

Supplementary material:

Temporal development of *Drosophila* embryos is highly robust across a wide temperature range

Jeronica Chong^{1,*}, Christopher Amourda^{1,*} and Timothy E. Saunders^{1,2,3,#}

¹ Mechanobiology Institute, National University of Singapore, Singapore

² Department of Biological Sciences, National University of Singapore, Singapore

³ Institute of Molecular and Cell Biology, A*Star, Proteos, Singapore

* These authors contributed equally to this paper

Correspondence: dbsste@nus.edu.sg

Supplementary figures

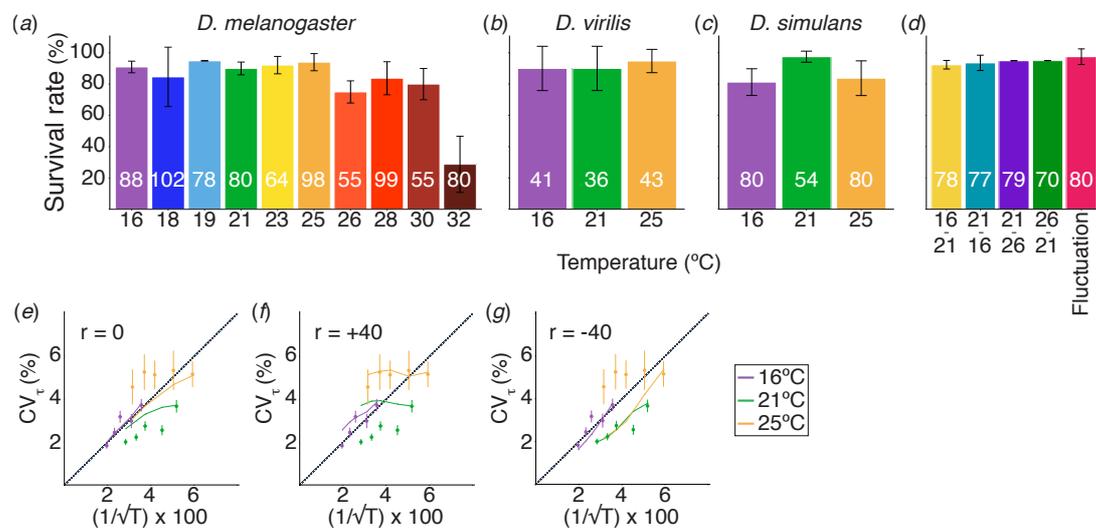


Figure S1: Experimental setup and controls

(a-d) Survival rate of *D. melanogaster* (a), *D. virilis* (b), *D. simulans* (c) and *D. melanogaster* with temperature perturbations (d). Numbers embedded in bars correspond to number of embryos. (e) Simulations of developmental trajectories in *D. melanogaster* without history dependence at any temperature. (f) Simulations of developmental trajectories in *D. melanogaster* with reinforcement of the history dependence at all temperatures. (g) Simulations of developmental trajectories in *D. melanogaster* with negative feedback in the history dependence at all temperatures. Errors on CV_τ are s.d., calculated using Bootstrapping as described in Methods.

