

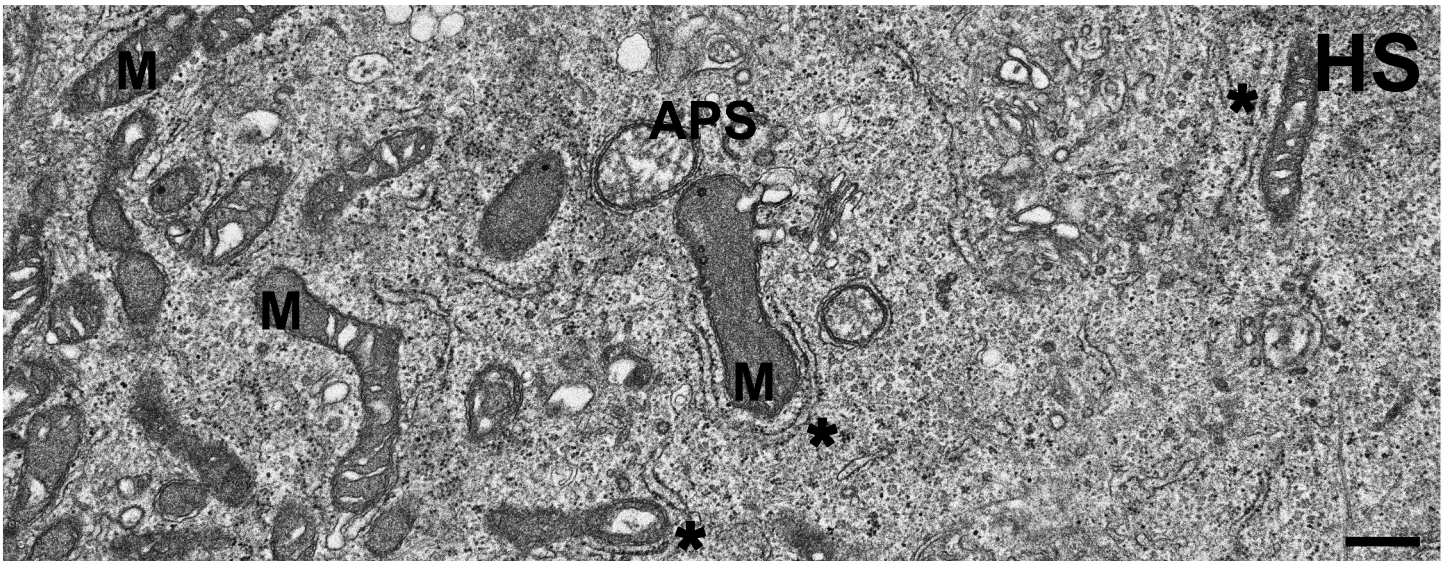
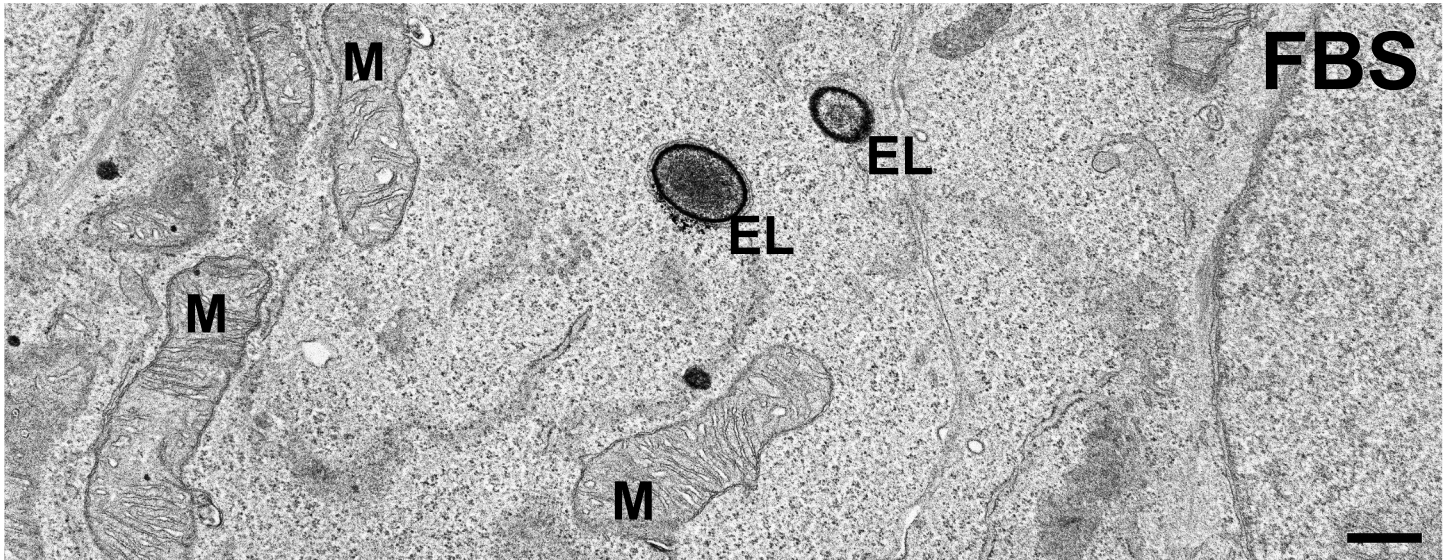
Supplemental data

Establishing normal metabolism and differentiation in hepatocellular carcinoma cells by culturing in adult human serum. Rineke Steenbergen,

Martin Oti, Rob ter Horst, Wilson Tat, Chris Neufeldt, Alexandr Belovodskiy, Tiing Tiing Chua, Woo Jung Cho, Michael Joyce, Bas E. Dutilh, D. Lorne Tyrrell

