



Figure S1. Deletion of *vpsA* from *V. cholerae* impairs biofilm formation independent of T6SS functionality. Data A-B represent the mean  $\pm$  standard deviation of two biological replicates. (A) Crystal violet staining of *vpsA* mutants in wild type V52 and V52 *vasK* (T6SS<sup>-</sup>). (B) Expression of indicated genes in *vpsA* mutant backgrounds were measured by qRT-PCR with respect to wild type V52. (C) Supernatant (S) and cell pellets (P) were collected for wild type and *vpsA* mutants in the presence and absence of a functional T6SS and analyzed by western blot using anti-Hcp antibody. Blot with anti-vipA is a loading control.

Table S1. Strains used in this study

Organism	Genotype	Plasmid	Relevant Characteristics	Reference
<i>V. cholerae</i>				
2740-80	ClpV-mCherry2		Parental strain, lacZ <sup>-</sup> , Str <sup>r</sup>	(1)
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔvipA ClpV-mCherry2		<i>vipA</i> (VCA0107) in frame deletion, lacZ <sup>-</sup> , Str <sup>r</sup>	(1)
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔvpsA ClpV-mCherry2		<i>vpsA</i> (VC1917) in frame deletion, lacZ <sup>-</sup> , Str <sup>r</sup>	this study
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔvpsA ΔvipA ClpV-mCherry2		<i>vpsA</i> (VC1917) in frame deletion in <i>vipA</i> background, lacZ <sup>-</sup> , Str <sup>r</sup>	this study
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔflaA ClpV-mCherry2		<i>flaA</i> (VC2188) in frame deletion, lacZ <sup>-</sup> , Str <sup>r</sup>	this study
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔmotX ClpV-mCherry2		<i>motX</i> (VC2601) in frame deletion, lacZ <sup>-</sup> , Str <sup>r</sup>	this study
	lacZ <sup>-</sup> , Str <sup>r</sup> , ΔflaA ΔvpsA ClpV-mCherry2		<i>vpsA</i> (VC1917) in frame deletion in <i>flaA</i> background, lacZ <sup>-</sup> , Str <sup>r</sup>	this study
V52	Δrhh		<i>rtxA hlyA hapA</i> parental strain, Str <sup>r</sup>	(2)
	Δrhh ΔvasK		<i>vasK</i> (VCA0120) in frame deletion, Str <sup>r</sup>	(2)
	Δrhh ΔvpsA		<i>vpsA</i> (VC1917) in frame deletion, Str <sup>r</sup>	this study
	Δrhh ΔvpsA ΔvasK		<i>vpsA</i> in frame deletion in <i>vasK</i> background, Str <sup>r</sup>	this study
	Δrhh ΔflaA		<i>flaA</i> (VC2188) in frame deletion, Str <sup>r</sup>	this study
	Δrhh ΔmotX		<i>motX</i> (VC2601) in frame deletion, Str <sup>r</sup>	this study
	Δrhh	pBAD33	parental strain with empty pBAD33 plasmid, Str <sup>r</sup> , Cm <sup>r</sup>	this study
	Δrhh ΔvpsA	pBAD33	<i>vpsA</i> deletion in parental strain with empty pBAD33 plasmid, Str <sup>r</sup> , Cm <sup>r</sup>	this study
	Δrhh ΔVgrG-3ΔVCA0124		<i>VgrG3-VCA0124</i> in frame deletion, Str <sup>r</sup>	(2)
	Δrhh ΔVC1417-VC1421		<i>VC1417-VC1421</i> in frame deletion, Str <sup>r</sup>	(2)
	Δrhh ΔVCA0019-VCA0021		<i>VCA0019-VCA0021</i> in frame deletion, Str <sup>r</sup>	(2)
	Δrhh ΔVgrG-3ΔVCA0124 ΔvpsA		<i>vpsA</i> in frame deletion in <i>VgrG3-VCA0124</i> background, Str <sup>r</sup>	this study
	Δrhh ΔVC1417-VC1421 ΔvpsA		<i>vpsA</i> in frame deletion in <i>VC1417-VC1421</i> background, Str <sup>r</sup>	this study
	Δrhh ΔVCA0019-VCA0021 ΔvpsA		<i>vpsA</i> in frame deletion in <i>VCA0019-VCA0021</i> background, Str <sup>r</sup>	this study
	Δrhh ΔVgrG-3ΔVCA0124	pCR-XL-TOPO	<i>VgrG3-VCA0124</i> in frame deletion with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study
	Δrhh ΔVC1417-VC1421	pCR-XL-TOPO	<i>VC1417-VC1421</i> in frame deletion with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study
	Δrhh ΔVCA0019-VCA0021	pCR-XL-TOPO	<i>VCA0019-VCA0021</i> in frame deletion with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study

	$\Delta$ rh $\Delta$ VgrG-3 $\Delta$ VCA0124 $\Delta$ vpsA	pCR-XL-TOPO	<i>vpsA</i> deletion in <i>VgrG3-VCA0124</i> background with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study
	$\Delta$ rh $\Delta$ VC1417-VC1421 $\Delta$ vpsA	pCR-XL-TOPO	<i>vpsA</i> deletion in <i>VC1417-VC1421</i> background with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study
	$\Delta$ rh $\Delta$ VCA0019-VCA0021 $\Delta$ vpsA	pCR-XL-TOPO	<i>vpsA</i> deletion in <i>VCA0019-VCA0021</i> background with empty pCR-XL-TOPO plasmid, Str <sup>r</sup> , Kan <sup>r</sup>	this study
	$\Delta$ rh $\Delta$ rbmA		<i>rbmA</i> (VC0928) in frame deletion, Str <sup>r</sup>	this study
	$\Delta$ rh $\Delta$ rbmC		<i>rbmC</i> (VC0930) in frame deletion, Str <sup>r</sup>	this study
	$\Delta$ rh $\Delta$ bap1		<i>rbmA</i> (VC1888) in frame deletion, Str <sup>r</sup>	this study
	$\Delta$ vgrG3	pNucSe1	<i>vgrG3</i> in frame deletion with NucSe1 expressed on pBAD33	(3)
	$\Delta$ vgrG3 $\Delta$ vipA	pNucSe1	<i>vgrG3 vipA</i> in frame deletion with NucSe1 expressed on pBAD33	(3)
<i>E. coli</i>	MG1655		Gm <sup>r</sup>	(1)
<i>A. baylyi</i>	ADP1		ATCC 33305 parental strain	(4)
	ADP1 $\Delta$ T6SS		T6SS deletion, Kan <sup>r</sup>	(4)
<i>P. aeruginosa</i>	PA01 $\Delta$ retS ClpV-GFP		parental strain, Irg <sup>r</sup>	(1)

Table S2. Plasmids used in this study

<b>Plasmid</b>	<b>Relevant Characteristics</b>	<b>Source</b>
pDS132	suicide vector used for allelic exchange, <i>sacB</i> , Cm <sup>r</sup>	(5)
pWM91	suicide vector used for allelic exchange, <i>sacB</i> , Carb <sup>r</sup>	(6)
pCR-XL-TOPO	TOPO cloning vector, used to make strains kanamycin resistant, Kan <sup>r</sup>	Invitrogen
pBAD33	expression vector, Cm <sup>r</sup>	(1)
pWM91- $\Delta$ vpsA	<i>vpsA</i> suicide vector, Carb <sup>r</sup>	this study
pDS132- $\Delta$ flaA	<i>flaA</i> suicide vector, Cm <sup>r</sup>	this study
pDS132- $\Delta$ motX	<i>motX</i> suicide vector, Cm <sup>r</sup>	this study

## References

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