

BMJ Open is committed to open peer review. As part of this commitment we make the peer review history of every article we publish publicly available.

When an article is published we post the peer reviewers' comments and the authors' responses online. We also post the versions of the paper that were used during peer review. These are the versions that the peer review comments apply to.

The versions of the paper that follow are the versions that were submitted during the peer review process. They are not the versions of record or the final published versions. They should not be cited or distributed as the published version of this manuscript.

BMJ Open is an open access journal and the full, final, typeset and author-corrected version of record of the manuscript is available on our site with no access controls, subscription charges or pay-per-view fees (<u>http://bmjopen.bmj.com</u>).

If you have any questions on BMJ Open's open peer review process please email <u>info.bmjopen@bmj.com</u>

BMJ Open

A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-Infected Women Living in Rwanda

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-020432
Article Type:	Protocol
Date Submitted by the Author:	04-Nov-2017
Complete List of Authors:	Murenzi, Gad; Rwanda Military Hospital Dusingize, Jean; Regional Alliance for Sustainable Development , Research and Scientific Capacity Building Rurangwa, Theogene; Rwanda Military Hospital Sinayobye, Jean d'Amour; Regional Alliance for Sustainable Development, ; Women's Equity in Access to Care & Treatment (WE-ACTx), Munyaneza, Athanase; Rwanda Military Hospital Murangwa, Anthere; Rwanda Military Hospital Zawadi, Thierry; Rwanda Military Hospital Hebert, Tiffany; Yeshiva University Albert Einstein College of Medicine Mugenzi, Pacifique; Rwanda Military Hospital Adedimeji, Adebola; Yeshiva University Albert Einstein College of Medicine Mutesa, Leon; Rwanda Military Hospital; University of Rwanda College of Medicine and Health Sciences Anastos, Kathryn; Albert Einstein College of Medicine Medical Center, Bronx, Castle, Philip; Yeshiva University Albert Einstein College of Medicine,
Keywords:	human papillomavirus (HPV), cervical cancer, HIV & AIDS < INFECTIOUS DISEASES, GYNAECOLOGY, cervical intraepithelial neoplasia

SCHOLARONE[™] Manuscripts Page 1 of 27

1	
2	
2	
⊿	
3 4 5 7 8 9 10 11	
5 6	
6	
7	
8	
9	
10	
11	
12	
13	
12 13 14 15	
15	
16	
17	
18	
14 15 16 17 18 19	
20	
∠∪ 21	
21	
22	
19 20 21 22 23 24 25 26 27 28 29 30	
24	
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
34 35 36 37	
0C 7C	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
52 53	
54	
55	
56	
57	
58	
59	
60	

1	A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-
2	Infected Women Living in Rwanda
3	Gad Murenzi, MD* ¹ , Jean-Claude Dusingize, MD, MS ¹ , Theogene Rurangwa, MD, MMed ¹ ,
4	Jean d'Amour Sinayobye, MD, MS ¹ , Athanase Munyaneza, RN ¹ , Anthere Murangwa, MS ¹ ,
5	Thierry Zawadi, MD ¹ , Tiffany Hebert, MD ² , Pacifique Mugenzi, MD ¹ , MMed, Adebola
6	Adedimeji, PhD, MPH ² , Leon Mutesa, MD, PhD ^{1,3} , Kathryn Anastos, MD ² , Philip E. Castle,
7	PhD, MPH ^{2,4}
8	¹ Rwanda Military Hospital, Kigali, Rwanda; ² Albert Einstein College of Medicine, Bronx, NY,
9	USA; ³ University of Rwanda, Kigali, Rwanda; ⁴ Global Coalition Against Cervical Cancer,
10	Arlington, VA, USA.
11	*Correspondence: <u>gadcollins@gmail.com</u> , +250788589085
12	Disclosures: This research study has received HPV tests for reduced or no cost from Cepheid,
13	Arbor Vita Corporation, and Roche.
14	Funding: This study was funded by NCI/NIH Grant 5U54CA19016304 and by a grant from the
15	Prevent Cancer Foundation.
16	

17 Abstract

Introduction. The optimal method(s) for screening human immunodeficiency virus-infected
 women, especially for those living in sub-Saharan Africa, for cervical precancer and early cancer
 has yet to be established.

Methods and analysis. A convenience sample of >5,000 Rwandan women, aged 30-54 years, living with HIV infection will be enrolled into a cross-sectional study of cervical cancer screening strategies. Eligible and consenting women will be enrolled into the study, complete a short risk-factor questionnaire, and screened for high-risk human papillomavirus (hrHPV) using the Xpert HPV assay (Cepheid, Sunnyvale, CA, USA), unaided visual inspection after acetic acid (VIA), and aided VIA using the EVA system (Mobile ODT, Tel Aviv, Israel). Women positive for hrHPV or by VIA will undergo colposcopy, which will include the collection of two cervical specimens prior to undergoing a 4-quadrant microbiopsy protocol. The colposcopy-collected specimens will be tested by dual immunocytochemical staining for p16^{INK4a} and Ki-67 (CINtec® PLUS Cytology, Ventana, Tucson, AZ, USA) and for E6 or E7 for 8 hrHPV genotypes (HPV16, 18, 31, 33, 35, 45, 52, and 58) using the next-generation AV Avantage hrHPV E6/E7 test (Arbor Vita Corporation, Freemont, CA, USA). Women with local pathology diagnosis of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) or pathology-review diagnosis of CIN grade 3 or more severe (CIN3+) will receive treatment. Clinical performance and cost-effectiveness (e.g., sensitivity, specificity, and predictive values)

- ²⁷ 36 of different screening strategies and algorithms will be evaluated.
- 30 37 *Ethics and dissemination.* The protocol has been approved by local and institutional review
- boards for human subjects research. Results will be disseminated to the scientific community
 - 39 through peer-reviewed publication and to the Rwandan stakeholders through an external
- 32 39 through peer-rev
 33 40 advisory panel.
 34

1		
2 3	42	Strengths and Weaknesses
4	12	Strengens and Weaknesses
5 6	43	• We will enroll a very large sample size of HIV-infected women living in Rwanda who
7	44	otherwise would probably not get cervical cancer screening.
8 9	45	• We will employ rigorous disease ascertainment protocols to minimize misclassification.
10	46 47	• Some tests, such as the Xpert HPV and the AV Avantage HPV E6/E7 assays, will be done on site in Rwanda using technologies that could feasibly be deployed there.
11	48	 A weakness of the study is that it will not be feasible to include cervical cytology as a
12 13	49	comparator due to financial and logistical constraints.
14	50	
15 16		
17		
18		
19 20		
21		
22		
23 24		
25		
26 27		
28		
29 30		
30 31		
32		
33 34		
35		
36 37		comparator due to financial and logistical constraints.
38		
39		
40 41		
42		
43 44		
45		
46 47		
47 48		
49		
50 51		
52		
53 54		
54 55		
56		
57 58		
59		

51 Introduction

Invasive cervical cancer (ICC) remains a significant cause of morbidity and mortality globally. Approximately 530,000 cases of and 270,000 deaths due to ICC occur annually, making it the 4th most common malignancy and cause of cancer-related death in women globally.¹ In many high-income countries (HIC), cervical cancer rates have declined by 50% or more² due to the introduction of effective, high-coverage Pap testing (cervical cytology)-based screening programs that include timely follow-up of screen positives, treatment of women with precursor lesions, and management of cancers. Now, almost 90% of ICC and ICC-related deaths occur in low- and middle income countries (LMICs) due to a lack of resources and healthcare infrastructure needed to provide preventive services. ICC and ICC-related mortality rates are particularly high in Sub-Saharan Africa, which also has the highest rates of HIV infection in the world. Now, over 12 million HIV-infected (HIV[+]) women in Sub-Saharan Africa are living longer because of anti-retroviral therapy, thus increasing their likelihood of dying from ICC.³ However, many of these women are already exposed to human papillomavirus (HPV), the viral cause of cervical cancer, and will not benefit

from or be targeted for prophylactic HPV vaccination. Thus, cervical cancer screening is neededfor the foreseeable future.

However, setting up effective cytology for cervical cancer screening is expensive and requires a complex clinical and lab infrastructure that generally does not exist in LMICs.^{4;5} Moreover, it is now well understood that cytology has only a low- to moderate one-time sensitivity for precursor lesions and therefore must be done repeatedly over many years to reduce cancer risk. Alternative

BMJ Open

strategies to address the cervical cancer burden in LMICs, especially in SSA, must be developedand validated.

Persistent cervical infections by high-risk HPV (hrHPV) types cause virtually all ICC and its immediate precursor lesions, e.g. cervical intraepithelial neoplasia grade 3 (CIN3) and adenocarcinoma *in situ* (AIS) everywhere in the world.^{6;7} hrHPV causes most anal and vaginal cancer and a significant proportion of vulvar, penile, and oropharyngeal cancers.⁸ HPV16 is the most important causal type, responsible for ~60% of ICC.⁹ HPV18 is the next most important, responsible for 10-15% of ICC, including 30-40% of adenocarcinoma of the cervix⁹, which is on the rise in Western Countries.^{10;11} Together, HPV16 and HPV18 account for ~70% of ICC, and the same 15 hrHPV types account for ~99% of ICC everywhere in the world.⁹

There is now overwhelming evidence to suggest that testing for hrHPV is more sensitive, albeit less specific, than high-quality cytology for identifying women with cervical precancer.¹²⁻¹⁶ Onetime hrHPV testing can reduce the risk of ICC incidence by approximately 40% in 6.5 years compared to cytology screening¹⁶, and ICC mortality by approximately 40% (approximately 50% overall) in 8 years compared to cytology.¹⁷ Importantly, a negative hrHPV test provides superior reassurance against CIN3+ ¹⁸ and against ICC^{16;17}, permitting safe extension of screening intervals.

The World Health Organization released cervical cancer screening and treatment guidelines in 2013, recommending two evidence-based approaches to ICC screening¹⁹: (I) Use either hrHPV testing or visual inspection after acetic acid (VIA), which involves the inspection of the cervix with a speculum in place and following the application of dilute acetic acid to help identify potential CIN by its characteristic white coloring in the presence of acetic acid (acetowhite), as

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

alternative initial screening tests instead of Pap, and (II) immediately treat those who screen
positive using the screening test, rather than require diagnostic verification through colposcopy
and biopsy. This approach is commonly called screen-and-treat (S&T), and is increasingly
thought to be more amenable to LMIC settings.

However, hrHPV testing is also a much more effective screen than VIA¹⁷, which on a large-scale
appears to only down-stage cancer rather than prevent it.²⁰ Thus, the recent American Society for
Clinical Oncology (ASC) resource-stratified guidelines for secondary cervical cancer
prevention^{21;22} emphasize that hrHPV testing is the preferred choice for screening, with VIA
only being used until hrHPV testing becomes available, and that HIV-infected women, because
of their higher risk, should be screened twice as frequently as the general (HIV-uninfected)
population.

Recent data in HIV[+] women living in the U.S. suggest that hrHPV testing may have clinical utility similar to that in HIV-negative (HIV[-]) women. Several observational studies have shown that an extended screening interval is safe in HIV[+] women who test hrHPV and Pap negative as it is for HIV[-] women.^{23;24} In a study of women enrolled in Women's Interagency Health Study (WIHS) in 2002, HIV[+] and HIV[-] women who tested hrHPV and Pap negative were at a similarly low risk of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) histology over a 5-year follow-up.²³ In addition, no cases of histologically confirmed CIN2+ were diagnosed in the follow-up of hrHPV- and Pap-negative HIV[+] women aged 30-64 years who underwent routine three-year hrHPV and cytology cotesting at Kaiser Permanente Northern California.²⁴ Thus, both studies found very high negative predictive values (NPV) >99% in HIV[+] women who test hrHPV negative. However, how hrHPV testing can

Page 7 of 27

BMJ Open

1	
2 3 4	116
5 6	117
7 8	
9 10	118
11 12	119
13 14	120
15 16	121
17 18 19	122
20 21	123
22 23	124
24 25	
26 27	125
28 29	126
30 31	127
32 33	128
34 35 36	129
37 38	130
39 40	131
41 42	132
43 44 45	133
46 47	134
48 49	135
50 51	136
52 53	100
54 55	
56 57	
58	
59 60	

best be used to screen HIV[+] women living in Sub-Saharan Africa to prevent cervical cancerremains to be determined.

Recent studies²⁵⁻²⁸ in HIV[+] women living in Sub-Saharan Africa have compared hrHPV, VIA, and/or Pap for the detection of cervical precancer/cancer. The results can be summarized as follows: 1) hrHPV detection was more sensitive but less specific than VIA and 2) surprisingly, cytology was equally or more sensitive but less specific than VIA and 2) surprisingly, cytology was equally or more sensitive but less specific (vs. the converse) than hrHPV testing. Results and conclusions are varied, leaving unanswered the question of what screening strategy in HIV[+] women living in Sub-Saharan Africa has the greatest effectiveness and cost effectiveness.

Regardless of the screening method, most screen-positive women who go to colposcopy or are treated immediately without diagnostic verification do not have cervical precancer and cancer (positive predictive value [PPV] for screening tests are typically 10%-20%). In places like Sub-Saharan Africa that lack necessary infrastructure and personnel such as pathologists²⁹, excessive referral to colposcopy is problematic. Although WHO recommendations for S&T will hopefully overcome this bottleneck and increase the number of women living in LMICs who get screened, many countries may not adopt current S&T strategies because of concerns of low specificity and overtreatment, resulting in increase that could otherwise be used to expand access to screening. Thus, methods to increase the accuracy of screening by reducing the numbers of women having colposcopy and biopsy or getting treated immediately in this context are highly desirable as they will likely increase the uptake of cervical cancer screening.

1 2 4 5 6 7	
7 8 9 10 11 12	
8 9 10 11 12 13 14 15 16 17 18	
19 20 21 22 23 24	
25 26 27 28 29 30	
31 32 33 34 35 36 37	
 37 38 39 40 41 42 43 	
44 45 46 47 48 49	
50 51 52 53 54 55	
55 56 57 58 59 60	

137	In order to improve the specificity of screening tests, secondary tests (biomarkers) are used
138	following a screen-positive result, with women who test positive for the triage undergoing
139	further management (e.g., colposcopy or immediate treatment) and those who test negative
140	typically being deferred to further evaluation in 6-18 months to allow hrHPV infections to clear.
141	There are several very promising biomarkers that might be used to improve the specificity and
142	positive predictive value (PPV) of the screening tests. ²⁴ Given that HIV-infected women are
143	more likely to test hrHPV positive ³⁰⁻³⁴ , it is important to validate a triage strategy of using a
144	secondary biomarker that sensitively and specifically rules-in women with cervical precancer
145	and cancer among the HIV-positive, hrHPV-positive women.
146	We are therefore conducting a cervical cancer screening study of >5,000 Rwandan women, aged
147	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA
148	and VIA) and different triage tests and biomarkers for screen-positive women (E6/E7
149	oncoprotein detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-
150	positive women undergo a rigorous colposcopic evaluation with multiple biopsies taken and the
151	biopsies will undergo pathology review, to minimize the misclassification of endpoints. The
152	primary objective of the study is to determine and compare clinical performance (Sensitivity
153	(Se), Specificity (Sp), PPV, and NPV) and cost-effectiveness for identifying HIV[+] women with
154	CIN3+ and CIN2+ of different cervical cancer screening and management algorithms.
155	Methods and Analysis
156	Study design and population: We are recruiting those women receiving care in health centers

157 (HC) and various hospitals operated by the Ministry of Health or Rwanda Military Hospital

during 2016-18 (Table 1). Sites were selected in collaboration with Rwanda Biomedical Center

Page 9 of 27

1	
2	
3	
4	
5	
6	
7	
, 8	
9	
10	
11	
12	
13	
13 14 15	
15	
16 17	
17	
18	
19	
20	
20	
י∠ בר	
22 23	
23	
24	
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
55 54	
54 55	
56	
57	
58	
59	
60	

	159	(RBC), which calculated estimated numbers of potentially eligible women using data from the
	160	HIV database (OpenMRS-Open Medical Records System; http://openmrs.org/). Sites were also
	161	selected from all provinces to ensure geographic representation. The total is the number of 30-54
0 1	162	year old women receiving care at all selected sites and our study population is an estimate of the
2 3 4	163	women who will actually be eligible according to all inclusion and exclusion criteria.
5 6	164	Inclusion criteria include 1) living and receiving HIV care in Rwanda, 2) aged 30-54 years, 3)
7 8 9	165	confirmed HIV+, 4) no prior cervical cancer screening, 5) no history of ICC, and 6) willing, able
0 1	166	and competent to provide written, informed consent. Exclusion criteria, in addition to not
2 3	167	meeting the inclusion criteria, include 1) pregnant, 2) signs of abnormal, non-menstrual bleeding
4 5	168	suggestive of ICC, 3) without a cervix due to hysterectomy, and 4) not sufficiently healthy to
6 7 8	169	participate in a research study based on the judgment of the clinicians. Excluded women will be
9 0 1	170	advised to seek routine cervical cancer screening through government programs.
2 3 4	171	Prior to initiation of enrollment at a specific clinic, the local HIV-care provider team identifies
4 5 6	172	potentially eligible women at their routine clinic visits and offer them enrollment. Women
7 8	173	indicating interest in the study are then registered by our research nurses using the eligibility
9 0	174	criteria checklist. We enroll women at one site until all the eligible women at that site are
1 2 2	175	screened for cervical cancer. Our team of at least two research nurses schedules 12 to 15 women
3 4 5	176	three to four days a week by calling them and confirming appointments over the telephone. We
6 7	177	have two teams in the field meaning that we run two cervical cancer screening clinics
8 9 0	178	simultaneously.
1 2 3	179	Enrollment Visit: The study participant flow is summarized in Figure 1. During their
4 5	180	appointment for screening, women are educated on cervical cancer risk factors, mainly HPV

181 infection, and why they are more at risk to develop cervical cancer than HIV-uninfected women. 182 They are also allowed to ask questions before they commit to participating in the study. Women 183 are asked to provide informed, written consent to participate in the study using a printed out 184 consent form. Those who provide consent complete a short nurse-administered questionnaire on 185 cervical cancer risk factors and sociodemographic characteristics using a data capture screen in 186 Microsoft Access.

Enrolled women then undergo a pelvic exam, with VIA and a single cervical exfoliated ("Pap")
specimen collected and placed into 20 ml PreservCyt (Hologic, Bedford, MA, USA) which is
then sent to the lab at RMH for hrHPV testing. Finally, a portable colposcope
(http://www.mobileodt.com/; MobleODT, Tel Aviv, Israel) is used for digital cervicography
(~VIA with magnification) and the image is captured and saved for quality control, research, and
to develop a digital library.

Colposcopy Visit: Screen-positive women are called using a telephone as soon as the hrHPV result is available and provided colposcopy. All women receiving colposcopy will have two additional specimens collected, one into PreservCyt for the evaluation of other molecular biomarkers (genotype-specific hrHPV viral methylation and load, and p16/Ki-67 immunocytochemistry CINtec® PLUS Cytology Kit [Roche, Tucson, AZ, USA]) and a second using a dry swab for HPV16, 18, 31, 33, 35, 45, 52, and 58 E6/E7 oncoprotein detection by the next generation lateral flow hrHPV oncoprotein test from Arbor Vita Corporation (Fremont, CA, USA) included in this study as a triage for screen-positive women to identify those women who are at higher risk of having CIN3+. The residual PreservCyt specimens from both the screening and colposcopy visits will be stored at -20°C, creating a biobank in Rwanda for future retrospective evaluations of promising new biomarkers and tests.

Page 11 of 27

1 2		
3 4	204	After specimen collection, a colposcopic evaluation of the cervix is done and a modified version
5 6	205	of the 4-quadrant microbiopsy procedure is performed. ³⁵ Compared to the standard biopsy, the
7 8	206	microbiopsy protocol improves disease ascertainment and reduces biases by selecting on the
9 10 11 12 13 14 15	207	most obvious acetowhite lesions while removing less tissue (~13 mm ² for 4 microbiopsies vs.
	208	~28mm ² for 1 standard biopsy). Modifications to the standard 4-quadrant microbiopsy procedure
	209	are: 1) endocervical curettage is taken only for those women whose squamocolumnar junction is
16 17	210	not entirely visible and the lesion extends into the endocervical canal; and 2) standard biopsies of
18 19 20	211	very large lesions can be taken to increase the likelihood that the most severe area is biopsied.
21 22	212	Buth a larger Dianging and managered in a single acception as that a single slide has a spatian from all
23 24	212	Pathology: Biopsies are processed in a single cassette so that a single slide has a section from all
24 25 26	213	biopsies taken. Biopsies read by a local pathologist at RMH and Dr. Hebert or another
27 28	214	pathologist at Montefiore Medical Center, Bronx, NY, USA. Women receiving a diagnosis of
29 30 31 32	215	CIN2+ by the local pathologist or CIN3+ diagnosis by consensus review will be referred for
	216	treatment. A slide with biopsies also will undergo p16 immunohistochemistry (IHC) using the
33 34 35 36	217	CINtec® Histology Kit (Roche).
30 37 38	218	Endpoints: The primary scientific endpoint of the study will be histologically confirmed,
39 40	219	consensus CIN3+. The secondary, clinical endpoint will be histologically confirmed \geq +
41 42 42	220	diagnosed by the Rwandan pathologist. Additional endpoints using pathology review and p16
43 44 45	221	IHC will be used but not for evaluating the performance of p16 immunocytochemistry due to the
46 47 48	222	possibility of p16-related autocorrelation.
49 50 51	223	Treatment: Women diagnosed with CIN2+ will be referred for treatment. Those precancerous
52 53	224	lesions will be treated by ablation if they meet WHO criteria for cryotherapy. ³⁶ Those who do
54 55 56	225	not meet those criteria will undergo an excision procedure (e.g., loop electrosurgical excision
50 57 58		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

_				 		

1 2									
3 4	226	procedure [LEEP] or cold-knife cone [CKC]) or, in the case of an ICC diagnosis, referred for							
5 6	227	cancer management. Screen-positive women with <cin2 advised="" be="" in<="" re-screening="" seek="" td="" to="" will=""></cin2>							
7 8 9	228	a year through the existing healthcare system.							
10 11 12 13	229	Data sources: Data are collected from the following sources:							
14 15	230	1. A nurse-administered questionnaire on sociodemographic characteristics and cervical							
16 17	231	cancer risk factors including age at first sexual intercourse, number of sexual partners,							
18 19 20	232	smoking, contraception, parity and socioeconomic status.							
21 22 23 24	233	2. Pelvic exam, VIA, Mobile ODT and colposcopy data capture forms							
25 26	234	3. Medical record data on HIV status (e.g.,							
27 28 29	235	(http://www.who.int/hiv/pub/guidelines/HIVstaging150307.pdf), CD4 count, viral load,							
30 31	236	antiretroviral therapy (ART) regimen(s)), care, and dates.							
32 33 34 35	237	Laboratory Testing: The following laboratory tests will be performed:							
36 37 38	238	Xpert HPV Testing—cervical Pap specimens in PreservCyt will be sent to the RMH laboratory							
39 40	239	in Kigali, Rwanda for hrHPV DNA testing using the Xpert HPV test (Cepheid, Sunnyvale, CA,							
41 42	240	USA). ³⁷⁻⁴² The Xpert HPV Assay is a new, qualitative, real-time PCR assay for the detection of							
43 44 45	241	hrHPV DNA. The Xpert HPV Assay includes simultaneous detection of 14 hrHPV types,							
45 46 47	242	hydroxymethylbilane synthase (HMBS), and an internal Probe Check Control (PCC). The 14							
48 49	243	targeted hrHPV types are detected in 5 fluorescent channels: 1) HPV16, 2) HPV18 and hrHPV							
50 51 52	244	45 (HPV18/45), 3) HPV31, 33, 35, 52, and 58, 4) HPV51 and HPV59, and 5) HPV39, 56, 66,							
52 53 54 55 56	245	and 68. HMBS (fluorescent channel 6) verifies specimen adequacy.							
57 58 59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml							

Page 13 of 27

1		
2 3 4	246	Specimens are mixed and a 1-mL pre-aliquot is removed using a disposable pipette and placed in
5 6	247	the testing cartridge per the manufacturer's instructions. Unsatisfactory results due to insufficient
7 8 9	248	cellular content are re-run. If the second test is also unsatisfactory, the final result will be
9 10 11 12	249	unsatisfactory but women are referred to colposcopy for safety.
13 14	250	HPV Viral Methylation—To identify single hrHPV type infections, we will select single-channel
15 16 17	251	positives from the Xpert HPV assay. For those that are hrHPV positive for a channel other than
17 18 19	252	HPV16, which is detected singly, we will test them to identify the single type infections using a
20 21	253	standard protocol for PCR amplification using MY09/11 L1 consensus primers and hrHPV
22 23	254	genotype detection using dot-hybridization for 39 individual type-specific probes and a mixture
24 25 26	255	of probes for 10 other uncommon hrHPV types as previously described. ^{43;44} To isolate the DNA,
27 28	256	ThinPrep specimens (1.5 mL) will be pelleted, re-suspended in STM, digested with Proteinase K,
29 30	257	precipitated overnight in ammonium acetate ethanol at -20°C, washed, and suspended and stored
31 32 33	258	in TE buffer.
34 35 36	259	The isolated DNA will then undergo bisulfite conversion. ⁴⁵ Following bisulfite conversion and
37 38	260	DNA purification and de-sulphonation, bisulfite-treated DNA will be used as template for Next-
39 40 41	261	Gen Sequencing (NGS) (HiSeq2000, Illumina, San Diego, CA) using barcoded-type specific
41 42 43	262	primers. Sequences for pads and barcodes are not found in the targeted genomic region. Use of
44 45	263	padding and barcodes enables reads to be identified by amplicon (forward or reverse) or by
46 47 48	264	sample during downstream bioinformatics analysis. ⁴⁶
49 50 51	265	All PCR products for NGS are pooled (by assay) and a single DNA band containing multiple
52 53	266	amplicons from different samples (with unique barcodes) is isolated from a gel for NGS. ⁴⁶
54 55 56 57 58	267	Briefly, equal concentrations of each barcoded PCR product (based on PCR band intensity) are

pooled and isolated. Upon confirmation of correct product size, all purified DNA pools are combined and submitted for library preparation and paired-end 100 base pair Illumina HiSeq2000 sequencing at the Einstein Genomics Core Facility. Methylation status will be determined in the lab of Dr. Robert Burk at Albert Einstein College of Medicine (USA). Prior to determination of methylation status, de-multiplexing based on the unique barcodes is performed using in-house generated scripts to obtain paired-end NGS reads of each sample. Reads are aligned with hrHPV reference genome sequences by bowtie v0.12.9.47 Methylation status of each CpG site is then determined by bismark v0.7.7⁴⁸ using the default quality score parameter set to Q30, and the formula of the methylation ratio of the number of C read by the number of C+T read. E6/E7 Oncoprotein Testing—Dry swab specimens collected at the time of colposcopy will be tested for individual E6/E7 oncoproteins as previously described^{49;50}, according to the manufacturer's instructions, at the RMH laboratory in Kigali, Rwanda. The only deviation from the previous methods is that 3 lateral flow strips will be used to detect 8 hrHPV types in this study vs. 1 lateral flow strip used to detect 3 types in the study in China. *Analyses*: We will evaluate combinations of the above mentioned screening strategies and tests to estimate the clinical performance (e.g., Se, Sp, PPV, and NPV) for the detection of consensus CIN3+ and community CIN2+. A log binomial model implemented with generalized estimating equations will be used to take into account correlation between different tests from the same subject. Note that these models are developed for the estimation and comparison of performance for two tests, but the model can be extended to allow more than two tests by including more indicator variables for test type.

Page 15 of 27

BMJ Open

2	
3 4	2
5 6	2
7 8	2
9 10	
11 12	2
13 14	2
9 10 11 12 13 14 15 16 17 18 19	2
17 18	2
20	2
21 22 23	2
24 25	7
26 27	-
28 29	Ĵ
30 31 32	3
33	3
34 35	2
36 37	3
38 39	3
40 41	3
42 43	3
44 45	3
46 47 48	3
40 49 50	3
50 51 52	3
53 54	3
55 56	
57 58	
59	
60	

290 Some analyses of biomarkers, such as viral methylation are restricted to hrHPV-positives. 291 Comparisons of hrHPV viral methylation to other triage biomarkers will be restricted to the 292 subset that gets tested for viral methylation as described. 293 Sample size calculations: We based our sample size on the ability to detect modest but minimally important differences in Se of 15%. We conservatively assume that the population prevalence of 294 CIN3+ is 2% based on our past study in Rwanda.^{33;51} We propose to enroll and have complete 295 follow-up of at least 5,000 HIV[+] women. A sample size of 5,000 HIV[+] women with 296 297 completed follow-up of the screen positives will yield 100 cases of CIN3+, which has at least 298 80% power (α =5%) to crudely detect a 15% difference in Se between two screening strategies for a range of 10%-25% discordance. With this sample size of 5,000 women, 4,900 will not have 299 CIN3+; we will also have at least 90% power (α =5%) to detect a difference in Sp of 3% for 300 discordance up to 40%. Finally, we will have 80% power (α =5%) to crudely detect an 8%, 10%, 301 or 11% difference in PPV if the reference PPV is 10%, 20%, or 30%, respectively.⁵² 302 303 Cost Effectiveness: We will conduct assessments of the costs and cost-effectiveness of the different combinations of screening and triage tests, i.e., algorithms, as well as those of the entire 304 305 community-based screening "system." Costs measurement will be conducted using a micro-306 costing (ingredients) approach in which resource use throughout each step in the screening process will be tracked and unit costs for each of the resources will be applied to generate an 307 308 average screening cost per woman to be compared against what the estimated costs are for a possible program based on hrHPV screening and VIA triage or VIA screening. For estimating 309 310 costs of the screening system and scale-up of screening to 100,000 women in a month, analyses

- 311 will distinguish financial costs, which reflect actual expenditures of the program, from economic
 - For peer review only http://bmjopen.bmj.com/site/about/guidelines.xhtml

costs, including the value of donated and shared resources to more fully assess opportunity costs.
Projections on budget impact and economic cost implications over time will be made under
varying assumptions of screening uptake, follow-up compliance, and scenarios of changing
disease burden.