Supplemental Data 1

Other morphological changes in HS cultured cells



Additional morphological changes that were observed in the electron microscopic analysis include:

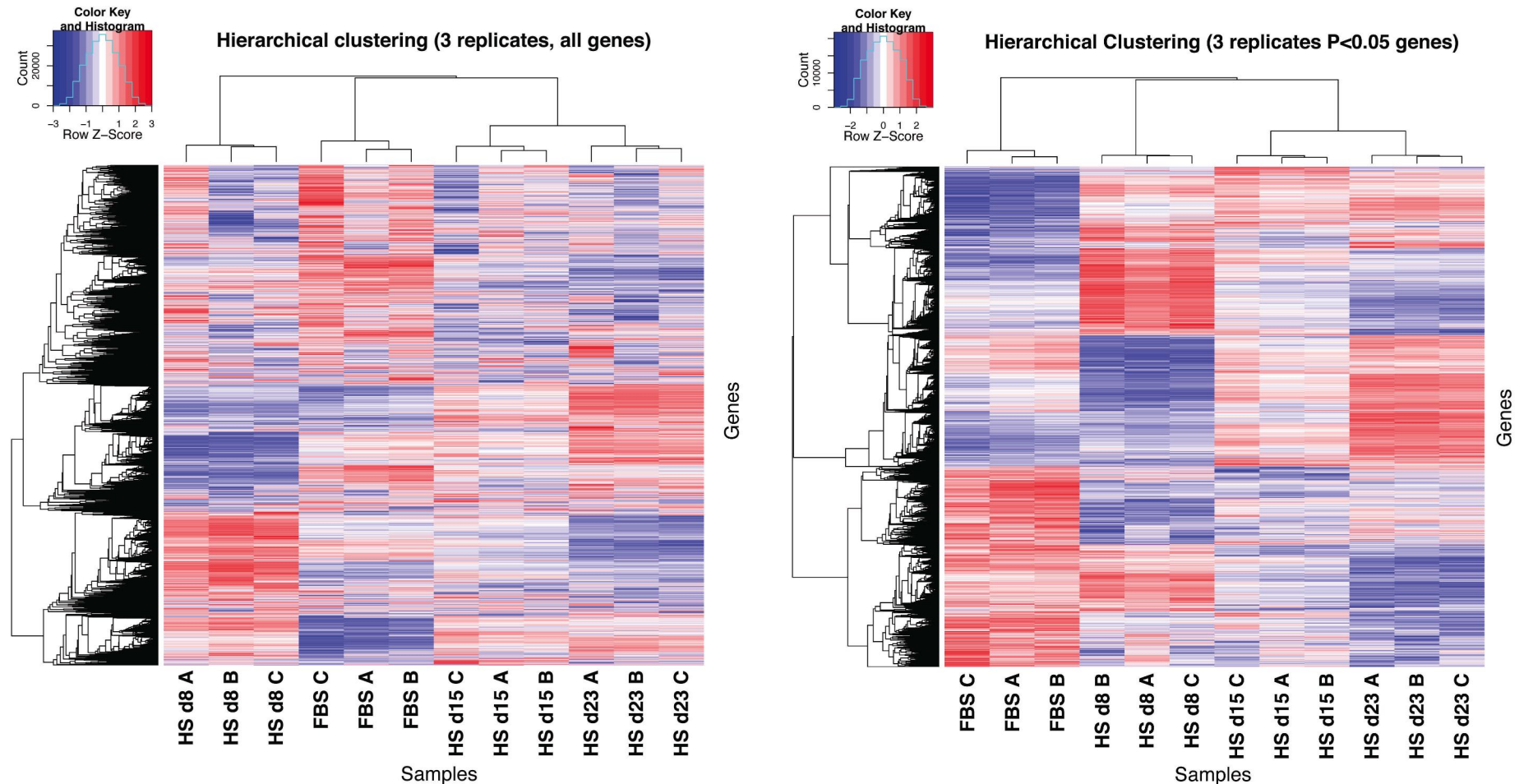
- The cytoplasm of HS cultured cells appears much more 'crowded' than the cytoplasm of FBS cultured cells.
- Mitochondrial (M) morphology has drastically changed, as outlined in figure 4 of the main document.
- In general, organelles appeared more structured and organized, for example, the endoplasmic reticulum (asterisks) shows a higher degree of organization, particularly around the mitochondria.
- An increase in vesicle transport and changes in autophagy and the lysosomal pathways were also observed on the electron micrographs. Whereas in FBS cells early lysosomes (EL) are abundant, other components of the endosomal/lysosomal pathway were harder to find, and no evidence was found of autophagy. In HS cultured cells late endosomes, multivesicular bodies and autophagosomes (APS) were abundant, potentially indicating a better functioning or more balanced endosomal/lysosomal/autophagosomal pathway is operational in HS cultured cells.

Supplemental Data 2

Comparison	Number of differentially expressed probes (multiple testing corrected, $p < 0.05$)	%
FBS - HSd8	16304	33
FBS - HSd15	11420	23
FBS - HSd23	16252	33

Number of differentially expressed probes for the different pairwise comparisons between days. The total number of probes on the Primeview Array was 49395.

Supplemental data 3, hierarchical clustering analysis



Clustering is a machine learning algorithm that groups data that have a high degree of similarity together in a cluster. In this analysis, data were clustered, initially, by using hierarchical clustering as shown above, based on gene expression pattern similarity of the entire dataset (y-axis) as well as on patterns of expression over time (x-axis). The z-scores of the gene expression levels were used in order to focus on the expression variation pattern across the samples rather than on their absolute expression levels, which can vary strongly from gene to gene.

