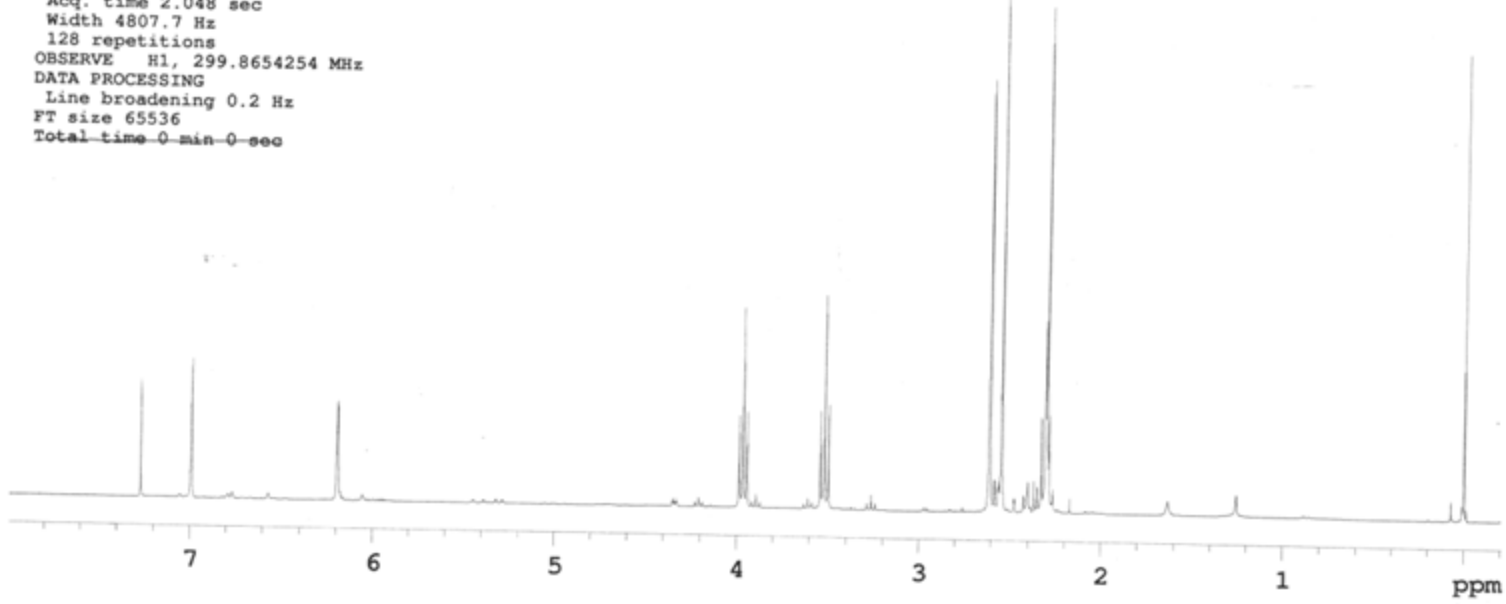
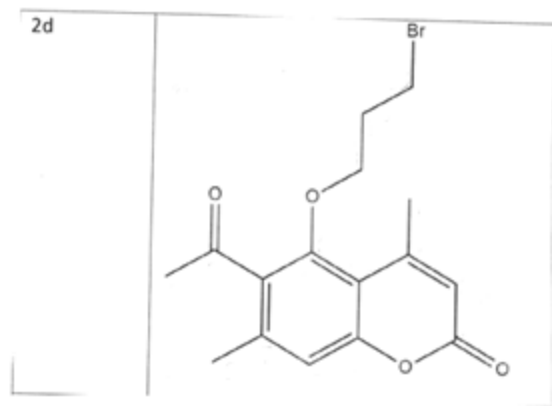
OBSERVE H1, 299.8654254 MHz

DATA PROCESSING

Line broadening 0.2 Hz

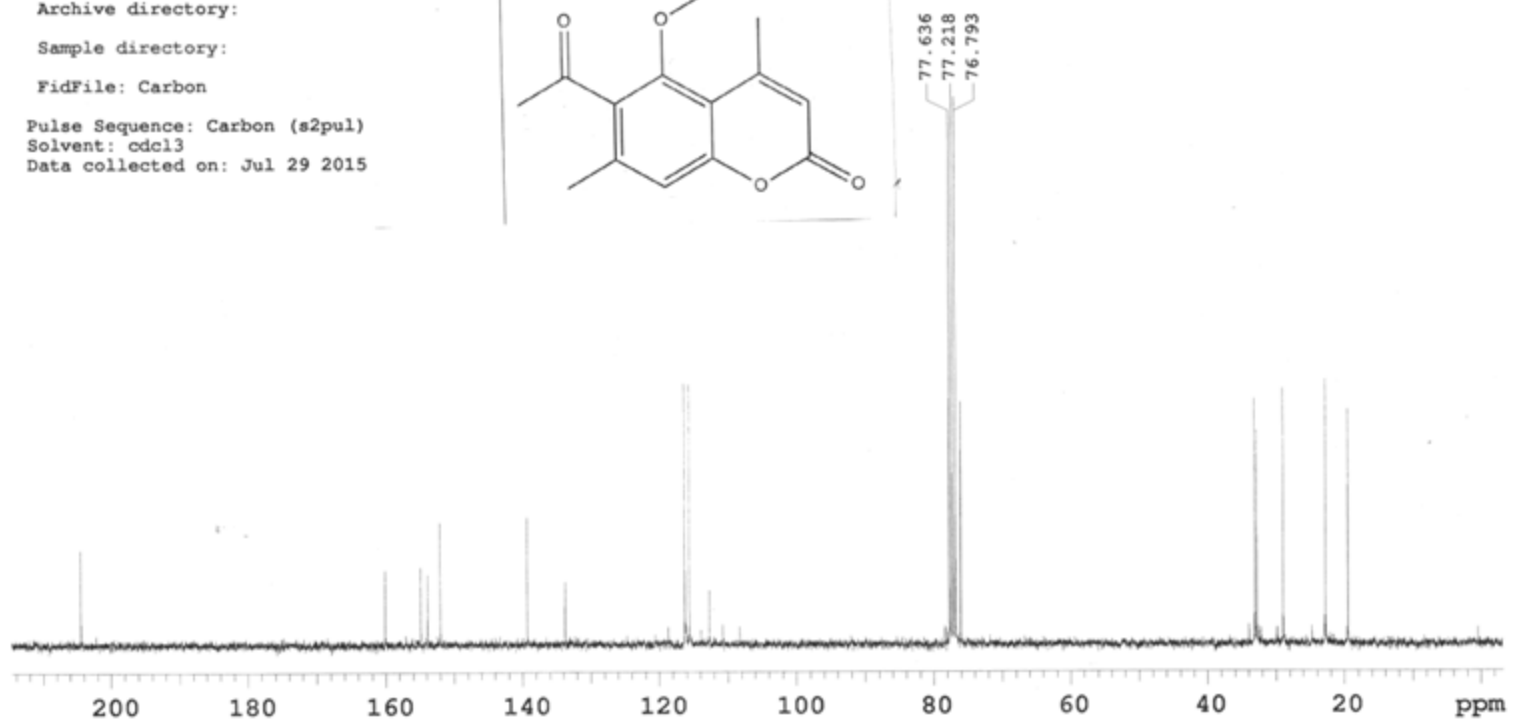
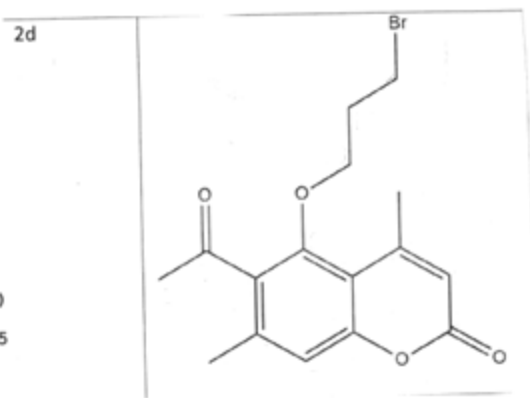
FT size 65536