BMJ Open

Clinical outcomes will include true positive, true negative, false negative, and false positive test results, number of colposcopies, incident cancer, and cancer death. Cost-effectiveness will be measured as cost/CIN2+ detected, cost/CIN3+ detected, cost/invasive cancer prevented, cost/cancer death prevented, cost/life-year saved, and cost/quality-adjusted life year (QALY) saved; in addition, we will calculate harm/benefit ratios, using varying definitions of harms (colposcopies, false positive results) to benefits (cancers prevented, deaths prevented, life years and QALYs saved). Costs and effectiveness will be discounted at a 3% annual rate, with the rate varied from 0-5% in sensitivity analysis. For assessment of value-of-information (VOI), we will use net monetary benefits (NMB), defined as a function of the willingness-to-pay threshold (WTP) for different costs and outcomes as: *NMB*=(*WTP* * *Effectiveness*) – *Costs*.

326 Ethics and Dissemination

Ethics: This study protocol was reviewed and approved by the Rwanda National Ethics
Committee (RNEC) as well as the Institutional Review Board for human subjects research at
Albert Einstein College of Medicine.

Confidentiality measures and protection against potential risks: The risks for those participating
331 in our study include:

1		
2 3 4	332	• Collection of Pap specimens/cervical swabs involves a modest risk of bleeding
5 6	333	which is typically very limited when it occurs. Testing positive for any test may
7 8 9	334	cause psychological distress (anxiety).
10 11	335	• Colposcopy and excisional treatments induce vaginal bleeding and may incur pain,
12 13 14	336	infection, and short-term psychological distress (anxiety). A diagnosis of CIN2 or
14 15 16	337	more severe may cause psychological distress (anxiety). A diagnosis of invasive
17 18	338	cervical cancer may cause severe psychological distress.
19 20 21	339	• Questions in the questionnaire, regarding sexual behavior and other matters of a
22 23	340	personal nature, may cause anxiety and embarrassment. Participants are advised
24 25 26	341	that they are free not to answer specific questions.
27 28	342	• There is also the risk of psycho-social stress which could occur if there was
29 30 31	343	inadvertent disclosure of confidential medical or other personal information.
32 33	344	Protection against the risk of inadvertent disclosure of confidential information is addressed by
34 35 36	345	the standard procedures at the Rwandan study site, including: (i) storing completed paper copies
37 38	346	of questionnaires and other hard copy information (described above), identified by study number
39 40 41	347	only, in a filing system separate from the name-address file of participants in the study; and (ii)
41 42 43	348	only the designated local personnel have access to cross-reference the files; (iii) all paper files,
44 45	349	including consent forms, will be maintained in locked cabinets in locked rooms, with access
46 47 48	350	restricted to specific research personnel.
48 49 50 51 52	351	In addition, we will include the following security measures to protect the data:
53 54 55 56	352	• Controlled access to project data;
57 58		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2		
3 4	353	• A tracking system for data forms and activities;
5 6	354	• Date and time of stamping of all data records with electronic signatures;
7 8	355	• Audit trails to track all changes made to data records; and
9 10 11	356	• Data kept on password-protected computers and in locked rooms.
12 13	257	Determined Development of the second statistic first the second statistic sector in se
14 15	357	Potential Patient Benefits: There are potential direct benefits to study participants. HIV[+]
16 17	358	women participating in the study are at very high risk of cervical cancer. They will be rigorously
18 19	359	screened and evaluated, more effectively than the standard of care anywhere in the world. As a
20 21	360	result, women with precancer who are at imminent risk of invasive cervical cancer will be
22 23 24	361	diagnosed sooner and treated more effectively than women receiving routine care and thereby
24 25 26	362	more likely averting the development of cervical cancer. Women with cervical cancer will be
27 28	363	diagnosed earlier thereby reducing the morbidity and the risk of mortality caused by cervical
29 30	364	cancer. Conversely, any pain, bleeding, or stress that might occur related to colposcopy or
31 32 33	365	cervical swab are typically modest and well tolerated.
34 35	366	There are also substantial potential societal benefits related to the study due to its implications to
36 37	367	improving cervical cancer screening practices and guidelines in HIV[+] women – changes in
38 39	368	practice which might also benefit the study participants themselves, if and when these changes
40 41 42	369	are enacted. There is a great need to identify more effective and practical methods for cervical
43		
44 45	370	cancer screening for HIV[+] women living in Africa, who are living longer than ever and are
46 47	371	therefore at potentially greater risk of cervical cancer.
48 49 50	372	Dissemination: We plan to publish a series of scientific reports in peer-reviewed scientific
50 51 52	373	journals. As building research capacity in Rwanda is a major goal of this research project, all
53 54	374	investigators of the research team will be asked and supported to lead at least one analysis and
55 56 57	375	one manuscript preparation, based on interests and expertise.
58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2		
2 3 4	376	In addition, an external advisory panel (EAP) composed of leaders from the Rwanda Ministry of
5 6	377	Health, University of Rwanda, and Rwanda medical community has been formed. The
7 8	378	responsibilities of the EAP include providing advice on the conduct of the project and
9 10 11	379	interpretation for and dissemination of the study results to Rwandan stakeholders. The latter is
12 13	380	important for the adoption of evidence-based best practices for cervical cancer screening as
14 15	381	warranted.
$\begin{array}{c} 16\\ 17\\ 18\\ 19\\ 20\\ 21\\ 22\\ 23\\ 24\\ 25\\ 26\\ 27\\ 28\\ 29\\ 30\\ 31\\ 32\\ 33\\ 35\\ 36\\ 37\\ 38\\ 39\\ 41\\ 42\\ 43\\ 44\\ 50\\ 51\\ 52\\ 54\\ 55\\ 56\end{array}$	382	important for the adoption of evidence-based best practices for cervical cancer screening as warranted.
57 58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

383 Contributor Statement:

KA, PEC, JCD, AA, and JDS conceived the original concept of the study and the interventions. GM, KA, JDS, and PEC drafted the protocol. PEC performed the sample size calculations, and PEC and GM will lead analysis of the results. GM, JDS, and Athanase Munyaneza are supporting patient recruitment. GM, TR, and Athanase Munyaneza are providing clinical care for patients. Anthere Murangwa and LM oversee laboratory testing, and TZ and TH are responsible for pathology. PM and LM oversee and administer the study activities at the clinical site in Rwanda. All authors (GM, JCD, TR, JDS, Athanase Munyaneza, Anthere Murangwa, TZ, TH, PM, AA, LM, KA, and PEC) contributed to the scientific design of the study and the protocol development, are involved in the implementation of the project, and have read and approved the final manuscript.

1		
2 3 4	394	Reference List
5 6 7	395	
8 9 10 11 12	396 397 398 399	 (1) Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C et al. GLOBOCAN 2012 v1.0, Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. 2013. Lyon, France, International Agency for Research on Cancer. 4-22-2014. Ref Type: Online Source
13 14 15 16	400 401 402	(2) Lonnberg S, Hansen BT, Haldorsen T, Campbell S, Schee K, Nygard M. Cervical cancer prevented by screening: Long-term incidence trends by morphology in Norway. <i>Int J Cancer</i> 2015;10.
17 18 19	403 404	(3) UNAIDS. UNAIDS Data 2017. 2017. Ref Type: Report
20 21 22 23	405 406	(4) Cervix Cancer Screening. [10]. 2005. IARC Press. IARC Handbooks of Cancer Prevention. Ref Type: Serial (Book,Monograph)
24 25 26	407 408	(5) Kitchener HC, Castle PE, Cox JT. Chapter 7: Achievements and limitations of cervical cytology screening. <i>Vaccine</i> 2006; 24 Suppl 3:S63-70.:S63-S70.
27 28 29	409 410	(6) Schiffman M, Castle PE, Jeronimo J, Rodriguez AC, Wacholder S. Human papillomavirus and cervical cancer. <i>Lancet</i> 2007; 370(9590):890-907.
30 31 32 33	411 412 413	(7) Schiffman M, Clifford G, Buonaguro FM. Classification of weakly carcinogenic human papillomavirus types: addressing the limits of epidemiology at the borderline. <i>Infect Agent Cancer</i> 2009; 4:8.
34 35 36 37 38	414 415 416	(8) Forman D, de MC, Lacey CJ, Soerjomataram I, Lortet-Tieulent J, Bruni L et al. Global burden of human papillomavirus and related diseases. <i>Vaccine</i> 2012; 30 Suppl 5:F12-23. doi: 10.1016/j.vaccine.2012.07.055.:F12-F23.
39 40 41 42	417 418 419	(9) de SS, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Lloveras B et al. Human papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study. <i>Lancet Oncol</i> 2010; 11(11):1048-1056.
43 44 45 46	420 421 422	(10) Bray F, Carstensen B, Moller H, Zappa M, Zakelj MP, Lawrence G et al. Incidence trends of adenocarcinoma of the cervix in 13 European countries. <i>Cancer Epidemiol Biomarkers Prev</i> 2005; 14(9):2191-2199.
47 48 49	423 424	(11) Adegoke O, Kulasingam S, Virnig B. Cervical cancer trends in the United States: a 35-year population-based analysis. <i>J Womens Health (Larchmt)</i> 2012; 21(10):1031-1037.
50 51 52 53 54	425 426 427	(12) Cuzick J, Clavel C, Petry KU, Meijer CJ, Hoyer H, Ratnam S et al. Overview of the European and North American studies on HPV testing in primary cervical cancer screening. <i>Int J Cancer</i> 2006; 119(5):1095-1101.
55 56 57 58		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

3 4 5 6	428 429 430	(13)	Naucler P, Ryd W, Tornberg S, Strand A, Wadell G, Elfgren K et al. Human papillomavirus and Papanicolaou tests to screen for cervical cancer. <i>N Engl J Med</i> 2007; 357(16):1589-1597.
7 8 9 10	431 432 433	(14)	Ronco G, Giorgi-Rossi P, Carozzi F, Confortini M, Palma PD, Del MA et al. Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised controlled trial. <i>Lancet Oncol</i> 2010.
11 12 13 14 15 16	434 435 436 437	(15)	Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkmans NW, Heideman DA et al. Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the POBASCAM randomised controlled trial. <i>Lancet</i> <i>Oncol</i> 2012; 13(1):78-88.
17 18 19 20	438 439 440	(16)	Ronco G, Dillner J, Elfstrom KM, Tunesi S, Snijders PJ, Arbyn M et al. Efficacy of HPV-based screening for prevention of invasive cervical cancer: follow-up of four European randomised controlled trials. <i>Lancet</i> 2013.
21 22 23	441 442	(17)	Sankaranarayanan R, Nene BM, Shastri SS, Jayant K, Muwonge R, Budukh AM et al. HPV screening for cervical cancer in rural India. <i>N Engl J Med</i> 2009; 360(14):1385-1394.
24 25 26 27	443 444 445	(18)	Dillner J, Rebolj M, Birembaut P, Petry KU, Szarewski A, Munk C et al. Long term predictive values of cytology and human papillomavirus testing in cervical cancer screening: joint European cohort study. <i>BMJ</i> 2008; 337:a1754. doi: 10.1136/bmj.a1754.:a1754.
28 29	446 447	(19)	New guidelines on screening and treatment for cervical cancer. 2013. South Africa, World Health Organization.
30 31 32	448	Ref Ty	pe: Pamphlet
33 34 35 36	449 450 451	(20)	Shastri SS, Mittra I, Mishra GA, Gupta S, Dikshit R, Singh S et al. Effect of VIA screening by primary health workers: randomized controlled study in Mumbai, India. <i>J Natl Cancer Inst</i> 2014; 106(3):dju009.
37 38 39 40	452 453 454	(21)	Jeronimo J, Castle PE, Temin S, Shastri SS. Secondary Prevention of Cervical Cancer: American Society of Clinical Oncology Resource-Stratified Clinical Practice Guideline Summary. <i>J Oncol Pract</i> 2016;JOP2016017889.
41 42 43	455 456	(22)	Castle PE, Jeronimo J, Temin S, Shastri SS. Screening to Prevent Invasive Cervical Cancer: ASCO Resource-Stratified Clinical Practice Guideline. <i>J Clin Oncol</i> 2017; 35(11):1250-1252.
44 45 46 47	457 458 459	(23)	Keller MJ, Burk RD, Xie X, Anastos K, Massad LS, Minkoff H et al. Risk of cervical precancer and cancer among HIV-infected women with normal cervical cytology and no evidence of oncogenic HPV infection. <i>JAMA</i> 2012; 308(4):362-369.
48 49 50 51	460 461 462	(24)	Castle PE, Fetterman B, Poitras N, Lorey T, Kinney W. Safety against cervical precancer and cancer following negative human papillomavirus and Papanicolaou test results in human immunodeficiency virus-infected women. <i>Arch Intern Med</i> 2012; 172(13):1041-1043.
52 53 54 55 56 57	463 464 465	(25)	Chung MH, McKenzie KP, De VH, Richardson BA, Rana F, Pamnani R et al. Comparing pap smear, via, and hpv cervical cancer screening methods among hiv-positive women by immune status and antiretroviral therapy. <i>AIDS</i> 2013.
58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

2			
3	466	(26)	Firnhaber C, Mayisela N, Mao L, Williams S, Swarts A, Faesen M et al. Validation of cervical
4	467		cancer screening methods in HIV positive women from Johannesburg South Africa. PLoS
5 6	468		<i>One</i> 2013; 8(1):e53494.
7			
8	469	(27)	Dartell MA, Rasch V, Iftner T, Kahesa C, Mwaiselage JD, Junge J et al. Performance of visual
9	470		inspection with acetic acid and human papillomavirus testing for detection of high-grade
10	471		cervical lesions in HIV positive and HIV negative Tanzanian women. <i>Int J Cancer</i> 2014;10.
11	472	(28)	Mabeya H, Khozaim K, Liu T, Orango O, Chumba D, Pisharodi L et al. Comparison of
12 13	473	(20)	conventional cervical cytology versus visual inspection with acetic acid among human
13 14	474		immunodeficiency virus-infected women in Western Kenya. J Low Genit Tract Dis 2012;
15	475		16(2):92-97.
16			
17	476	(29)	Adesina A, Chumba D, Nelson AM, Orem J, Roberts DJ, Wabinga H et al. Improvement of
18	477		pathology in sub-Saharan Africa. <i>Lancet Oncol</i> 2013; 14(4):e152-e157.
19			
20	478	(30)	Dartell M, Rasch V, Kahesa C, Mwaiselage J, Ngoma T, Junge J et al. Human papillomavirus
21 22	479		prevalence and type distribution in 3603 HIV-positive and HIV-negative women in the
23	480		general population of Tanzania: the PROTECT study. <i>Sex Transm Dis</i> 2012; 39(3):201-208.
24	481	(31)	D'Souza G, Burk RD, Zhong Y, Minkoff H, Massad LS, Xue X et al. Cervicovaginal human
25	482	(31)	papillomavirus (HPV)-infection before and after hysterectomy: evidence of different tissue
26	483		tropism for oncogenic and nononcogenic HPV types in a cohort of HIV-positive and HIV-
27	484		negative women. Int J Cancer 2012; 131(6):1472-1478.
28 29	101		
29 30	485	(32)	Ng'andwe C, Lowe JJ, Richards PJ, Hause L, Wood C, Angeletti PC. The distribution of
31	486		sexually-transmitted Human Papillomaviruses in HIV positive and negative patients in
32	487		Zambia, Africa. BMC Infect Dis 2007; 7:77.:77.
33			
34	488	(33)	Singh DK, Anastos K, Hoover DR, Burk RD, Shi Q, Ngendahayo L et al. Human papillomavirus
35	489		infection and cervical cytology in HIV-infected and HIV-uninfected Rwandan women. J Infect
36 37	490		Dis 2009; 199(12):1851-1861.
37 38	491	(24)	Margia DI Vardag E. Damiga C. Allan P. Kay D. Daga PC at al. The impact of human
39	491	(34)	Marais DJ, Vardas E, Ramjee G, Allan B, Kay P, Rose RC et al. The impact of human immunodeficiency virus type 1 status on human papillomavirus (HPV) prevalence and HPV
40	493		antibodies in serum and cervical secretions. J Infect Dis 2000; 182(4):1239-1242.
41	175		
42	494	(35)	Pretorius RG, Zhang WH, Belinson JL, Huang MN, Wu LY, Zhang X et al. Colposcopically
43	495	()	directed biopsy, random cervical biopsy, and endocervical curettage in the diagnosis of
44 45	496		cervical intraepithelial neoplasia II or worse. Am J Obstet Gynecol 2004; 191(2):430-434.
43 46			
47	497	(36)	World Health Organization. WHO guidelines for treatment of cervical intraepithelial
48	498	neopla	sia 2-3 and adenocarcinoma in situ. World Health Organization [2014 Available from:
49	499		URL: <u>http://apps.who.int/iris/bitstream/10665/104174/1/9789241506779_eng.pdf</u>
50	500	(27)	Castle DE Smith KM Davis TE Schmeler KM Earris DC Savers All at al Delichility of the
51 52	500 501	(37)	Castle PE, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Reliability of the Xpert HPV assay to detect high-risk human papillomavirus DNA in a colposcopy referral
52 53	501		population. Am J Clin Pathol 2015; 143(1):126-133.
55 54	502		population. <i>All j olil i activi 2013</i> , 143(1).120 ⁻ 133.
55			
56			
57			
58			
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml
00			- Ferrer and a state an

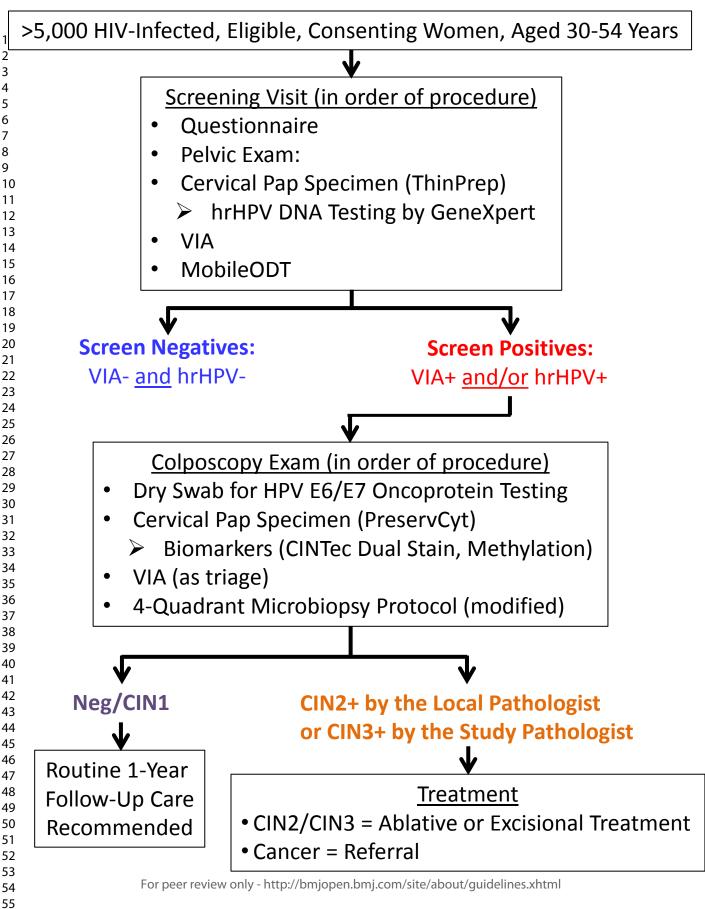
(38) Cuzick J, Cuschieri K, Denton K, Hopkins M, Thorat MA, Wright C et al. Performance of the Xpert HPV assay in women attending for cervical screening. *Papillomavirus Research* 2015; 1:32-37. (39) Einstein MH, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Clinical Evaluation of the Cartridge-Based GeneXpert Human Papillomavirus Assay in Women Referred for Colposcopy. J Clin Microbiol 2014. (40) Toliman P, Badman SG, Gabuzzi J, Silim S, Forereme L, Kumbia A et al. Field Evaluation of Xpert HPV Point-of-Care Test for Detection of Human Papillomavirus Infection by Use of Self-Collected Vaginal and Clinician-Collected Cervical Specimens. *J Clin Microbiol* 2016; 54(7):1734-1737. (41) Cuschieri K, Geraets D, Cuzick J, Cadman L, Moore C, Vanden Broeck D et al. Performance of a Cartridge-Based Assay for Detection of Clinically Significant Human Papillomavirus (HPV) Infection: Lessons from VALGENT (Validation of HPV Genotyping Tests). *J Clin Microbiol* 2016; 54(9):2337-2342. (42) Kunckler M, Schumacher F, Kenfack B, Catarino R, Viviano M, Tincho E et al. Cervical cancer screening in a low-resource setting: a pilot study on an HPV-based screen-and-treat approach. Cancer Med 2017;10. (43) Guimaraes MD, Grinsztein B, Melo VH, Rocha GM, Campos LN, Pilotto JH et al. Anal HPV prevalence and associated factors among HIV-seropositive men under antiretroviral treatment in Brazil. J Acquir Immune Defic Syndr 2011; 57 Suppl 3:S217-S224. (44) Castle PE, Schiffman M, Gravitt PE, Kendall H, Fishman S, Dong H et al. Comparisons of HPV DNA detection by MY09/11 PCR methods. J Med Virol 2002; 68(3):417-423. (45) Hayatsu H, Shiraishi M, Negishi K. Bisulfite modification for analysis of DNA methylation. Curr Protoc Nucleic Acid Chem 2008; Chapter 6:Unit. (46) Smith BC, McAndrew T, Chen Z, Harari A, Barris DM, Viswanathan S et al. The cervical microbiome over 7 years and a comparison of methodologies for its characterization. PLoS One 2012; 7(7):e40425. (47) Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. *Genome Biol* 2009; 10(3):R25. (48) Krueger F, Andrews SR. Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. *Bioinformatics* 2011; 27(11):1571-1572. (49) Qiao YL, Jeronimo J, Zhao FH, Schweizer J, Chen W, Valdez M et al. Lower cost strategies for triage of human papillomavirus DNA-positive women. Int J Cancer 2013. (50) Zhao FH, Jeronimo J, Qiao YL, Schweizer J, Chen W, Valdez M et al. An evaluation of novel, lower-cost molecular screening tests for human papillomavirus in rural China. Cancer Prev Res (Phila) 2013; 6(9):938-948.

1 2			
3 4 5 6	539 540 541	(51)	Anastos K, Hoover DR, Burk RD, Cajigas A, Shi Q, Singh DK et al. Risk factors for cervical precancer and cancer in HIV-infected, HPV-positive Rwandan women. <i>PLoS One</i> 2010; %20;5(10):e13525.
7 8 9 10 11 12	542 543 544 545	(52)	Leisenring W, Alonzo T, Pepe MS. Comparisons of predictive values of binary medical diagnostic tests for paired designs. <i>Biometrics</i> 2000; 56(2):345-351.
	546		
51 52 53 54 55 56			
57 58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

Table 1: Recruitment sites and estimated population to be recruited from each site.

Province	Site	Type of site	Potential participants po
			site (approximate)
Kigali	Cor-unum HC	Public Health Center	1,405
Kigali	Kimironko HC	Public Health Center	1,227
Kigali	Rwampara HC	Public Referral Hospital	1,098
Kigali	Kicukiro HC	Public Health Center	1,054
Kigali	Kacyiru HC	Public Health Center	905
Kigali	Gikondo HC	Public Health Center	925
Kigali	Rwanda Military Hospital	Public Referral Hospital	300
Kigali	WEACTx for Hope	Private HIV Clinic	500
Kigali	Busanza HC	Public Health Center	100
Kigali	Nyarugunga HC	Public Health Center	100
West	Gisenyi DH	Public District Hospital	810
Total			8,424

```
Figure_12Study diagram
```



BMJ Open

A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-Infected Women Living in Rwanda

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-020432.R1
Article Type:	Protocol
Date Submitted by the Author:	19-Mar-2018
Complete List of Authors:	Murenzi, Gad; Rwanda Military Hospital Dusingize, Jean; Regional Alliance for Sustainable Development , Research and Scientific Capacity Building Rurangwa, Theogene; Rwanda Military Hospital Sinayobye, Jean d'Amour; Regional Alliance for Sustainable Development, ; Women's Equity in Access to Care & Treatment (WE-ACTx), Munyaneza, Athanase; Rwanda Military Hospital Murangwa, Anthere; Rwanda Military Hospital Zawadi, Thierry; Rwanda Military Hospital Hebert, Tiffany; Yeshiva University Albert Einstein College of Medicine Mugenzi, Pacifique; Rwanda Military Hospital Adedimeji, Adebola; Yeshiva University Albert Einstein College of Medicine Mutesa, Leon; Rwanda Military Hospital; University of Rwanda College of Medicine and Health Sciences Anastos, Kathryn; Albert Einstein College of Medicine Medical Center, Bronx, Castle, Philip; Yeshiva University Albert Einstein College of Medicine,
Primary Subject Heading :	Global health
Secondary Subject Heading:	Obstetrics and gynaecology, Infectious diseases, HIV/AIDS, Epidemiology, Diagnostics
Keywords:	human papillomavirus (HPV), cervical cancer, HIV & AIDS < INFECTIOUS DISEASES, GYNAECOLOGY, cervical intraepithelial neoplasia

SCHOLARONE[™] Manuscripts Page 1 of 34

1
2
2
4
5
6
7
8
9
10
10
11
12
13
14
15
16
17
18
10
19
20
2 3 4 5 6 7 8 9 10 112 13 14 15 16 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 4 5 6 7 8 9 10 112 3 3 4 5 6 7 8 9 10 112 3 3 4 5 6 7 8 9 10 112 3 3 4 5 6 7 8 9 10 112 3 3 4 5 6 7 8 9 10 112 3 3 3 4 5 5 6 7 7 8 9 10 3 1 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3
22
23
24
25
26
20
27
28
29
30
31
32
33
24
24
35
36
37
38
39
40
41
41
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60

1	A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-
2	Infected Women Living in Rwanda
3	Gad Murenzi, MD* ¹ , Jean-Claude Dusingize, MD, MS ¹ , Theogene Rurangwa, MD, MMed ¹ ,
4	Jean d'Amour Sinayobye, MD, MS ¹ , Athanase Munyaneza, RN ¹ , Anthere Murangwa, MS ¹ ,
5	Thierry Zawadi, MD ¹ , Tiffany Hebert, MD ² , Pacifique Mugenzi, MD ¹ , MMed, Adebola
6	Adedimeji, PhD, MPH ² , Leon Mutesa, MD, PhD ^{1,3} , Kathryn Anastos, MD ² , Philip E. Castle,
7	PhD, MPH ^{2,4}
8	¹ Rwanda Military Hospital, Kigali, Rwanda; ² Albert Einstein College of Medicine, Bronx, NY,
9	USA; ³ University of Rwanda, Kigali, Rwanda; ⁴ Global Coalition Against Cervical Cancer,
10	Arlington, VA, USA.
11	*Correspondence: <u>gadcollins@gmail.com</u> , +250788589085
12	Disclosures: This research study has received HPV tests for reduced or no cost from Cepheid,
13	Arbor Vita Corporation, and Roche.
14	Funding: This study was funded by NCI/NIH Grant 5U54CA19016304 and by a grant from the
15	Prevent Cancer Foundation.
16	

17 Abstract

Introduction. The optimal method(s) for screening human immunodeficiency virus-infected
 women, especially for those living in sub-Saharan Africa, for cervical precancer and early cancer
 has yet to be established.