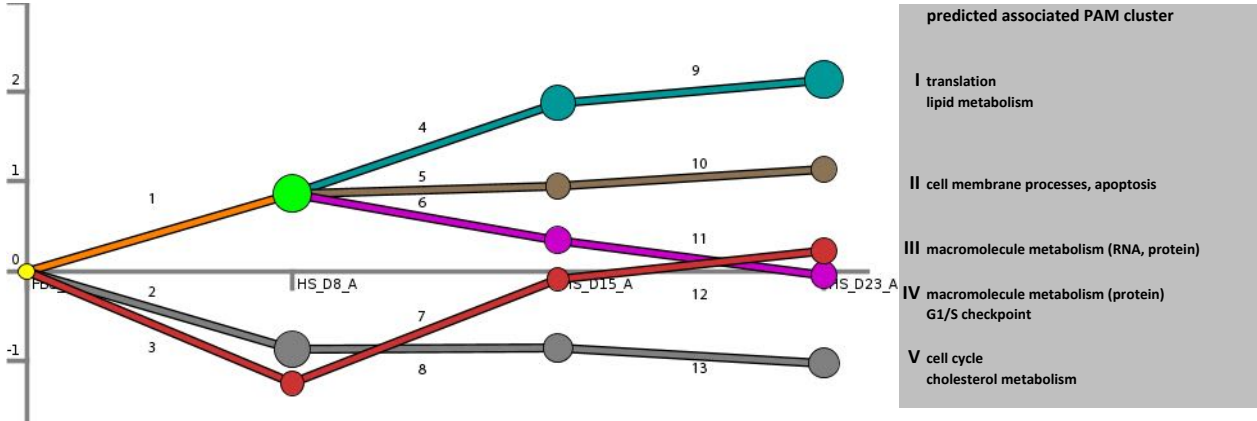
Hierarchical clustering of the entire data set is shown on the left and of transcripts that showed a significant change ($p < 0.05$) is shown on the right. In both analyses the replicates clustered together (x-axis), as was also shown by the PCA analysis (figure 1), and 6 patterns emerged (y-axis) when the data with significant changes were considered (right panel). To further analyze the genes or processes that were associated with these 6 clusters, a variant of k-means clustering was used, PAM clustering (partitioning around medoids), to generate more clearly delineated clusters (see figure 3 in the main document).

Supplemental Data 5: DREM analysis

The Dynamic Regulatory Events Miner (DREM) allows one to model, analyze, and visualize transcriptional gene regulation dynamics. The method of DREM takes as input time series gene expression data and known transcription factor-gene interaction data, and produces as output a dynamic regulatory map. The dynamic regulatory map highlights major branching events in the time series expression data and described the transcription factors (TFs) potentially responsible for them.

A: TFs that were significantly upregulated and predicted to be associated with upregulated pathways ($p < 10e-12$).

B: TFs predicted to be associated with upregulated pathways ($p < 10e-12$) (in order of significance for the associated processes).



FBS to Day 8

DAY8 TO DAY 15

DAY15 TO DAY 23

A

1	2	3

4	5	6	7	8
	AHR MYC EGR1 DDIT3	AHR	AHR	MYC AHR

9	10	11	12	13
	AHR MYC DDIT3	AHR	AHR	MYC AHR

B

RXRA	POU2F1	CUX1
POU2F1	HNF1A	POU2F1
CUX1	CUX1	MEF2A
MEF2A	POU3F2	TCF3
HNF1A	RXRA	STAT1
STAT1	NR3C1	STAT4
TCF3	STAT1	RXRA
SRF	MEF2A	CEBPA
PPARG	NFYA	SRF
IRF1	PGR	PDX1
POU3F2	CEBPA	FOX1L
STAT5A	NFYB	ARNT
CREB1	IRF1	AIRE
NR1H2	IRF7	CD40
PBX1	TFDP1	STAT6
ARNT	JUN	RORA
CD40	DSP	STAT2
GATA1	E2F4	AHR
NFKB1	E2F1	EGR3
NR3C1	PAX6	NFKB1
TBP	GATA1	E2F4
CEBPA	CREB1	STAT5B
AR	PBX1	TFDP1
NFYA	IRF2	TBX5
IRF8	FOXO4	DSP
SRY	MYC	
IRF2	MAX	
STAT3	ZEB1	
NFYB	AIRE	
HNF4A	PPARG	
AHR	E2F3	
GABPA	FOX1D	
PAX6	TBP	
PGR	SOX9	
HNF1B	TP53	
NR2F2	ATF2	
MEIS1	NFKB1	
JUN	NRF1	
PDX1	FOXC1	
RORA	GATA3	
SOX9	LMO2	