Total time 0 min 0 sec



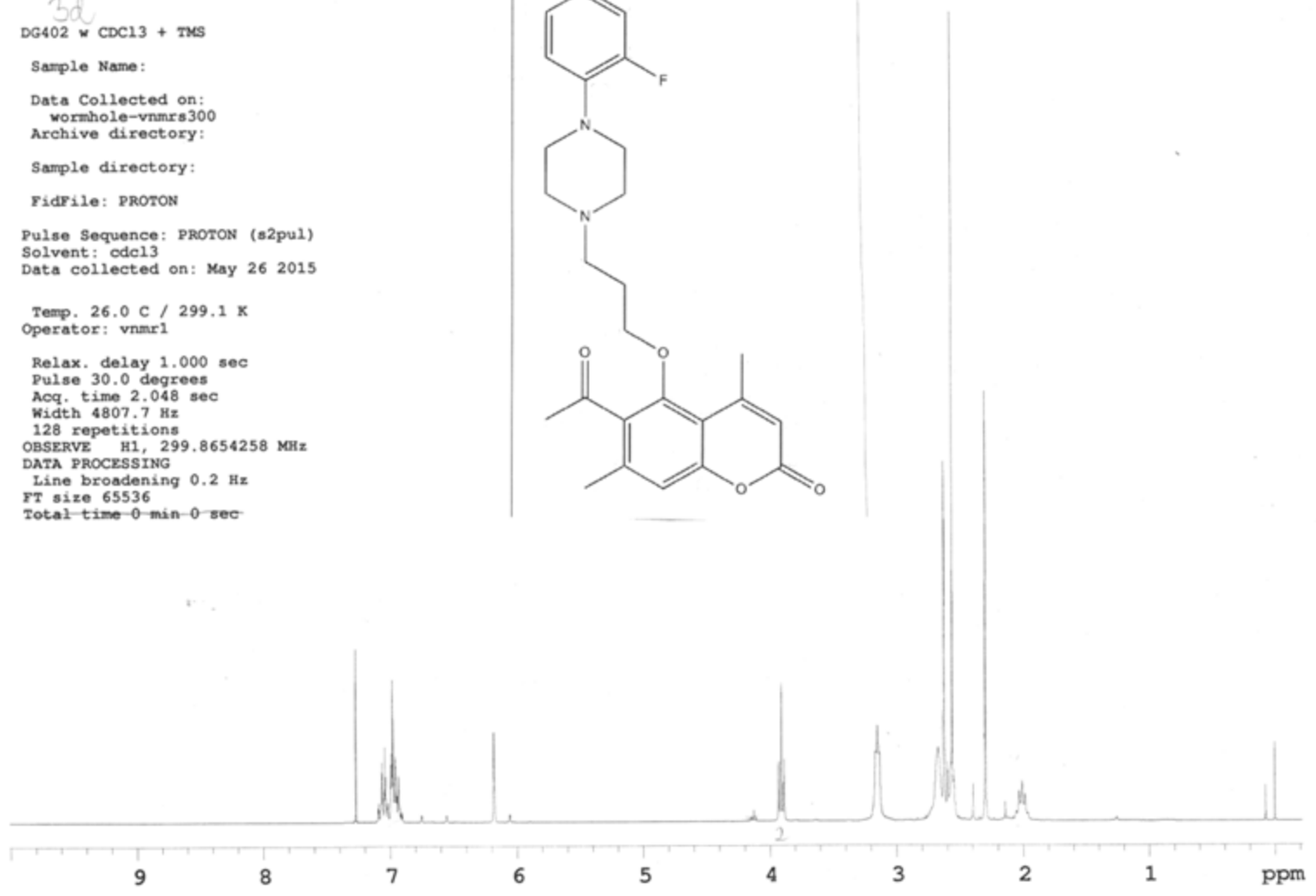
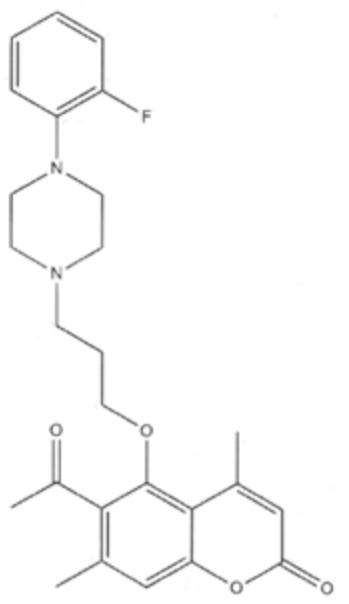
DG401a
~~XXXX~~ 13C

Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: Carbon
Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Jul 29 2015



3d
DG402 w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: May 26 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654258 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

3d



DG 402 13C w CDCl3

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

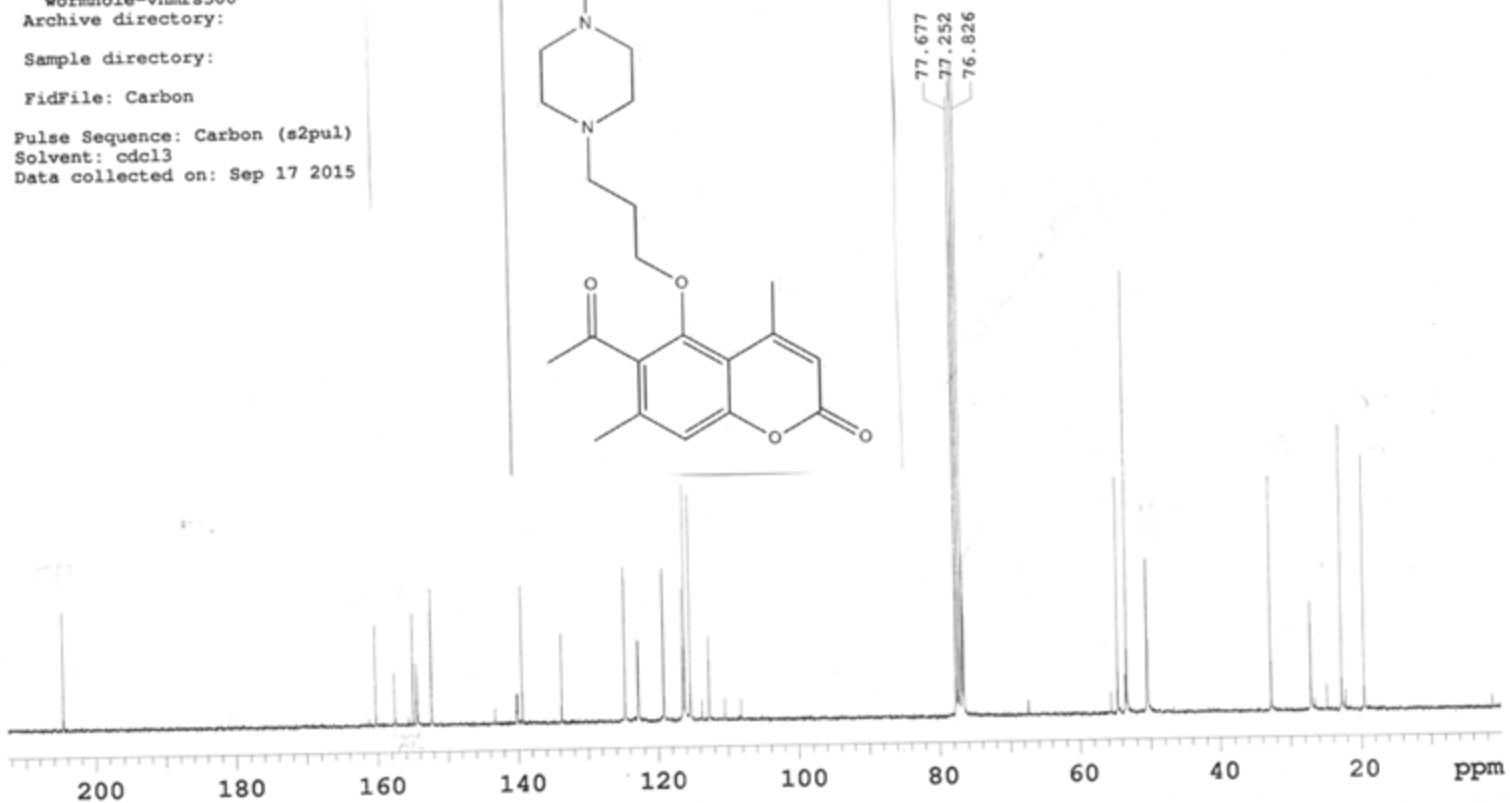
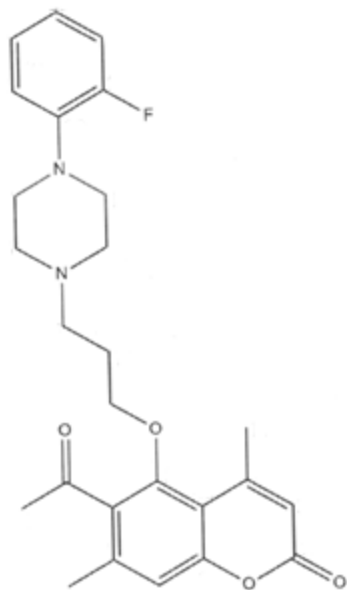
FidFile: Carbon