Methods and analysis. A convenience sample of >5,000 Rwandan women, aged 30-54 years, living with HIV infection will be enrolled into a cross-sectional study of cervical-cancer screening strategies. Eligible and consenting women will be enrolled into the study, complete a short risk-factor questionnaire, and screened for high-risk human papillomavirus (hrHPV) using the Xpert HPV assay (Cepheid, Sunnyvale, CA, USA), unaided visual inspection after acetic acid (VIA), and aided VIA using the EVA system (Mobile ODT, Tel Aviv, Israel). Women positive for hrHPV or by VIA will undergo colposcopy, which will include the collection of two cervical specimens prior to undergoing a 4-quadrant microbiopsy protocol. The colposcopy-collected specimens will be tested by dual immunocytochemical staining for p16^{INK4a} and Ki-67 (CINtec® PLUS Cytology, Ventana, Tucson, AZ, USA) and for E6 or E7 for 8 hrHPV genotypes (HPV16, 18, 31, 33, 35, 45, 52, and 58) using the next-generation AV Avantage hrHPV E6/E7 test (Arbor Vita Corporation, Freemont, CA, USA). Women with local pathology diagnosis of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) or pathology-review diagnosis of CIN grade 3 or more severe (CIN3+) will receive treatment. Clinical performance and cost-effectiveness (e.g., sensitivity, specificity, and predictive values)

- ²⁷ 36 of different screening strategies and algorithms will be evaluated.
- 30 37 *Ethics and dissemination.* The protocol has been approved by local and institutional review
- boards for human subjects research. Results will be disseminated to the scientific community
 - 39 through peer-reviewed publication and to the Rwandan stakeholders through an external
- 33 40 advisory panel.

1		
2 3	42	Strengths and Weaknesses
4 5	43	• We will enroll a very large sample size of HIV-infected women living in Rwanda who
6 7	44	otherwise would probably not get cervical-cancer screening.
8	45	• We will employ rigorous disease ascertainment protocols to minimize misclassification.
9	46	• Some tests, such as the Xpert HPV and the AV Avantage HPV E6/E7 assays, will be
10	47	done on site in Rwanda using technologies that could feasibly be deployed there.
11 12	48	
12	49	comparator due to financial and logistical constraints.
14	50	
15		
16		
17 18		
19		
20		
21 22		
22		
24		
25		
26 27		
28		
29		
30 31		
32		A weakness of the study is that it will not be feasible to include cervical cytology as a comparator due to financial and logistical constraints.
33		
34		
35 36		
37		
38		
39 40		
40		
42		
43		
44 45		
46		
47		
48 49		
49 50		
51		
52		
53 54		
55		
56		
57		
58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

51 Introduction

Invasive cervical cancer (ICC) remains a significant cause of morbidity and mortality globally. Approximately 530,000 cases of and 270,000 deaths due to ICC occur annually, making it the 4th most common malignancy and cause of cancer-related death in women globally.¹ In many high-income countries (HIC), cervical cancer rates have declined by 50% or more² due to the introduction of effective, high-coverage Pap testing (cervical cytology)-based screening programs that include timely follow-up of screen positives, treatment of women with precursor lesions, and management of cancers. Now, almost 90% of ICC and ICC-related deaths occur in low- and middle income countries (LMICs) due to a lack of resources and healthcare infrastructure needed to provide preventive services. ICC and ICC-related mortality rates are particularly high in Sub-Saharan Africa, which also has the highest rates of HIV infection in the world. Now, over 12 million HIV-infected (HIV[+]) women in Sub-Saharan Africa are living longer because of anti-retroviral therapy, thus increasing their likelihood of dying from ICC.³ However, many of these women are already exposed to human papillomavirus (HPV), the viral cause of cervical cancer, and will not benefit

from or be targeted for prophylactic HPV vaccination. Thus, cervical-cancer screening is neededfor the foreseeable future.

However, setting up effective cytology for cervical-cancer screening is expensive and requires a complex clinical and lab infrastructure that generally does not exist in LMICs.^{4;5} Moreover, it is now well understood that cytology has only a low- to moderate one-time sensitivity for precursor lesions and therefore must be done repeatedly over many years to reduce cancer risk. Alternative

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

BMJ Open

strategies to address the cervical cancer burden in LMICs, especially in SSA, must be developedand validated.

Persistent cervical infections by high-risk HPV (hrHPV) types cause virtually all ICC and its immediate precursor lesions, e.g. cervical intraepithelial neoplasia grade 3 (CIN3) and adenocarcinoma *in situ* (AIS) everywhere in the world.^{6;7} hrHPV causes most anal and vaginal cancer and a significant proportion of vulvar, penile, and oropharyngeal cancers.⁸ HPV16 is the most important causal type, responsible for ~60% of ICC.⁹ HPV18 is the next most important, responsible for 10-15% of ICC, including 30-40% of adenocarcinoma of the cervix⁹, which is on the rise in Western Countries.^{10;11} Together, HPV16 and HPV18 account for ~70% of ICC, and the same 15 hrHPV types account for ~99% of ICC everywhere in the world.⁹

There is now overwhelming evidence to suggest that testing for hrHPV is more sensitive, albeit less specific, than high-quality cytology for identifying women with cervical precancer.¹²⁻¹⁶ Onetime hrHPV testing can reduce the risk of ICC incidence by approximately 40% in 6.5 years compared to cytology screening¹⁶, and ICC mortality by approximately 40% (approximately 50% overall) in 8 years compared to cytology.¹⁷ Importantly, a negative hrHPV test provides superior reassurance against CIN3+ ¹⁸ and against ICC^{16;17}, permitting safe extension of screening intervals.

The World Health Organization released cervical-cancer screening and treatment guidelines in 2013, recommending two evidence-based approaches to cervical-cancer screening¹⁹: (I) Use either hrHPV testing or visual inspection after acetic acid (VIA), which involves the inspection of the cervix with a speculum in place and following the application of dilute acetic acid to help identify potential CIN by its characteristic white coloring in the presence of acetic acid

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

(acetowhite), as alternative initial screening tests instead of Pap, and (II) immediately treat those who screen positive using the screening test, rather than require diagnostic verification through colposcopy and biopsy. This approach is commonly called screen-and-treat (S&T), and is increasingly thought to be more amenable to LMIC settings.

However, hrHPV testing is also a much more effective screen than VIA¹⁷, which on a large-scale appears to only down-stage cancer rather than prevent it.²⁰ Thus, the recent American Society for Clinical Oncology (ASC) resource-stratified guidelines for secondary cervical cancer prevention^{21;22} emphasize that hrHPV testing is the preferred choice for screening, with VIA only being used until hrHPV testing becomes available, and that HIV-infected women, because of their higher risk, should be screened twice as frequently as the general (HIV-uninfected) population.

Recent data in HIV[+] women living in the U.S. suggest that hrHPV testing may have clinical utility similar to that in HIV-negative (HIV[-]) women. Several observational studies have shown that an extended screening interval is safe in HIV[+] women who test hrHPV and Pap negative as it is for HIV[-] women.^{23;24} In a study of women enrolled in Women's Interagency Health Study (WIHS) in 2002, HIV[+] and HIV[-] women who tested hrHPV and Pap negative were at a similarly low risk of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) histology over a 5-year follow-up.²³ In addition, no cases of histologically confirmed CIN2+ were diagnosed in the follow-up of hrHPV- and Pap-negative HIV[+] women aged 30-64 years who underwent routine three-year hrHPV and cytology cotesting at Kaiser Permanente Northern California.²⁴ Thus, both studies found very high negative predictive values (NPV) >99% in HIV[+] women who test hrHPV negative. However, how hrHPV testing can

Page 7 of 34

BMJ Open

1		
2 3 4	116	best be use
5 6 7	117	remains to
8 9 10	118	Recent stud
10 11 12	119	and/or Pap
13 14	120	follows: 1)
15 16	121	cytology w
17 18 19	122	was equally
20 21	123	conclusion
22 23	124	women liv
24 25		
25 26 27	125	Regardless
28 29	126	treated imr
30 31	127	(positive p
32 33	128	Saharan At
34 35 36	129	referral to
37 38	130	overcome 1
39 40	131	many coun
41 42	132	overtreatm
43 44 45	133	wasting va
46 47	134	screening.
48 49	135	women hav
50 51 52	136	desirable a
53		
54		
55 56		
56 57		
58		
59		
60		

best be used to screen HIV[+] women living in Sub-Saharan Africa to prevent cervical cancer remains to be determined.

Recent studies²⁵⁻²⁸ in HIV[+] women living in Sub-Saharan Africa have compared hrHPV, VIA, and/or Pap for the detection of cervical precancer/cancer. The results can be summarized as follows: 1) hrHPV detection was more sensitive but less specific than VIA; 2) surprisingly, cytology was equally or more sensitive but less specific than VIA; and 3) surprisingly, cytology was equally or more sensitive but less specific (vs. the converse) than hrHPV testing. Results and conclusions are varied, leaving unanswered the question of what screening strategy in HIV[+] women living in Sub-Saharan Africa has the greatest effectiveness and cost effectiveness.

Regardless of the screening method, most screen-positive women who go to colposcopy or are treated immediately without diagnostic verification do not have cervical precancer and cancer (positive predictive value [PPV] for screening tests are typically 10%-20%). In places like Sub-Saharan Africa that lack necessary infrastructure and personnel such as pathologists²⁹, excessive referral to colposcopy is problematic. Although WHO recommendations for S&T will hopefully overcome this bottleneck and increase the number of women living in LMICs who get screened, many countries may not adopt current S&T strategies because of concerns of low specificity and overtreatment, resulting in increase that could otherwise be used to expand access to screening. Thus, methods to increase the accuracy of screening by reducing the numbers of women having colposcopy and biopsy or getting treated immediately in this context are highly desirable as they will likely increase the uptake of cervical-cancer screening.

2
3
4
4 5
-
6
7
8
9
10
11
12
13
13 14
15
16
16 17 18
18
10
19 20
21
22
23
24
25
26
26 27
28
29
30
31
32
33
33 34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
40 49
50
51
52
53
54
55
56
57
58
59
60
00

1

137	In order to improve the specificity of screening tests, secondary tests (biomarkers) are used
138	following a screen-positive result, with women who test positive for the triage undergoing
139	further management (e.g., colposcopy or immediate treatment) and those who test negative
140	typically being deferred to further evaluation in 6-18 months to allow hrHPV infections to clear.
141	There are several very promising biomarkers that might be used to improve the specificity and
142	positive predictive value (PPV) of the screening tests. ²⁴ Given that HIV-infected women are
143	more likely to test hrHPV positive ³⁰⁻³⁴ , it is important to validate a triage strategy of using a
144	secondary biomarker that sensitively and specifically rules-in women with cervical precancer
145	and cancer among the HIV-positive, hrHPV-positive women.
146	We are therefore conducting a cervical-cancer screening study of >5,000 Rwandan women, aged
146 147	We are therefore conducting a cervical-cancer screening study of >5,000 Rwandan women, aged 30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA
147	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA
147 148	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage
147 148 149	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage tests and biomarkers for screen-positive women (E6/E7 oncoprotein detection, p16INK4a
147 148 149 150	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage tests and biomarkers for screen-positive women (E6/E7 oncoprotein detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-positive women undergo a
147 148 149 150 151	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage tests and biomarkers for screen-positive women (E6/E7 oncoprotein detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-positive women undergo a rigorous colposcopic evaluation with multiple biopsies taken and the biopsies will undergo
147 148 149 150 151 152	30-54 years, living with HIV infection. We will evaluate different screening tests (hrHPV DNA and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage tests and biomarkers for screen-positive women (E6/E7 oncoprotein detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-positive women undergo a rigorous colposcopic evaluation with multiple biopsies taken and the biopsies will undergo pathology review, to minimize the misclassification of endpoints. The primary objective of the

- 155 different cervical-cancer screening and management algorithms.
- 156 Methods and Analysis

Study design and population: We are recruiting those women receiving care for confirmed HIV
infection at health centers (HC) and various hospitals operated by the Ministry of Health or

Page 9 of 34

1

BMJ Open

2
3
1
4
5
6
7
8
9
10
11
12
13
14
15
16
10
17
18
19
20
21
22
23
23 24
24
25
26
27
28
29
30
31
32
33
34
35
36
50
37
38
39
40
41
42
43
44
45
46
47
48
49
5 0
51
52
53
54
55
56
57
58
59

60

159	Rwanda Military Hospital during 2016-18 (Table 1). Sites were selected in collaboration with
160	Rwanda Biomedical Center (RBC), which calculate estimated numbers of potentially eligible
161	women using data from the HIV database (OpenMRS-Open Medical Records System;
162	http://openmrs.org/). Sites were also selected from all provinces to ensure geographic
163	representation. The total of 8,424 is the estimated number of women eligible for the study
164	according to the inclusion/exclusion criteria (see below) at the beginning for the study. From
165	each site, a convenience sample of women were recruited to participate.
166	Inclusion criteria include 1) living and receiving HIV care in Rwanda, 2) aged 30-54 years, 3)
167	confirmed HIV+ based on medical records, 4) no prior cervical-cancer screening, 5) no history of
168	ICC, and 6) willing, able and competent to provide written, informed consent. We extended age
169	range beyond that of age range (30-49 years) recommended by the WHO for cervical-cancer
170	screening ¹⁹ because there is limited evidence for the optimal upper age for cervical-cancer
171	screening of HIV-infected women. Exclusion criteria, in addition to not meeting the inclusion
172	criteria, include 1) pregnant, 2) signs of abnormal, non-menstrual bleeding suggestive of ICC, 3)
173	without a cervix due to hysterectomy, and 4) not sufficiently healthy to participate in a research
174	study based on the judgment of the clinicians. Excluded women are advised to seek routine
175	cervical-cancer screening through government programs.
176	Prior to initiation of enrollment at a specific clinic, the local HIV-care provider team identifies
177	potentially eligible women at their routine clinic visits and offers them enrollment. Women

indicating interest in the study are then registered by our research nurses using the eligibility criteria checklist. Women at one site are enrolled until all the eligible and willing women at that site are screened for cervical cancer. The study team of at least two research nurses schedules 12 to 15 women three to four days a week by calling them and confirming appointments over the

telephone. Two teams of nurses are in the field, meaning that two cervical-cancer screeningclinics can be run simultaneously.

Enrollment Visit: Enrollment visits, including pelvic exams with VIA and specimen collection, are done entirely by a team of two study nurses. During their enrollment visit, women are educated on cervical cancer risk factors, mainly HPV infection, and why they are more at risk to develop cervical cancer than HIV-uninfected women. They are also allowed to ask questions before they commit to participating in the study. Women are asked to provide informed, written consent to participate in the study using a printed out consent form. Those who provide consent complete a short nurse-administered questionnaire (Appendix I) on cervical cancer risk factors and sociodemographic characteristics using a data capture screen in Microsoft Access. The questionnaire collects information on basic sociodemographics, factors associated with acquiring HPV (e.g., marital status and recent and lifetime number of sexual partners), factors associated with increased risk of progression of hrHPV infection to precancer and cancer (e.g., smoking and other tobacco use, parity, and oral and other contraceptive use), and other infections common in Rwanda such as malaria³⁵ and tuberculosis³⁶ that have been previously reported to be associated with precancer among hrHPV-infected women. The questionnaire was not pretested.

Enrolled women then undergo a pelvic exam, with VIA and a single cervical exfoliated ("Pap")
specimen collected and placed into 20 ml PreservCyt (Hologic, Bedford, MA, USA) which is
then sent to the lab at RMH for hrHPV testing. Finally, a portable colposcope
(<u>http://www.mobileodt.com/</u>; MobleODT, Tel Aviv, Israel) is used for digital cervicography
(~VIA with magnification) and the image is captured and saved for quality control, research, and

to develop a digital library.

Page 11 of 34

BMJ Open

Colposcopy Visit: Screen-positive women (women who test hrHPV and/or VIA positive) are called using a telephone as soon as the hrHPV result is available and invited to return for colposcopy within one month. All women receiving colposcopy will have two additional specimens collected, one into PreservCyt for the evaluation of other molecular biomarkers (genotype-specific hrHPV viral methylation and load, and p16/Ki-67 immunocytochemistry CINtec® PLUS Cytology Kit [Roche, Tucson, AZ, USA]) and a second using a dry swab for HPV16, 18, 31, 33, 35, 45, 52, and 58 E6/E7 oncoprotein detection by the next generation lateral flow hrHPV oncoprotein test from Arbor Vita Corporation (Fremont, CA, USA) included in this study as a triage for screen-positive women to identify those women who are at higher risk of having CIN3+. The residual PreservCyt specimens from both the screening and colposcopy visits will be stored at -20°C, creating a biobank in Rwanda for future retrospective evaluations of promising new biomarkers and tests. After specimen collection, a colposcopic evaluation of the cervix is done and a modified version

of the 4-quadrant microbiopsy procedure is performed.³⁷ Compared to the standard biopsy, the microbiopsy protocol improves disease ascertainment and reduces biases by selecting on the most obvious acetowhite lesions while removing less tissue (\sim 13 mm² for 4 microbiopsies vs. \sim 28mm² for 1 standard biopsy). Modifications to the standard 4-quadrant microbiopsy procedure are: 1) endocervical curettage is taken only for those women whose squamocolumnar junction is not entirely visible and the lesion extends into the endocervical canal; and 2) standard biopsies of very large lesions can be taken to increase the likelihood that the most severe area is biopsied.

Pathology: Biopsies are processed in a single cassette so that a single slide has a section from all
biopsies taken. Biopsies read by a local pathologist at RMH and Dr. Hebert or another

226 pathologist at Montefiore Medical Center, Bronx, NY, USA. Women receiving a diagnosis of

1

59

2		
3 4	227	CIN2+ by the Rwandan pathologist (T.Z.) or, as a safety precaution, CIN3+ diagnosis by
5 6	228	Montefiore pathologist (T.H.) will receive treatment ³⁸ : 1) CIN2, CIN3, or AIS will be referred to
7 8 9	229	study doctors to undergo an excision procedure (e.g., loop electrosurgical excision procedure
10 11	230	[LEEP] or cold-knife cone [CKC]) and 2) ICC will be referred to RMH Hospital for care.
12 13 14	231	Women with <cin2 a="" advised="" be="" in="" re-screening="" seek="" td="" to="" will="" year.<=""></cin2>
15 16 17	232	A slide with biopsies also will undergo p16 immunohistochemistry (IHC) using the CINtec®
18 19 20	233	Histology Kit (Roche) for research purposes only.
21 22	234	Endpoints: The primary scientific endpoint of the study will be histologically confirmed,
23 24 25	235	consensus CIN2+ i.e., both pathologists diagnosed CIN2+ (without adjudication). The
25 26 27	236	secondary, clinical endpoint will be histologically confirmed CIN2+ diagnosed by the Rwandan
28 29	237	pathologist. Additional endpoints using pathology review and p16 IHC will be used but not for
30 31 32	238	evaluating the performance of p16 immunocytochemistry due to the possibility of p16-related
33 34 35	239	autocorrelation.
36 37	240	Treatment: Women diagnosed with CIN2+ will be referred for treatment. Those precancerous
38 39	241	lesions will be treated by ablation if they meet WHO criteria for cryotherapy. ³⁹ Those who do
40 41 42	242	not meet those criteria will undergo an excision procedure (e.g., loop electrosurgical excision
43 44	243	procedure [LEEP] or cold-knife cone [CKC]) or, in the case of an ICC diagnosis, referred for
45 46	244	cancer management. Screen-positive women with <cin2 advised="" be="" in<="" re-screening="" seek="" td="" to="" will=""></cin2>
47 48 49	245	a year through the existing healthcare system.
50 51 52 53 54 55	246	Data sources: Data are collected from the following sources:
55 56 57		
58 50		

BMJ Open

1 2		
2 3 4	247	1. A nurse-administered questionnaire on sociodemographic characteristics and cervical
5 6	248	cancer risk factors including age at first sexual intercourse, number of sexual partners,
7 8 9	249	smoking, contraception, parity and socioeconomic status.
9 10 11 12 13	250	2. Pelvic exam, VIA, Mobile ODT and colposcopy data capture forms
14 15	251	3. Medical record data on HIV status (e.g.,
16 17	252	(http://www.who.int/hiv/pub/guidelines/HIVstaging150307.pdf), CD4 count, viral load,
18 19 20	253	antiretroviral therapy (ART) regimen(s)), care, and dates.
21		
22 23	254	Laboratory Testing: The following laboratory tests will be performed:
24 25		
25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40	255	Xpert HPV Testing—cervical Pap specimens in PreservCyt will be sent to the RMH laboratory
	256	in Kigali, Rwanda for hrHPV DNA testing using the Xpert HPV test (Cepheid, Sunnyvale, CA,
	257	USA). ⁴⁰⁻⁴⁵ The Xpert HPV Assay is a new, qualitative, real-time PCR assay for the detection of
	258	hrHPV DNA. The Xpert HPV Assay includes simultaneous detection of 14 hrHPV types,
	259	hydroxymethylbilane synthase (HMBS), and an internal Probe Check Control (PCC). The 14
	260	targeted hrHPV types are detected in 5 fluorescent channels: 1) HPV16, 2) HPV18 and hrHPV
	261	45 (HPV18/45), 3) HPV31, 33, 35, 52, and 58, 4) HPV51 and HPV59, and 5) HPV39, 56, 66,
41 42 43	262	and 68. HMBS (fluorescent channel 6) verifies specimen adequacy.
44 45 46	263	Specimens are mixed and a 1-mL pre-aliquot is removed using a disposable pipette and placed in
47 48	264	the testing cartridge per the manufacturer's instructions. Unsatisfactory results due to insufficient
49 50	265	cellular content are re-run. If the second test is also unsatisfactory, the final result will be
51 52 53 54 55 56	266	unsatisfactory but women are referred to colposcopy for safety.
57 58 59		

HPV Viral Methylation—To identify single hrHPV type infections, we will select single-channel positives from the Xpert HPV assay. For those that are hrHPV positive for a channel other than HPV16, which is detected singly, we will test them to identify the single type infections using a standard protocol for PCR amplification using MY09/11 L1 consensus primers and hrHPV genotype detection using dot-hybridization for 39 individual type-specific probes and a mixture of probes for 10 other uncommon hrHPV types as previously described.^{46;47} To isolate the DNA, ThinPrep specimens (1.5 mL) will be pelleted, re-suspended in STM, digested with Proteinase K, precipitated overnight in ammonium acetate ethanol at -20°C, washed, and suspended and stored in TE buffer. The isolated DNA will then undergo bisulfite conversion.⁴⁸ Following bisulfite conversion and DNA purification and de-sulphonation, bisulfite-treated DNA will be used as template for Next-Gen Sequencing (NGS) (HiSeq2000, Illumina, San Diego, CA) using barcoded-type specific primers. Sequences for pads and barcodes are not found in the targeted genomic region. Use of padding and barcodes enables reads to be identified by amplicon (forward or reverse) or by sample during downstream bioinformatics analysis.⁴⁹ All PCR products for NGS are pooled (by assay) and a single DNA band containing multiple amplicons from different samples (with unique barcodes) is isolated from a gel for NGS.⁴⁹ Briefly, equal concentrations of each barcoded PCR product (based on PCR band intensity) are pooled and isolated. Upon confirmation of correct product size, all purified DNA pools are

combined and submitted for library preparation and paired-end 100 base pair Illumina

287 HiSeq2000 sequencing at the Einstein Genomics Core Facility.

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

Page 15 of 34

BMJ Open

י כ		
2 3		
4		
5		
6		
7		
7 8		
0		
9 10	`	
1(
11		
	2	
13		
14	+	
15) -	
16	2	
17		
18		
19)	
20)	
20 21 22 22 22 22 22 22 22 22 22 22 22 22		
22	2	
23	3	
24	1	
25	5	
26	5	
27	7	
28	3	
29)	
3()	
3	I	
	2	
	3	
34	1	
35	5	
36	5	
37 38	7	
38	3	
39)	
40		
41		
42	2	
	3	
44	1	
45	5	
46		
47	7	
48	3	
49		
50		
51		
	2	
	3	
52		
55		
56		
57		
57 58		
59 59		
55 6(

288 Methylation status will be determined in the lab of Dr. Robert Burk at Albert Einstein College of 289 Medicine (USA). Prior to determination of methylation status, de-multiplexing based on the 290 unique barcodes is performed using in-house generated scripts to obtain paired-end NGS reads of each sample. Reads are aligned with hrHPV reference genome sequences by bowtie v0.12.9.⁵⁰ 291 Methylation status of each CpG site is then determined by bismark $v0.7.7^{51}$ using the default 292 293 quality score parameter set to Q30, and the formula of the methylation ratio of the number of C 294 read by the number of C+T read. E6/E7 Oncoprotein Testing—Dry swab specimens collected at the time of colposcopy will be 295 tested for individual E6/E7 oncoproteins as previously described^{52;53}, according to the 296 297 manufacturer's instructions, at the RMH laboratory in Kigali, Rwanda. The E6/E7 oncoprotein test uses three lateral flow strips to detect 8 hrHPV types whereas the E6 oncoprotein test used a 298 299 single lateral flow strip to detect 3 hrHPV types. Analyses: We will evaluate combinations of the above mentioned screening strategies and tests 300 301 to estimate the clinical performance (e.g., Se, Sp, PPV, and NPV) for the detection of consensus 302 CIN3+ and community CIN2+. A log binomial model implemented with generalized estimating 303 equations will be used to take into account correlation between different tests from the same 304 subject. Note that these models are developed for the estimation and comparison of performance 305 for two tests, but the model can be extended to allow more than two tests by including more indicator variables for test type. 306 307 Some analyses of biomarkers, such as viral methylation are restricted to hrHPV-positives. 308 Comparisons of hrHPV viral methylation to other triage biomarkers will be restricted to the 309 subset that gets tested for viral methylation as described.

Sample size calculations: We based our sample size on the ability to detect modest but minimally important differences in Se of 15%. We conservatively assume that the population prevalence of CIN3+ is 2% based on our past study in Rwanda.^{33,35} We propose to enroll and have complete follow-up of at least 5,000 HIV[+] women. A sample size of 5,000 HIV[+] women with completed follow-up of the screen positives will yield 100 cases of CIN3+, which has at least 80% power (α =5%) to crudely detect a 15% difference in Se between two screening strategies for a range of 10%-25% discordance. With this sample size of 5,000 women, 4,900 will not have CIN3+; we will also have at least 90% power (α =5%) to detect a difference in Sp of 3% for discordance up to 40%. Finally, we will have 80% power (α =5%) to crudely detect an 8%, 10%, or 11% difference in PPV if the reference PPV is 10%, 20%, or 30%, respectively.⁵⁴ Cost Effectiveness: We will conduct assessments of the costs and cost-effectiveness of the different combinations of screening and triage tests, i.e., algorithms, as well as those of the entire community-based screening "system." Costs measurement will be conducted using a micro-costing (ingredients) approach in which resource use throughout each step in the screening process will be tracked and unit costs for each of the resources will be applied to generate an average screening cost per woman to be compared against what the estimated costs are for a possible program based on hrHPV screening and VIA triage or VIA screening. For estimating costs of the screening system and scale-up of screening to 100,000 women in a month, analyses will distinguish financial costs, which reflect actual expenditures of the program, from economic costs, including the value of donated and shared resources to more fully assess opportunity costs. Projections on budget impact and economic cost implications over time will be made under varying assumptions of screening uptake, follow-up compliance, and scenarios of changing disease burden.