MEF2A	RXRA	POU2F1	CUX1	POU2F1
POU2F1	MEF2A	CUX1	POU2F1	HNF1A
RXRA	POU2F1	GABPA	MEF2A	CUX1
IRF1	PPARG	GATA1	TCF3	POU3F2
SRF	STAT1	HNF1A	STAT1	RXRA
HNF1A	CUX1	TCF3	STAT4	NR3C1
CUX1	HNF1A	RXRA	RXRA	STAT1
NR3C1	TCF3	STAT1	CEBPA	MEF2A
TCF3	POU3F2	TBP	SRF	NFYA
PPARG	CREB1	ELK4	PDX1	PGR
NRIH4	STAT5A	TEAD1	FOX1L	CEBPA
PBX1	NR1H2	IRF1	ARNT	NFYB
NR2F2	SRF	SRF	AIRE	IRF1
CEBPA	NFKB1	ARNT	CD40	IRF7
STAT1	CD40	CREB1	STAT6	TFDP1
	NFYA	JUN	RORA	JUN
	PBX1	POU1F1	STAT2	DSP
	ARNT	NFE2L1	EGR3	E2F4
	NFYB	STAT5A	NFKB1	E2F1
	PGR	RORA	E2F4	PAX6
	NR3C1	TP53	STAT5B	GATA1
	AR	IL6	TFDP1	CREB1
	IRF1	TFAP2A	TBX5	PBX1
	FOX1D	NFYA	DSP	IRF2
	MEIS1	CEBPB		FOXO4
	SRY	FOX1L		MAX
	CEBPA	ATF2		AIRE
	PAX5	FOS		PPARG
	MAX	EFNA2		E2F3
	VDR	STAT4		FOX1D
	PAX6			TBP
	REST			SOX9
	IRF8			TP53
	JUN			ATF2
	SOX9			NFKB1
	FOX1J			NRF1
	PDX1			FOX1C
	STAT3			GATA3
	NR1H3			LMO2
	RORA			RXR8
	IRF2			HNF1B

MEF2A	RXRA	POU2F1	CUX1	POU2F1
POU2F1	MEF2A	CUX1	POU2F1	HNF1A
RXRA	POU2F1	GABPA	MEF2A	CUX1
IRF1	PPARG	GATA1	TCF3	POU3F2
SRF	STAT1	HNF1A	STAT1	RXRA
HNF1A	CUX1	TCF3	STAT4	NR3C1
CUX1	HNF1A	RXRA	RXRA	STAT1
NR3C1	TCF3	STAT1	CEBPA	MEF2A
TCF3	POU3F2	TBP	SRF	NFYA
PPARG	CREB1	ELK4	PDX1	PGR
NRIH4	STAT5A	TEAD1	FOX1L	CEBPA
PBX1	NR1H2	IRF1	ARNT	NFYB
NR2F2	SRF	SRF	AIRE	IRF1
CEBPA	NFKB1	ARNT	CD40	IRF7
STAT1	CD40	CREB1	STAT6	TFDP1
	NFYA	JUN	RORA	JUN
	PBX1	POU1F1	STAT2	DSP
	ARNT	NFE2L1	EGR3	E2F4
	NFYB	STAT5A	NFKB1	E2F1
	PGR	RORA	E2F4	PAX6
	NR3C1	TP53	STAT5B	GATA1
	AR	IL6	TFDP1	CREB1
	IRF1	TFAP2A	TBX5	PBX1
	FOX1D	NFYA	DSP	IRF2
	MEIS1	CEBPB		FOXO4
	SRY	FOX1L		MAX
	CEBPA	ATF2		AIRE
	PAX5	FOS		PPARG
	MAX	EFNA2		E2F3
	VDR	STAT4		FOX1D
	PAX6			TBP
	REST			SOX9
	IRF8			TP53
	JUN			ATF2
	SOX9			NFKB1
	FOX1J			NRF1
	PDX1			FOX1C
	STAT3			GATA3
	NR1H3			LMO2
	RORA			RXR8
	IRF2			HNF1B

NR1H4 AHR
VDR RXRB
IRF7 HNF1B
SOX5 CEBPB
FOXF2 GATA2
RELA ELK1
MAX HIF1A
EGR1 SETD2
IRF3 HNF4A
IRF4 RFX1
IRF5 CD40
LEF1 FOS
NR2F1 RUNX1
POU1F1 PDX1
ARID5B BACH1
CEBPD STAT6
E2F1 CASR
EGR2 NR1I2
PAX5 FOXF2
FOS POU3F1
ATF6 MZF1
FOXA1 SPIB
ZBTB16 STAT5A
TP53 IRF3
NKX3-1 IRF4
FKBP4 IRF5
FOXO1 IRF8
REST STAT3
CNTN2 FOXJ2
MYC POU2F2
RFX1 GATA4
STAT5B NR1H2
ELK4 BACH2
ONECUT1 USF2
ONECUT2 ARNT
CEBPB ATF1
SREBF1 GABPA
FOXO3 AR
HOXA9 SPI
STAT4 NFE2L1
STAT6 CEBPG
IRF9 HSF1
EGR3 NR1H4
NFE2L1 UBE4A
TEAD1 VDR
ELSPBP1 NR2F2
FOXJ1 STAT4
FOXA2 ETS2
FOXA3 PLAU
FOXM1 GATA6
FOXC1 SRY
PLAU SRF
ATF2 ESR1
NR1H3 NR2F1
ETS1 SREBF1
FOXD1 SP3
ZEB1 NR1H3
FOXO4 FOXO1
SOX10 NHLH1
FOXL1 PAX2
ELF2 RUNX2
POU3F1 TCF3
BACH1 ATF3
MYOD1 GABPB1
RREB1 GABPB2
ALX1 RFX5
NR1I2 RFXANK
ESR1 RFXAP
LHX3 FOSL1
ETS2 JUNB
ATF3 JUND
CASR TFAP2A
FOSL1 TEAD1
JUNB TBX5
JUND GFI1
TFAP2A GFI1B
USF1 CREM
SP1 HOXA7
YY1 LEF1
DSP FOSL2
RXRB ARID5B
CREM FOXO3
SMAD4 FOXI1
ALX4 NR5A2
FOSL2 ATF7
STAT2 STAT5B
HLF LHX3
ATF1 ETS1
TCF4 RARA

HNF1B
GATA1
FOXC1
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MZF1
SPIB
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irf4
irf5
irf8
stat3
IRF7
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pou2f2
gata4
nr1h2
bach2
usf2
arnt
atf1
gabpa
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cebpb
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hsf1
nr1h4
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JUND
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HLF
FOXI1
TFAP4
PAX2