Pulse Sequence: Carbon (s2pul)

Solvent: cdcl3

Data collected on: Sep 17 2015

3d



4d

DG403 w CDC13 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)

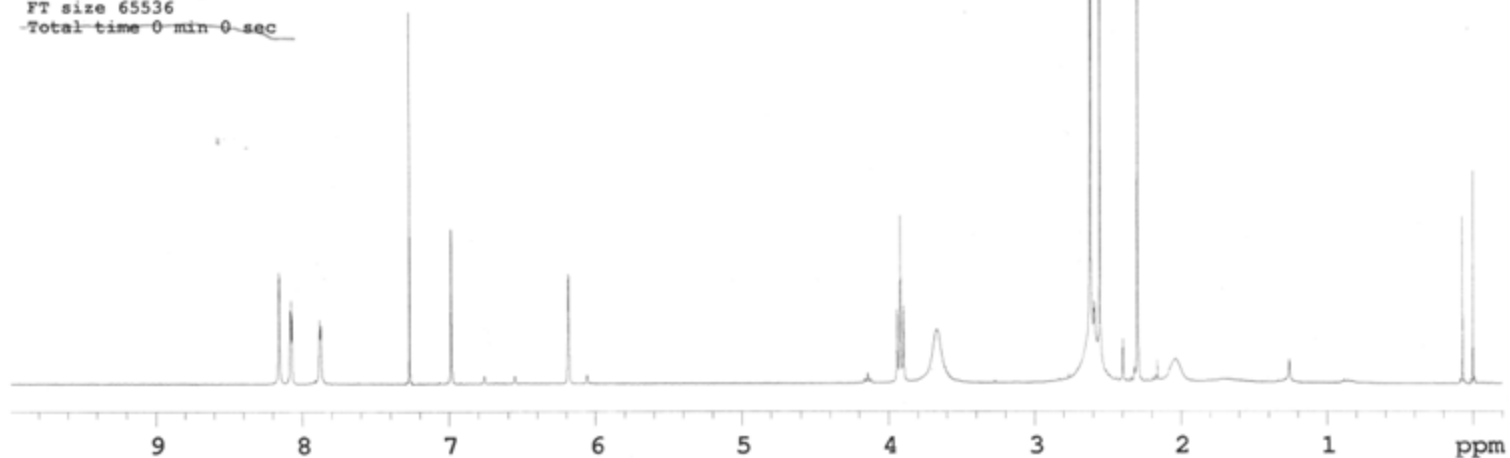
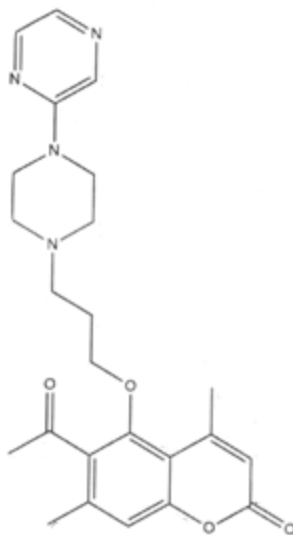
Solvent: cdcl3

Data collected on: May 26 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654261 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

4d



DG 403 13C

4d

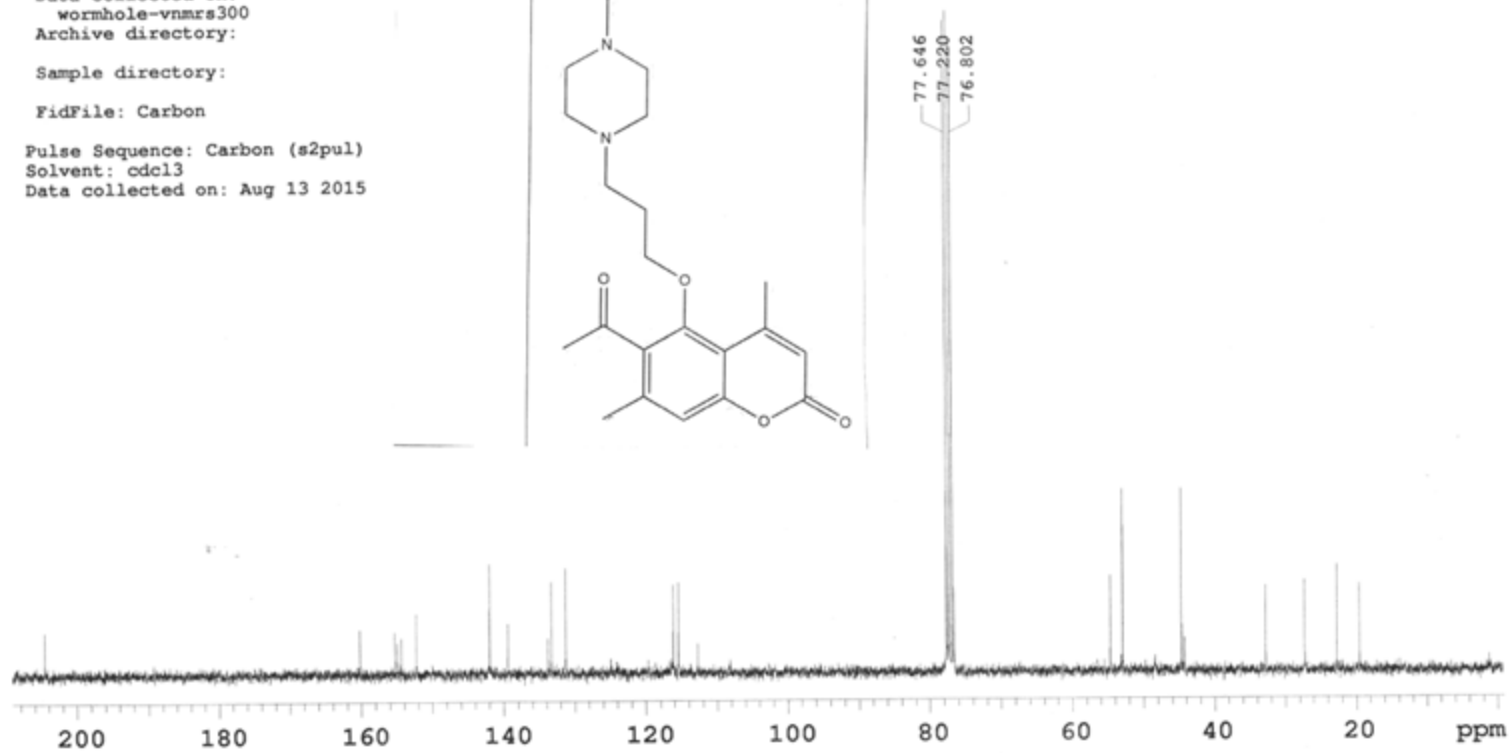
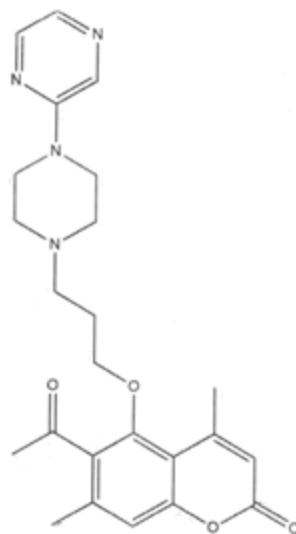
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Aug 13 2015