Page 17 of 34

ge 17 of 34		BMJ Open			
	333	Clinical outcomes will include true positive, true negative, false negative, and false positive test			
	334	results, number of colposcopies, incident cancer, and cancer death. Cost-effectiveness will be			
	335	measured as cost/CIN2+ detected, cost/CIN3+ detected, cost/invasive cancer prevented,			
	336	cost/cancer death prevented, cost/life-year saved, and cost/quality-adjusted life year (QALY)			
	337	saved; in addition, we will calculate harm/benefit ratios, using varying definitions of harms			
	338	(colposcopies, false positive results) to benefits (cancers prevented, deaths prevented, life years			
	339	and QALYs saved). Costs and effectiveness will be discounted at a 3% annual rate, with the rate			
	340	varied from 0-5% in sensitivity analysis. For assessment of value-of-information (VOI), we will			
	341	use net monetary benefits (NMB), defined as a function of the willingness-to-pay threshold			
	342	(WTP) for different costs and outcomes as: <i>NMB=(WTP * Effectiveness) – Costs</i> .			
	343	Patient and Public Involvement			
	344	• How was the development of the research question and outcome measures informed by			
	345	patients' priorities, experience, and preferences? There was no patient engagement in the			
	346	development of the study.			
	347	• How did you involve patients in the design of this study? There was no patient			
	348	engagement in the design of the study.			
	349	• Were patients involved in the recruitment to and conduct of the study? There was no			
	350	patient involvement in the recruitment and conduct of the study.			
	351	• How will the results be disseminated to study participants? Participants will receive their			
	352	results directly since it is related to their care.			
		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml			

3 4	353	• For randomized controlled trials, was the burden of the intervention assessed by patient.		
5 6 7	354	themselves? Not applicable		
8 9 10	355	• Patient advisers should also be thanked in the contributorship		
11 12	356	statement/acknowledgements. Not applicable		
13 14 15	357	Ethics and Dissemination		
16 17				
17 18 19	358	Ethics: This study protocol was reviewed and approved by the Rwanda National Ethics		
20 21	359	Committee (RNEC) as well as the Institutional Review Board for human subjects research at		
22 23 24	360	Albert Einstein College of Medicine.		
25 26 27	361	Confidentiality measures and protection against potential risks: The risks for those participating		
28 29	362	in our study include:		
30 31 32 33 34	363	• Collection of Pap specimens/cervical swabs involves a modest risk of bleeding		
	364	which is typically very limited when it occurs. Testing positive for any test may		
 35 36 365 cause psychological distress (anxiety). 37 		cause psychological distress (anxiety).		
38 39	366	• Colposcopy and excisional treatments induce vaginal bleeding and may incur pain,		
40 41 42	367	7 infection, and short-term psychological distress (anxiety). A diagnosis of CIN		
43 44	368	more severe may cause psychological distress (anxiety). A diagnosis of invasive		
45 46 47	369 cervical cancer may cause severe psychological distress.			
 48 49 370 Questions in the questionnaire 		• Questions in the questionnaire, regarding sexual behavior and other matters of a		
50 51 52	371	personal nature, may cause anxiety and embarrassment. Participants are advised		
53 54 55 56 57 58	372	that they are free not to answer specific questions.		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml		

1 2				
3 4	373	• There is also the risk of psycho-social stress which could occur if there was		
5 6 7	374	inadvertent disclosure of confidential medical or other personal information.		
9 10 11 12 13 14	375	Protection against the risk of inadvertent disclosure of confidential information is addressed by		
	376	the standard procedures at the Rwandan study site, including: (i) storing completed paper copies		
	377	of questionnaires and other hard copy information (described above), identified by study number		
15 16	378	only, in a filing system separate from the name-address file of participants in the study; and (ii)		
17 18	379	only the designated local personnel have access to cross-reference the files; (iii) all paper files,		
19 20 21	380	including consent forms, will be maintained in locked cabinets in locked rooms, with access		
22 23	381	restricted to specific research personnel.		
24 25 26 27	382	In addition, we will include the following security measures to protect the data:		
28 29 30	383	Controlled access to project data;		
31 32 33 34 35 36 37 38 39 40 41 42 43	384	• A tracking system for data forms and activities;		
	385	• Date and time of stamping of all data records with electronic signatures;		
	386	• Audit trails to track all changes made to data records; and		
	387	• Data kept on password-protected computers and in locked rooms.		
	388	Potential Patient Benefits: There are potential direct benefits to study participants. HIV[+]		
44 45	389	women participating in the study are at very high risk of cervical cancer. They will be rigorously		
46 47 48 49 50 51 52 53 54	390	screened and evaluated, more effectively than the standard of care anywhere in the world. As a		
	391	result, women with precancer who are at imminent risk of invasive cervical cancer will be		
	392	diagnosed sooner and treated more effectively than women receiving routine care and thereby		
	393	more likely averting the development of cervical cancer. Women with cervical cancer will be		
55 56 57	394	diagnosed earlier thereby reducing the morbidity and the risk of mortality caused by cervical		
58 59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml		

cancer. Conversely, any pain, bleeding, or stress that might occur related to colposcopy orcervical swab are typically modest and well tolerated.

There are also substantial potential societal benefits related to the study due to its implications to improving cervical-cancer screening practices and guidelines in HIV[+] women – changes in practice which might also benefit the study participants themselves, if and when these changes are enacted. There is a great need to identify more effective and practical methods for cervicalcancer screening for HIV[+] women living in Africa, who are living longer than ever and are therefore at potentially greater risk of cervical cancer.

Dissemination: We plan to publish a series of scientific reports in peer-reviewed scientific 404 journals. As building research capacity in Rwanda is a major goal of this research project, all 405 investigators of the research team will be asked and supported to lead at least one analysis and 406 one manuscript preparation, based on interests and expertise.

407 In addition, an external advisory panel (EAP) composed of leaders from the Rwanda Ministry of
408 Health, University of Rwanda, and Rwanda medical community has been formed. The
409 responsibilities of the EAP include providing advice on the conduct of the project and
410 interpretation for and dissemination of the study results to Rwandan stakeholders. The latter is
411 important for the adoption of evidence-based best practices for cervical-cancer screening as
412 warranted.

413 Limitations

There are several limitations to the study that bear mentioning. First, cervical cytology was not
included in the study. There is limited cervical cytology services available locally and of
unknown quality and it is unlikely that cytology will be widely available in Rwanda, making its

estimated from this study.

inclusion as a comparator test of limited value. Moreover, there are significant costs and
logistical challenges in shipping PreservCyt specimens to and having cytology slides made and
read in the U.S. Second, we did not conduct biopsies in screen-negative women, which would
have allowed us to estimate absolute clinical performance. The burden of sending screennegative women to colposcopy was deemed too great and it was impractical to send a sufficient
numbers of screen-negative women to colposcopy to accurately estimate the false-negative
disease (CIN3+) fraction. Thus, only relative clinical performance of the screening tests can be

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

this study.

Contributor Statement:

KA, PEC, JCD, AA, and JDS conceived the original concept of the study and the interventions. GM, KA, JDS, and PEC drafted the protocol. PEC performed the sample size calculations, and PEC and GM will lead analysis of the results. GM, JDS, and Athanase Munyaneza are supporting patient recruitment. GM, TR, and Athanase Munyaneza are providing clinical care for patients. Anthere Murangwa and LM oversee laboratory testing, and TZ and TH are responsible for pathology. PM and LM oversee and administer the study activities at the clinical site in Rwanda. All authors (GM, JCD, TR, JDS, Athanase Munyaneza, Anthere Murangwa, TZ, TH, PM, AA, LM, KA, and PEC) contributed to the scientific design of the study and the protocol development, are involved in the implementation of the project, and have read and approved the Stopper texter only final manuscript.

1 2			
3 4	436	Reference List	
5 6	437		
7 8 9 10 11	438 439 440 441	(1) Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mather Cancer Incidence and Mortality Worldwide: IARC CancerBas France, International Agency for Research on Cancer. 4-22-2 Ref Type: Online Source	se No. 11 [Internet]. 2013. Lyon,
12 13 14 15 16	442 443 444	(2) Lonnberg S, Hansen BT, Haldorsen T, Campbell S, Schee K, N prevented by screening: Long-term incidence trends by mor 2015;10.	
17 18 19	445 446	(3) UNAIDS. UNAIDS Data 2017. 2017. Ref Type: Report	
20 21 22	447 448	(4) Cervix Cancer Screening. [10]. 2005. IARC Press. IARC Hand Ref Type: Serial (Book,Monograph)	books of Cancer Prevention.
23 24 25	449 450	(5) Kitchener HC, Castle PE, Cox JT. Chapter 7: Achievements an cytology screening. <i>Vaccine</i> 2006; 24 Suppl 3:S63-70.:S63-S	
26 27 28	451 452	(6) Schiffman M, Castle PE, Jeronimo J, Rodriguez AC, Wacholde cervical cancer. <i>Lancet</i> 2007; 370(9590):890-907.	r S. Human papillomavirus and
 453 453 453 454 454 455 455 455 456 (8) Forman D, de MC, Lacey CJ, Soerjomataram I, Lortet-Tieulent J, Bru of human papillomavirus and related diseases. <i>Vaccine</i> 2012; 30 St 10.1016/j.vaccine.2012.07.055.:F12-F23. 459 (9) de SS, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Llovera papillomavirus genotype attribution in invasive cervical cancer: a sectional worldwide study. <i>Lancet Oncol</i> 2010; 11(11):1048-1056. 460 461 462 (10) Bray F, Carstensen B, Moller H, Zappa M, Zakelj MP, Lawrence G et adenocarcinoma of the cervix in 13 European countries. <i>Cancer Ep</i> 2005; 14(9):2191-2199. 	454	papillomavirus types: addressing the limits of epidemiology	
	of human papillomavirus and related diseases. Vaccine 2012	-	
	ncer: a retrospective cross-		
	463	adenocarcinoma of the cervix in 13 European countries. Car	
46 47 48	465 466	(11) Adegoke O, Kulasingam S, Virnig B. Cervical cancer trends in population-based analysis. <i>J Womens Health (Larchmt)</i> 201	-
49 50 51 52	467 468 469	(12) Cuzick J, Clavel C, Petry KU, Meijer CJ, Hoyer H, Ratnam S et and North American studies on HPV testing in primary cerv <i>Cancer</i> 2006; 119(5):1095-1101.	-
53 54 55 56 57	470 471 472	(13) Naucler P, Ryd W, Tornberg S, Strand A, Wadell G, Elfgren K and Papanicolaou tests to screen for cervical cancer. <i>N Engl</i> 1597.	
58 59 60		For peer review only - http://bmjopen.bmj.com/site/abou	t/guidelines.xhtml

2			
3	473	(14)	Ronco G, Giorgi-Rossi P, Carozzi F, Confortini M, Palma PD, Del MA et al. Efficacy of human
4	474		papillomavirus testing for the detection of invasive cervical cancers and cervical
5	475		intraepithelial neoplasia: a randomised controlled trial. <i>Lancet Oncol</i> 2010.
6 7			
8	476	(15)	Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkmans NW, Heideman DA et al.
9	477		Human papillomavirus testing for the detection of high-grade cervical intraepithelial
10	478		neoplasia and cancer: final results of the POBASCAM randomised controlled trial. Lancet
11	479		Oncol 2012; 13(1):78-88.
12			
13	480	(16)	Ronco G, Dillner J, Elfstrom KM, Tunesi S, Snijders PJ, Arbyn M et al. Efficacy of HPV-based
14	481		screening for prevention of invasive cervical cancer: follow-up of four European
15	482		randomised controlled trials. <i>Lancet</i> 2013.
16			
17	483	(17)	Sankaranarayanan R, Nene BM, Shastri SS, Jayant K, Muwonge R, Budukh AM et al. HPV
18	484		screening for cervical cancer in rural India. <i>N Engl J Med</i> 2009; 360(14):1385-1394.
19			
20	485	(18)	Dillner J, Rebolj M, Birembaut P, Petry KU, Szarewski A, Munk C et al. Long term predictive
21	486		values of cytology and human papillomavirus testing in cervical cancer screening: joint
22 23	487		European cohort study. <i>BMJ</i> 2008; 337:a1754. doi: 10.1136/bmj.a1754.:a1754.
25 24			
25	488	(19)	New guidelines on screening and treatment for cervical cancer. 2013. South Africa, World
26	489	_	Health Organization.
27	490	Ref Ty	pe: Pamphlet
28		(2.2)	
29	491	(20)	Shastri SS, Mittra I, Mishra GA, Gupta S, Dikshit R, Singh S et al. Effect of VIA screening by
30	492		primary health workers: randomized controlled study in Mumbai, India. J Natl Cancer Inst
31	493		2014; 106(3):dju009.
32		(24)	
33	494	(21)	Jeronimo J, Castle PE, Temin S, Shastri SS. Secondary Prevention of Cervical Cancer:
34	495		American Society of Clinical Oncology Resource-Stratified Clinical Practice Guideline
35	496		Summary. J Oncol Pract 2016; JOP2016017889.
36 37	407	(22)	Castle DE Janonime I. Temin C. Chastri CC. Concening to Drevent Investive Corrigal Concerv
38	497	(22)	Castle PE, Jeronimo J, Temin S, Shastri SS. Screening to Prevent Invasive Cervical Cancer:
39	498		ASCO Resource-Stratified Clinical Practice Guideline. <i>J Clin Oncol</i> 2017; 35(11):1250-1252.
40	499	(23)	Keller MJ, Burk RD, Xie X, Anastos K, Massad LS, Minkoff H et al. Risk of cervical precancer
41	500	(23)	and cancer among HIV-infected women with normal cervical cytology and no evidence of
42	501		oncogenic HPV infection. JAMA 2012; 308(4):362-369.
43	301		oncogenic HPV infection. JAMA 2012; 506(4):502-509.
44	502	(24)	Castle PE, Fetterman B, Poitras N, Lorey T, Kinney W. Safety against cervical precancer and
45	502	(24)	cancer following negative human papillomavirus and Papanicolaou test results in human
46	504		immunodeficiency virus-infected women. Arch Intern Med 2012; 172(13):1041-1043.
47	304		$\frac{1}{10}$
48	505	(25)	Chung MH, McKenzie KP, De VH, Richardson BA, Rana F, Pamnani R et al. Comparing pap
49 50	506	(20)	smear, via, and hpv cervical cancer screening methods among hiv-positive women by
50 51	507		immune status and antiretroviral therapy. <i>AIDS</i> 2013.
51 52	207		
52	508	(26)	Firnhaber C, Mayisela N, Mao L, Williams S, Swarts A, Faesen M et al. Validation of cervical
54	509	(==)	cancer screening methods in HIV positive women from Johannesburg South Africa. <i>PLoS</i>
55	510		One 2013; 8(1):e53494.
56			
57			
58			
59			For poor roviou only http://hmiopon.hmi.com/rite/about/ruidalines.yhtml
60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
3 4 5 6	511 512 513	(27)	Dartell MA, Rasch V, Iftner T, Kahesa C, Mwaiselage JD, Junge J et al. Performance of visual inspection with acetic acid and human papillomavirus testing for detection of high-grade cervical lesions in HIV positive and HIV negative Tanzanian women. <i>Int J Cancer</i> 2014;10.
7 8 9 10 11	514 515 516 517	(28)	Mabeya H, Khozaim K, Liu T, Orango O, Chumba D, Pisharodi L et al. Comparison of conventional cervical cytology versus visual inspection with acetic acid among human immunodeficiency virus-infected women in Western Kenya. <i>J Low Genit Tract Dis</i> 2012; 16(2):92-97.
12 13 14	518 519	(29)	Adesina A, Chumba D, Nelson AM, Orem J, Roberts DJ, Wabinga H et al. Improvement of pathology in sub-Saharan Africa. <i>Lancet Oncol</i> 2013; 14(4):e152-e157.
15 16 17 18 19	520 521 522	(30)	Dartell M, Rasch V, Kahesa C, Mwaiselage J, Ngoma T, Junge J et al. Human papillomavirus prevalence and type distribution in 3603 HIV-positive and HIV-negative women in the general population of Tanzania: the PROTECT study. <i>Sex Transm Dis</i> 2012; 39(3):201-208.
20 21 22 23 24	523 524 525 526	(31)	D'Souza G, Burk RD, Zhong Y, Minkoff H, Massad LS, Xue X et al. Cervicovaginal human papillomavirus (HPV)-infection before and after hysterectomy: evidence of different tissue tropism for oncogenic and nononcogenic HPV types in a cohort of HIV-positive and HIV-negative women. <i>Int J Cancer</i> 2012; 131(6):1472-1478.
25 26 27 28	527 528 529	(32)	Ng'andwe C, Lowe JJ, Richards PJ, Hause L, Wood C, Angeletti PC. The distribution of sexually-transmitted Human Papillomaviruses in HIV positive and negative patients in Zambia, Africa. <i>BMC Infect Dis</i> 2007; 7:77.:77.
29 30 31 32	530 531 532	(33)	Singh DK, Anastos K, Hoover DR, Burk RD, Shi Q, Ngendahayo L et al. Human papillomavirus infection and cervical cytology in HIV-infected and HIV-uninfected Rwandan women. <i>J Infect Dis</i> 2009; 199(12):1851-1861.
33 34 35 36 37	533 534 535	(34)	Marais DJ, Vardas E, Ramjee G, Allan B, Kay P, Rose RC et al. The impact of human immunodeficiency virus type 1 status on human papillomavirus (HPV) prevalence and HPV antibodies in serum and cervical secretions. <i>J Infect Dis</i> 2000; 182(4):1239-1242.
38 39 40 41	536 537 538	(35)	Anastos K, Hoover DR, Burk RD, Cajigas A, Shi Q, Singh DK et al. Risk factors for cervical precancer and cancer in HIV-infected, HPV-positive Rwandan women. <i>PLoS One</i> 2010; %20;5(10):e13525.
42 43 44 45	539 540 541	(36)	Zhao FH, Varanasi AP, Cunningham CA, Graubard BI, Hu SY, Chen F et al. Tuberculosis and oncogenic HPV: potential co-infections in women at high-risk of cervical cancer in rural China. <i>Asian Pac J Cancer Prev</i> 2011; 12(6):1409-1415.
46 47 48 49	542 543 544	(37)	Pretorius RG, Zhang WH, Belinson JL, Huang MN, Wu LY, Zhang X et al. Colposcopically directed biopsy, random cervical biopsy, and endocervical curettage in the diagnosis of cervical intraepithelial neoplasia II or worse. <i>Am J Obstet Gynecol</i> 2004; 191(2):430-434.
50 51 52	545 546	(38)	Schiffman M, Adrianza ME. ASCUS-LSIL Triage Study. Design, methods and characteristics of trial participants. <i>Acta Cytol</i> 2000; 44(5):726-742.
53 54 55 56 57	547 548 549	(39) neopla	World Health Organization. WHO guidelines for treatment of cervical intraepithelial isia 2-3 and adenocarcinoma in situ. World Health Organization [2014 Available from: URL: <u>http://apps.who.int/iris/bitstream/10665/104174/1/9789241506779_eng.pdf</u>
58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

3 4 5 6	550 551 552	(40)	Castle PE, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Reliability of the Xpert HPV assay to detect high-risk human papillomavirus DNA in a colposcopy referral population. <i>Am J Clin Pathol</i> 2015; 143(1):126-133.
7 8 9 10	553 554 555	(41)	Cuzick J, Cuschieri K, Denton K, Hopkins M, Thorat MA, Wright C et al. Performance of the Xpert HPV assay in women attending for cervical screening. <i>Papillomavirus Research</i> 2015; 1:32-37.
11 12 13 14	556 557 558	(42)	Einstein MH, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Clinical Evaluation of the Cartridge-Based GeneXpert Human Papillomavirus Assay in Women Referred for Colposcopy. <i>J Clin Microbiol</i> 2014.
15 16 17 18 19 20	559 560 561 562	(43)	Toliman P, Badman SG, Gabuzzi J, Silim S, Forereme L, Kumbia A et al. Field Evaluation of Xpert HPV Point-of-Care Test for Detection of Human Papillomavirus Infection by Use of Self-Collected Vaginal and Clinician-Collected Cervical Specimens. <i>J Clin Microbiol</i> 2016; 54(7):1734-1737.
21 22 23 24 25	563 564 565 566	(44)	Cuschieri K, Geraets D, Cuzick J, Cadman L, Moore C, Vanden Broeck D et al. Performance of a Cartridge-Based Assay for Detection of Clinically Significant Human Papillomavirus (HPV) Infection: Lessons from VALGENT (Validation of HPV Genotyping Tests). <i>J Clin Microbiol</i> 2016; 54(9):2337-2342.
26 27 28 29	567 568 569	(45)	Kunckler M, Schumacher F, Kenfack B, Catarino R, Viviano M, Tincho E et al. Cervical cancer screening in a low-resource setting: a pilot study on an HPV-based screen-and-treat approach. <i>Cancer Med</i> 2017;10.
30 31 32 33 34	570 571 572	(46)	Guimaraes MD, Grinsztejn B, Melo VH, Rocha GM, Campos LN, Pilotto JH et al. Anal HPV prevalence and associated factors among HIV-seropositive men under antiretroviral treatment in Brazil. <i>J Acquir Immune Defic Syndr</i> 2011; 57 Suppl 3:S217-S224.
35 36 37	573 574	(47)	Castle PE, Schiffman M, Gravitt PE, Kendall H, Fishman S, Dong H et al. Comparisons of HPV DNA detection by MY09/11 PCR methods. <i>J Med Virol</i> 2002; 68(3):417-423.
38 39 40	575 576	(48)	Hayatsu H, Shiraishi M, Negishi K. Bisulfite modification for analysis of DNA methylation. <i>Curr Protoc Nucleic Acid Chem</i> 2008; Chapter 6:Unit.
41 42 43 44	577 578 579	(49)	Smith BC, McAndrew T, Chen Z, Harari A, Barris DM, Viswanathan S et al. The cervical microbiome over 7 years and a comparison of methodologies for its characterization. <i>PLoS One</i> 2012; 7(7):e40425.
45 46 47	580 581	(50)	Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. <i>Genome Biol</i> 2009; 10(3):R25.
48 49 50	582 583	(51)	Krueger F, Andrews SR. Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. <i>Bioinformatics</i> 2011; 27(11):1571-1572.
51 52 53 54 55 56 57 58	584 585	(52)	Qiao YL, Jeronimo J, Zhao FH, Schweizer J, Chen W, Valdez M et al. Lower cost strategies for triage of human papillomavirus DNA-positive women. <i>Int J Cancer</i> 2013.
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
3 4 5 6	586 587 588	(53)	Zhao FH, Jeronimo J, Qiao YL, Schweizer J, Chen W, Valdez M et al. An evaluation of novel, lower-cost molecular screening tests for human papillomavirus in rural China. <i>Cancer Prev Res (Phila)</i> 2013; 6(9):938-948.
7 8 9 10 11 12	589 590 591 592	(54)	Leisenring W, Alonzo T, Pepe MS. Comparisons of predictive values of binary medical diagnostic tests for paired designs. <i>Biometrics</i> 2000; 56(2):345-351.
	593		
55 56 57 58 59			
60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

Table 1: Recruitment sites and estimated eligible population.

Province	Site	Type of site	Potential participants I
			site (approximate)
Kigali	Cor-unum HC	Public Health Center	1,405
Kigali	Kimironko HC	Public Health Center	1,227
Kigali	Rwampara HC	Public Referral Hospital	1,098
Kigali	Kicukiro HC	Public Health Center	1,054
Kigali	Kacyiru HC	Public Health Center	905
Kigali	Gikondo HC	Public Health Center	925
Kigali	Rwanda Military Hospital	Public Referral Hospital	300
Kigali	WEACTx for Hope	Private HIV Clinic	500
Kigali	Busanza HC	Public Health Center	100
Kigali	Nyarugunga HC	Public Health Center	100
West	Gisenyi DH	Public District Hospital	810
Total			8,424
			1

29 of 34	BMJ Open
	Appendix I. Study Questionnaire
	A. <u>Socio-demographics</u>
	1. What is your date of birth? <u>D</u> <u>D</u> / <u>M</u> <u>M</u> / <u>Y</u> <u>Y</u> <u>Y</u> (enter all 0 if not remembered) (If date of birth or year of birth is given, skip to Question 3)
	2. What is your age?(Years)
	3. What is your marital status?
	Married/Cohabiting
	Divorced
	Widowed
	Separated
	□ Single
	Choose not to answer
	4. Do you live in Kigali?
	☐ Yes
	□ No {Skip to Question A5}
	Choose not to answer {Skip to Question A5}
	5. In what area of Kigali City Province do you live?
	Nyarugenge
	☐ Kicukiro
	 Kicukiro Gasabo urban (e.g., Kacyiru, Kimironko, Kinyinya, etc.) Gasabo rural (e.g., Nduba, Jabana, Gikomero, etc.)
	Gasabo rural (e.g., Nduba, Jabana, Gikomero, etc.)
	Do not know
	Choose not to answer
	6. What is your household income per month in FRW?
	□ < 5,000
	5,000 – 9,999
	10,000 - 34,999
	35,000 - 59,999

60,000 - 84,999

- 85,000 109,999
- 110,000 134,999
- 135,000 269,999
- 270,000 or more

7. How many people live in your household? _____ {Enter 00 if refuse to answer}

8. What is your occupation?

Employed by government, another institution, or company

Self-employed (Small and medium enterprises)

Self-employed (High income earnings)

Farming (peasants)

Unemployed/Does not work

Other (specify)

Choose not to answer {Skip to Question B1}

The following questions are sensitive and personal in nature. Your answers will be kept confidential. You may choose not to answer certain questions. Answering any question is voluntary}

B. Sexual Behaviors

<u>Sexual Behaviors</u>
How old were you when you first had sex? (Years) {Enter 00 if refuse to answer}

2. How many sexual partners have you had in your lifetime?

- No Partners (Ineligible)
- 1 Partner
- 2-3 Partners
- **4-5** Partners
- 6-9 Partners
- 10 or more partners
- Choose not to answer

3. How many sexual partners have you had in the last 6 months?

No Partners

	or more Partners
	Choose not to answer
C.	<u>Parity</u>
1	What age did you have your first child? (Years) {Enter 00 if ref answer, Enter 99 if never pregnant} (if 00 or 99, skip to Question D1)
2	2. How many live births have you had in your lifetime?
3	3. Have you given birth in the last year?
<u> </u>	Zes O
	Jo
	Choose not to answer
D.	Tobacco Use
1	. Have you ever smoked cigarettes?
<u> </u>	/es
<u> </u>	No (skip to Question D3)
	Choose not to answer (skip to Question D3)
2	2. Do you currently smoke cigarettes?
<u> </u>	/es
	Jo O
	Choose not to answer
3	3. Have you ever chewed/used tobacco orally (Ubugoro)?
<u> </u>	les
<u> </u>	No (skip to Question D5)
	Choose not to answer (skip to Question D5)
4	Are you currently chewing/using tobacco orally (Ubugoro)?
<u> </u>	les
<u> </u>	lo
$\Box c$	Choose not to answer

1
2
3
4
5
6
7
8
9
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
42
44
45
46
47
48
49
50
51
52
53
55 54
54 55
56
57
58
59

5.	Have you ever chewed/used tobacco ora	llv ((Tobacco	leaves-Igikamba	a)?
.	Huve you ever enewed, used tobacco of a		UDucco	icutos isinuinot	•,•

Yes

No (skip to Question E1)

Choose not to answer (skip to Question E1)

6. Are you currently chewing/using tobacco orally (Tobacco leaves-Igikamba)?

Yes Yes

No

Choose not to answer

E. <u>Contraceptive Use</u>

1. Have you ever used oral contraceptives?

- Yes
- No (Skip to Question E3)

Choose not to answer (Skip to Question E3)

2. Do you currently use oral contraceptives?

- Yes
- 🗌 No

Choose not to answer

3. Have you ever used Depo Provera (contraceptive)?

Yes

No (Skip to Question E5)

Choose not to answer (Skip to Question E5)

4. Do you currently use Depo Provera (contraceptive)?

- Yes
- No

Choose not to answer

5. Have you ever used Jadell (contraceptive)?

Yes

No (Skip to Question E7)

Choose not to answer (Skip to Qu	estion E7)
6. Do you currently use Jadell	(contraceptive)?
Yes	
🗌 No	
Choose not to answer	
7. Have you ever used IUD (co	ntraceptive)?
Yes	
No (Skip to Question E9)	
Choose not to answer (Skip to Qu	estion E9)
8. Do you currently use IUD (contraceptive)?
🗌 Yes	
🗌 No	
Choose not to answer	
9. Do use condoms as a contra	ceptive?
Yes	
No (skip to Question F1)	
Choose not to answer (skip to Qu	estion F1)
10. Have you used condoms as	a contraceptive in the last 6 months?
Yes	
No	
Choose not to answer	
F. <u>Infections</u>	
1. Have you ever had Malari	a
Yes	
☐ No (skip to Question F8)	
Choose not to answer (skip to Qu	estion EQ
	estion F8)

4	. Have you had your Malaria treated using drugs?
_	
∐ Ye	
∐ No	
	oose not to answer 5. If Yes, how was it treated?
_	eated only with traditional medicine
	eated only with drugs (e.g. Coartem)
	eated with both (traditional and modern)
	t treated
_	oose not to answer
6	5. Was your last episode of Malaria treated using drugs?
Ye	s
No	
Ch	oose not to answer
7	7. If Yes, how was it treated?
Tre	eated only with traditional medicine
Tre	eated only with drugs (e.g. Coartem)
Tre	eated with both (traditional and modern)
🗌 No	t treated
Ch	oose not to answer
8	8. Have you ever had Tuberculosis (TB)?
Ye	S
🗌 No	(skip to end)
Ch	oose not to answer (skip to end)
9	. If Yes, how many times have you had TB in your lifetime?
1	0. What year did you last have Tuberculosis (TB)? <u>Y</u> <u>Y</u> <u>Y</u> <u>Y</u>
	END OF QUESTIONNAIRE

BMJ Open

A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-Infected Women Living in Rwanda

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-020432.R2
Article Type:	Protocol
Date Submitted by the Author:	22-May-2018
Complete List of Authors:	Murenzi, Gad; Rwanda Military Hospital Dusingize, Jean; Regional Alliance for Sustainable Development , Research and Scientific Capacity Building Rurangwa, Theogene; Rwanda Military Hospital Sinayobye, Jean d'Amour; Regional Alliance for Sustainable Development, ; Women's Equity in Access to Care & Treatment (WE-ACTx), Munyaneza, Athanase; Rwanda Military Hospital Murangwa, Anthere; Rwanda Military Hospital Zawadi, Thierry; Rwanda Military Hospital Hebert, Tiffany; Yeshiva University Albert Einstein College of Medicine Mugenzi, Pacifique; Rwanda Military Hospital Adedimeji, Adebola; Yeshiva University Albert Einstein College of Medicine Mutesa, Leon; Rwanda Military Hospital; University of Rwanda College of Medicine and Health Sciences Anastos, Kathryn; Albert Einstein College of Medicine Medical Center, Bronx, Castle, Philip; Yeshiva University Albert Einstein College of Medicine,
Primary Subject Heading :	Global health
Secondary Subject Heading:	Obstetrics and gynaecology, Infectious diseases, HIV/AIDS, Epidemiology, Diagnostics
Keywords:	human papillomavirus (HPV), cervical cancer, HIV & AIDS < INFECTIOUS DISEASES, GYNAECOLOGY, cervical intraepithelial neoplasia

SCHOLARONE[™] Manuscripts Page 1 of 35

1

BMJ Open

r	
2	
3	
4	
4 5	
c	
6 7 8	
7	
8	
9	
9 10	
10	
11	
12	
11	
14	
13 14 15 16 17 18	
16	
17	
18	
10	
19	
20	
21	
22	
23	
24	
25	
26	
27	
20	
28	
29	
30	
31	
32	
52	
33	
34	
35	
36	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

1	A Study of Cervical Cancer Screening Technologies in Human Immunodeficiency Virus-
2	Infected Women Living in Rwanda
3	Gad Murenzi, MD* ¹ , Jean-Claude Dusingize, MD, MS ¹ , Theogene Rurangwa, MD, MMed ¹ ,
4	Jean d'Amour Sinayobye, MD, MS ¹ , Athanase Munyaneza, RN ¹ , Anthere Murangwa, MS ¹ ,
5	Thierry Zawadi, MD ¹ , Tiffany Hebert, MD ² , Pacifique Mugenzi, MD ¹ , MMed, Adebola
6	Adedimeji, PhD, MPH ² , Leon Mutesa, MD, PhD ^{1,3} , Kathryn Anastos, MD ² , Philip E. Castle,
7	PhD, MPH ²
8	¹ Rwanda Military Hospital, Kigali, Rwanda; ² Albert Einstein College of Medicine, Bronx, NY,
9	USA; ³ University of Rwanda, Kigali, Rwanda
10	*Correspondence: <u>gadcollins@gmail.com</u> , +250788589085
11	Disclosures: This research study has received HPV tests for reduced or no cost from Cepheid,
12	Arbor Vita Corporation, and Roche.
13	Funding: This study was funded by NCI/NIH Grant 5U54CA19016304 and by a grant from the
14	Prevent Cancer Foundation.
15	

16 Abstract

Introduction. The optimal method(s) for screening human immunodeficiency virus-infected
 women, especially for those living in sub-Saharan Africa, for cervical precancer and early cancer
 has yet to be established.