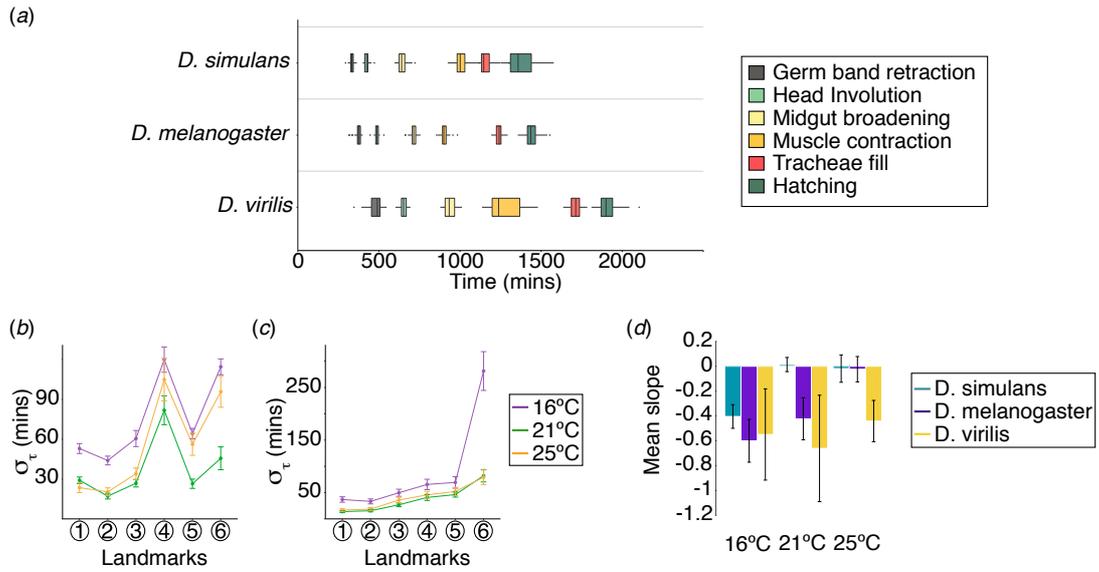


Figure S2: Temporal development of *Drosophila* species at 16°C, 21°C and 25°C
 (a) Distribution of developmental times at each landmark scored for *D. simulans*, *D. melanogaster* and *D. virilis* at 25°C. (b,c) Absolute error in developmental time at each landmark for *D. virilis* (b) and *D. simulans* (c) at 16, 21 and 25°C. (d) Fitting $CV_\tau = a\tau^s$ for each species at each temperature. Mean slope corresponds to value of s for fitting between germband retraction to trachea filling. All error bars are s.d.

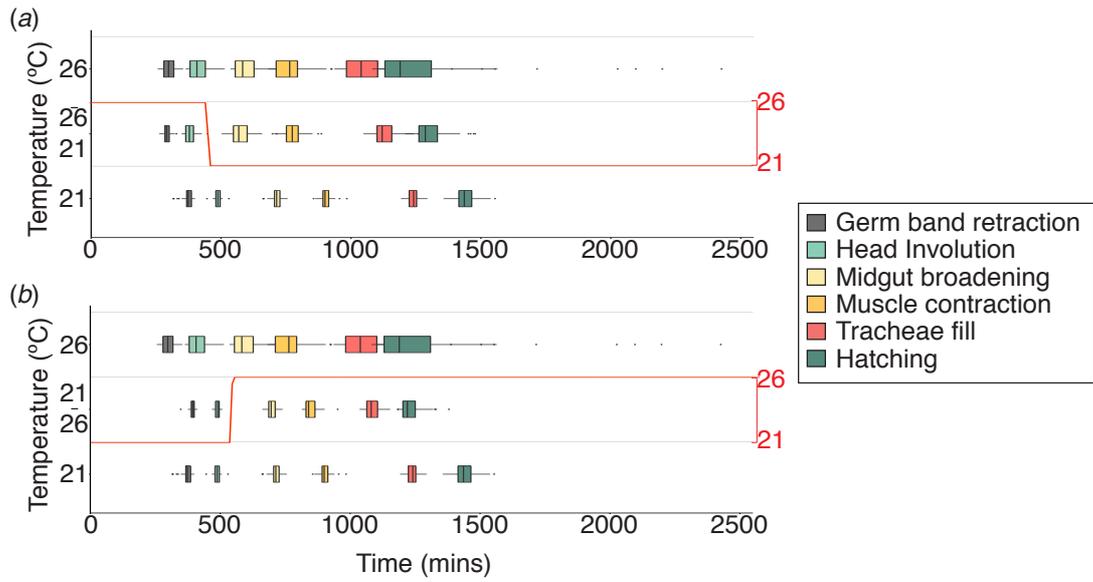


Figure S3: Further analysis of temperature shift

(a,b) Distribution of developmental times at each landmark scored for *D. melanogaster* under 21 to/from 26°C temperature shift. Temperature was shifted after head involution and is represented by a red curve.

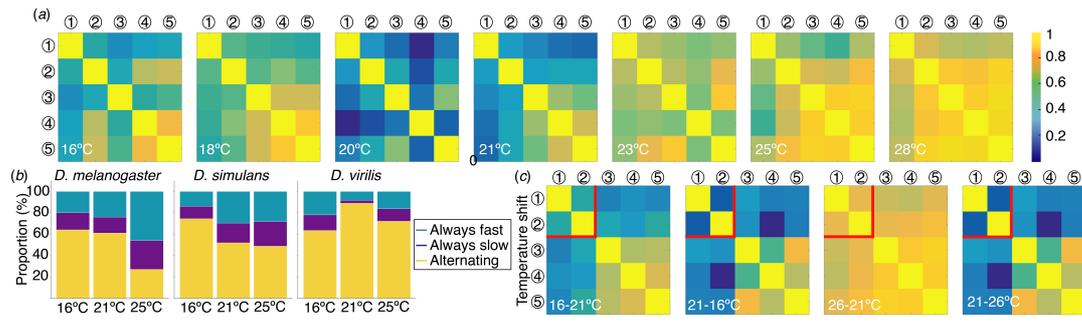


Figure S4: History dependence in *D. melanogaster*

(a) Covariance matrices for all temperatures recorded in *D. melanogaster*. (b) Proportion of embryos always faster than the mean (turquoise), slower than the mean (purple) or alternating between being faster or slower (yellow) when excluding germband retraction from analysis. (c) Covariance matrices for temperature shift experiments recorded in *D. melanogaster*.