CEBPB
GATA2
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irf8
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bach2
usf2
arnt
atf1
gabpa
AR
SPI1
nfe2l1
cebpb
nr2f1
hsf1
nr1h4
ube4a
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FOXO3
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LHX3
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RARA
RARB
RARG
CEBPD
SMAD3

FOXD3	RARB
ZNF238	RARG
CEBPG	RXRG
NFIL3	CEBPD
IRF6	SMAD3
BACH2	USF1
SP3	GATA5
PPARA	ATF6
FOXJ2	FOXJ1
POU2F2	ATF4
TFAP4	EFNA2
GATA2	NFATC1
NKX2-1	NFATC2
NKX2-2	NFATC3
NR6A1	NFATC4
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POU5F1	NFIL3
POU5F1B	SMAD4
FOXI1	NFIC
NRF1	MEIS1
PAX2	TFE3
TFDP1	TFEB
CUZD1	YY1
FLI1	CEBPE
HSF1	PAX8

FOXL1
POU2AF1
POU4F1
POU5F1
POU5F1B
NFKB2
IRF6
SP2
AIRE
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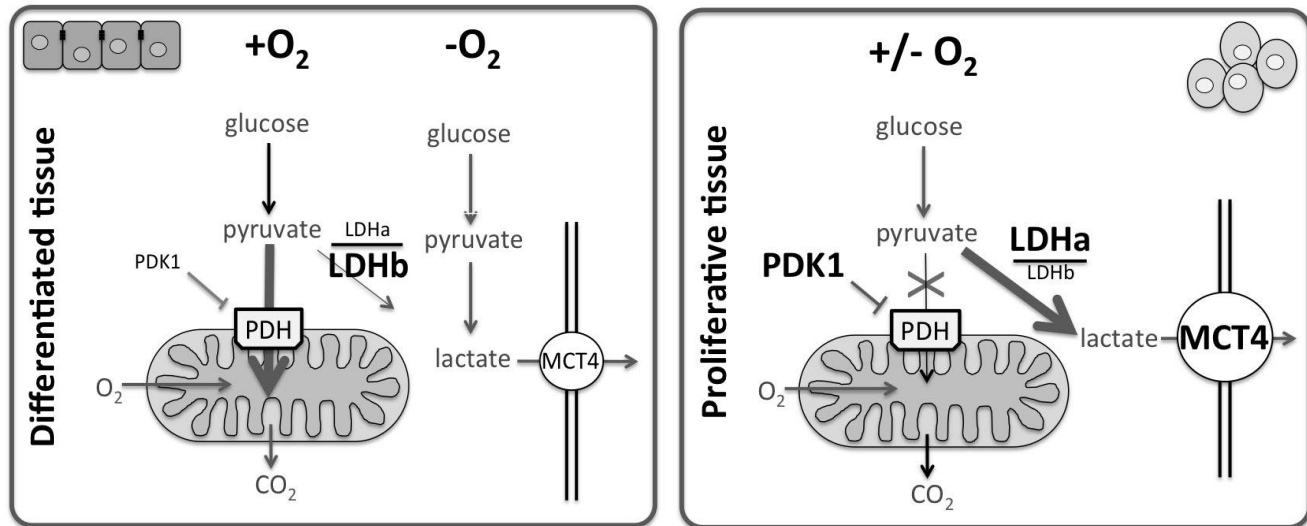
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FOXL1
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TFDP1
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EGR4
GATA4
FOSL2
MTF1
REL

USF1
GATA5
ATF6
FOXJ1
ATF4
EFNA2
NFATC1
NFATC2
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NFATC4
SOX5
BRCA1
ETV4
NKX2-1
FOXL1
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SMAD4
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TFE3
TFEB
YY1
CEBPE
PAX8
ELF2

Supplemental data 6

regulation of the Warburg effect



Proliferating cells often display a metabolic profile that is referred to as ‘cancer metabolism’, first described by Otto Warburg in 1924. Cancer metabolism is typically characterized by reduced levels of oxidative phosphorylation and mitochondrial activity, higher dependence on aerobic glycolysis and glutaminolysis for ATP production, and increased generation of biosynthetic intermediates that are essential for the production of macromolecules (phospholipids, nucleotides, proteins) to support cell proliferation (reviewed in^{3, 5-7}). The metabolic reprogramming that occurs during the Warburg effect is tightly regulated. The likely benefit for proliferative cells to adopt a Warburg-like metabolic profile, despite the much lower yield of ATP, is the conservation of pyruvate for the synthesis of lipids, nucleotides and amino acids, as building blocks for new cells¹⁻⁴.

Key regulators of the Warburg effect include:

* **PDK1** (pyruvate dehydrogenase kinase 1): in proliferative cells PDK1 is up-regulated, inhibiting PDH (pyruvate dehydrogenase), which transport pyruvate into the mitochondria, PDK1 upregulation results in limiting the uptake of pyruvate into the mitochondria. *PDK1 is down-regulated in HS-cultured cells, in line with a non-proliferative character. PDH did not change.*

* **LDHA and LDHB**: lactate dehydrogenases A and B catalyze the conversion of pyruvate to lactate (LDHA), vice versa (LDHB) High LDHA levels direct the conversion of pyruvate to lactate, whereas low LDHA levels, with higher LDHB levels favour the reverse reaction. *LDHA is decreased in HS cultured cells, whereas LDHB is increased, consistent with the ratio in differentiated tissue*

* **MCT4** (monocarboxylic acid transporter 4): MCT4 removes lactate from the cells, and is increased during aerobic glycolysis (right panel). *MCT4 is decreased in HS cultured cells, consistent with the reversal of the Warburg effect.*

1. Cairns, R.A., Harris, I.S. & Mak, T.W. Regulation of cancer cell metabolism. *Nat Rev Cancer* 11, 85-95 (2011).
2. Lunt, S.Y. & Vander Heiden, M.G. Aerobic glycolysis: meeting the metabolic requirements of cell proliferation. *Annu Rev Cell Dev Biol* 27, 441-464 (2011).
3. Pavlova, N.N. & Thompson, C.B. The Emerging Hallmarks of Cancer Metabolism. *Cell Metab* 23, 27-47 (2016).
4. Vander Heiden, M.G., Cantley, L.C. & Thompson, C.B. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 324, 1029-1033 (2009).