Methods and analysis. A convenience sample of >5,000 Rwandan women, ages 30-54 years and living with HIV infection, are being consented and enrolled into a cross-sectional study of cervical-cancer screening strategies. Participants are completing an administered short risk-factor questionnaire and being screened for high-risk human papillomavirus (hrHPV) using the Xpert HPV assay (Cepheid, Sunnyvale, CA, USA), unaided visual inspection after acetic acid (VIA), and aided VIA using the EVA system (Mobile ODT, Tel Aviv, Israel). Women positive for hrHPV and/or by VIA undergo colposcopy, which includes the collection of two cervical specimens prior to undergoing a 4-quadrant microbiopsy protocol. The colposcopy-collected specimens are being tested by dual immunocytochemical staining for p16^{INK4a} and Ki-67 (CINtec® PLUS Cytology, Ventana, Tucson, AZ, USA) and for E6 or E7 for 8 hrHPV genotypes (HPV16, 18, 31, 33, 35, 45, 52, and 58) using the next-generation AV Avantage hrHPV E6/E7 test (Arbor Vita Corporation, Freemont, CA, USA). Women with local pathology diagnosis of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) or pathology-review diagnosis of CIN grade 3 or more severe (CIN3+) receive treatment. Clinical performance and cost-effectiveness (e.g., sensitivity, specificity, and predictive values) of different screening strategies and algorithms will be evaluated.

- *Ethics and dissemination.* The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination.
- 33 39 through an external advisory panel.

1 2 3	41	Strengths and Weaknesses
2 3 2 3 4 5 6 7 2 8 2 2 9 10 2 10 12 2 13 14 2 16 17 18 19 20 21 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 45 46 47 48 49 49	41 42 43 44 45 46 47 48 49 50	<text><list-item><list-item><list-item></list-item></list-item></list-item></text>
44 45 46 47 48		

51 Introduction

Invasive cervical cancer (ICC) remains a significant cause of morbidity and mortality globally. Approximately 530,000 cases of and 270,000 deaths due to ICC occur annually, making it the 4th most common malignancy and cause of cancer-related death in women globally.¹ In many high-income countries (HIC), ICC rates have declined by 50% or more² due to the introduction of effective, high-coverage Pap testing (cervical cytology)-based screening programs that include timely follow-up of screen positives, treatment of women with precursor lesions, and management of cancers. Now, almost 90% of ICC and ICC-related deaths occur in low- and middle income countries (LMICs) due to a lack of resources and healthcare infrastructure needed to provide preventive services.

ICC and ICC-related mortality rates are particularly high in Sub-Saharan Africa, which also has the highest rates of HIV infection in the world. Now, over 12 million HIV-infected (HIV[+]) women in Sub-Saharan Africa are living longer because of anti-retroviral therapy, thus increasing their likelihood of dying from ICC.³ However, many of these women are already exposed to human papillomavirus (HPV), the viral cause of ICC, and will not benefit from or be targeted for prophylactic HPV vaccination. Thus, cervical-cancer screening is needed for the foreseeable future.

However, setting up effective cytology for cervical-cancer screening is expensive and requires a complex clinical and lab infrastructure that generally does not exist in LMICs.^{4;5} Moreover, it is now well understood that cytology has only a low- to moderate one-time sensitivity for precursor lesions and therefore must be done repeatedly over many years to reduce cancer risk. Alternative

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

BMJ Open

2	
3	
4	
5	
6	
7	
/	
8	
9	
10	
11	
12	
13	
14	
15	
16	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
27 28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
50 51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

strategies to address the ICC burden in LMICs, especially in SSA, must be developed andvalidated.

74 Persistent cervical infections by high-risk HPV (hrHPV) types cause virtually all ICC and its 75 immediate precursor lesions, e.g. cervical intraepithelial neoplasia grade 3 (CIN3) and adenocarcinoma *in situ* (AIS) everywhere in the world.^{6;7} hrHPV causes most anal and vaginal 76 cancer and a significant proportion of vulvar, penile, and oropharyngeal cancers.⁸ HPV16 is the 77 most important causal type, responsible for ~60% of ICC.⁹ HPV18 is the next most important, 78 responsible for 10-15% of ICC, including 30-40% of adenocarcinoma of the cervix⁹, which is on 79 the rise in Western Countries.^{10;11} Together, HPV16 and HPV18 account for ~70% of ICC, and 80 the same 15 hrHPV types account for ~99% of ICC everywhere in the world.⁹ 81

There is now overwhelming evidence to suggest that testing for hrHPV is more sensitive, albeit less specific, than high-quality cytology for identifying women with cervical precancer.¹²⁻¹⁶ Onetime hrHPV testing can reduce the risk of ICC incidence by approximately 40% in 6.5 years compared to cytology screening¹⁶, and ICC mortality by approximately 40% (approximately 50% overall) in 8 years compared to cytology.¹⁷ Importantly, a negative hrHPV test provides superior reassurance against CIN3+ ¹⁸ and against ICC^{16;17}, permitting safe extension of screening intervals.

The World Health Organization released cervical-cancer screening and treatment guidelines in 2013, recommending two evidence-based approaches to cervical-cancer screening¹⁹: (I) Use either hrHPV testing or visual inspection after acetic acid (VIA), which involves the inspection of the cervix with a speculum in place and following the application of dilute acetic acid to help identify potential CIN by its characteristic white coloring in the presence of acetic acid

94 (acetowhite), as alternative initial screening tests instead of Pap, and (II) immediately treat those
95 who screen positive using the screening test, rather than require diagnostic verification through
96 colposcopy and biopsy. This approach is commonly called screen-and-treat (S&T), and is
97 increasingly thought to be more amenable to LMIC settings.
98 However, hrHPV testing is also a much more effective screen than VIA¹⁷, which on a large-scale

appears to only down-stage cancer rather than prevent it.²⁰ Thus, the recent American Society for
Clinical Oncology (ASC) resource-stratified guidelines for secondary cervical-cancer
prevention^{21;22} emphasize that hrHPV testing is the preferred choice for screening, with VIA
only being used until hrHPV testing becomes available, and that HIV-infected women, because
of their higher risk, should be screened twice as frequently as the general (HIV-uninfected)
population.

Recent data in HIV[+] women living in the U.S. suggest that hrHPV testing may have clinical utility similar to that in HIV-negative (HIV[-]) women. Several observational studies have shown that an extended screening interval is safe in HIV[+] women who test hrHPV and Pap negative as it is for HIV[-] women.^{23;24} In a study of women enrolled in Women's Interagency Health Study (WIHS) in 2002, HIV[+] and HIV[-] women who tested hrHPV and Pap negative were at a similarly low risk of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) histology over a 5-year follow-up.²³ In addition, no cases of histologically confirmed CIN2+ were diagnosed in the follow-up of hrHPV- and Pap-negative HIV[+] women aged 30-64 years who underwent routine three-year hrHPV and cytology cotesting at Kaiser Permanente Northern California.²⁴ Thus, both studies found very high negative predictive values (NPV) >99% in HIV[+] women who test hrHPV negative. However, how hrHPV testing can

BMJ Open

2		
3 4	116	best be used to screen HIV[+] women living in Sub-Saharan Africa to prevent ICC remains to be
5 6 7	117	determined.
8 9 10	118	Recent studies ²⁵⁻²⁸ in HIV[+] women living in Sub-Saharan Africa have compared hrHPV, VIA,
11 12	119	and/or Pap for the detection of cervical precancer/cancer. The results can be summarized as
13 14 15	120	follows: 1) hrHPV detection was more sensitive but less specific than VIA; 2) surprisingly,
15 16 17	121	cytology was equally or more sensitive but less specific than VIA; and 3) surprisingly, cytology
18 19	122	was equally or more sensitive but less specific (vs. the converse) than hrHPV testing. Results and
20 21 22	123	conclusions are varied, leaving unanswered the question of what screening strategy in HIV[+]
22 23 24 25	124	women living in Sub-Saharan Africa has the greatest effectiveness and cost effectiveness.
25 26 27	125	Regardless of the screening method, most screen-positive women who go to colposcopy or are
28 29	126	treated immediately without diagnostic verification do not have cervical precancer and cancer
30 31 32	127	(positive predictive value [PPV] for screening tests are typically 10%-20%). In places like Sub-
33 34	128	Saharan Africa that lack necessary infrastructure and personnel such as pathologists ²⁹ , excessive
35 36	129	referral to colposcopy is problematic. Although WHO recommendations for S&T will hopefully
37 38 30	130	overcome this bottleneck and increase the number of women living in LMICs who get screened,
39 40 41	131	many countries may not adopt current S&T strategies because of concerns of low specificity and
42 43	132	overtreatment, resulting in increased costs, unnecessary patient discomfort and concern, and
44 45	133	wasting valuable healthcare resources that could otherwise be used to expand access to
46 47 48	134	screening. Thus, methods to increase the accuracy of screening by reducing the numbers of
49 50	135	women having colposcopy and biopsy or getting treated immediately in this context are highly
51 52 53 54 55 56	136	desirable as they will likely increase the uptake of cervical-cancer screening.

2
3
4
5
6
7
8
9
10
11
12 13
13 14
14 15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30 31
32
33
34
35
36
37
38
39
40
41
42
43
44 45
45 46
40 47
48
49
50
51
52
53
54
55
56
57
58
59 60
60

137	In order to improve the specificity of screening tests, secondary tests (biomarkers) are used
138	following a screen-positive result, with women who test positive for the triage undergoing
139	further management (e.g., colposcopy or immediate treatment) and those who test negative
140	typically being deferred to further evaluation in 6-18 months to allow hrHPV infections to clear.
141	There are several very promising biomarkers that might be used to improve the specificity and
142	positive predictive value (PPV) of the screening tests. ²⁴ Given that HIV-infected women are
143	more likely to test hrHPV positive ³⁰⁻³⁴ , it is important to validate a triage strategy of using a
144	secondary biomarker that sensitively and specifically rules-in women with cervical precancer
145	and cancer among the HIV-positive, hrHPV-positive women.
146	We are therefore conducting a cervical-cancer screening study of >5,000 Rwandan women, ages
147	30-54 years, living with HIV infection. We are evaluating different screening tests (hrHPV DNA
148	and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage
149	tests and biomarkers for the management of screen-positive women (E6/E7 oncoprotein
150	detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-positive
151	women undergo a rigorous colposcopic evaluation with multiple biopsies taken and the biopsies
152	will undergo pathology review, to minimize the misclassification of endpoints. The primary
153	objective of the study is to determine and compare clinical performance (Sensitivity (Se),
154	Specificity (Sp), PPV, and NPV) and cost-effectiveness for identifying HIV[+] women with
155	CIN3+ and CIN2+ of different cervical-cancer screening and management algorithms.
156	Methods and Analysis
157	Study design and population: We are recruiting those women receiving care for confirmed HIV
158	infection at health centers (HC) and various hospitals operated by the Ministry of Health or

Page 9 of 35

1

BMJ Open

2
3
4
5
6
7
0
8
9
10
11
10
12
13
14
15
10
16
17
18
12 13 14 15 16 17 18 19 20
20
20
20 21
22
23 24 25 26 27 28
25
24
25
26
20
27
28
29
30
21
31
32
33
34 35
25
35
36
36 37 38
20
20
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59

179

60

159	Rwanda Military Hospital during 2016-18 (Table 1). Sites were selected in collaboration with
160	Rwanda Biomedical Center (RBC), which calculate estimated numbers of potentially eligible
161	women using data from their HIV database (OpenMRS-Open Medical Records System;
162	http://openmrs.org/). Sites were also selected from all provinces to ensure geographic
163	representation. The total of 8,424 was the estimated number of women eligible for the study
164	according to the inclusion/exclusion criteria (see below) at the beginning for the study in 2016.
165	From each site, a convenience sample of women were/are being recruited to participate.
166	Inclusion criteria include 1) living and receiving HIV care in Rwanda, 2) ages 30-54 years, 3)
167	confirmed HIV+ based on medical records, 4) no prior cervical-cancer screening, 5) no history of
168	ICC, and 6) willing, able and competent to provide written, informed consent. We extended age
169	range beyond that of age range (30-49 years) recommended by the WHO for cervical-cancer
170	screening ¹⁹ because there is limited evidence for the optimal upper age for cervical-cancer
171	screening of HIV-infected women. Exclusion criteria, in addition to not meeting the inclusion
172	criteria, include 1) pregnant, 2) signs of abnormal, non-menstrual bleeding suggestive of ICC, 3)
173	without a cervix due to hysterectomy, and 4) not sufficiently healthy to participate in a research
174	study based on the judgment of the clinicians. Excluded women are being advised to seek routine
175	cervical-cancer screening through government programs.
176	Prior to initiation of enrollment at a specific clinic, the local HIV-care provider team identifies
177	potentially eligible women at their routine clinic visits and offers them enrollment. Women
178	indicating interest in the study then are being registered by our research nurses using the

180 willing women at that site are screened. The study team of at least two research nurses schedules

eligibility criteria checklist. Women at one site are being enrolled until all the eligible and

181 12 to 15 women three to four days a week by calling them and confirming appointments over the

telephone. Two teams of nurses are in the field, meaning that two cervical-cancer screeningclinics can be run simultaneously.

Enrollment Visit: The study participant flow is summarized in **Figure 1**. Enrollment visits, including pelvic exams with VIA and specimen collection, are being done entirely by a team of two study nurses. During their enrollment visit, women are being educated on cervical-cancer risk factors, mainly HPV infection, and why they are more at risk to develop ICC than HIVuninfected women. They also are being allowed to ask questions before they commit to participating in the study. Women are then being asked to provide informed, written consent to participate in the study using a printed out consent form. Those who provide consent complete a short nurse-administered questionnaire (Appendix I) on cervical cancer risk factors and sociodemographic characteristics using a data capture screen in Microsoft Access. The questionnaire collects information on basic sociodemographics, factors associated with acquiring HPV (e.g., marital status and recent and lifetime number of sexual partners), factors associated with increased risk of progression of hrHPV infection to precancer and cancer (e.g., smoking and other tobacco use, parity, and oral and other contraceptive use), and other infections common in Rwanda such as malaria³⁵ and tuberculosis³⁶ that have been previously reported to be associated with precancer among hrHPV-infected women. The questionnaire was not pretested. Enrolled women then undergo a pelvic exam, with VIA and a single cervical exfoliated ("Pap") specimen collected and placed into 20 ml PreservCyt (Hologic, Bedford, MA, USA) which is

201 then being sent to the lab at RMH for hrHPV testing. Finally, a portable colposcope

202 (<u>http://www.mobileodt.com/;</u> MobleODT, Tel Aviv, Israel) is being used for digital

203 cervicography (comparable to VIA with magnification) and the image is being captured and

saved for quality control, research, and to develop a digital library.

Page 11 of 35

BMJ Open

2		
- 3 4	205	Colposcopy Visit: Screen-positive women (women who test hrHPV and/or VIA positive) are
5	206	being called using a telephone as soon as the hrHPV result is available and being invited to
/ 3 9	207	return for colposcopy within one month. All screen-positive women receiving colposcopy will
10 11	208	are having two additional specimens collected, one into PreservCyt for the evaluation of other
12 13	209	molecular biomarkers (genotype-specific hrHPV viral methylation and load, and p16/Ki-67
14 15 16	210	immunocytochemistry CINtec® PLUS Cytology Kit [Roche, Tucson, AZ, USA]) and a second
17 18	211	as a dry swab for HPV16, 18, 31, 33, 35, 45, 52, and 58 E6/E7 oncoprotein detection by the next
19 20	212	generation lateral flow hrHPV oncoprotein test from Arbor Vita Corporation (Fremont, CA,
21 22	213	USA) being included in this study as a triage for screen-positive women to identify those women
23 24 25	214	who are at higher risk of having CIN3+. The residual PreservCyt specimens from both the
26 27	215	screening and colposcopy visits are being stored at -20°C, creating a biobank in Rwanda for
28 29 30	216	future retrospective evaluations of promising new biomarkers and tests.
31 32 33	217	After specimen collection, a colposcopic evaluation of the cervix is being done with a modified
33 34 35	218	version of the 4-quadrant microbiopsy procedure being performed. ³⁷ Compared to the standard
36 37	219	biopsy, the microbiopsy protocol improves disease ascertainment and reduces biases related to
38 39	220	selecting the most visually obvious acetowhite lesions while removing less tissue ($\sim 13 \text{ mm}^2$ for 4
40 41 42	221	microbiopsies vs. ~28mm ² for 1 standard biopsy). Modifications to the standard 4-quadrant
43 44	222	microbiopsy procedure are: 1) endocervical curettage is being taken only for those women whose
45 46	223	squamocolumnar junction is not entirely visible and the lesion extends into the endocervical
47 48 49	224	canal; and 2) standard-size biopsies of very large lesions are being taken to increase the
50	225	likelihood that the most severe area is being biopsied.
51 52 53 54 55	226	Pathology: Biopsies are being processed in a single cassette so that a single slide has a section

from all biopsies taken. Biopsies are being read by a local pathologist at RMH and Dr. Hebert or

1	
2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 22 23 24 25 26 27 28 29 30 31 20 21 22 23 24 25 26 27 28 29 30 31 20 21 22 23 24 25 26 27 28 29 30 31 20 20 20 20 20 20 20 20 20 20	
5 4	
5	
6	
/ 8	
9	
10	
11	
12	2
14	
15	
16	
17 18	
19	4
20	
21	-
22 23	
24	
25	
26	
27 28	
29	
30	
31	4
32 33	
34	4
35	:
36	
37 38	
39	
40	
41	4
42 43	
44	
45	
46	
47 48	2
49	
50	2
51	
52 53	
55 54	
55	
56	
57 58	
58 59	
60	

228	another pathologist at Montefiore Medical Center, Bronx, NY, USA. Women receiving a
229	diagnosis of CIN2+ by the Rwandan pathologist (T.Z.) or, as a safety precaution, CIN3+
230	diagnosis by Montefiore pathologist (T.H.) are receiving treatment ³⁸ : 1) CIN2, CIN3, or AIS are
231	being referred to study doctors to undergo an excision procedure (e.g., loop electrosurgical
232	excision procedure [LEEP] or cold-knife cone [CKC]) and 2) ICC are being referred to RMH
233	Hospital for care. Women with <cin2 a="" advised="" are="" being="" in="" re-screening="" seek="" td="" to="" year.<=""></cin2>
234	A slide with biopsies also are undergoing p16 immunohistochemistry (IHC) using the CINtec®
235	Histology Kit (Roche) for research purposes only.
236	Endpoints: The primary scientific endpoints of the study are histologically confirmed, consensus
237	CIN2+ i.e., both pathologists diagnose CIN2+ (without adjudication) or CIN3+ by the study
238	pathologist. The secondary, clinical endpoint is histologically confirmed CIN2+ diagnosed by
239	the Rwandan pathologist. Additional endpoints using pathology review and p16 IHC are being
240	used but not for evaluating the performance of p16 immunocytochemistry due to the possibility
241	of p16-related autocorrelation.
242	<i>Treatment</i> : Women diagnosed with CIN2+ are being referred for treatment. Those precancerous
243	lesions are being treated by ablation if they meet WHO criteria for cryotherapy. ³⁹ Those who do
244	not meet those criteria undergo an excision procedure (e.g., loop electrosurgical excision
245	procedure [LEEP] or cold-knife cone [CKC]) or, in the case of an ICC diagnosis, referred for
246	cancer management. Screen-positive women with <cin2 advised="" are="" being="" re-screening<="" seek="" td="" to=""></cin2>
247	in a year through the existing healthcare system.
248	Data sources: Data are being collected from the following sources:

60

BMJ Open

1 2		
3 4	249	1. A nurse-administered questionnaire on sociodemographic characteristics and cervical
5 6	250	cancer risk factors including age at first sexual intercourse, number of sexual partners,
7 8 9	251	smoking, contraception, parity and socioeconomic status.
10 11 12 13	252	2. Pelvic exam, VIA, Mobile ODT and colposcopy data capture forms
14 15	253	3. Medical record data on HIV status (e.g.,
16 17	254	(http://www.who.int/hiv/pub/guidelines/HIVstaging150307.pdf), CD4 count, viral load,
18 19 20	255	antiretroviral therapy (ART) regimen(s)), care, and dates.
21 22 23 24	256	Laboratory Testing: The following laboratory tests are being performed:
25 26	257	Xpert HPV Testing—cervical Pap specimens in PreservCyt are being sent to the RMH
27 28 29	258	laboratory in Kigali, Rwanda for hrHPV DNA testing using the Xpert HPV test (Cepheid,
30 31	259	Sunnyvale, CA, USA). ⁴⁰⁻⁴⁵ The Xpert HPV Assay is a new, qualitative, real-time PCR assay for
32 33	260	the detection of hrHPV DNA. The Xpert HPV Assay includes simultaneous detection of 14
34 35 36	261	hrHPV types, hydroxymethylbilane synthase (HMBS), and an internal Probe Check Control
37 38	262	(PCC). The 14 targeted hrHPV types are detected in 5 fluorescent channels: 1) HPV16, 2)
39 40	263	HPV18 and hrHPV 45 (HPV18/45), 3) HPV31, 33, 35, 52, and 58, 4) HPV51 and HPV59, and
41 42 43	264	5) HPV39, 56, 66, and 68. HMBS (fluorescent channel 6) verifies specimen adequacy.
44 45 46	265	Specimens are being mixed and a 1-mL pre-aliquot is being removed using a disposable pipette
47 48	266	and placed in the testing cartridge per the manufacturer's instructions. Unsatisfactory results due
49 50	267	to insufficient cellular content are being re-tested. If the second test is also unsatisfactory, the
51 52 53	268	final result are being recorded as unsatisfactory but women with unsatisfactory results are being
54 55 56 57 58	269	referred to colposcopy for safety.

HPV Viral Methylation—We will conduct a retrospective analysis of HPV viral methylation and its association with CIN2+. To identify single hrHPV type infections, we will select singlechannel positives from the Xpert HPV assay. For those that are hrHPV positive for a channel other than HPV16, which is detected singly, we will test them to identify the single type infections using a standard protocol for PCR amplification using MY09/11 L1 consensus primers and hrHPV genotype detection using dot-hybridization for 39 individual type-specific probes and a mixture of probes for 10 other uncommon hrHPV types as previously described.^{46;47} To isolate the DNA, ThinPrep specimens (1.5 mL) will be pelleted, re-suspended in STM, digested with Proteinase K, precipitated overnight in ammonium acetate ethanol at -20°C, washed, and suspended and stored in TE buffer. The isolated DNA then will undergo bisulfite conversion.⁴⁸ Following bisulfite conversion and DNA purification and de-sulphonation, bisulfite-treated DNA will be used as template for Next-Gen Sequencing (NGS) (HiSeq2000, Illumina, San Diego, CA) using barcoded-type specific primers. Sequences for pads and barcodes are not found in the targeted genomic region. Use of padding and barcodes will enable reads to be identified by amplicon (forward or reverse) or by sample during downstream bioinformatics analysis.⁴⁹ All PCR products for NGS will be pooled (by assay) and a single DNA band containing multiple amplicons from different samples (with unique barcodes) will be isolated from a gel for NGS.⁴⁹

288 Briefly, equal concentrations of each barcoded PCR product (based on PCR band intensity) will

289 be pooled and isolated. Upon confirmation of correct product size, all purified DNA pools will

be combined and submitted for library preparation and paired-end 100 base pair Illumina

HiSeq2000 sequencing at the Einstein Genomics Core Facility.

Page 15 of 35

BMJ Open

2	
2 3 4 5 6 7	2
4 5	
6	2
7 8	-
9	4
10 11	2
12	
13 14	4
9 10 11 12 13 14 15 16 17 18 19 20	2
16 17	
18	4
19 20	
21	4
22 23	3
21 22 23 24 25 26	
25 26	Ċ
27 28	3
27 28 29 30 31 32 33 34 35 36	-
30 31	
32	
33 34	
35	2
36 37 38	
	÷
39 40	
41 42	
42 43	
44 45	2
46	
47 48	
49	
50 51	3
52	
53 54	
55	
56 57	
58	
59 60	

Methylation status are being determined in the lab of Dr. Robert Burk at Albert Einstein College of Medicine (USA). Prior to determination of methylation status, de-multiplexing based on the unique barcodes is being performed using in-house generated scripts to obtain paired-end NGS reads of each sample. Reads are being aligned with hrHPV reference genome sequences by bowtie v0.12.9.⁵⁰ Methylation status of each CpG site is then determined by bismark v0.7.7⁵¹ using the default quality score parameter set to Q30, and the formula of the methylation ratio of the number of C read by the number of C+T read.

E6/E7 Oncoprotein Testing—Dry swab specimens, collected at the time of colposcopy, are being
 tested for individual E6/E7 oncoproteins as previously described^{52;53}, according to the
 manufacturer's instructions, at the RMH laboratory in Kigali, Rwanda. The E6/E7 oncoprotein
 test uses three lateral flow strips to detect 8 hrHPV types whereas the E6 oncoprotein test used a
 single lateral flow strip to detect 3 hrHPV types.

Analyses: We will evaluate combinations of the above mentioned screening strategies and tests to estimate the clinical performance (e.g., Se, Sp, PPV, and NPV) for the detection of consensus CIN3+ and community CIN2+. A log binomial model using generalized estimating equations will be used to take into account correlation between different tests from the same subject. Note while these models will be developed for the estimation and comparison of performance for two tests, the model can be extended to allow more than two tests by including more indicator variables for test type.

Some analyses of biomarkers, such as viral methylation will be restricted to hrHPV-positives.
Comparisons of hrHPV viral methylation to other triage biomarkers will be restricted to the
subset that gets tested for viral methylation as described.

