G1 phase					S phase				
control	merge	centrin1	ADD1 ^{pS726}	γ-tubulin	control	merge	centrin1	ADD1 ^{p8726}	γ-tubulin
ADD1 ^{pS726} peptide					ADD1 ^{pS726} peptide				
Centrin1 ADD 195726 DAP1	merge	centrin1	ADD1 ^{pS726}	γ-tubulin	Centrin1 ADD15528 DAPI	merge	centrin1	ADD1 ^{pS726}	γ-tubulin
G2 phase	merge	centrin1	ADD1pS726	γ-tubulin	Mitosis	merge	centrin1	ADD1 ^{pS726}	γ-tubulin
control	-		10		control	8			
	5	•		٠		*			٠
ADD1 ^{pS726} peptide		۰.			ADD1 ^{p\$726} peptide	2			
		:			Centrin1				

Expanded View Figures

Figure EV1. Verification of the specificity of the anti-ADD1 pS726 antibody in immunofluorescence staining.

RPE1 cells at the indicated cell cycle stages were stained for centrin1 (green), ADD1 pS726 (red), and γ -tubulin (blue). For phosphopeptide competition, the anti-ADD1 pS726 antibody was pre-mixed with a phosphopeptide (ADD1 pS726 peptide) before staining. DNA was visualized by DAPI staining. High magnification images of each centrosome are shown on the right. Scale bars, 10 μ m (main image) and 1 μ m (zoomed images).



Figure EV2.

Figure EV2. FLAG-ADD1 S726A restores the defects of spindle distortion and elongation, but not multipolar spindle formation, induced by ADD1 depletion.

- A HeLa cells were infected with lentiviruses expressing shRNAs to ADD1 (sh-ADD1) or luciferase (sh-Luc.) as a control. FLAG-ADD1 WT or the S726A mutant was reexpressed in the cells whose endogenous ADD1 had been depleted. Equal amounts of whole-cell lysates were analyzed by immunoblotting (IB) with the indicated antibodies.
- B The percentage of distorted spindles in the total number of mitotic cells was measured (367–408 mitotic cells were counted in each group).
- C The ratio of spindle length to cell diameter was measured (80-112 mitotic cells were counted in each group).
- D The percentage of multipolar spindles in the total number of mitotic cells was measured (405–509 mitotic cells were counted in each group).

Data information: In (B–D), values (mean \pm s.d.) are from three independent experiments. Statistical significance of differences is assessed with a Student's *t*-test: for (B), ****P* = 0.0009; for (C), ***P* = 0.0092; for (D), ***P* = 0.0018, ****P* = 0.0002.

Source data are available online for this figure.



Figure EV3. Increased number of centrosome foci during mitosis in ADD1-depleted cells is not caused by centrosome overduplication or centriole disengagement at interphase.

- A HeLa cells were infected with lentiviruses expressing shRNAs to ADD1 (sh-ADD1 #1 and sh-ADD1 #2) or luciferase (sh-Luc.) as a control. The expression levels of ADD1 and β -actin (loading control) were analyzed by immunoblotting with the indicated antibodies.
- B The percentage of interphase cells with 1, 2, or > 2 γ -tubulin foci was assessed (1,085–1,311 cells were counted in each group).
- C Schematic representation of the engaged and disengaged centriole pairs. Engaged centriole pairs have one C-Nap1 focus and two centrin2 foci, whereas disengaged centriole pairs have both foci of C-Nap1 and centrin2.
- D HeLa cells were infected with lentiviruses expressing shRNAs to ADD1 (sh-ADD1 #1 and sh-ADD1 #2) or luciferase (sh-Luc.) as a control. In the control experiment, the cells were treated with a CDK1 inhibitor, RO-3306, for 16 h to induce arrest at the G2 phase. The cells were stained for centrin2 (red), c-Nap1 (green), and DNA (blue). Quantification of the interphase cells with disengaged centrioles (700–1,690 cells were counted in each group). Scale bars, 5 μm (main image) and 1 μm (zoomed images).

Data information: In (B and D), values (mean \pm s.d.) are from three independent experiments. **P = 0.001 (Student's t-test). Source data are available online for this figure.



2:1 ratio = engaged



= disengaged

disengaged





Figure EV4. Mitotic arrest contributes to, but does not fully explain ADD1 depletion-induced spindle multipolarity.

A–F HeLa cells stably expressing mCherry-tubulin were infected with lentiviruses expressing shRNAs to (A) luciferase (sh-Luc.) as a control, (B, C) ADD1 (sh-ADD1), (D, E) TPX2 (sh-TPX2), or (F) Myo10 (sh-Myo10). The cells were monitored with time-lapse microscopy. Images were captured every 3 min for 18 h. Arrowheads indicate the multiple spindle poles occurs within 1 h upon nuclear envelope breakdown. Arrows indicate the multiple spindle poles occurs more than 1 h after entering mitosis. Scale bars, 10 µm. The percentages of these two categories in the total counted samples (53 for sh-ADD1 and 57 for sh-TPX2) are indicated to the right of the micrographs.



Figure EV5. TPX2 deletion mutants retain their capability to associate with mitotic spindles.

HeLa cells stably expressing HA-TPX2 WT and the mutants were stained for HA-TPX2, α -tubulin, and DNA. Scale bars, 5 μ m.