Supplemental Data 7: 25 genes with highest increase or decrease in expression

Increased expression (Top 25)

Gene Title	Gene Symbol	fold increase		
		d8	d15	d23
1 sulfotransferase family 1E, estrogen-preferring, member 1	SULT1E1	1.60980851	10.074521	17.2660059
2 apolipoprotein A-IV	APOA4	1.46445382	3.96256758	12.9515673
3 urothelial cancer associated 1 (non-protein coding)	UCA1	13.0403278	14.1624532	12.653981
4 S100 calcium binding protein A14	S100A14	1.10835538	2.47252987	12.1360695
5 anterior gradient 2 homolog (Xenopus laevis)	AGR2	4.09387803	7.3550794	11.4152296
6 phospholipase A1 member A	PLA1A	1.72761844	6.24060598	11.0728783
7 Kruppel-like factor 4 (gut)	KLF4	4.8156499	6.25374168	10.9723147
8 epithelial splicing regulatory protein 1	ESRP1	1.72298527	4.746049	10.2605455
9 guanylate binding protein 2, interferon-inducible	GBP2	4.90674414	7.71908765	10.2079568
10 transcription factor EC	TFEC	2.9324839	5.22814844	9.95592446
11 leucine rich repeat containing 19	LRRC19	1.10196646	2.89413995	9.76210338
12 zinc finger protein 114	ZNF114	2.96517484	4.5009058	9.35384665
13 cholinergic receptor, nicotinic, alpha 1 (muscle)	CHRNA1	3.89463693	4.89524151	8.91442928
14 discoidin domain receptor tyrosine kinase 1 /// microRNA 4640	DDR1	1.01238471	3.17424196	8.03071659
15 chromosome 19 open reading frame 69	C19orf69	1.08588648	2.76599503	7.98168685
16 UDP glucuronosyltransferase 2 family, polypeptide A3	UGT2A3	1.85090717	6.82158978	7.87140438
17 protease, serine, 23	PRSS23	3.95576075	6.46029912	7.76669156
18 chromosome 12 open reading frame 39	C12orf39	4.53196492	6.43655253	7.71021042
19 proline/histidine/glycine-rich 1	PHGR1	1.80191957	5.29692291	7.02058702
20 anoctamin 1, calcium activated chloride channel	ANO1	0.97804907	0.94345729	6.98110729
21 vanin 2	VNN2	11.4370228	10.9074657	6.93981919
22 insulin-like growth factor binding protein 3	IGFBP3	0.93963018	2.63802643	6.76395235
23 hephaestin	HEPH	2.6201261	4.21156181	6.52502213
24 cell death-inducing DFFA-like effector c	CIDEC	4.24499095	4.38011981	6.48955863
25 family with sequence similarity 47, member E	FAM47E	4.80857272	7.66127981	6.25362451

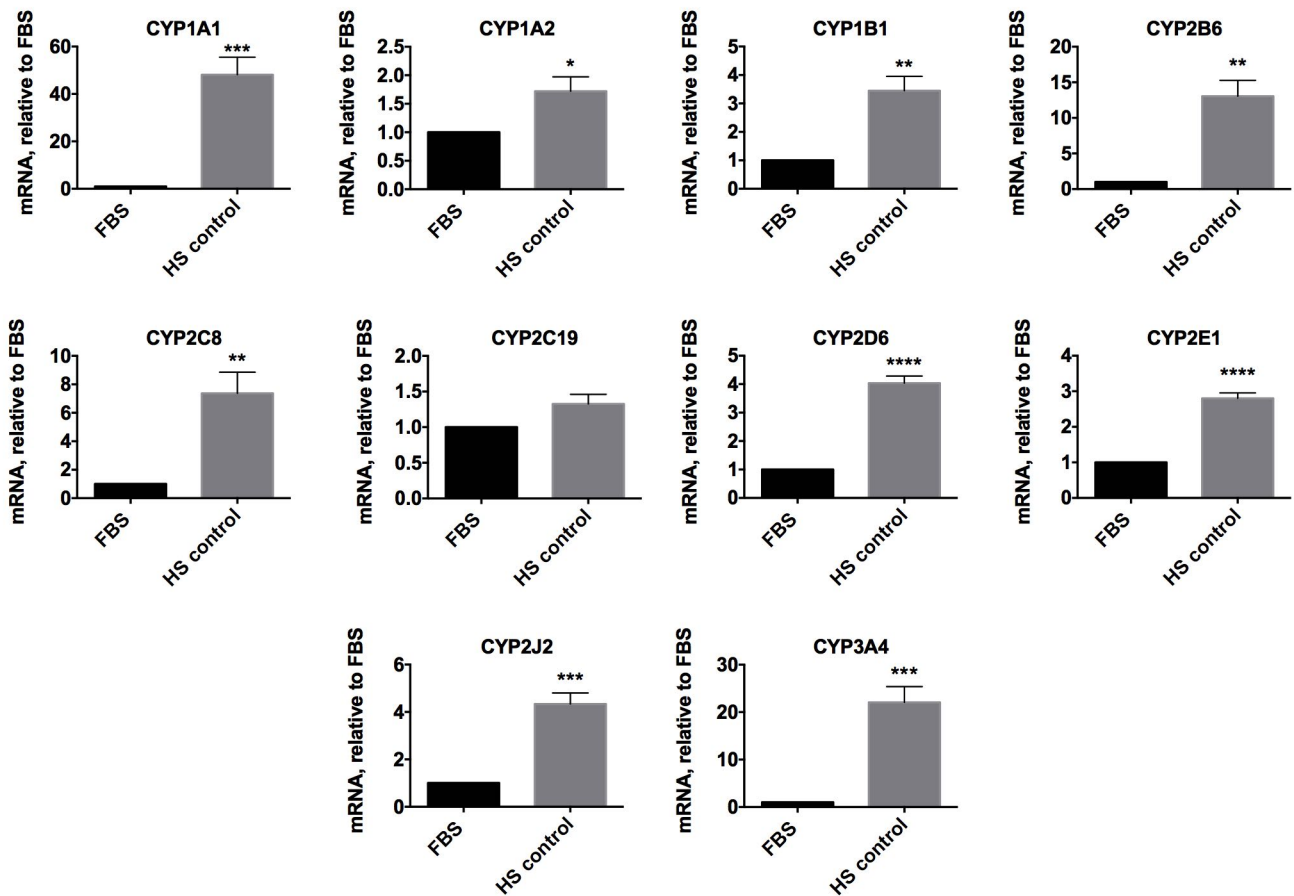
Decreased expression (Top 25)