1	
2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
27	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
40 41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
55 54	
54 57	
55	
56	
57	
58	
59	

60

314	Sample size calculations: We are basing our sample size on the ability to detect modest but
315	minimally important differences in Se of 15%. We conservatively assume that the population
316	prevalence of CIN3+ is 2% based on our past study in Rwanda. ^{33;35} We will enroll and have
317	complete follow-up of at least 5,000 HIV[+] women. A sample size of 5,000 HIV[+] women
318	with completed follow-up of the screen positives will yield 100 cases of CIN3+, which will have
319	at least 80% power (α =5%) to crudely detect a 15% difference in Se between two screening
320	strategies for a range of 10%-25% discordance. With this sample size of 5,000 women, 4,900
321	will not have CIN3+; we will also have at least 90% power (α =5%) to detect a difference in Sp
322	of 3% for discordance up to 40%. Finally, we will have 80% power (α =5%) to crudely detect an
323	8%, 10%, or 11% difference in PPV if the reference PPV is 10%, 20%, or 30%, respectively. ⁵⁴
324	Cost Effectiveness: We will conduct assessments of the costs and cost-effectiveness of the
325	different combinations of screening and triage tests, i.e., algorithms, as well as those of the entire
326	community-based screening "system." Costs measurement will be conducted using a micro-
327	costing (ingredients) approach in which resource use throughout each step in the screening
328	process will be tracked and unit costs for each of the resources will be applied to generate an
329	average screening cost per woman to be compared against what the estimated costs are for a
330	possible program based on hrHPV screening and VIA triage or VIA screening. For estimating
331	costs of the screening system and scale-up of screening to 100,000 women in a month, analyses
332	will distinguish financial costs, which reflect actual expenditures of the program, from economic
333	costs, including the value of donated and shared resources to more fully assess opportunity costs.
334	Projections on budget impact and economic cost implications over time will be made under
335	varying assumptions of screening uptake, follow-up compliance, and scenarios of changing
336	disease burden.

Page 17 of 35

BMJ Open

Clinical outcomes will include true positive, true negative, false negative, and false positive test results, number of colposcopies, incident cancer, and cancer death. Cost-effectiveness will be measured as cost/CIN2+ detected, cost/CIN3+ detected, cost/invasive cancer prevented, cost/cancer death prevented, cost/life-year saved, and cost/quality-adjusted life year (QALY) saved: in addition, we will calculate harm/benefit ratios, using varying definitions of harms (colposcopies, false positive results) to benefits (cancers prevented, deaths prevented, life years and QALYs saved). Costs and effectiveness will be discounted at a 3% annual rate, with the rate varied from 0-5% in sensitivity analysis. For assessment of value-of-information (VOI), we will use net monetary benefits (NMB), defined as a function of the willingness-to-pay threshold (WTP) for different costs and outcomes as: NMB = (WTP * Effectiveness) - Costs.

Patient and Public Involvement

There was no patient engagement in the development or design of the study, recruitment, and the conduct of the study. Participants are receiving their results directly since it is related to their care. As this was not a randomized controlled trial, the burden of the intervention was not assessed by patients themselves. There were no patient advisors to acknowledge.

352 Ethics and Dissemination

Ethics: This study protocol was reviewed and approved by the Rwanda National Ethics
Committee (RNEC) as well as the Institutional Review Board for human subjects research at
Albert Einstein College of Medicine.

Confidentiality measures and protection against potential risks: The risks for those participating
357 in our study include:

1 2

2 3 4	250	Collection of Dan anazimana/comviced awaha involves a modest risk of blooding
	358	Collection of Pap specimens/cervical swabs involves a modest risk of bleeding
5 6	359	which is typically very limited when it occurs. Testing positive for any test may
7 8	360	cause psychological distress (anxiety).
9	500	eause psychological distress (anxiety).
10 11	361	• Colposcopy and excisional treatments induce vaginal bleeding and may incur pain,
12	262	
13 14	362	infection, and short-term psychological distress (anxiety). A diagnosis of CIN2 or
15 16	363	more severe may cause psychological distress (anxiety). A diagnosis of ICC may
17	364	cause severe psychological distress.
18 19	501	eduse severe psychological distress.
20 21	365	• Questions in the questionnaire, regarding sexual behavior and other matters of a
22	200	nerconal nature, may appear any intra and emberragement. Participants are advised
23 24	366	personal nature, may cause anxiety and embarrassment. Participants are advised
25 26	367	that they are free not to answer specific questions.
27		
28 29	368	• There is also the risk of psycho-social stress which could occur if there was
30 31	369	inadvertent disclosure of confidential medical or other personal information.
32		
33 34	370	Protection against the risk of inadvertent disclosure of confidential information is being
35	371	addressed by the standard procedures at the Rwandan study site, including: (i) storing completed
36 37		
38 39	372	paper copies of questionnaires and other hard copy information (described above), identified by
40	373	study number only, in a filing system separate from the name-address file of participants in the
41 42	374	study; and (ii) only the designated local personnel have access to cross-reference the files; (iii)
43 44	571	
45	375	all paper files, including consent forms, are being maintained in locked cabinets in locked rooms,
46 47	376	with access restricted to specific research personnel.
48 49		
50	377	In addition, we will include the following security measures to protect the data:
51 52	077	
53 54	378	• Controlled access to project data;
55	570	• Controlled access to project data,
56 57		
58		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

60

BMJ Open

379	• A tracking system for data forms and activities;
380	• Date and time of stamping of all data records with electronic signatures;
381	• Audit trails to track all changes made to data records; and
382	• Data kept on password-protected computers and in locked rooms.
383	Potential Patient Benefits: There are potential direct benefits to study participants. HIV[+]
384	women participating in the study are at very high risk of ICC. They are being rigorously
385	screened and evaluated, more effectively than the standard of care anywhere in the world. As a
386	result of the study, women with precancer who are at imminent risk of ICC are being diagnosed
387	sooner and treated more effectively than women receiving routine care and thereby more likely
388	averting the development of ICC. Women with cervical cancer are being diagnosed earlier
389	thereby reducing the morbidity and the risk of mortality caused by ICC. Conversely, any pain,
390	bleeding, or stress that might occur related to colposcopy or cervical swab are typically modest
391	and well tolerated.
302	There are also substantial potential societal benefits related to the study due to its implications to
393	improving cervical-cancer screening practices and guidelines in HIV[+] women – changes in
394	practice which might also benefit the study participants themselves, if and when these changes
395	are enacted. There is a great need to identify more effective and practical methods for cervical-
396	cancer screening for HIV[+] women living in Africa, who are living longer than ever and are
397	therefore at potentially greater risk of ICC.
398	Dissemination: We plan to publish a series of scientific reports in peer-reviewed scientific
399	journals. As building research capacity in Rwanda is a major goal of this research project, all
400	investigators of the research team are being asked and supported to lead at least one analysis and
401	one manuscript preparation, based on interests and expertise.
	 380 381 382 383 384 385 386 387 388 389 390 391 392 393 394 395 396 397 398 397 398 397 398 397 398 397 398 397 398 399 400

In addition, an external advisory panel (EAP) composed of leaders from the Rwanda Ministry of
Health, University of Rwanda, and Rwanda medical community has been formed. The
responsibilities of the EAP include providing advice on the conduct of the project and
interpretation for and dissemination of the study results to Rwandan stakeholders. The latter is
important for the adoption of evidence-based best practices for cervical-cancer screening as
warranted.

408 Limitations

There are several limitations to the study that bear mentioning. First, cervical cytology was not included in the study. There is limited cervical cytology services available locally and of unknown quality and it is unlikely that cytology are being widely available in Rwanda, making its inclusion as a comparator test of limited value. Moreover, there are significant costs and logistical challenges in shipping PreservCyt specimens to and having cytology slides made and read in the U.S. Second, we did not conduct biopsies in screen-negative women, which would have allowed us to estimate absolute clinical performance. The burden of sending screen-negative women to colposcopy was deemed too great and it was impractical to send a sufficient numbers of screen-negative women to colposcopy to accurately estimate the false-negative disease (CIN3+) fraction. Thus, only relative clinical performance of the screening tests can be estimated from this study.

BMJ Open

KA, PEC, JCD, AA, and JDS conceived the original concept of the study and the interventions.

GM, KA, JDS, and PEC drafted the protocol. PEC performed the sample size calculations, and

supporting patient recruitment. GM, TR, and Athanase Munyaneza are providing clinical care for

patients. Anthere Murangwa and LM oversee laboratory testing, and TZ and TH are responsible

Rwanda. All authors (GM, JCD, TR, JDS, Athanase Munyaneza, Anthere Murangwa, TZ, TH,

PM, AA, LM, KA, and PEC) contributed to the scientific design of the study and the protocol

development, are involved in the implementation of the project, and have read and approved the

VOIV

for pathology. PM and LM oversee and administer the study activities at the clinical site in

PEC and GM will lead analysis of the results. GM, JDS, and Athanase Munyaneza are

Contributor Statement:

final manuscript.

1		
2 3	432	Figure Legends:
2 3 4 5 6	433	
6 7 8	434	Figure 1. Study Design
8 9 10	435	
10 11 12	100	
12 13 14		
15 16		
17 18		
19 20		
21 22 23		
24 25		
26 27		
28 29		
30 31 32		
33 34		
35 36		
37 38 39		
40 41		
42 43		
44 45 46		
40 47 48		
49 50		
51 52		
53 54		
55 56 57		
57 58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
2 3	436	Refere	nce List
4	150	Refere	
5 6	437		
7	438	(1)	Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C et al. GLOBOCAN 2012 v1.0,
8	439	ĊĴ	Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. 2013. Lyon,
9 10	440		France, International Agency for Research on Cancer. 4-22-2014.
11	441	Ref Ty	pe: Online Source
12			
13	442	(2)	Lonnberg S, Hansen BT, Haldorsen T, Campbell S, Schee K, Nygard M. Cervical cancer
14	443		prevented by screening: Long-term incidence trends by morphology in Norway. Int J Cancer
15	444		2015;10.
16	445	(2)	
17	445		UNAIDS. UNAIDS Data 2017. 2017.
18 19	446	Refity	pe: Report
20	447	(4)	Cervix Cancer Screening. [10]. 2005. IARC Press. IARC Handbooks of Cancer Prevention.
21	448		pe: Serial (Book,Monograph)
22	110	ner ry	
23	449	(5)	Kitchener HC, Castle PE, Cox JT. Chapter 7: Achievements and limitations of cervical
24	450		cytology screening. Vaccine 2006; 24 Suppl 3:S63-70.:S63-S70.
25			
26 27	451	(6)	Schiffman M, Castle PE, Jeronimo J, Rodriguez AC, Wacholder S. Human papillomavirus and
27 28	452		cervical cancer. <i>Lancet</i> 2007; 370(9590):890-907.
29			
30	453	(7)	Schiffman M, Clifford G, Buonaguro FM. Classification of weakly carcinogenic human
31	454		papillomavirus types: addressing the limits of epidemiology at the borderline. <i>Infect Agent</i>
32	455		<i>Cancer</i> 2009; 4:8.
33	456	(8)	Forman D, de MC, Lacey CJ, Soerjomataram I, Lortet-Tieulent J, Bruni L et al. Global burden
34	450 457	(0)	of human papillomavirus and related diseases. <i>Vaccine</i> 2012; 30 Suppl 5:F12-23. doi:
35 36	458		10.1016/j.vaccine.2012.07.055.:F12-F23.
37	430		10.1010/j.vaccinc.2012.07.055112-125.
38	459	(9)	de SS, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Lloveras B et al. Human
39	460	(1)	papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-
40	461		sectional worldwide study. <i>Lancet Oncol</i> 2010; 11(11):1048-1056.
41			
42	462	(10)	Bray F, Carstensen B, Moller H, Zappa M, Zakelj MP, Lawrence G et al. Incidence trends of
43	463		adenocarcinoma of the cervix in 13 European countries. Cancer Epidemiol Biomarkers Prev
44 45	464		2005; 14(9):2191-2199.
46		(1.1)	
47	465	(11)	Adegoke O, Kulasingam S, Virnig B. Cervical cancer trends in the United States: a 35-year
48	466		population-based analysis. J Womens Health (Larchmt) 2012; 21(10):1031-1037.
49	467	(12)	Cuzick J, Clavel C, Petry KU, Meijer CJ, Hoyer H, Ratnam S et al. Overview of the European
50	467	(12)	and North American studies on HPV testing in primary cervical cancer screening. Int J
51	469		<i>Cancer</i> 2006; 119(5):1095-1101.
52	409		Cancer 2000, 117(5).1075-1101.
53 54	470	(13)	Naucler P, Ryd W, Tornberg S, Strand A, Wadell G, Elfgren K et al. Human papillomavirus
55	471	()	and Papanicolaou tests to screen for cervical cancer. N Engl J Med 2007; 357(16):1589-
56	472		1597.
57			
58			
59			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml
60			i or peer review only - http://binjopen.binj.com/site/about/guidennes.xhtml

1 2								
2 3 4 5	473 474 475	(14)	Ronco G, Giorgi-Rossi P, Carozzi F, Confortini M, Palma PD, Del MA et al. Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised controlled trial. <i>Lancet Oncol</i> 2010.					
6 7 8 9 10	476 477 478	(15)	Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkmans NW, Heideman DA et al. Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the POBASCAM randomised controlled trial. <i>Lancet</i>					
10 11 12	479		Oncol 2012; 13(1):78-88.					
13 14 15 16	480 481 482	(16)	Ronco G, Dillner J, Elfstrom KM, Tunesi S, Snijders PJ, Arbyn M et al. Efficacy of HPV-based screening for prevention of invasive cervical cancer: follow-up of four European randomised controlled trials. <i>Lancet</i> 2013.					
17 18 19	483 484	(17)	Sankaranarayanan R, Nene BM, Shastri SS, Jayant K, Muwonge R, Budukh AM et al. HPV screening for cervical cancer in rural India. <i>N Engl J Med</i> 2009; 360(14):1385-1394.					
20 21 22 23	485 486 487	(18)	Dillner J, Rebolj M, Birembaut P, Petry KU, Szarewski A, Munk C et al. Long term predictive values of cytology and human papillomavirus testing in cervical cancer screening: joint European cohort study. <i>BMJ</i> 2008; 337:a1754. doi: 10.1136/bmj.a1754.:a1754.					
24 25	488	(19)	New guidelines on screening and treatment for cervical cancer. 2013. South Africa, World					
26	489 490	Ref Tv	Health Organization. ef Type: Pamphlet					
27 28	170	ner ry						
29	491	(20)	Shastri SS, Mittra I, Mishra GA, Gupta S, Dikshit R, Singh S et al. Effect of VIA screening by					
30 31 32	492 493		primary health workers: randomized controlled study in Mumbai, India. <i>J Natl Cancer Inst</i> 2014; 106(3):dju009.					
33	494	(21)	Jeronimo J, Castle PE, Temin S, Shastri SS. Secondary Prevention of Cervical Cancer:					
34 35 36	495 496		American Society of Clinical Oncology Resource-Stratified Clinical Practice Guideline Summary. <i>J Oncol Pract</i> 2016; JOP2016017889.					
37	497	(22)	Castle PE, Jeronimo J, Temin S, Shastri SS. Screening to Prevent Invasive Cervical Cancer:					
38 39	498		ASCO Resource-Stratified Clinical Practice Guideline. <i>J Clin Oncol</i> 2017; 35(11):1250-1252.					
40	499	(23)	Keller MJ, Burk RD, Xie X, Anastos K, Massad LS, Minkoff H et al. Risk of cervical precancer					
41 42 43	500 501		and cancer among HIV-infected women with normal cervical cytology and no evidence of oncogenic HPV infection. <i>JAMA</i> 2012; 308(4):362-369.					
44 45 46 47	502 503 504	(24)	Castle PE, Fetterman B, Poitras N, Lorey T, Kinney W. Safety against cervical precancer and cancer following negative human papillomavirus and Papanicolaou test results in human immunodeficiency virus-infected women. <i>Arch Intern Med</i> 2012; 172(13):1041-1043.					
48 49 50 51	505 506 507	(25)	Chung MH, McKenzie KP, De VH, Richardson BA, Rana F, Pamnani R et al. Comparing pap smear, via, and hpv cervical cancer screening methods among hiv-positive women by immune status and antiretroviral therapy. <i>AIDS</i> 2013.					
52 53 54 55 56 57	508 509 510	(26)	Firnhaber C, Mayisela N, Mao L, Williams S, Swarts A, Faesen M et al. Validation of cervical cancer screening methods in HIV positive women from Johannesburg South Africa. <i>PLoS One</i> 2013; 8(1):e53494.					
58								
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml					

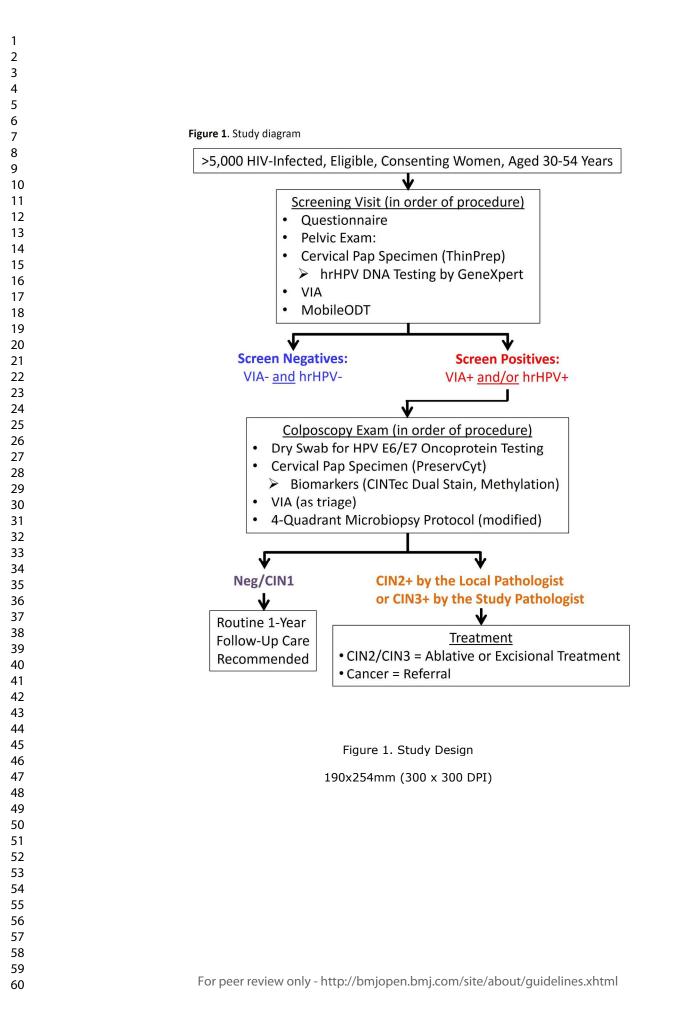
1 2			
3 4 5 6	511 512 513	(27)	Dartell MA, Rasch V, Iftner T, Kahesa C, Mwaiselage JD, Junge J et al. Performance of visual inspection with acetic acid and human papillomavirus testing for detection of high-grade cervical lesions in HIV positive and HIV negative Tanzanian women. <i>Int J Cancer</i> 2014;10.
7 8 9 10 11	514 515 516 517	(28)	Mabeya H, Khozaim K, Liu T, Orango O, Chumba D, Pisharodi L et al. Comparison of conventional cervical cytology versus visual inspection with acetic acid among human immunodeficiency virus-infected women in Western Kenya. <i>J Low Genit Tract Dis</i> 2012; 16(2):92-97.
12 13 14	518 519	(29)	Adesina A, Chumba D, Nelson AM, Orem J, Roberts DJ, Wabinga H et al. Improvement of pathology in sub-Saharan Africa. <i>Lancet Oncol</i> 2013; 14(4):e152-e157.
15 16 17 18 19	520 521 522	(30)	Dartell M, Rasch V, Kahesa C, Mwaiselage J, Ngoma T, Junge J et al. Human papillomavirus prevalence and type distribution in 3603 HIV-positive and HIV-negative women in the general population of Tanzania: the PROTECT study. <i>Sex Transm Dis</i> 2012; 39(3):201-208.
20 21 22 23 24	523 524 525 526	(31)	D'Souza G, Burk RD, Zhong Y, Minkoff H, Massad LS, Xue X et al. Cervicovaginal human papillomavirus (HPV)-infection before and after hysterectomy: evidence of different tissue tropism for oncogenic and nononcogenic HPV types in a cohort of HIV-positive and HIV-negative women. <i>Int J Cancer</i> 2012; 131(6):1472-1478.
25 26 27 28	527 528 529	(32)	Ng'andwe C, Lowe JJ, Richards PJ, Hause L, Wood C, Angeletti PC. The distribution of sexually-transmitted Human Papillomaviruses in HIV positive and negative patients in Zambia, Africa. <i>BMC Infect Dis</i> 2007; 7:77.:77.
29 30 31 32	530 531 532	(33)	Singh DK, Anastos K, Hoover DR, Burk RD, Shi Q, Ngendahayo L et al. Human papillomavirus infection and cervical cytology in HIV-infected and HIV-uninfected Rwandan women. <i>J Infect Dis</i> 2009; 199(12):1851-1861.
33 34 35 36 37	533 534 535	(34)	Marais DJ, Vardas E, Ramjee G, Allan B, Kay P, Rose RC et al. The impact of human immunodeficiency virus type 1 status on human papillomavirus (HPV) prevalence and HPV antibodies in serum and cervical secretions. <i>J Infect Dis</i> 2000; 182(4):1239-1242.
38 39 40 41	536 537 538	(35)	Anastos K, Hoover DR, Burk RD, Cajigas A, Shi Q, Singh DK et al. Risk factors for cervical precancer and cancer in HIV-infected, HPV-positive Rwandan women. <i>PLoS One</i> 2010; %20;5(10):e13525.
42 43 44 45	539 540 541	(36)	Zhao FH, Varanasi AP, Cunningham CA, Graubard BI, Hu SY, Chen F et al. Tuberculosis and oncogenic HPV: potential co-infections in women at high-risk of cervical cancer in rural China. <i>Asian Pac J Cancer Prev</i> 2011; 12(6):1409-1415.
46 47 48 49	542 543 544	(37)	Pretorius RG, Zhang WH, Belinson JL, Huang MN, Wu LY, Zhang X et al. Colposcopically directed biopsy, random cervical biopsy, and endocervical curettage in the diagnosis of cervical intraepithelial neoplasia II or worse. <i>Am J Obstet Gynecol</i> 2004; 191(2):430-434.
50 51 52	545 546	(38)	Schiffman M, Adrianza ME. ASCUS-LSIL Triage Study. Design, methods and characteristics of trial participants. <i>Acta Cytol</i> 2000; 44(5):726-742.
53 54 55 56 57	547 548 549	(39) neopla	World Health Organization. WHO guidelines for treatment of cervical intraepithelial isia 2-3 and adenocarcinoma in situ. World Health Organization [2014 Available from: URL: <u>http://apps.who.int/iris/bitstream/10665/104174/1/9789241506779_eng.pdf</u>
58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

3 4 5 6	550 551 552	(40)	Castle PE, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Reliability of the Xpert HPV assay to detect high-risk human papillomavirus DNA in a colposcopy referral population. <i>Am J Clin Pathol</i> 2015; 143(1):126-133.
7 8 9 10	553 554 555	(41)	Cuzick J, Cuschieri K, Denton K, Hopkins M, Thorat MA, Wright C et al. Performance of the Xpert HPV assay in women attending for cervical screening. <i>Papillomavirus Research</i> 2015; 1:32-37.
11 12 13 14 15	556 557 558	(42)	Einstein MH, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Clinical Evaluation of the Cartridge-Based GeneXpert Human Papillomavirus Assay in Women Referred for Colposcopy. <i>J Clin Microbiol</i> 2014.
16 17 18 19 20	559 560 561 562	(43)	Toliman P, Badman SG, Gabuzzi J, Silim S, Forereme L, Kumbia A et al. Field Evaluation of Xpert HPV Point-of-Care Test for Detection of Human Papillomavirus Infection by Use of Self-Collected Vaginal and Clinician-Collected Cervical Specimens. <i>J Clin Microbiol</i> 2016; 54(7):1734-1737.
21 22 23 24 25	563 564 565 566	(44)	Cuschieri K, Geraets D, Cuzick J, Cadman L, Moore C, Vanden Broeck D et al. Performance of a Cartridge-Based Assay for Detection of Clinically Significant Human Papillomavirus (HPV) Infection: Lessons from VALGENT (Validation of HPV Genotyping Tests). <i>J Clin Microbiol</i> 2016; 54(9):2337-2342.
26 27 28 29	567 568 569	(45)	Kunckler M, Schumacher F, Kenfack B, Catarino R, Viviano M, Tincho E et al. Cervical cancer screening in a low-resource setting: a pilot study on an HPV-based screen-and-treat approach. <i>Cancer Med</i> 2017;10.
30 31 32 33	570 571 572	(46)	Guimaraes MD, Grinsztejn B, Melo VH, Rocha GM, Campos LN, Pilotto JH et al. Anal HPV prevalence and associated factors among HIV-seropositive men under antiretroviral treatment in Brazil. <i>J Acquir Immune Defic Syndr</i> 2011; 57 Suppl 3:S217-S224.
34 35 36 37	573 574	(47)	Castle PE, Schiffman M, Gravitt PE, Kendall H, Fishman S, Dong H et al. Comparisons of HPV DNA detection by MY09/11 PCR methods. <i>J Med Virol</i> 2002; 68(3):417-423.
38 39 40	575 576	(48)	Hayatsu H, Shiraishi M, Negishi K. Bisulfite modification for analysis of DNA methylation. <i>Curr Protoc Nucleic Acid Chem</i> 2008; Chapter 6:Unit.
41 42 43 44	577 578 579	(49)	Smith BC, McAndrew T, Chen Z, Harari A, Barris DM, Viswanathan S et al. The cervical microbiome over 7 years and a comparison of methodologies for its characterization. <i>PLoS One</i> 2012; 7(7):e40425.
45 46 47	580 581	(50)	Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. <i>Genome Biol</i> 2009; 10(3):R25.
48 49 50	582 583	(51)	Krueger F, Andrews SR. Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. <i>Bioinformatics</i> 2011; 27(11):1571-1572.
51 52 53 54 55 56 57 58	584 585	(52)	Qiao YL, Jeronimo J, Zhao FH, Schweizer J, Chen W, Valdez M et al. Lower cost strategies for triage of human papillomavirus DNA-positive women. <i>Int J Cancer</i> 2013.
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
3 4 5 6	586 587 588	(53)	Zhao FH, Jeronimo J, Qiao YL, Schweizer J, Chen W, Valdez M et al. An evaluation of novel, lower-cost molecular screening tests for human papillomavirus in rural China. <i>Cancer Prev Res (Phila)</i> 2013; 6(9):938-948.
7 8 9 10 11 12	589 590 591 592	(54)	Leisenring W, Alonzo T, Pepe MS. Comparisons of predictive values of binary medical diagnostic tests for paired designs. <i>Biometrics</i> 2000; 56(2):345-351.
$\begin{array}{c} 12\\ 13\\ 14\\ 15\\ 16\\ 17\\ 18\\ 19\\ 20\\ 21\\ 22\\ 32\\ 42\\ 52\\ 62\\ 7\\ 28\\ 29\\ 30\\ 31\\ 32\\ 33\\ 34\\ 35\\ 36\\ 37\\ 38\\ 39\\ 40\\ 41\\ 43\\ 44\\ 56\\ 47\\ 48\\ 49\\ 50\\ 51\\ 52\\ 53\\ 54\\ 55\end{array}$	593		
56 57 58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml
50			

Table 1: Recruitment sites and estimated eligible population.