5d

DG404 w CDCl3 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

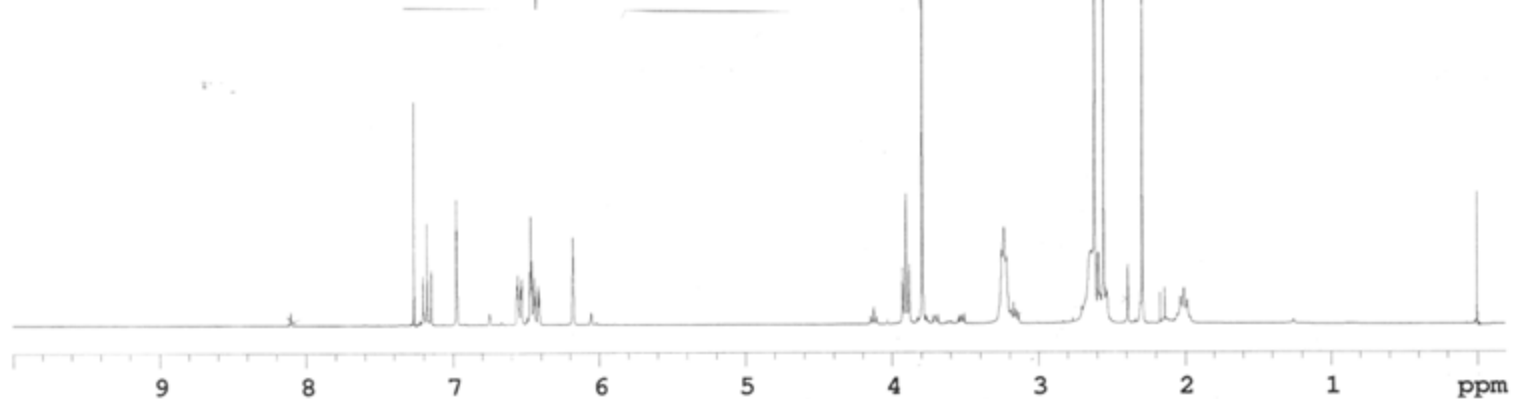
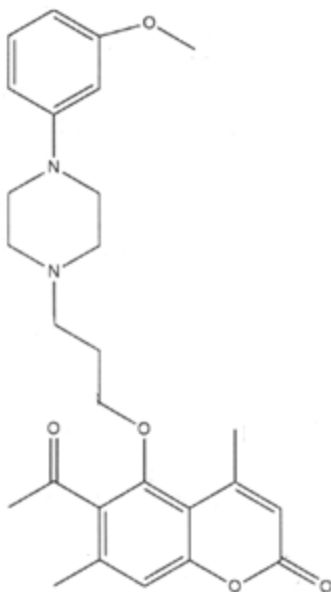
FidFile: PROTON

Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: May 26 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654270 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

5d



DG 404 13C

5d

Sample Name:

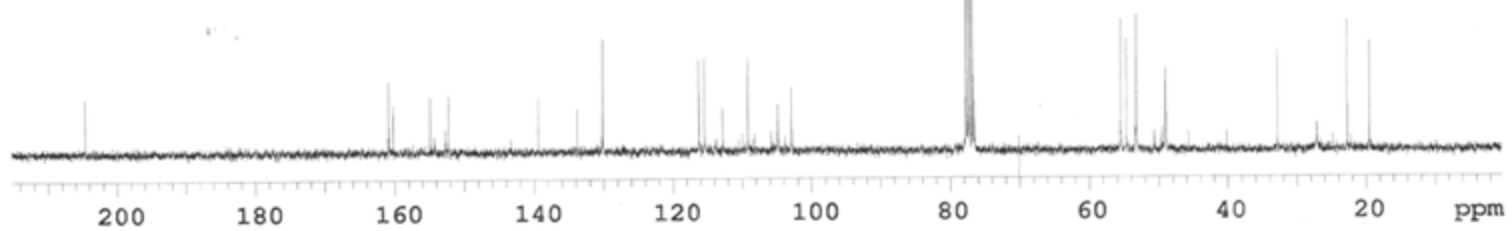
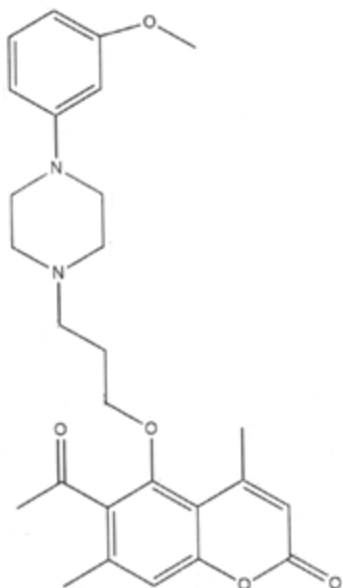
Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Aug 11 2015



6d

DG 405 1H w CDC13

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

Sample directory:

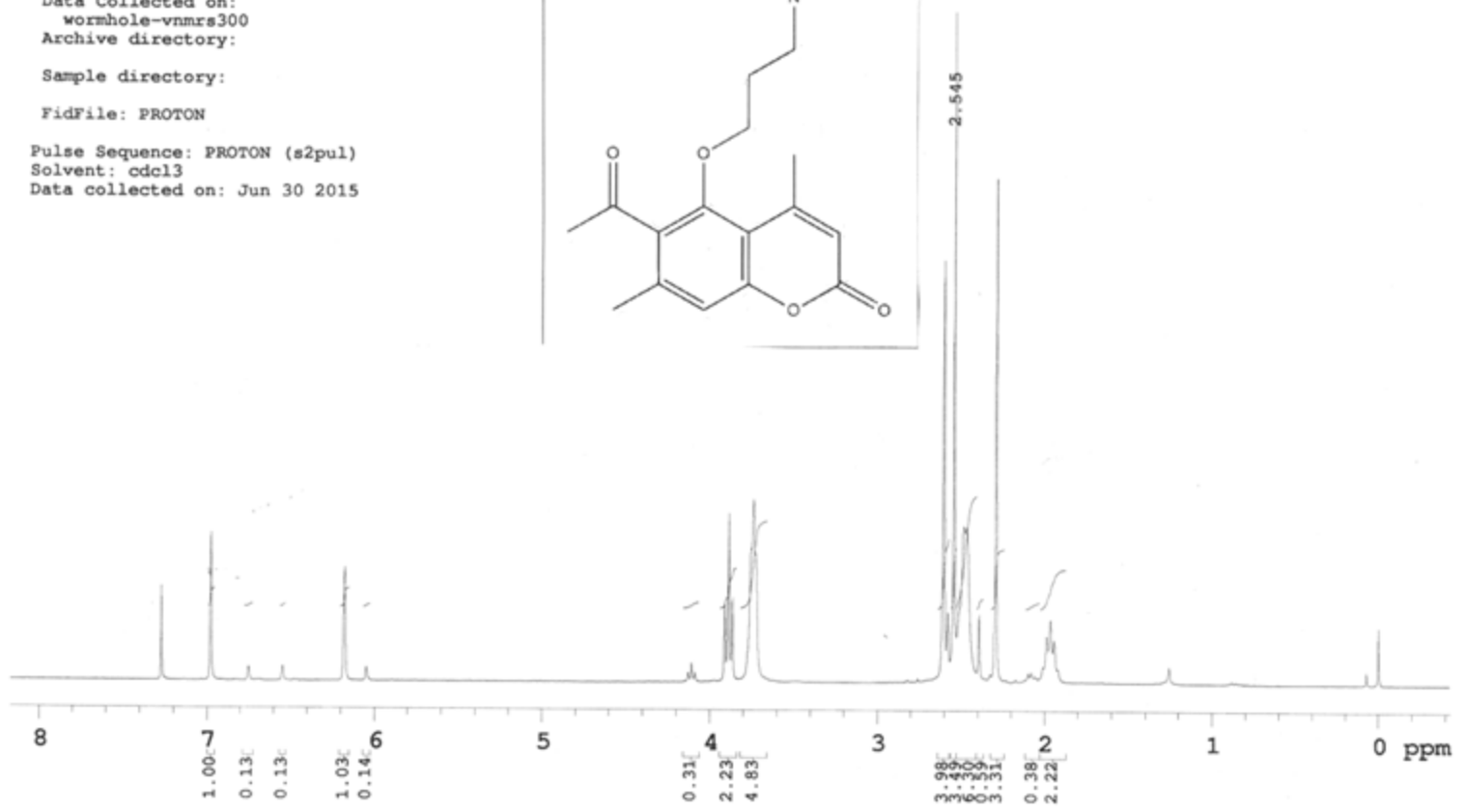
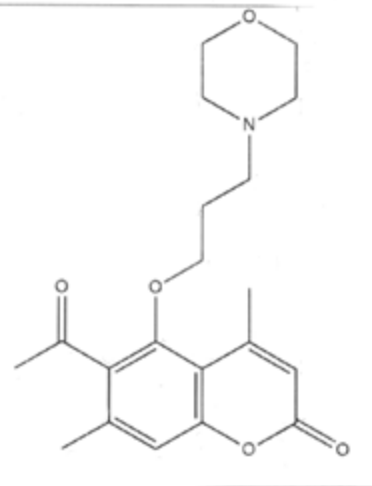
FidFile: PROTON

Pulse Sequence: PROTON (s2pul)