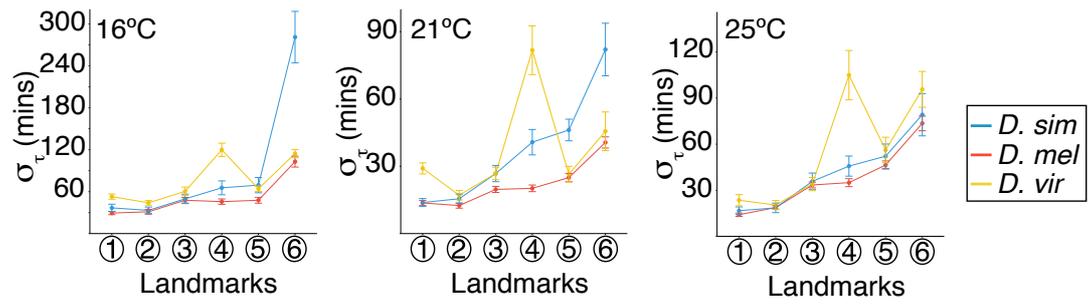


Figure S5: Absolute standard error of the different species

Absolute error in developmental time at each landmark for *D. melanogaster*, *D. virilis* and *D. simulans* at 16, 21 and 25°C. Errors are s.d.

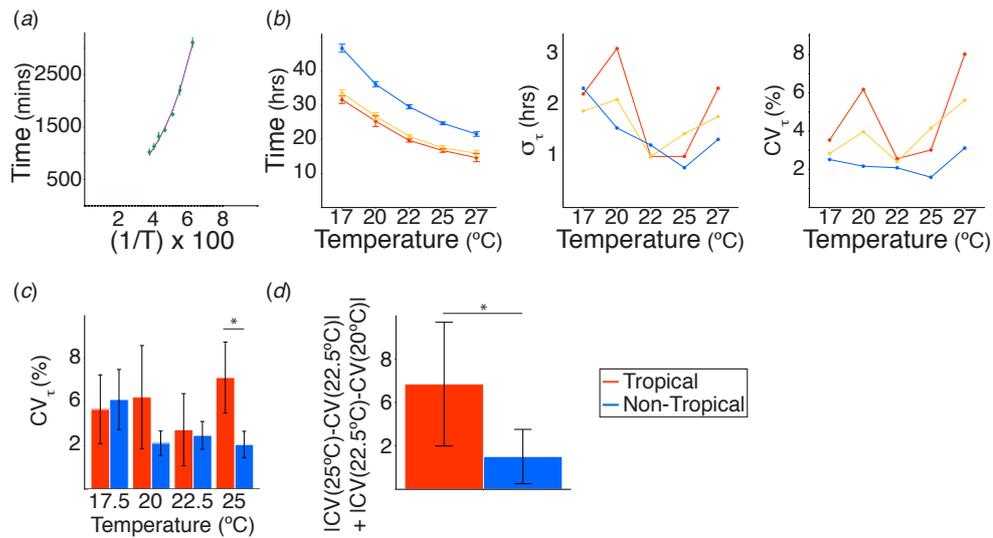


Figure S6: Comparison to previous measurements of temporal trajectories

(a) Time to hatching from cephalic furrow formation for *D. melanogaster* at different temperatures. Purple curve is a fit to the Arrhenius equation, $\tau = \tau_0 e^{T_0/T}$. (b) All data extracted from figure 5 of Kuntz and Eisen, PLoS Genetics 2014. Left: Measured time to tracheae filling at different temperatures in *D. simulans* (red), *D. melanogaster* (yellow) and *D. virilis* (blue). Centre: Measured temporal error, and right: coefficient of variation. Error bars are standard deviation. (c) CV_{τ} for the variability in times to trachea filling from the data set in Kuntz and Eisen, PLoS Genetics 2014. Data is separated into tropical (red, n=6) and non-tropical (blue, n=4) species, as described in main text. Error bars are s.d.. (d) Comparing the variability in CV_{τ} from 20°C to 25°C between tropical and non-tropical species. Non-tropical species (blue) have small change in CV_{τ} (<2% on average) across 20°C to 25°C, whereas tropical species (red) display larger variability in the recorded CV_{τ} with temperature. Error bars s.d., and significance determined by t-test comparison of the means.