Gene Title	Gene Symbol	fold decrease		
		d8	d15	d23
1 fibrinogen beta chain	FGB	1.01799746	2.61994629	5.23852945
2 leukemia inhibitory factor	LIF	5.24506451	5.02385697	4.77031089
3 tubulin, beta 1 class VI	TUBB1	2.94554978	3.25901429	4.70925991
4 glucan (1,4-alpha-), branching enzyme 1	GBE1	2.20827505	3.07756181	4.11316342
5 coagulation factor XIII, B polypeptide	F13B	2.234701	3.08244714	3.96939829
6 reelin	RELN	2.70351405	2.92224273	4.83406186
7 alcohol dehydrogenase 6 (class V)	ADH6	3.36367076	3.17448993	4.49200411
8 protein phosphatase 1, regulatory (inhibitor) subunit 1A	PPP1R1A	3.86675208	3.40475789	3.49474836
9 cancer susceptibility candidate 5	CASC5	2.32440917	2.14682899	3.47556406
10 F-box and leucine-rich repeat protein 21 (gene/pseudogene)	FBXL21	2.56581742	1.99000289	3.40485515
11 forkhead box N4	FOXN4	2.82919142	3.00748786	3.30598489
12 natriuretic peptide B	NPPB	0.73137249	1.77084611	3.2991722
13 claudin 14	CLDN14	3.44809761	3.16915066	3.28154663
14 fucosyltransferase 11 (alpha (1,3) fucosyltransferase)	FUT11	1.55861651	2.67772711	3.17790018
15 ribonucleotide reductase M2	RRM2	1.22693238	1.67626212	3.57449496
16 SPC25, NDC80 kinetochore complex component, homolog (S. cerevisiae)	SPC25	1.69842724	1.92727121	3.13577253
17 E2F transcription factor 7	E2F7	2.29203247	2.27124832	3.05634284
18 alanine-glyoxylate aminotransferase 2-like 1	AGXT2L1	1.11519966	1.67205374	3.40744429
19 fibrinogen alpha chain	FGA	2.0769312	3.00503011	3.4734301
20 6-phosphofructo-2-kinase/fructose-2,6-biphosphatase 4	PFKFB4	3.22650034	3.78462628	3.42101327
21 pyruvate dehydrogenase kinase, isozyme 1	PDK1	2.32540542	2.45489876	3.07676876
22 non-SMC condensin I complex, subunit G	NCAPG	1.98587391	2.51523109	3.02975911
23 hypoxia inducible lipid droplet-associated	HILPDA	1.63404113	2.40863783	3.15806134
24 kinesin family member 14	KIF14	1.26819019	1.80179264	3.40913024
25 zinc finger protein 789	ZNF789	1.45259678	2.17329004	2.86947321

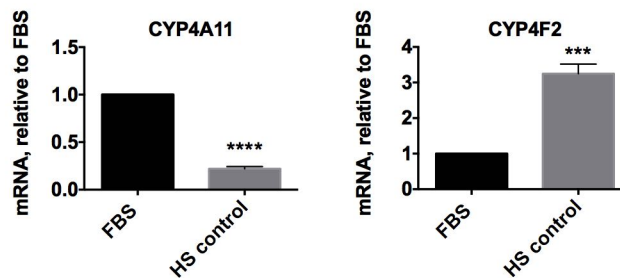
Supplemental Data 9

mRNA levels determined by quantitative PCR of different cytochrome P450 genes.