Province	Site	Type of site	Potential participants p
			site (approximate)
Kigali	Cor-unum HC	Public Health Center	1,405
Kigali	Kimironko HC	Public Health Center	1,227
Kigali	Rwampara HC	Public Referral Hospital	1,098
Kigali	Kicukiro HC	Public Health Center	1,054
Kigali	Kacyiru HC	Public Health Center	905
Kigali	Gikondo HC	Public Health Center	925
Kigali	Rwanda Military Hospital	Public Referral Hospital	300
Kigali	WEACTx for Hope	Private HIV Clinic	500
Kigali	Busanza HC	Public Health Center	100
Kigali	Nyarugunga HC	Public Health Center	100
West	Gisenyi DH	Public District Hospital	810
Total			8,424
			1



BMJ Open	Pa
Appendix I. Study Questionnaire	
A. <u>Socio-demographics</u>	
1. What is your date of birth? <u>D</u> <u>D</u> / <u>M</u> <u>M</u> / <u>Y</u> <u>Y</u> <u>Y</u> (enter all 0 if not remembered) (If date of birth or year of birth is given, skip to Question 3)	
2. What is your age?(Years)	
3. What is your marital status?	
 Married/Cohabiting Divorced Widowed Separated Single Choose not to answer 	
4. Do you live in Kigali?	
 Yes No {Skip to Question A5} Choose not to answer {Skip to Question A5} 	
5. In what area of Kigali City Province do you live?	
 Nyarugenge Kicukiro Gasabo urban (e.g., Kacyiru, Kimironko, Kinyinya, etc.) Gasabo rural (e.g., Nduba, Jabana, Gikomero, etc.) Do not know Choose not to answer 	
6. What is your household income per month in FRW?	
$ < 5,000 \\ 5,000 - 9,999 \\ 10,000 - 34,999 \\ 35,000 - 59,999 $	

1 2	
2 3	
4	60,000 - 84,999
5	85,000 - 109,999
6 7	110,000 – 134,999
8	135,000 - 269,999
9 10	270,000 or more
11	
12 13	7. How many people live in your household? {Enter 00 if refuse to answer}
14	8. What is your occupation?
15 16	
17	Employed by government, another institution, or company
18 19	Self-employed (Small and medium enterprises)
20	Self-employed (High income earnings)
21	Farming (peasants)
22 23	Unemployed/Does not work
24	Other (specify)
25 26	
27	Choose not to answer {Skip to Question B1}
28	{The following questions are sensitive and personal in nature. Your answers will be kept
29 30	confidential. You may choose not to answer certain questions. Answering any question is
31	voluntary}
32 33	B. <u>Sexual Behaviors</u>
34	
35	1. How old were you when you first had sex? (Years) {Enter 00 if refuse to
36 37	answer}
38	2. How many sexual partners have you had in your lifetime?
39 40	
41	No Partners (Ineligible)
42	1 Partner
43 44	2-3 Partners
45	4-5 Partners
46 47	6-9 Partners
48	
49 50	10 or more partners
51	Choose not to answer
52	3. How many sexual partners have you had in the last 6 months?
53 54	
55	No Partners
56 57	
57 58	
59	
60	For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
13 14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24 25	
25 26	
20	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43 44	
44 45	
45 46	
40 47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	

1

1 Partner

2 or more Partners

Choose not to answer

C. Parity

1. What age did you have your first child? ____ (Years) {Enter 00 if refuse to answer, Enter 99 if never pregnant} (if 00 or 99, skip to Question D1)

2. How many live births have you had in your lifetime? _____

3. Have you given birth in the last year?

Yes

No

Choose not to answer

D. <u>Tobacco Use</u>

1. Have you ever smoked cigarettes?

Yes

No (skip to Question D3)

Choose not to answer (skip to Question D3)

2. Do you currently smoke cigarettes?

Yes

No

Choose not to answer

3. Have you ever chewed/used tobacco orally (Ubugoro)?

Yes

No (skip to Question D5)

Choose not to answer (skip to Question D5)

4. Are you currently chewing/using tobacco orally (Ubugoro)?

Yes

] No

Choose not to answer

5.	Have you ever chewed/used tobacco orally (Tobacco leaves-Igikamba)?
Y	es
N	o (skip to Question E1)
	noose not to answer (skip to Question E1)
6.	Are you currently chewing/using tobacco orally (Tobacco leaves-Igikamba)?
Y	es
N)
	noose not to answer
E.	Contraceptive Use
1.	Have you ever used oral contraceptives?
Y	es
N	o (Skip to Question E3)
	noose not to answer (Skip to Question E3)
2.	Do you currently use oral contraceptives?
Y	es
N	2
	noose not to answer
3.	Have you ever used Depo Provera (contraceptive)?
Y	es
N	o (Skip to Question E5)
	noose not to answer (Skip to Question E5)
4.	Do you currently use Depo Provera (contraceptive)?
Y	es
N)
	noose not to answer
5.	Have you ever used Jadell (contraceptive)?
$\Box \mathbf{v}$	es
1 1 1 1	

1	
2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
16 17	
18	
19	
20	
21	
22	
22 23	
24	
25	
26	
27	
28	
29	
30	
31 32	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

Choose not to answer (Skip to Question E7)
6. Do you currently use Jadell (contraceptive)?
Yes
No
Choose not to answer
7. Have you ever used IUD (contraceptive)?
Yes
No (Skip to Question E9)
Choose not to answer (Skip to Question E9)
8. Do you currently use IUD (contraceptive)?
☐ Yes
□ No
Choose not to answer
9. Do use condoms as a contraceptive?
Yes
□ No (skip to Question F1)
Choose not to answer (skip to Question F1)
10. Have you used condoms as a contraceptive in the last 6 months?
☐ Yes
□ No
Choose not to answer
F. <u>Infections</u>
1. Have you ever had Malaria
Yes
No (skip to Question F8)
Choose not to answer (skip to Question F8)
2. How many times have you had Malaria?

4.	Have you had your Malaria treated using drugs?
Yes	
No	
Cho	ose not to answer
5.	If Yes, how was it treated?
Trea	ted only with traditional medicine
Trea	tted only with drugs (e.g. Coartem)
Trea	ted with both (traditional and modern)
Not	treated
Cho	ose not to answer
6.	Was your last episode of Malaria treated using drugs?
Yes	
No	
Cho	ose not to answer
7.	If Yes, how was it treated?
Trea	tted only with traditional medicine
Trea	tted only with drugs (e.g. Coartem)
Trea	tted with both (traditional and modern)
Not	treated
Cho	ose not to answer
8.	Have you ever had Tuberculosis (TB)?
Yes	
🗌 No ((skip to end)
Cho	ose not to answer (skip to end)
9.	If Yes, how many times have you had TB in your lifetime?
1(). What year did you last have Tuberculosis (TB)? <u>Y</u> Y <u>Y</u>
	END OF QUESTIONNAIRE

BMJ Open

A Protocol for the Study of Cervical-Cancer Screening Technologies in Human Immunodeficiency Virus-Infected Women Living in Rwanda

Journal:	BMJ Open
Manuscript ID	bmjopen-2017-020432.R3
Article Type:	Protocol
Date Submitted by the Author:	19-Jun-2018
Complete List of Authors:	Murenzi, Gad; Rwanda Military Hospital Dusingize, Jean; Regional Alliance for Sustainable Development , Research and Scientific Capacity Building Rurangwa, Theogene; Rwanda Military Hospital Sinayobye, Jean d'Amour; Regional Alliance for Sustainable Development, ; Women's Equity in Access to Care & Treatment (WE-ACTx), Munyaneza, Athanase; Rwanda Military Hospital Murangwa, Anthere; Rwanda Military Hospital Zawadi, Thierry; Rwanda Military Hospital Hebert, Tiffany; Yeshiva University Albert Einstein College of Medicine Mugenzi, Pacifique; Rwanda Military Hospital Adedimeji, Adebola; Yeshiva University Albert Einstein College of Medicine Mutesa, Leon; Rwanda Military Hospital; University of Rwanda College of Medicine and Health Sciences Anastos, Kathryn; Albert Einstein College of Medicine Medical Center, Bronx, Castle, Philip; Yeshiva University Albert Einstein College of Medicine,
Primary Subject Heading :	Global health
Secondary Subject Heading:	Obstetrics and gynaecology, Infectious diseases, HIV/AIDS, Epidemiology, Diagnostics
Keywords:	human papillomavirus (HPV), cervical cancer, HIV & AIDS < INFECTIOUS DISEASES, GYNAECOLOGY, cervical intraepithelial neoplasia

SCHOLARONE[™] Manuscripts Page 1 of 35

BMJ Open

1		
2 3 4	1	A Protocol for the Study of Cervical-Cancer Screening Technologies in Human
5 6 7	2	Immunodeficiency Virus-Infected Women Living in Rwanda
8 9 10	3	Gad Murenzi, MD* ¹ , Jean-Claude Dusingize, MD, MS ¹ , Theogene Rurangwa, MD, MMed ¹ ,
11 12	4	Jean d'Amour Sinayobye, MD, MS ¹ , Athanase Munyaneza, RN ¹ , Anthere Murangwa, MS ¹ ,
13 14	5	Thierry Zawadi, MD ¹ , Tiffany Hebert, MD ² , Pacifique Mugenzi, MD ¹ , MMed, Adebola
15 16 17	6	Adedimeji, PhD, MPH ² , Leon Mutesa, MD, PhD ^{1,3} , Kathryn Anastos, MD ² , Philip E. Castle,
17 18 19 20	7	PhD, MPH ²
21 22	8	¹ Rwanda Military Hospital, Kigali, Rwanda; ² Albert Einstein College of Medicine, Bronx, NY,
23 24 25	9	USA; ³ University of Rwanda, Kigali, Rwanda
26 27 28	10	*Correspondence: <u>gadcollins@gmail.com</u> , +250788589085
29 30 31	11	Disclosures: This research study has received HPV tests for reduced or no cost from Cepheid,
32 33 34	12	Arbor Vita Corporation, and Roche.
35 36 37	13	Funding: This study was funded by NCI/NIH Grant 5U54CA19016304 and by a grant from the
38 39 40	14	Prevent Cancer Foundation.
41 42 43	15	Prevent Cancer Foundation.
44 45		
46 47		
48 49		
50 51		
52 53		
54 55		
56 57		
58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

16 Abstract

Introduction. The optimal method(s) for screening human immunodeficiency virus-infected
 women, especially for those living in sub-Saharan Africa, for cervical precancer and early cancer
 has yet to be established.

Methods and analysis. A convenience sample of >5,000 Rwandan women, ages 30-54 years and living with HIV infection, are being consented and enrolled into a cross-sectional study of cervical-cancer screening strategies. Participants are completing an administered short risk-factor questionnaire and being screened for high-risk human papillomavirus (hrHPV) using the Xpert HPV assay (Cepheid, Sunnyvale, CA, USA), unaided visual inspection after acetic acid (VIA), and aided VIA using the EVA system (Mobile ODT, Tel Aviv, Israel). Women positive for hrHPV and/or by VIA undergo colposcopy, which includes the collection of two cervical specimens prior to undergoing a 4-quadrant microbiopsy protocol. The colposcopy-collected specimens are being tested by dual immunocytochemical staining for p16^{INK4a} and Ki-67 (CINtec® PLUS Cytology, Ventana, Tucson, AZ, USA) and for E6 or E7 for 8 hrHPV genotypes (HPV16, 18, 31, 33, 35, 45, 52, and 58) using the next-generation AV Avantage hrHPV E6/E7 test (Arbor Vita Corporation, Freemont, CA, USA). Women with local pathology diagnosis of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) or pathology-review diagnosis of CIN grade 3 or more severe (CIN3+) receive treatment. Clinical performance and cost-effectiveness (e.g., sensitivity, specificity, and predictive values) of different screening strategies and algorithms will be evaluated.

- *Ethics and dissemination.* The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
 Ethics and dissemination. The protocol was approved by local and institutional review boards for
- 33 39 through an external advisory panel.

1 2 3	41	Strengths and Weaknesses
2 3 2 3 4 5 6 7 2 8 2 2 9 10 2 10 12 2 13 14 2 16 17 18 19 20 21 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 45 46 47 48 49 49	41 42 43 44 45 46 47 48 49 50	<text><list-item><list-item><list-item></list-item></list-item></list-item></text>
44 45 46 47 48		

51 Introduction

Invasive cervical cancer (ICC) remains a significant cause of morbidity and mortality globally. Approximately 530,000 cases of and 270,000 deaths due to ICC occur annually, making it the 4th most common malignancy and cause of cancer-related death in women globally.¹ In many high-income countries (HIC), ICC rates have declined by 50% or more² due to the introduction of effective, high-coverage Pap testing (cervical cytology)-based screening programs that include timely follow-up of screen positives, treatment of women with precursor lesions, and management of cancers. Now, almost 90% of ICC and ICC-related deaths occur in low- and middle income countries (LMICs) due to a lack of resources and healthcare infrastructure needed to provide preventive services.

ICC and ICC-related mortality rates are particularly high in Sub-Saharan Africa, which also has the highest rates of HIV infection in the world. Now, over 12 million HIV-infected (HIV[+]) women in Sub-Saharan Africa are living longer because of anti-retroviral therapy, thus increasing their likelihood of dying from ICC.³ However, many of these women are already exposed to human papillomavirus (HPV), the viral cause of ICC, and will not benefit from or be targeted for prophylactic HPV vaccination. Thus, cervical-cancer screening will be needed for the foreseeable future.

However, setting up effective cytology for cervical-cancer screening is expensive and requires a complex clinical and lab infrastructure that generally does not exist in LMICs.^{4;5} Moreover, it is now well understood that cytology has only a low- to moderate one-time sensitivity for precursor lesions and therefore must be done repeatedly over many years to reduce cancer risk. Alternative

For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

BMJ Open

2	
3	
4	
5	
6	
7	
/	
8	
9	
10	
11	
12	
13	
14	
15	
16	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
27 28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
50 51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

strategies to address the ICC burden in LMICs, especially in SSA, must be developed andvalidated.

74 Persistent cervical infections by high-risk HPV (hrHPV) types cause virtually all ICC and its 75 immediate precursor lesions, e.g. cervical intraepithelial neoplasia grade 3 (CIN3) and adenocarcinoma *in situ* (AIS) everywhere in the world.^{6;7} hrHPV causes most anal and vaginal 76 cancer and a significant proportion of vulvar, penile, and oropharyngeal cancers.⁸ HPV16 is the 77 most important causal type, responsible for ~60% of ICC.⁹ HPV18 is the next most important, 78 responsible for 10-15% of ICC, including 30-40% of adenocarcinoma of the cervix⁹, which is on 79 the rise in Western Countries.^{10;11} Together, HPV16 and HPV18 account for ~70% of ICC, and 80 the same 15 hrHPV types account for ~99% of ICC everywhere in the world.⁹ 81

There is now overwhelming evidence to suggest that testing for hrHPV is more sensitive, albeit less specific, than high-quality cytology for identifying women with cervical precancer.¹²⁻¹⁶ Onetime hrHPV testing can reduce the risk of ICC incidence by approximately 40% in 6.5 years compared to cytology screening¹⁶, and ICC mortality by approximately 40% (approximately 50% overall) in 8 years compared to cytology.¹⁷ Importantly, a negative hrHPV test provides superior reassurance against CIN3+ ¹⁸ and against ICC^{16;17}, permitting safe extension of screening intervals.

The World Health Organization released cervical-cancer screening and treatment guidelines in 2013, recommending two evidence-based approaches to cervical-cancer screening¹⁹: (I) Use either hrHPV testing or visual inspection after acetic acid (VIA), which involves the inspection of the cervix with a speculum in place and following the application of dilute acetic acid to help identify potential CIN by its characteristic white coloring in the presence of acetic acid

94 (acetowhite), as alternative initial screening tests instead of Pap, and (II) immediately treat those
95 who screen positive using the screening test, rather than require diagnostic verification through
96 colposcopy and biopsy. This approach is commonly called screen-and-treat (S&T), and is
97 increasingly thought to be more amenable to LMIC settings.
98 However, hrHPV testing is also a much more effective screen than VIA¹⁷, which on a large-scale

appears to only down-stage cancer rather than prevent it.²⁰ Thus, the recent American Society for
Clinical Oncology (ASC) resource-stratified guidelines for secondary cervical-cancer
prevention^{21;22} emphasize that hrHPV testing is the preferred choice for screening, with VIA
only being used until hrHPV testing becomes available, and that HIV-infected women, because
of their higher risk, should be screened twice as frequently as the general (HIV-uninfected)
population.

Recent data in HIV[+] women living in the U.S. suggest that hrHPV testing may have clinical utility similar to that in HIV-negative (HIV[-]) women. Several observational studies have shown that an extended screening interval is safe in HIV[+] women who test hrHPV and Pap negative as it is for HIV[-] women.^{23;24} In a study of women enrolled in Women's Interagency Health Study (WIHS) in 2002, HIV[+] and HIV[-] women who tested hrHPV and Pap negative were at a similarly low risk of cervical intraepithelial neoplasia (CIN) grade 2 (CIN2) or more severe (CIN2+) histology over a 5-year follow-up.²³ In addition, no cases of histologically confirmed CIN2+ were diagnosed in the follow-up of hrHPV- and Pap-negative HIV[+] women aged 30-64 years who underwent routine three-year hrHPV and cytology cotesting at Kaiser Permanente Northern California.²⁴ Thus, both studies found very high negative predictive values (NPV) >99% in HIV[+] women who test hrHPV negative. However, how hrHPV testing can

BMJ Open

2		
3 4	116	best be used to screen HIV[+] women living in Sub-Saharan Africa to prevent ICC remains to be
5 6 7	117	determined.
8 9 10	118	Recent studies ²⁵⁻²⁸ in HIV[+] women living in Sub-Saharan Africa have compared hrHPV, VIA,
11 12	119	and/or Pap for the detection of cervical precancer/cancer. The results can be summarized as
13 14 15	120	follows: 1) hrHPV detection was more sensitive but less specific than VIA; 2) surprisingly,
15 16 17	121	cytology was equally or more sensitive but less specific than VIA; and 3) surprisingly, cytology
18 19	122	was equally or more sensitive but less specific (vs. the converse) than hrHPV testing. Results and
20 21 22	123	conclusions are varied, leaving unanswered the question of what screening strategy in HIV[+]
22 23 24 25	124	women living in Sub-Saharan Africa has the greatest effectiveness and cost effectiveness.
25 26 27	125	Regardless of the screening method, most screen-positive women who go to colposcopy or are
28 29	126	treated immediately without diagnostic verification do not have cervical precancer and cancer
30 31 32	127	(positive predictive value [PPV] for screening tests are typically 10%-20%). In places like Sub-
33 34	128	Saharan Africa that lack necessary infrastructure and personnel such as pathologists ²⁹ , excessive
35 36	129	referral to colposcopy is problematic. Although WHO recommendations for S&T will hopefully
37 38 30	130	overcome this bottleneck and increase the number of women living in LMICs who get screened,
39 40 41	131	many countries may not adopt current S&T strategies because of concerns of low specificity and
42 43	132	overtreatment, resulting in increased costs, unnecessary patient discomfort and concern, and
44 45	133	wasting valuable healthcare resources that could otherwise be used to expand access to
46 47 48	134	screening. Thus, methods to increase the accuracy of screening by reducing the numbers of
49 50	135	women having colposcopy and biopsy or getting treated immediately in this context are highly
51 52 53 54 55 56	136	desirable as they will likely increase the uptake of cervical-cancer screening.

2
3
4
5
6
7
8
9
10
11
12 13
13 14
14 15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30 31
32
33
34
35
36
37
38
39
40
41
42
43
44 45
45 46
40 47
48
49
50
51
52
53
54
55
56
57
58
59 60
60

137	In order to improve the specificity of screening tests, secondary tests (biomarkers) are used
138	following a screen-positive result, with women who test positive for the triage undergoing
139	further management (e.g., colposcopy or immediate treatment) and those who test negative
140	typically being deferred to further evaluation in 6-18 months to allow hrHPV infections to clear.
141	There are several very promising biomarkers that might be used to improve the specificity and
142	positive predictive value (PPV) of the screening tests. ²⁴ Given that HIV-infected women are
143	more likely to test hrHPV positive ³⁰⁻³⁴ , it is important to validate a triage strategy of using a
144	secondary biomarker that sensitively and specifically rules-in women with cervical precancer
145	and cancer among the HIV-positive, hrHPV-positive women.
146	We are therefore conducting a cervical-cancer screening study of >5,000 Rwandan women, ages
147	30-54 years, living with HIV infection. We are evaluating different screening tests (hrHPV DNA
148	and VIA), those recommended by the WHO for cervical-cancer screening ¹⁹ , and different triage
149	tests and biomarkers for the management of screen-positive women (E6/E7 oncoprotein
150	detection, p16INK4a immunocytochemistry, and hrHPV viral methylation). Screen-positive
151	women undergo a rigorous colposcopic evaluation with multiple biopsies taken and the biopsies
152	will undergo pathology review, to minimize the misclassification of endpoints. The primary
153	objective of the study is to determine and compare clinical performance (Sensitivity (Se),
154	Specificity (Sp), PPV, and NPV) and cost-effectiveness for identifying HIV[+] women with
155	CIN3+ and CIN2+ of different cervical-cancer screening and management algorithms.
156	Methods and Analysis
157	Study design and population: We are recruiting those women receiving care for confirmed HIV
158	infection at health centers (HC) and various hospitals operated by the Ministry of Health or

Page 9 of 35

BMJ Open

2	
3	
4	
4 5	
4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 20 21 23 24 25 26 27 28 29 30	
6	
7	
8	
9	
10	
11	
12	
12	
13	
14	
15	
16	
17	
18	
19	
20	
20	
21	
22	
23	
24	
25	
26	
27	
28	
20	
29	
30	
וכ	
32 33 34 35	
33	
34	
35	
36	
36 37 38	
27	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
55	
54	
55	
56	
57	
58	
59	
60	
50	

159	Rwanda Military Hospital during 2016-18 (Table 1). Sites were selected in collaboration with
160	Rwanda Biomedical Center (RBC), which calculate estimated numbers of potentially eligible
161	women using data from their HIV database (OpenMRS-Open Medical Records System;
162	http://openmrs.org/). Sites were also selected from all provinces to ensure geographic
163	representation. The total of 8,424 was the estimated number of women eligible for the study
164	according to the inclusion/exclusion criteria (see below) at the beginning for the study in 2016.
165	From each site, a convenience sample of women are being recruited to participate.
166	Inclusion criteria include 1) living and receiving HIV care in Rwanda, 2) ages 30-54 years, 3)
167	confirmed HIV+ based on medical records, 4) no prior cervical-cancer screening, 5) no history of
168	ICC, and 6) willing, able and competent to provide written, informed consent. We are extending
169	age range beyond that of the age range (30-49 years) recommended by the WHO for cervical-
170	cancer screening ¹⁹ because there is limited evidence for the optimal upper age for cervical-cancer
171	screening of HIV-infected women. Exclusion criteria, in addition to not meeting the inclusion
172	criteria, include 1) pregnant, 2) signs of abnormal, non-menstrual bleeding suggestive of ICC, 3)
173	without a cervix due to hysterectomy, and 4) not sufficiently healthy to participate in a research
174	study based on the judgment of the clinicians. Excluded women are being advised to seek routine
175	cervical-cancer screening through government programs.
176	Prior to initiation of enrollment at a specific clinic, the local HIV-care provider team identifies
177	potentially eligible women at their routine clinic visits and offers them enrollment. Women
178	indicating interest in the study then are then registered by our research nurses using the eligibility
179	criteria checklist. All eligible and willing women at that site enroll and receive screening before

180 moving to another site. The study team of at least two research nurses schedules 12 to 15 women

181 three to four days a week by calling them and confirming appointments over the telephone. Two

teams of nurses are in the field, meaning that two cervical-cancer screening clinics can be runsimultaneously.

Enrollment Visit: The study participant flow is summarized in **Figure 1**. Enrollment visits, including pelvic exams with VIA and specimen collection, are being done entirely by a team of two study nurses. During their enrollment visit, women are being educated on cervical-cancer risk factors, mainly HPV infection, and why they are more at risk to develop ICC than HIVuninfected women. They also are being allowed to ask questions before they commit to participating in the study. Women are then being asked to provide informed, written consent to participate in the study using a printed out consent form. Those who provide consent complete a short nurse-administered questionnaire (Appendix I) on cervical cancer risk factors and sociodemographic characteristics using a data capture screen in Microsoft Access. The questionnaire collects information on basic sociodemographics, factors associated with acquiring HPV (e.g., marital status and recent and lifetime number of sexual partners), factors associated with increased risk of progression of hrHPV infection to precancer and cancer (e.g., smoking and other tobacco use, parity, and oral and other contraceptive use), and other infections common in Rwanda such as malaria³⁵ and tuberculosis³⁶ that have been previously reported to be associated with precancer among hrHPV-infected women. The questionnaire was not pretested. Enrolled women then undergo a pelvic exam, with VIA and a single cervical exfoliated ("Pap") specimen collected and placed into 20 ml PreservCyt (Hologic, Bedford, MA, USA) which is then being sent to the lab at RMH for hrHPV testing. Finally, a portable colposcope

202 (<u>http://www.mobileodt.com/;</u> MobleODT, Tel Aviv, Israel) is being used for digital

203 cervicography (comparable to VIA with magnification) and the image is being captured and

saved for quality control, research, and to develop a digital library.

Page 11 of 35

BMJ Open

2		
- 3 4	205	Colposcopy Visit: Screen-positive women (women who test hrHPV and/or VIA positive) are
5	206	being called using a telephone as soon as the hrHPV result is available and being invited to
/ 3 9	207	return for colposcopy within one month. All screen-positive women receiving colposcopy will
10 11	208	are having two additional specimens collected, one into PreservCyt for the evaluation of other
12 13	209	molecular biomarkers (genotype-specific hrHPV viral methylation and load, and p16/Ki-67
14 15 16	210	immunocytochemistry CINtec® PLUS Cytology Kit [Roche, Tucson, AZ, USA]) and a second
17 18	211	as a dry swab for HPV16, 18, 31, 33, 35, 45, 52, and 58 E6/E7 oncoprotein detection by the next
19 20	212	generation lateral flow hrHPV oncoprotein test from Arbor Vita Corporation (Fremont, CA,
21 22	213	USA) being included in this study as a triage for screen-positive women to identify those women
23 24 25	214	who are at higher risk of having CIN3+. The residual PreservCyt specimens from both the
26 27	215	screening and colposcopy visits are being stored at -20°C, creating a biobank in Rwanda for
28 29 30	216	future retrospective evaluations of promising new biomarkers and tests.
31 32 33	217	After specimen collection, a colposcopic evaluation of the cervix is being done with a modified
33 34 35	218	version of the 4-quadrant microbiopsy procedure being performed. ³⁷ Compared to the standard
36 37	219	biopsy, the microbiopsy protocol improves disease ascertainment and reduces biases related to
38 39	220	selecting the most visually obvious acetowhite lesions while removing less tissue ($\sim 13 \text{ mm}^2$ for 4
40 41 42	221	microbiopsies vs. ~28mm ² for 1 standard biopsy). Modifications to the standard 4-quadrant
43 44	222	microbiopsy procedure are: 1) endocervical curettage is being taken only for those women whose
45 46	223	squamocolumnar junction is not entirely visible and the lesion extends into the endocervical
47 48 49	224	canal; and 2) standard-size biopsies of very large lesions are being taken to increase the
50	225	likelihood that the most severe area is being biopsied.
51 52 53 54 55	226	Pathology: Biopsies are being processed in a single cassette so that a single slide has a section

from all biopsies taken. Biopsies are being read by a local pathologist at RMH and Dr. Hebert or

1	
2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 37 37 37 37 37 37 37 37 37 37	
5 4	
5	
6	
/ 8	
9	
10	
11	
12	2
14	
15	
16	
17 18	
19	4
20	
21	-
22 23	
24	
25	
26	
27 28	
29	
30	
31	4
32 33	
34	4
35	:
36	
37 38	
39	
40	
41	4
42 43	
44	
45	
46	
47 48	2
49	
50	2
51	
52 53	
55 54	
55	
56	
57 58	
58 59	
60	

228	another pathologist at Montefiore Medical Center, Bronx, NY, USA. Women receiving a
229	diagnosis of CIN2+ by the Rwandan pathologist (T.Z.) or, as a safety precaution, CIN3+
230	diagnosis by Montefiore pathologist (T.H.) are receiving treatment ³⁸ : 1) CIN2, CIN3, or AIS are
231	being referred to study doctors to undergo an excision procedure (e.g., loop electrosurgical
232	excision procedure [LEEP] or cold-knife cone [CKC]) and 2) ICC are being referred to RMH
233	Hospital for care. Women with <cin2 a="" advised="" are="" being="" in="" re-screening="" seek="" td="" to="" year.<=""></cin2>
234	A slide with biopsies also are undergoing p16 immunohistochemistry (IHC) using the CINtec®
235	Histology Kit (Roche) for research purposes only.
236	Endpoints: The primary scientific endpoints of the study are histologically confirmed, consensus
237	CIN2+ i.e., both pathologists diagnose CIN2+ (without adjudication) or CIN3+ by the study
238	pathologist. The secondary, clinical endpoint is histologically confirmed CIN2+ diagnosed by
239	the Rwandan pathologist. Additional endpoints using pathology review and p16 IHC are being
240	used but not for evaluating the performance of p16 immunocytochemistry due to the possibility
241	of p16-related autocorrelation.
242	<i>Treatment</i> : Women diagnosed with CIN2+ are being referred for treatment. Those precancerous
243	lesions are being treated by ablation if they meet WHO criteria for cryotherapy. ³⁹ Those who do
244	not meet those criteria undergo an excision procedure (e.g., loop electrosurgical excision
245	procedure [LEEP] or cold-knife cone [CKC]) or, in the case of an ICC diagnosis, referred for
246	cancer management. Screen-positive women with <cin2 advised="" are="" being="" re-screening<="" seek="" td="" to=""></cin2>
247	in a year through the existing healthcare system.
248	Data sources: Data are being collected from the following sources:

60

BMJ Open

1 2		
3 4	249	1. A nurse-administered questionnaire on sociodemographic characteristics and cervical
5 6	250	cancer risk factors including age at first sexual intercourse, number of sexual partners,
7 8 9	251	smoking, contraception, parity and socioeconomic status.
10 11 12 13	252	2. Pelvic exam, VIA, Mobile ODT and colposcopy data capture forms
14 15	253	3. Medical record data on HIV status (e.g.,
16 17	254	(http://www.who.int/hiv/pub/guidelines/HIVstaging150307.pdf), CD4 count, viral load,
18 19 20	255	antiretroviral therapy (ART) regimen(s)), care, and dates.
21 22 23 24	256	Laboratory Testing: The following laboratory tests are being performed:
25 26	257	Xpert HPV Testing—cervical Pap specimens in PreservCyt are being sent to the RMH
27 28 29	258	laboratory in Kigali, Rwanda for hrHPV DNA testing using the Xpert HPV test (Cepheid,
30 31	259	Sunnyvale, CA, USA). ⁴⁰⁻⁴⁵ The Xpert HPV Assay is a new, qualitative, real-time PCR assay for
32 33	260	the detection of hrHPV DNA. The Xpert HPV Assay includes simultaneous detection of 14
34 35 36	261	hrHPV types, hydroxymethylbilane synthase (HMBS), and an internal Probe Check Control
37 38	262	(PCC). The 14 targeted hrHPV types are detected in 5 fluorescent channels: 1) HPV16, 2)
39 40	263	HPV18 and hrHPV 45 (HPV18/45), 3) HPV31, 33, 35, 52, and 58, 4) HPV51 and HPV59, and
41 42 43	264	5) HPV39, 56, 66, and 68. HMBS (fluorescent channel 6) verifies specimen adequacy.
44 45 46	265	Specimens are being mixed and a 1-mL pre-aliquot is being removed using a disposable pipette
47 48	266	and placed in the testing cartridge per the manufacturer's instructions. Unsatisfactory results due
49 50	267	to insufficient cellular content are being re-tested. If the second test is also unsatisfactory, the
51 52 53	268	final result are being recorded as unsatisfactory but women with unsatisfactory results are being
54 55 56 57 58	269	referred to colposcopy for safety.