Solvent: cdcl3

Data collected on: Jun 30 2015

6d



DG 405 13C

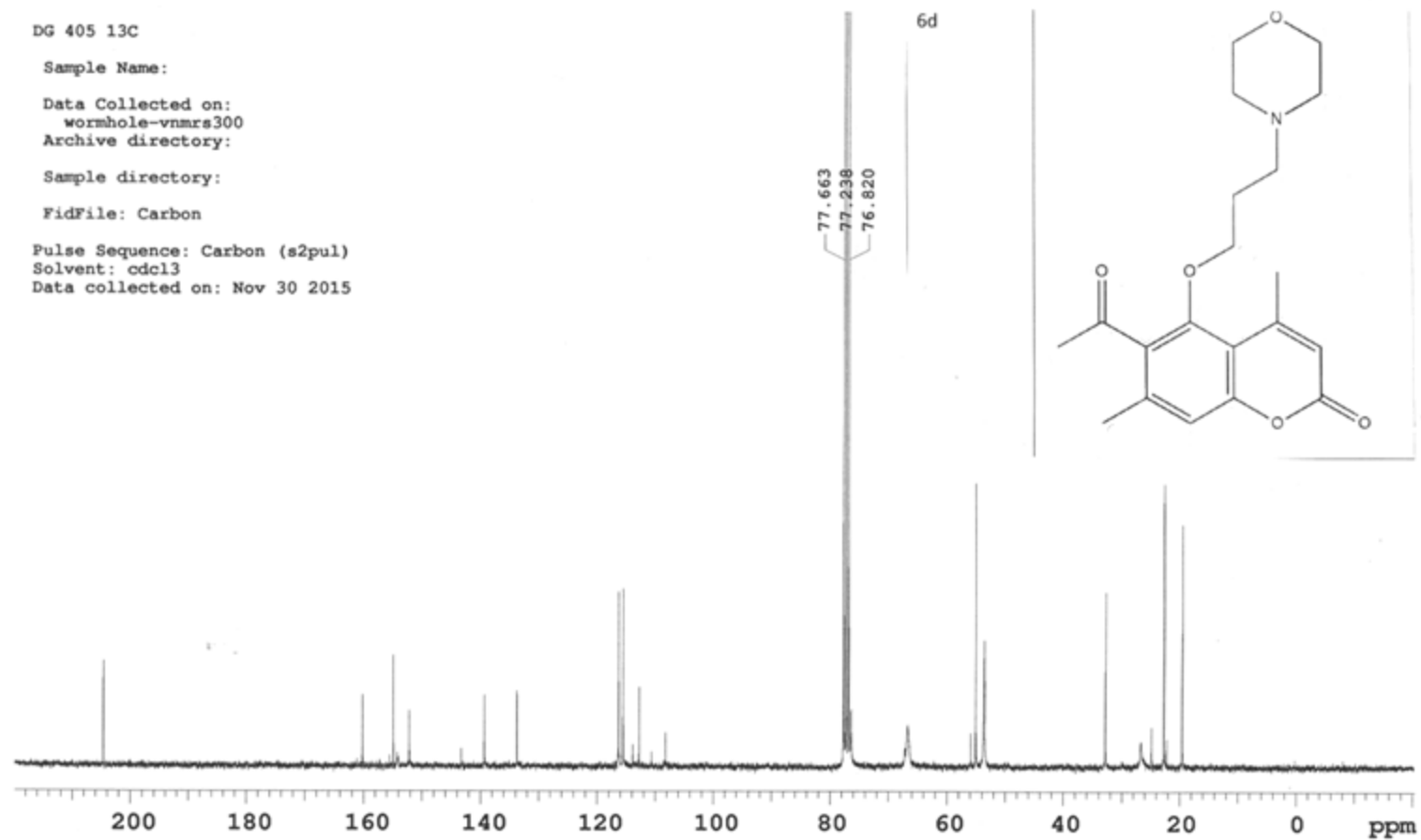
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

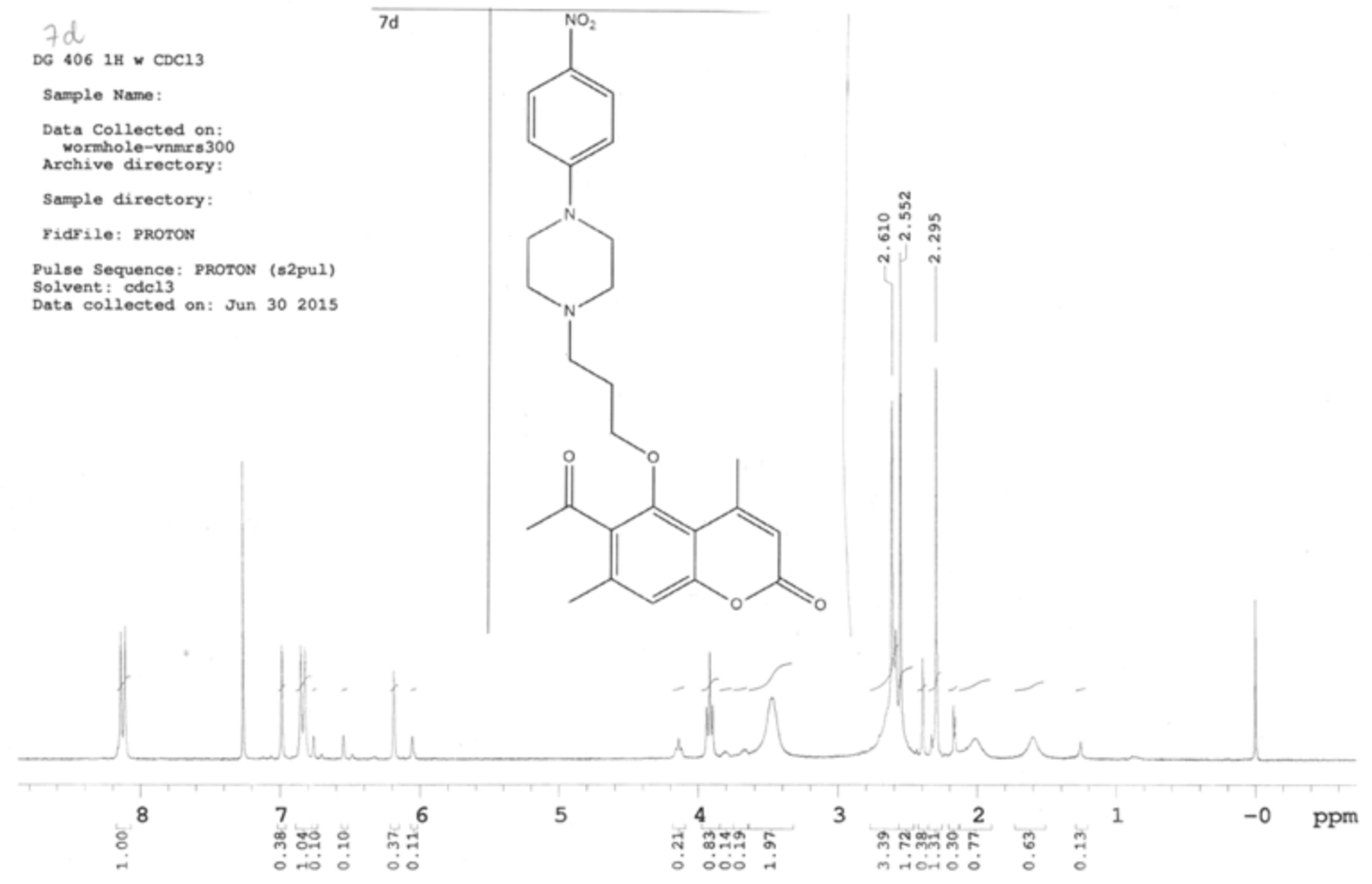
Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Nov 30 2015



7d
DG 406 1H w CDCl3
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Jun 30 2015



