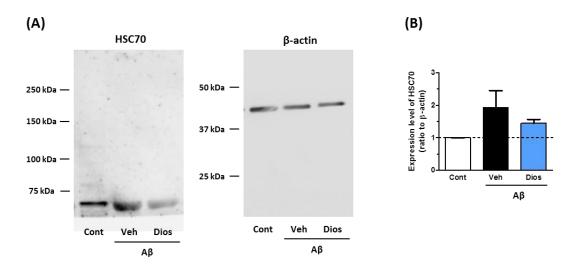
Supplementary Information

Diosgenin restores Aβ-induced axonal degeneration by reducing the expression of heat shock cognate 70 (HSC70)

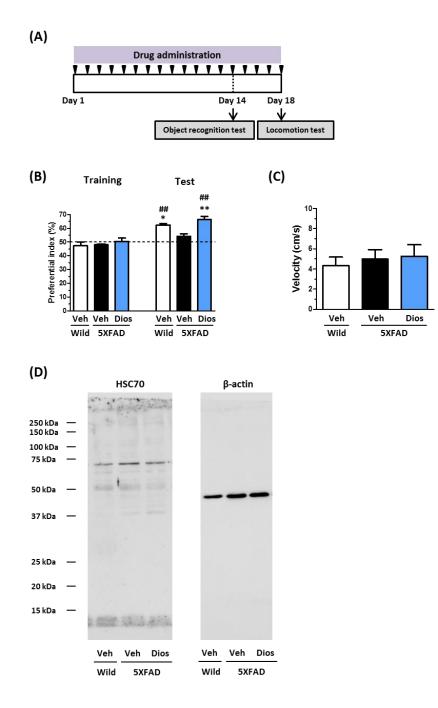
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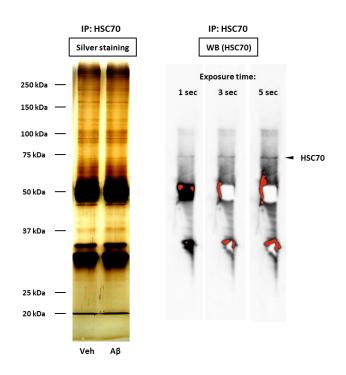
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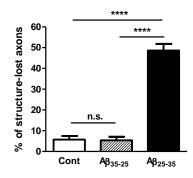
Supplementary Figure 1. Diosgenin reduced the expression level of HSC70 in neuron lysates. Mouse cortical neurons (ddY, E14) were cultured for three days and then treated with or without $A\beta_{25-35}$ (10 µM) for three days. After the three days of $A\beta_{25-35}$ treatment, neurons were treated with diosgenin (1 µM) or vehicle solution (0.1% EtOH) for four days. After the cells were washed with PBS, they were lysed and loaded on SDS-PAGE for WB analysis. (A) Proteins were detected by anti-HSC70 antibody or β -actin antibody, respectively. A whole membrane was cropped around 60 kDa into two pieces, upper one (for anti-HSC70) and lower one (for anti- β -actin) before WB. (B) Quantitative value for the expression levels of HSC70 (ratio to β -actin), n = 3 samples (2–4 lanes per a sample were quantified for the analysis).



Supplementary Figure 2. Diosgenin administration recovered memory deficits in the 5XFAD mice. Wild-type and 5XFAD mice (Female, 7–8 months old) were treated with diosgenin (0.1 µmol/kg/day, p.o.) or vehicle solution (sesame oil) for 18 days. (A) The time schedule of this study. On administration day 14, an object recognition memory test was performed. On the last day of administration, locomotion test was performed. (B) The preferential indexes of the training and test session are shown. *p < 0.05, **p < 0.01 vs 5XFAD/Veh, one-way ANOVA *post hoc* Dunnett's test, ##p < 0.01 vs 50%, one sample *t*-test, n = 4 mice. (C) The velocity (cm/s) within 10 min is shown. One-way ANOVA *post hoc* Dunnett's test, n = 4 mice. (D) The full-length WB images for the Figure 2C in the main text.



Supplementary Figure 3. The full-lengths gels for the Figure 5 in the main text.



Supplementary Figure 4. The effects of A β_{35-25} (a negative control peptide) and A β_{25-35} on axonal structures. Mouse cortical neurons were cultured for two days and treated with A β_{35-25} (10 µM) or A β_{25-35} (10 µM) for one day. After the medium was removed, the neurons were treated with fresh medium for four days. Then, neurons were fixed and double-immunostained for α -tubulin and pNF-H or observed with DIC. The percentage of structure-lost (low expression of α -tubulin) axons were quantified for each treatment. ****p < 0.0001, two-tailed one-way ANOVA *post hoc* Bonferroni's multiple comparison test. n = 3 photos were quantified for the analysis.