A: CYP1, 2 and 3 families: drug and steroid metabolism



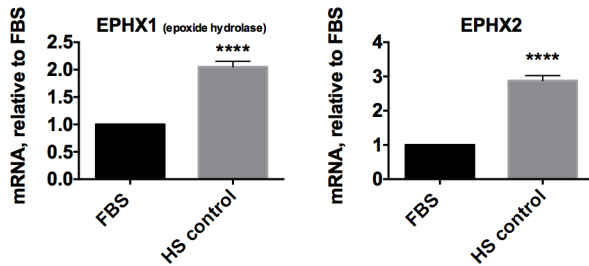
B: CYP4 family: arachidonic acid or fatty acid metabolism



mRNA levels determined by quantitative PCR of other factors involved in degradation or removal of xenobiotics

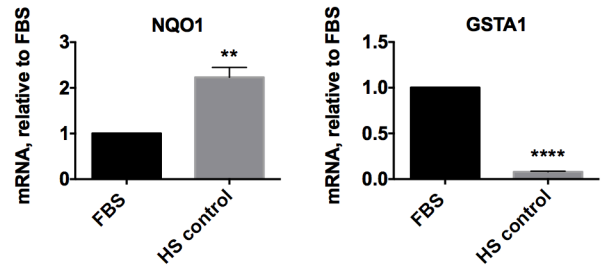
Epoxide hydrolases:

detoxification of exogenous chemicals such as polycyclic aromatic hydrocarbons



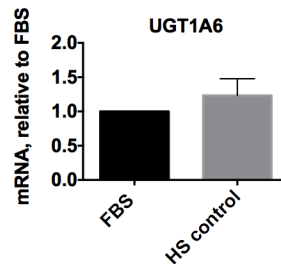
NQO1, GSTA1:

removal of lipid peroxidation by-products, carcinogens, bilirubin



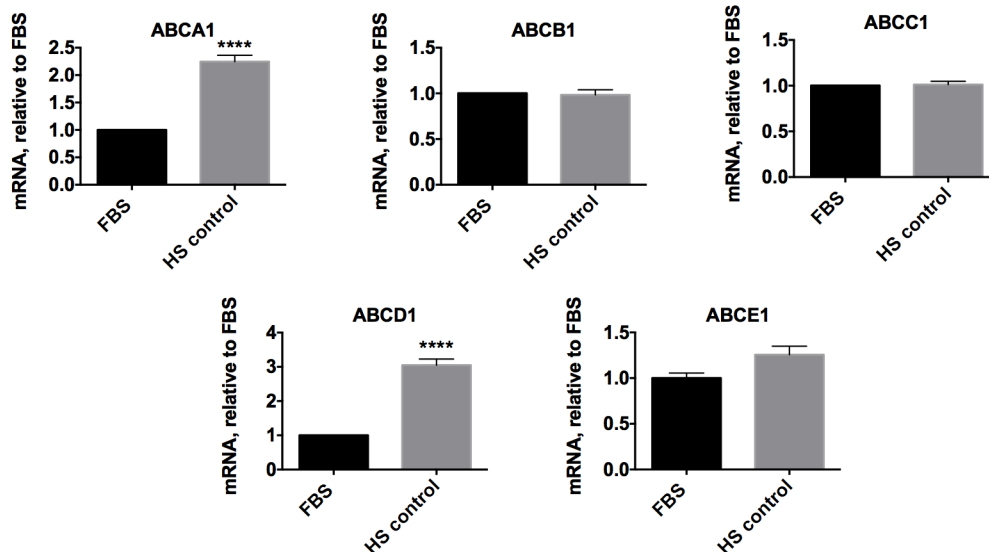
UDP-glucuronosulfotransferases:

transforms lipophilic molecules such as steroids, bilirubin, hormones, analgesic and other drugs to water soluble compounds



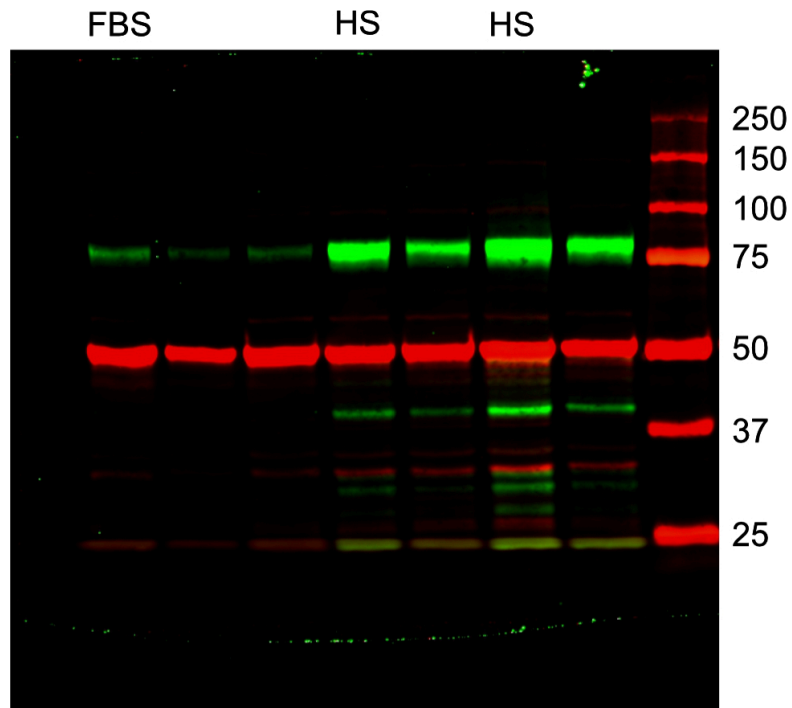
ABC transporters:

transport of a wide variety of substrates out of the cell, including metabolic products, lipids, steroids, and drugs. ABCA1 removes excess cholesterol from the cell, ABCB1 is also known as MDR-1, ABCC1 is also known as MRP1, ABCD1 transports fatty acids into the peroxisome,



Supplemental data 10

Full length blot of figure 5C



Full length Licor blot of the cropped images of figure 5C. CPT-1 has a predicted size of 78 kDa and is stained green. Tubulin and the molecular weight markers are stained red. Additional, unlabeled lanes on the blot represent other conditions that are not relevant for this study.