DG 406 13C

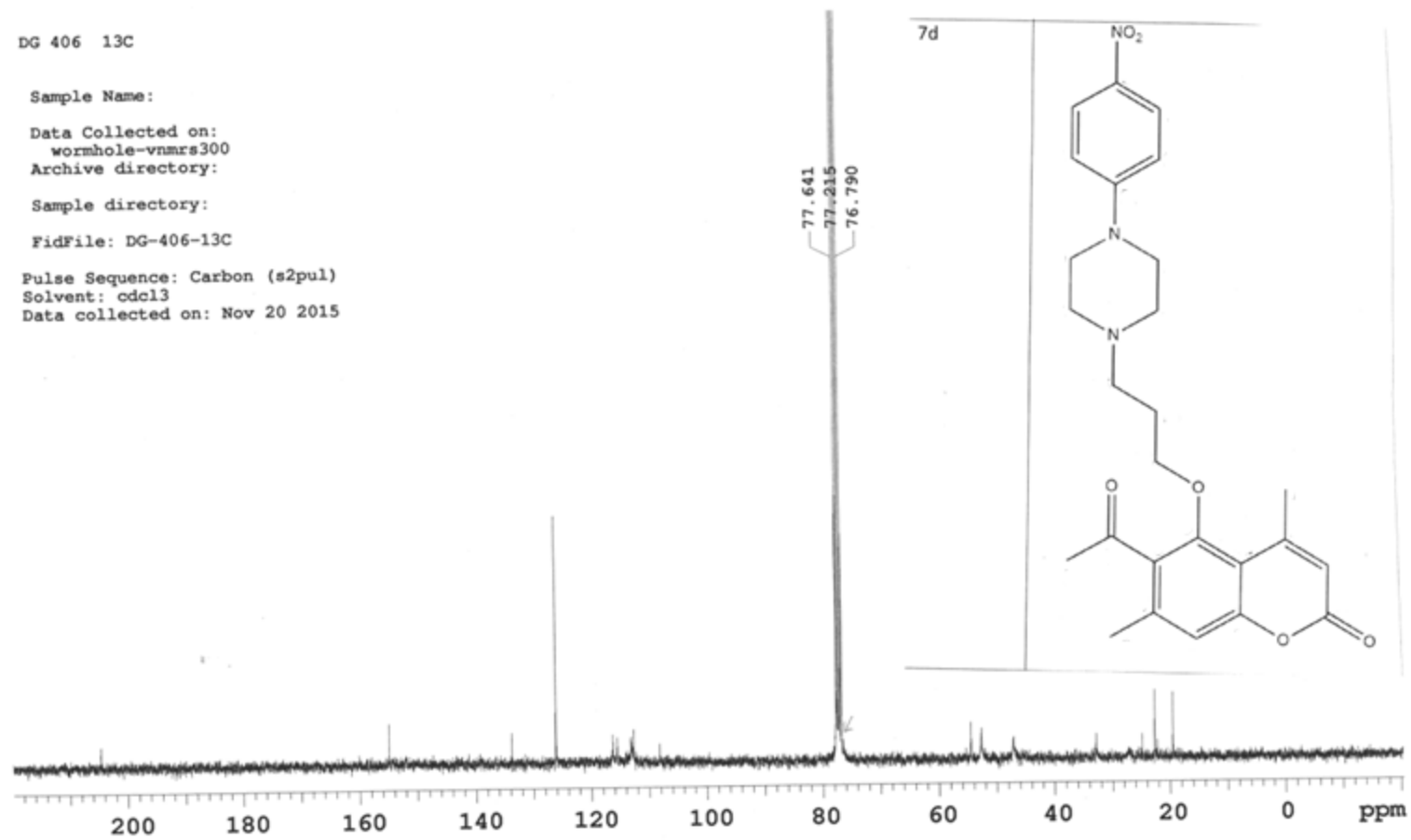
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

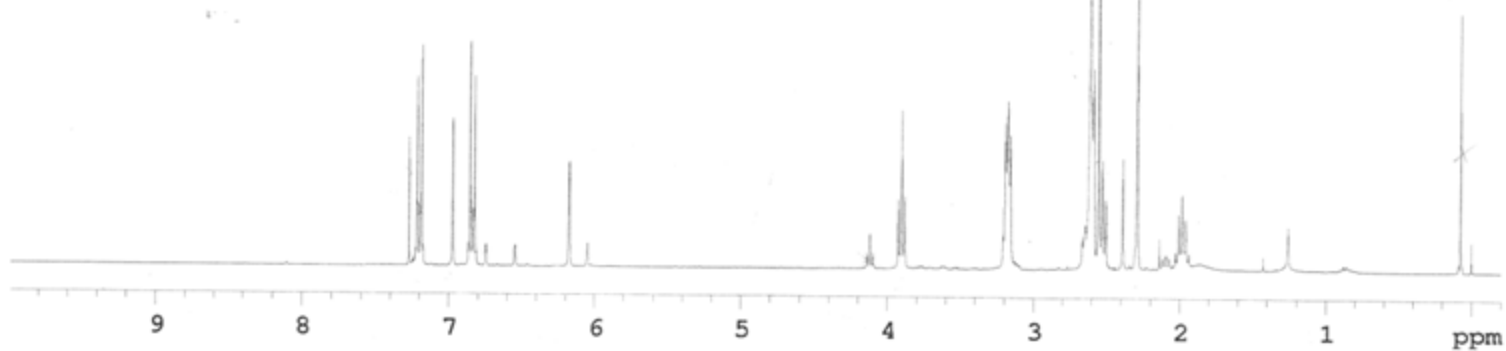
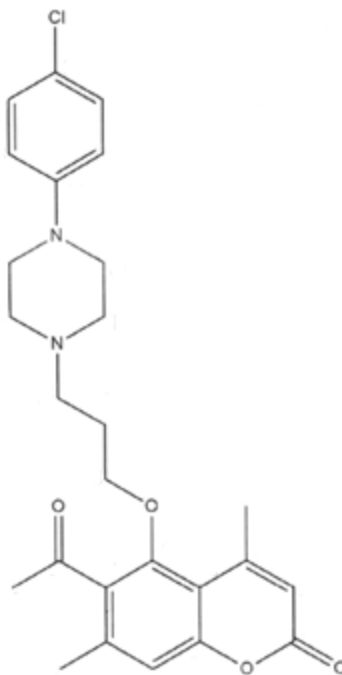
FidFile: DG-406-13C

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Nov 20 2015



8d
DG408 w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Sep 16 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654257 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec.

8d



DG 408 13C

Sample Name:

Data Collected on:
wormhole-vnmrs300

Archive directory:

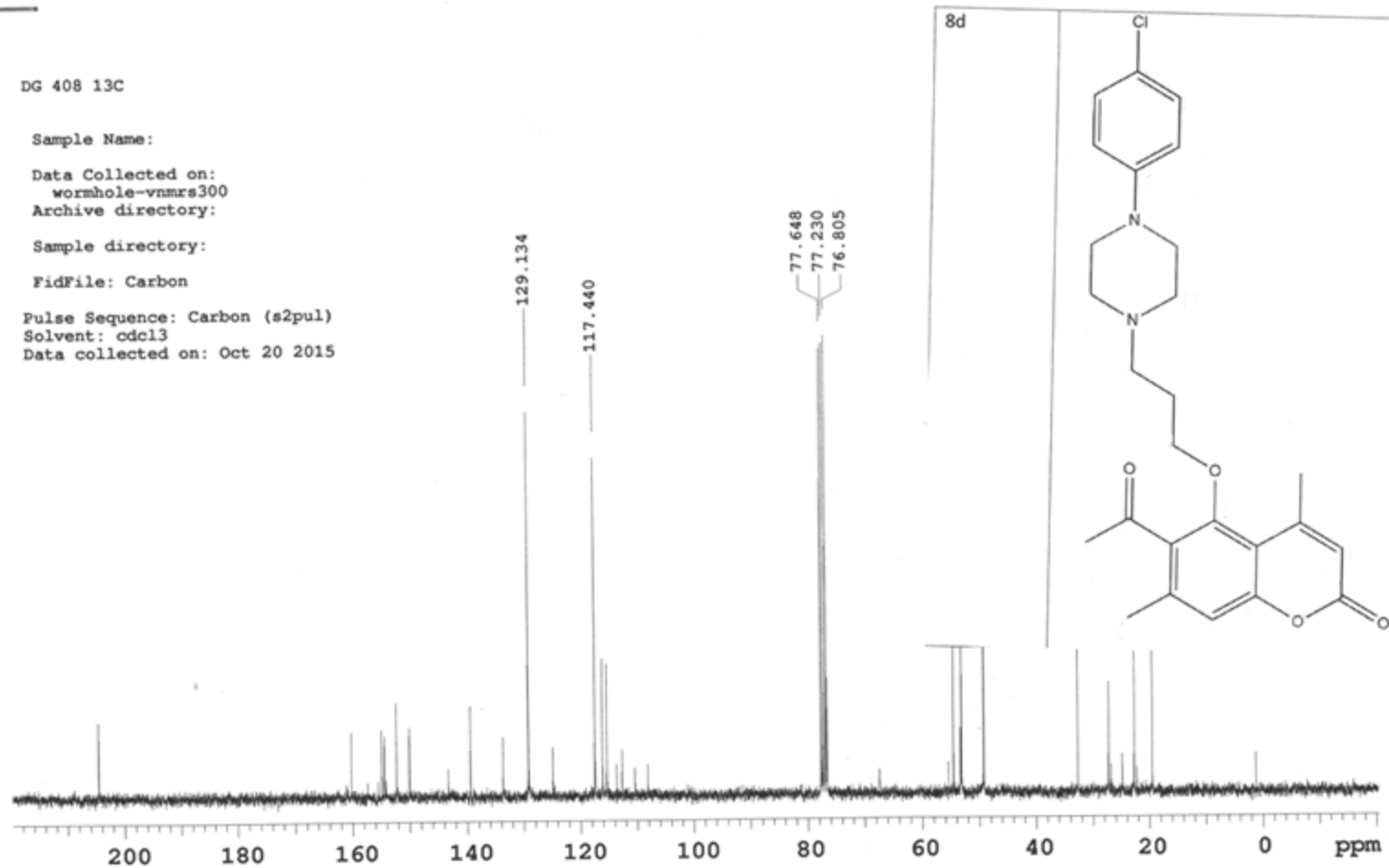
Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)

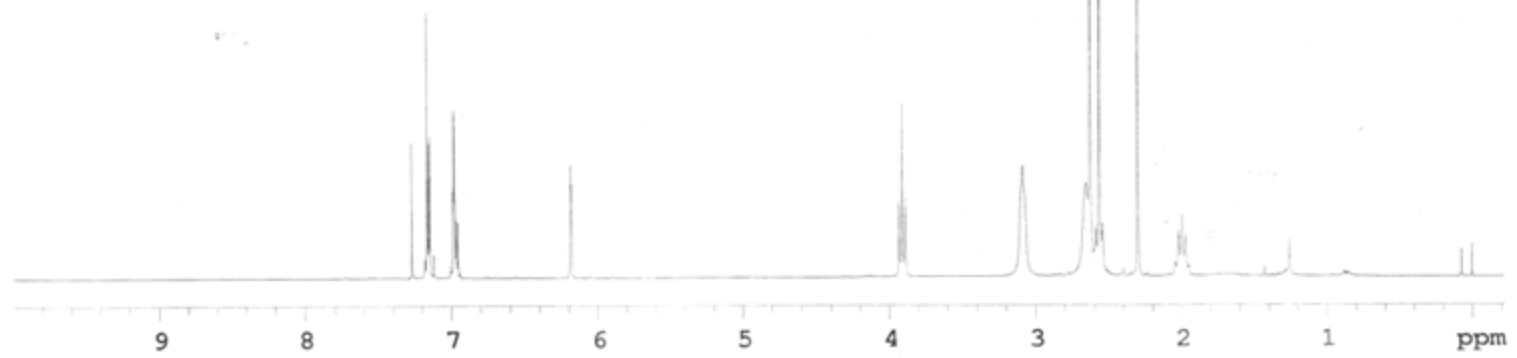
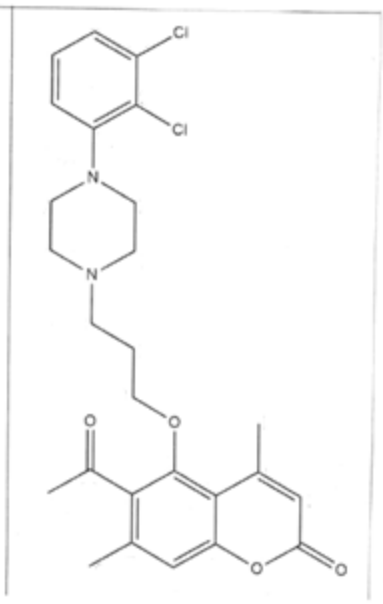
Solvent: cdcl3

Data collected on: Oct 20 2015



gd
DG412 w CDCl3 + TMS
Sample Name:
Data Collected on:
wormhole-vnmrs300
Archive directory:
Sample directory:
FidFile: PROTON
Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Sep 21 2015
Temp. 26.0 C / 299.1 K
Operator: vnmr1
Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654260 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
Total time 0 min 0 sec

9d



9d

DG 412 13C

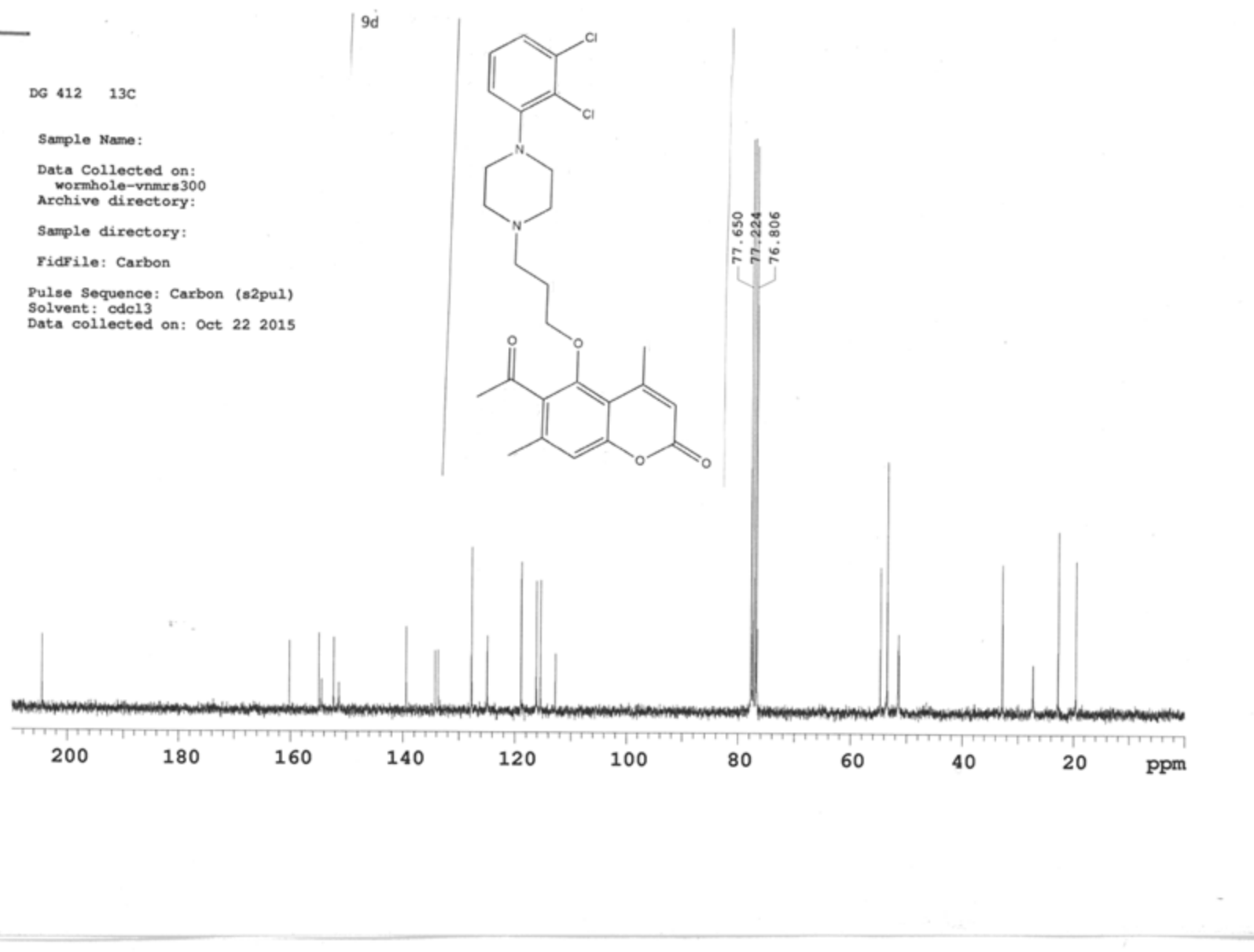
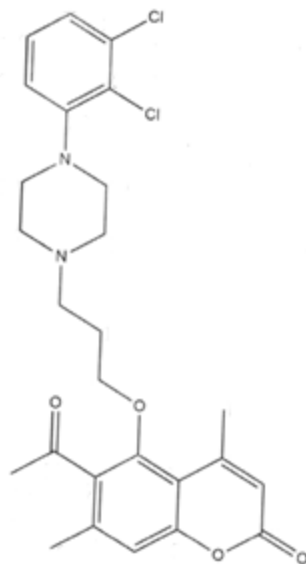
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Oct 22 2015



10d

DG 407 1H w CDCl3

Sample Name:

Data Collected on:
wormhole-vnmrs300

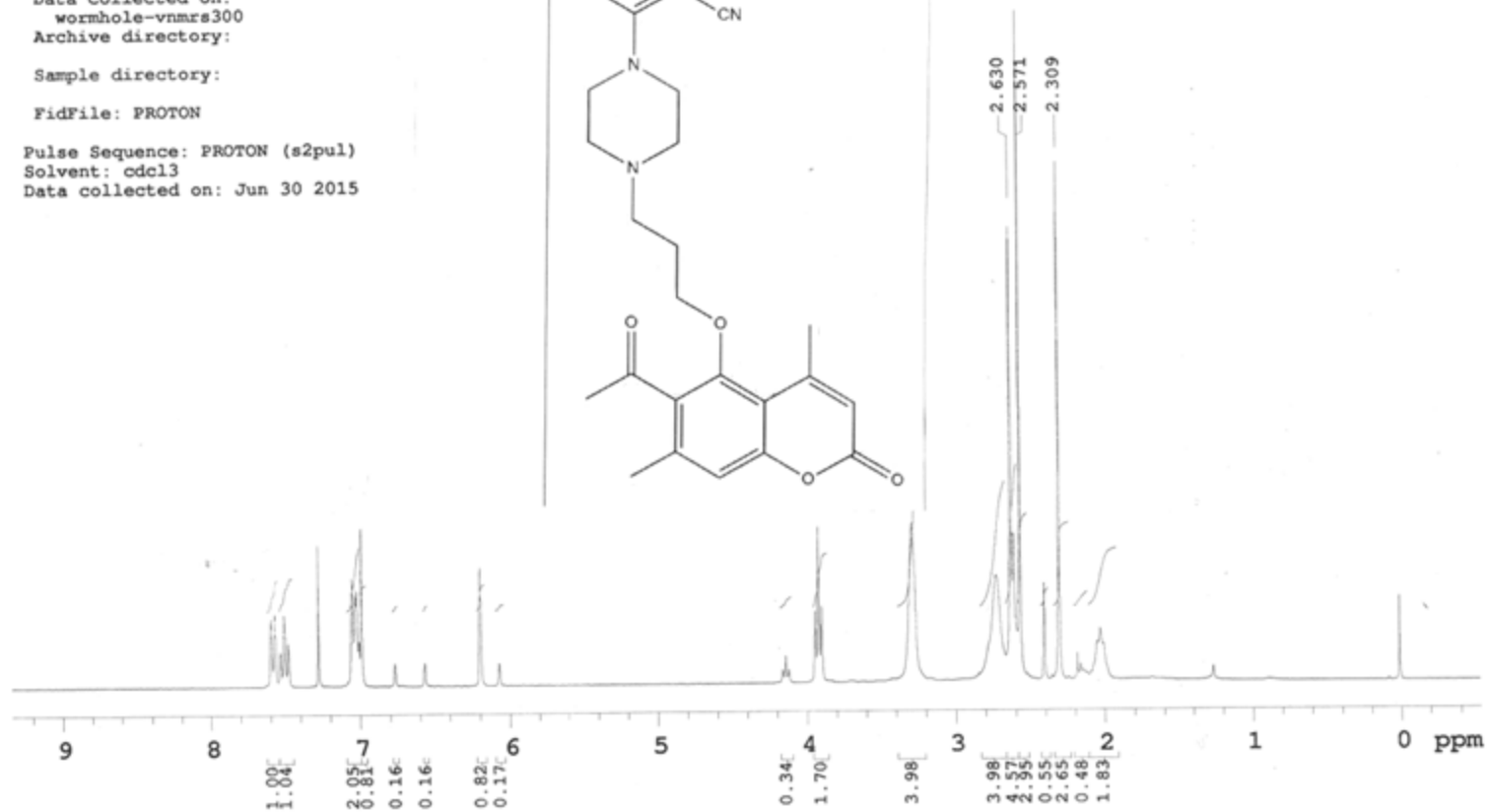
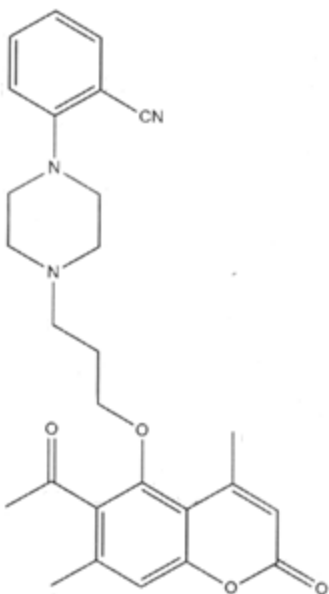
Archive directory:

Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Jun 30 2015

10d



DG 407 13C

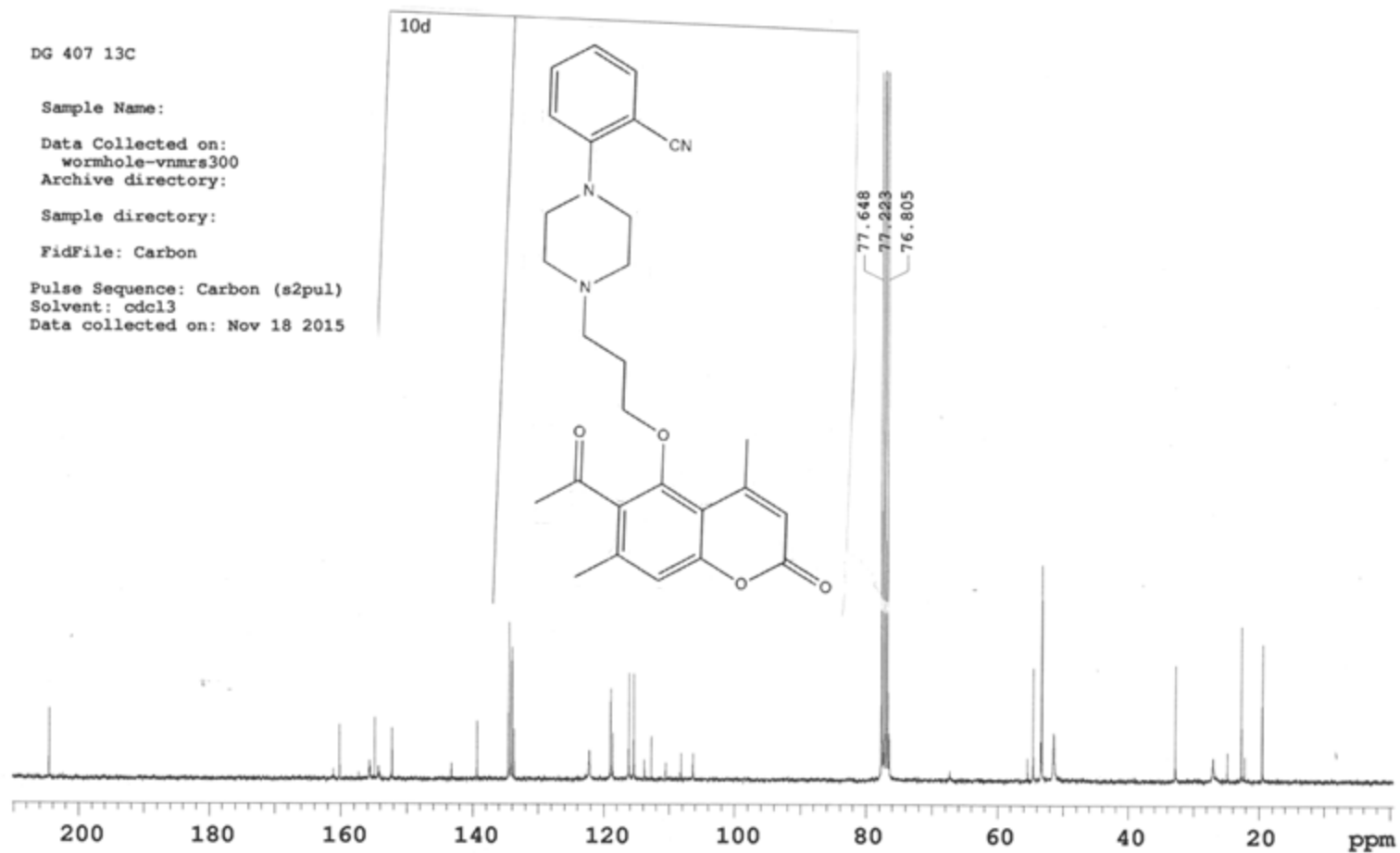
Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: Carbon

Pulse Sequence: Carbon (s2pul)
Solvent: cdcl3
Data collected on: Nov 18 2015



11d

DG411 w CDC13 + TMS

Sample Name:

Data Collected on:
wormhole-vnmrs300
Archive directory:

Sample directory:

FidFile: PROTON

Pulse Sequence: PROTON (s2pul)
Solvent: cdcl3
Data collected on: Sep 21 2015

Temp. 26.0 C / 299.1 K
Operator: vnmr1

Relax. delay 1.000 sec
Pulse 30.0 degrees
Acq. time 2.048 sec
Width 4807.7 Hz
128 repetitions
OBSERVE H1, 299.8654218 MHz
DATA PROCESSING
Line broadening 0.2 Hz
FT size 65536
~~Total time 0 min 0 sec~~