HPV Viral Methylation—We will conduct a retrospective analysis of HPV viral methylation and its association with CIN2+. To identify single hrHPV type infections, we will select singlechannel positives from the Xpert HPV assay. For those that are hrHPV positive for a channel other than HPV16, which is detected singly, we will test them to identify the single type infections using a standard protocol for PCR amplification using MY09/11 L1 consensus primers and hrHPV genotype detection using dot-hybridization for 39 individual type-specific probes and a mixture of probes for 10 other uncommon hrHPV types as previously described.^{46;47} To isolate the DNA, ThinPrep specimens (1.5 mL) will be pelleted, re-suspended in STM, digested with Proteinase K, precipitated overnight in ammonium acetate ethanol at -20°C, washed, and suspended and stored in TE buffer. The isolated DNA then will undergo bisulfite conversion.⁴⁸ Following bisulfite conversion and DNA purification and de-sulphonation, bisulfite-treated DNA will be used as template for Next-Gen Sequencing (NGS) (HiSeq2000, Illumina, San Diego, CA) using barcoded-type specific primers. Sequences for pads and barcodes are not found in the targeted genomic region. Use of padding and barcodes will enable reads to be identified by amplicon (forward or reverse) or by sample during downstream bioinformatics analysis.⁴⁹ All PCR products for NGS will be pooled (by assay) and a single DNA band containing multiple amplicons from different samples (with unique barcodes) will be isolated from a gel for NGS.⁴⁹

288 Briefly, equal concentrations of each barcoded PCR product (based on PCR band intensity) will

289 be pooled and isolated. Upon confirmation of correct product size, all purified DNA pools will

be combined and submitted for library preparation and paired-end 100 base pair Illumina

HiSeq2000 sequencing at the Einstein Genomics Core Facility.

Page 15 of 35

BMJ Open

2	
2 3 4 5 6 7	2
4 5	
6	2
7 8	-
9	4
10 11	2
12	
13 14	4
9 10 11 12 13 14 15 16 17 18 19 20	2
16 17	
18	4
19 20	
21	4
22 23	3
21 22 23 24 25 26	
25 26	Ċ
27 28	3
27 28 29 30 31 32 33 34 35 36	-
30 31	
32	
33 34	
35	2
36 37 38	
	÷
39 40	
41 42	
42 43	
44 45	2
46	
47 48	
49	
50 51	3
52	
53 54	
55	
56 57	
58	
59 60	

Methylation status are being determined in the lab of Dr. Robert Burk at Albert Einstein College of Medicine (USA). Prior to determination of methylation status, de-multiplexing based on the unique barcodes is being performed using in-house generated scripts to obtain paired-end NGS reads of each sample. Reads are being aligned with hrHPV reference genome sequences by bowtie v0.12.9.⁵⁰ Methylation status of each CpG site is then determined by bismark v0.7.7⁵¹ using the default quality score parameter set to Q30, and the formula of the methylation ratio of the number of C read by the number of C+T read.

E6/E7 Oncoprotein Testing—Dry swab specimens, collected at the time of colposcopy, are being
 tested for individual E6/E7 oncoproteins as previously described^{52;53}, according to the
 manufacturer's instructions, at the RMH laboratory in Kigali, Rwanda. The E6/E7 oncoprotein
 test uses three lateral flow strips to detect 8 hrHPV types whereas the E6 oncoprotein test used a
 single lateral flow strip to detect 3 hrHPV types.

Analyses: We will evaluate combinations of the above mentioned screening strategies and tests to estimate the clinical performance (e.g., Se, Sp, PPV, and NPV) for the detection of consensus CIN3+ and community CIN2+. A log binomial model using generalized estimating equations will be used to take into account correlation between different tests from the same subject. Note while these models will be developed for the estimation and comparison of performance for two tests, the model can be extended to allow more than two tests by including more indicator variables for test type.

Some analyses of biomarkers, such as viral methylation will be restricted to hrHPV-positives.
Comparisons of hrHPV viral methylation to other triage biomarkers will be restricted to the
subset that gets tested for viral methylation as described.

1	
2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
27	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
40 41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
55 54	
54 57	
55	
56	
57	
58	
59	

60

314	Sample size calculations: We are basing our sample size on the ability to detect modest but
315	minimally important differences in Se of 15%. We conservatively assume that the population
316	prevalence of CIN3+ is 2% based on our past study in Rwanda. ^{33;35} We will enroll and have
317	complete follow-up of at least 5,000 HIV[+] women. A sample size of 5,000 HIV[+] women
318	with completed follow-up of the screen positives will yield 100 cases of CIN3+, which will have
319	at least 80% power (α =5%) to crudely detect a 15% difference in Se between two screening
320	strategies for a range of 10%-25% discordance. With this sample size of 5,000 women, 4,900
321	will not have CIN3+; we will also have at least 90% power (α =5%) to detect a difference in Sp
322	of 3% for discordance up to 40%. Finally, we will have 80% power (α =5%) to crudely detect an
323	8%, 10%, or 11% difference in PPV if the reference PPV is 10%, 20%, or 30%, respectively. ⁵⁴
324	Cost Effectiveness: We will conduct assessments of the costs and cost-effectiveness of the
325	different combinations of screening and triage tests, i.e., algorithms, as well as those of the entire
326	community-based screening "system." Costs measurement will be conducted using a micro-
327	costing (ingredients) approach in which resource use throughout each step in the screening
328	process will be tracked and unit costs for each of the resources will be applied to generate an
329	average screening cost per woman to be compared against what the estimated costs are for a
330	possible program based on hrHPV screening and VIA triage or VIA screening. For estimating
331	costs of the screening system and scale-up of screening to 100,000 women in a month, analyses
332	will distinguish financial costs, which reflect actual expenditures of the program, from economic
333	costs, including the value of donated and shared resources to more fully assess opportunity costs.
334	Projections on budget impact and economic cost implications over time will be made under
335	varying assumptions of screening uptake, follow-up compliance, and scenarios of changing
336	disease burden.

Page 17 of 35

BMJ Open

Clinical outcomes will include true positive, true negative, false negative, and false positive test results, number of colposcopies, incident cancer, and cancer death. Cost-effectiveness will be measured as cost/CIN2+ detected, cost/CIN3+ detected, cost/invasive cancer prevented, cost/cancer death prevented, cost/life-year saved, and cost/quality-adjusted life year (QALY) saved: in addition, we will calculate harm/benefit ratios, using varying definitions of harms (colposcopies, false positive results) to benefits (cancers prevented, deaths prevented, life years and QALYs saved). Costs and effectiveness will be discounted at a 3% annual rate, with the rate varied from 0-5% in sensitivity analysis. For assessment of value-of-information (VOI), we will use net monetary benefits (NMB), defined as a function of the willingness-to-pay threshold (WTP) for different costs and outcomes as: NMB = (WTP * Effectiveness) - Costs.

Patient and Public Involvement

There was no patient engagement in the development or design of the study, recruitment, and the conduct of the study. Participants are receiving their results directly since it is related to their care. As this was not a randomized controlled trial, the burden of the intervention was not assessed by patients themselves. There were no patient advisors to acknowledge.

352 Ethics and Dissemination

Ethics: This study protocol was reviewed and approved by the Rwanda National Ethics
Committee (RNEC) as well as the Institutional Review Board for human subjects research at
Albert Einstein College of Medicine.

Confidentiality measures and protection against potential risks: The risks for those participating
357 in our study include:

1 2

2 3 4	250	Collection of Dan anazimana/comviced awaha involves a modest risk of blooding
	358	Collection of Pap specimens/cervical swabs involves a modest risk of bleeding
5 6	359	which is typically very limited when it occurs. Testing positive for any test may
7 8	360	cause psychological distress (anxiety).
9	500	eause psychological distress (anxiety).
10 11	361	• Colposcopy and excisional treatments induce vaginal bleeding and may incur pain,
12	262	
13 14	362	infection, and short-term psychological distress (anxiety). A diagnosis of CIN2 or
15 16	363	more severe may cause psychological distress (anxiety). A diagnosis of ICC may
17	364	cause severe psychological distress.
18 19	501	eduse severe psychological distress.
20 21	365	• Questions in the questionnaire, regarding sexual behavior and other matters of a
22	200	nerconal nature, may appear any intra and emberragement. Participants are advised
23 24	366	personal nature, may cause anxiety and embarrassment. Participants are advised
25 26	367	that they are free not to answer specific questions.
27		
28 29	368	• There is also the risk of psycho-social stress which could occur if there was
30 31	369	inadvertent disclosure of confidential medical or other personal information.
32		
33 34	370	Protection against the risk of inadvertent disclosure of confidential information is being
35	371	addressed by the standard procedures at the Rwandan study site, including: (i) storing completed
36 37		
38 39	372	paper copies of questionnaires and other hard copy information (described above), identified by
40	373	study number only, in a filing system separate from the name-address file of participants in the
41 42	374	study; and (ii) only the designated local personnel have access to cross-reference the files; (iii)
43 44	571	
45	375	all paper files, including consent forms, are being maintained in locked cabinets in locked rooms,
46 47	376	with access restricted to specific research personnel.
48 49		
50	377	In addition, we will include the following security measures to protect the data:
51 52	077	
53 54	378	• Controlled access to project data;
55	570	• Controlled access to project data,
56 57		
58		
59 60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

60

BMJ Open

379	• A tracking system for data forms and activities;
380	• Date and time of stamping of all data records with electronic signatures;
381	• Audit trails to track all changes made to data records; and
382	• Data kept on password-protected computers and in locked rooms.
383	Potential Patient Benefits: There are potential direct benefits to study participants. HIV[+]
384	women participating in the study are at very high risk of ICC. They are being rigorously
385	screened and evaluated, more effectively than the standard of care anywhere in the world. As a
386	result of the study, women with precancer who are at imminent risk of ICC are being diagnosed
387	sooner and treated more effectively than women receiving routine care and thereby more likely
388	averting the development of ICC. Women with cervical cancer are being diagnosed earlier
389	thereby reducing the morbidity and the risk of mortality caused by ICC. Conversely, any pain,
390	bleeding, or stress that might occur related to colposcopy or cervical swab are typically modest
391	and well tolerated.
302	There are also substantial potential societal benefits related to the study due to its implications to
393	improving cervical-cancer screening practices and guidelines in HIV[+] women – changes in
394	practice which might also benefit the study participants themselves, if and when these changes
395	are enacted. There is a great need to identify more effective and practical methods for cervical-
396	cancer screening for HIV[+] women living in Africa, who are living longer than ever and are
397	therefore at potentially greater risk of ICC.
398	Dissemination: We plan to publish a series of scientific reports in peer-reviewed scientific
399	journals. As building research capacity in Rwanda is a major goal of this research project, all
400	investigators of the research team are being asked and supported to lead at least one analysis and
401	one manuscript preparation, based on interests and expertise.
	 380 381 382 383 384 385 386 387 388 389 390 391 392 393 394 395 396 397 398 397 398 397 398 397 398 397 398 397 398 399 400

In addition, an external advisory panel (EAP) composed of leaders from the Rwanda Ministry of
Health, University of Rwanda, and Rwanda medical community has been formed. The
responsibilities of the EAP include providing advice on the conduct of the project and
interpretation for and dissemination of the study results to Rwandan stakeholders. The latter is
important for the adoption of evidence-based best practices for cervical-cancer screening as
warranted.

408 Limitations

There are several limitations to the study that bear mentioning. First, cervical cytology is not being included in the study. There is limited cervical cytology services available locally and of unknown quality and it is unlikely that cytology will be widely available in Rwanda, making its inclusion as a comparator test of limited value. Moreover, there are significant costs and logistical challenges in shipping PreservCyt specimens to and having cytology slides made and read in the U.S. Second, we are not conducting colposcopy and taking biopsies in screen-negative women, which would have allowed us to estimate absolute clinical performance. The burden of sending screen-negative women to colposcopy is deemed too great and it is impractical to send a sufficient numbers of screen-negative women to colposcopy to accurately estimate the false-negative disease (CIN3+) fraction. Thus, only relative clinical performance of the screening tests will be estimated from this study.

BMJ Open

KA, PEC, JCD, AA, and JDS conceived the original concept of the study and the interventions.

GM, KA, JDS, and PEC drafted the protocol. PEC performed the sample size calculations, and

supporting patient recruitment. GM, TR, and Athanase Munyaneza are providing clinical care for

patients. Anthere Murangwa and LM oversee laboratory testing, and TZ and TH are responsible

Rwanda. All authors (GM, JCD, TR, JDS, Athanase Munyaneza, Anthere Murangwa, TZ, TH,

PM, AA, LM, KA, and PEC) contributed to the scientific design of the study and the protocol

development, are involved in the implementation of the project, and have read and approved the

VOIV

for pathology. PM and LM oversee and administer the study activities at the clinical site in

PEC and GM will lead analysis of the results. GM, JDS, and Athanase Munyaneza are

Contributor Statement:

final manuscript.

1		
2 3	432	Figure Legends:
2 3 4 5 6	433	
6 7 8	434	Figure 1. Study Design
8 9 10	435	
10 11 12	100	
12 13 14		
15 16		
17 18		
19 20		
21 22 23		
24 25		
26 27		
28 29		
30 31 32		
33 34		
35 36		
37 38 39		
40 41		
42 43		
44 45 46		
40 47 48		
49 50		
51 52		
53 54		
55 56 57		
57 58 59		
60		For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
2 3	436	Refere	nce List
4	150	Refere	
5 6	437		
7	438	(1)	Ferlay J, Soerjomataram I, Ervik M, Dikshit R, Eser S, Mathers C et al. GLOBOCAN 2012 v1.0,
8	439	ĊĴ	Cancer Incidence and Mortality Worldwide: IARC CancerBase No. 11 [Internet]. 2013. Lyon,
9 10	440		France, International Agency for Research on Cancer. 4-22-2014.
11	441	Ref Ty	pe: Online Source
12			
13	442	(2)	Lonnberg S, Hansen BT, Haldorsen T, Campbell S, Schee K, Nygard M. Cervical cancer
14	443		prevented by screening: Long-term incidence trends by morphology in Norway. Int J Cancer
15	444		2015;10.
16	445	(2)	
17	445		UNAIDS. UNAIDS Data 2017. 2017.
18 19	446	Refity	pe: Report
20	447	(4)	Cervix Cancer Screening. [10]. 2005. IARC Press. IARC Handbooks of Cancer Prevention.
21	448		pe: Serial (Book,Monograph)
22	110	ner ry	
23	449	(5)	Kitchener HC, Castle PE, Cox JT. Chapter 7: Achievements and limitations of cervical
24	450		cytology screening. Vaccine 2006; 24 Suppl 3:S63-70.:S63-S70.
25			
26 27	451	(6)	Schiffman M, Castle PE, Jeronimo J, Rodriguez AC, Wacholder S. Human papillomavirus and
27 28	452		cervical cancer. <i>Lancet</i> 2007; 370(9590):890-907.
29			
30	453	(7)	Schiffman M, Clifford G, Buonaguro FM. Classification of weakly carcinogenic human
31	454		papillomavirus types: addressing the limits of epidemiology at the borderline. <i>Infect Agent</i>
32	455		<i>Cancer</i> 2009; 4:8.
33	456	(8)	Forman D, de MC, Lacey CJ, Soerjomataram I, Lortet-Tieulent J, Bruni L et al. Global burden
34	450 457	(0)	of human papillomavirus and related diseases. <i>Vaccine</i> 2012; 30 Suppl 5:F12-23. doi:
35 36	458		10.1016/j.vaccine.2012.07.055.:F12-F23.
37	430		10.1010/j.vaccinc.2012.07.055112-125.
38	459	(9)	de SS, Quint WG, Alemany L, Geraets DT, Klaustermeier JE, Lloveras B et al. Human
39	460	(1)	papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-
40	461		sectional worldwide study. <i>Lancet Oncol</i> 2010; 11(11):1048-1056.
41			
42	462	(10)	Bray F, Carstensen B, Moller H, Zappa M, Zakelj MP, Lawrence G et al. Incidence trends of
43	463		adenocarcinoma of the cervix in 13 European countries. Cancer Epidemiol Biomarkers Prev
44 45	464		2005; 14(9):2191-2199.
46		(1.1)	
47	465	(11)	Adegoke O, Kulasingam S, Virnig B. Cervical cancer trends in the United States: a 35-year
48	466		population-based analysis. J Womens Health (Larchmt) 2012; 21(10):1031-1037.
49	467	(12)	Cuzick J, Clavel C, Petry KU, Meijer CJ, Hoyer H, Ratnam S et al. Overview of the European
50	467	(12)	and North American studies on HPV testing in primary cervical cancer screening. Int J
51	469		<i>Cancer</i> 2006; 119(5):1095-1101.
52	409		Cancer 2000, 117(5).1075-1101.
53 54	470	(13)	Naucler P, Ryd W, Tornberg S, Strand A, Wadell G, Elfgren K et al. Human papillomavirus
55	471	()	and Papanicolaou tests to screen for cervical cancer. N Engl J Med 2007; 357(16):1589-
56	472		1597.
57			
58			
59			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml
60			i or peer review only - http://binjopen.binj.com/site/about/guidennes.xhtml

1 2									
2 3 4 5	473 474 475	(14)	Ronco G, Giorgi-Rossi P, Carozzi F, Confortini M, Palma PD, Del MA et al. Efficacy of human papillomavirus testing for the detection of invasive cervical cancers and cervical intraepithelial neoplasia: a randomised controlled trial. <i>Lancet Oncol</i> 2010.						
6 7 8 9 10	476 477 478	(15)	Rijkaart DC, Berkhof J, Rozendaal L, van Kemenade FJ, Bulkmans NW, Heideman DA et al. Human papillomavirus testing for the detection of high-grade cervical intraepithelial neoplasia and cancer: final results of the POBASCAM randomised controlled trial. <i>Lancet</i>						
10 11 12	479		Oncol 2012; 13(1):78-88.						
13 14 15 16	480 481 482	(16)	Ronco G, Dillner J, Elfstrom KM, Tunesi S, Snijders PJ, Arbyn M et al. Efficacy of HPV-based screening for prevention of invasive cervical cancer: follow-up of four European randomised controlled trials. <i>Lancet</i> 2013.						
17 18 19	483 484	(17)	Sankaranarayanan R, Nene BM, Shastri SS, Jayant K, Muwonge R, Budukh AM et al. HPV screening for cervical cancer in rural India. <i>N Engl J Med</i> 2009; 360(14):1385-1394.						
20 21 22 23	485 486 487	(18)	Dillner J, Rebolj M, Birembaut P, Petry KU, Szarewski A, Munk C et al. Long term predictive values of cytology and human papillomavirus testing in cervical cancer screening: joint European cohort study. <i>BMJ</i> 2008; 337:a1754. doi: 10.1136/bmj.a1754.:a1754.						
24 25	488	(19)	New guidelines on screening and treatment for cervical cancer. 2013. South Africa, World						
26	489 490	Ref Tv	Health Organization.						
27 28	170	ner ry	ef Type: Pamphlet						
29	491	(20)	Shastri SS, Mittra I, Mishra GA, Gupta S, Dikshit R, Singh S et al. Effect of VIA screening by						
30 31 32	492 493		primary health workers: randomized controlled study in Mumbai, India. <i>J Natl Cancer Inst</i> 2014; 106(3):dju009.						
33	494	(21)	Jeronimo J, Castle PE, Temin S, Shastri SS. Secondary Prevention of Cervical Cancer:						
34 35 36	495 496		American Society of Clinical Oncology Resource-Stratified Clinical Practice Guideline Summary. <i>J Oncol Pract</i> 2016; JOP2016017889.						
37	497	(22)	Castle PE, Jeronimo J, Temin S, Shastri SS. Screening to Prevent Invasive Cervical Cancer:						
38 39	498		ASCO Resource-Stratified Clinical Practice Guideline. <i>J Clin Oncol</i> 2017; 35(11):1250-1252.						
40	499	(23)	Keller MJ, Burk RD, Xie X, Anastos K, Massad LS, Minkoff H et al. Risk of cervical precancer						
41 42 43	500 501		and cancer among HIV-infected women with normal cervical cytology and no evidence of oncogenic HPV infection. <i>JAMA</i> 2012; 308(4):362-369.						
44 45 46 47	502 503 504	(24)	Castle PE, Fetterman B, Poitras N, Lorey T, Kinney W. Safety against cervical precancer and cancer following negative human papillomavirus and Papanicolaou test results in human immunodeficiency virus-infected women. <i>Arch Intern Med</i> 2012; 172(13):1041-1043.						
48 49 50 51	505 506 507	(25)	Chung MH, McKenzie KP, De VH, Richardson BA, Rana F, Pamnani R et al. Comparing pap smear, via, and hpv cervical cancer screening methods among hiv-positive women by immune status and antiretroviral therapy. <i>AIDS</i> 2013.						
52 53 54 55 56 57	508 509 510	(26)	Firnhaber C, Mayisela N, Mao L, Williams S, Swarts A, Faesen M et al. Validation of cervical cancer screening methods in HIV positive women from Johannesburg South Africa. <i>PLoS One</i> 2013; 8(1):e53494.						
58									
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml						

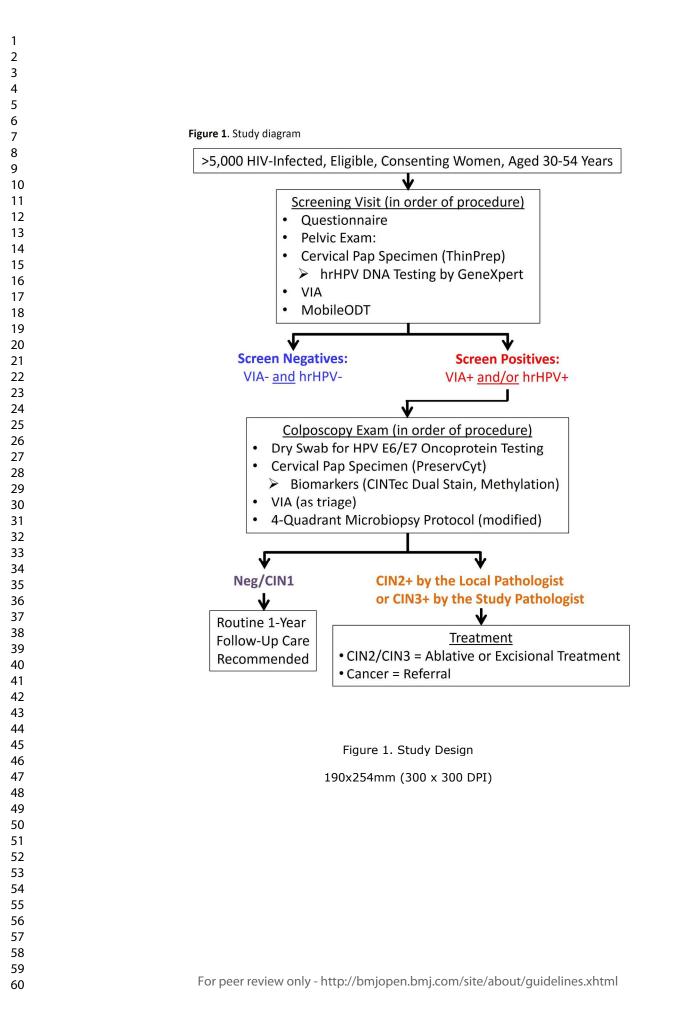
1 2			
3 4 5 7 8 9 10 11	511 512 513	(27)	Dartell MA, Rasch V, Iftner T, Kahesa C, Mwaiselage JD, Junge J et al. Performance of visual inspection with acetic acid and human papillomavirus testing for detection of high-grade cervical lesions in HIV positive and HIV negative Tanzanian women. <i>Int J Cancer</i> 2014;10.
	514 515 516 517	(28)	Mabeya H, Khozaim K, Liu T, Orango O, Chumba D, Pisharodi L et al. Comparison of conventional cervical cytology versus visual inspection with acetic acid among human immunodeficiency virus-infected women in Western Kenya. <i>J Low Genit Tract Dis</i> 2012; 16(2):92-97.
12 13 14	518 519	(29)	Adesina A, Chumba D, Nelson AM, Orem J, Roberts DJ, Wabinga H et al. Improvement of pathology in sub-Saharan Africa. <i>Lancet Oncol</i> 2013; 14(4):e152-e157.
15 16 17 18 19	520 521 522	(30)	Dartell M, Rasch V, Kahesa C, Mwaiselage J, Ngoma T, Junge J et al. Human papillomavirus prevalence and type distribution in 3603 HIV-positive and HIV-negative women in the general population of Tanzania: the PROTECT study. <i>Sex Transm Dis</i> 2012; 39(3):201-208.
20 21 22 23 24	523 524 525 526	(31)	D'Souza G, Burk RD, Zhong Y, Minkoff H, Massad LS, Xue X et al. Cervicovaginal human papillomavirus (HPV)-infection before and after hysterectomy: evidence of different tissue tropism for oncogenic and nononcogenic HPV types in a cohort of HIV-positive and HIV-negative women. <i>Int J Cancer</i> 2012; 131(6):1472-1478.
25 26 27 28	527 528 529	(32)	Ng'andwe C, Lowe JJ, Richards PJ, Hause L, Wood C, Angeletti PC. The distribution of sexually-transmitted Human Papillomaviruses in HIV positive and negative patients in Zambia, Africa. <i>BMC Infect Dis</i> 2007; 7:77.:77.
 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 	530 531 532	(33)	Singh DK, Anastos K, Hoover DR, Burk RD, Shi Q, Ngendahayo L et al. Human papillomavirus infection and cervical cytology in HIV-infected and HIV-uninfected Rwandan women. <i>J Infect Dis</i> 2009; 199(12):1851-1861.
	533 534 535	(34)	Marais DJ, Vardas E, Ramjee G, Allan B, Kay P, Rose RC et al. The impact of human immunodeficiency virus type 1 status on human papillomavirus (HPV) prevalence and HPV antibodies in serum and cervical secretions. <i>J Infect Dis</i> 2000; 182(4):1239-1242.
	536 537 538	(35)	Anastos K, Hoover DR, Burk RD, Cajigas A, Shi Q, Singh DK et al. Risk factors for cervical precancer and cancer in HIV-infected, HPV-positive Rwandan women. <i>PLoS One</i> 2010; %20;5(10):e13525.
	539 540 541	(36)	Zhao FH, Varanasi AP, Cunningham CA, Graubard BI, Hu SY, Chen F et al. Tuberculosis and oncogenic HPV: potential co-infections in women at high-risk of cervical cancer in rural China. <i>Asian Pac J Cancer Prev</i> 2011; 12(6):1409-1415.
	542 543 544	(37)	Pretorius RG, Zhang WH, Belinson JL, Huang MN, Wu LY, Zhang X et al. Colposcopically directed biopsy, random cervical biopsy, and endocervical curettage in the diagnosis of cervical intraepithelial neoplasia II or worse. <i>Am J Obstet Gynecol</i> 2004; 191(2):430-434.
	545 546	(38)	Schiffman M, Adrianza ME. ASCUS-LSIL Triage Study. Design, methods and characteristics of trial participants. <i>Acta Cytol</i> 2000; 44(5):726-742.
53 54 55 56 57	547 548 549	(39) neopla	World Health Organization. WHO guidelines for treatment of cervical intraepithelial isia 2-3 and adenocarcinoma in situ. World Health Organization [2014 Available from: URL: <u>http://apps.who.int/iris/bitstream/10665/104174/1/9789241506779_eng.pdf</u>
58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

3 4 5 6	550 551 552	(40)	Castle PE, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Reliability of the Xpert HPV assay to detect high-risk human papillomavirus DNA in a colposcopy referral population. <i>Am J Clin Pathol</i> 2015; 143(1):126-133.
7 8 9 10	553 554 555	(41)	Cuzick J, Cuschieri K, Denton K, Hopkins M, Thorat MA, Wright C et al. Performance of the Xpert HPV assay in women attending for cervical screening. <i>Papillomavirus Research</i> 2015; 1:32-37.
11 12 13 14 15	556 557 558	(42)	Einstein MH, Smith KM, Davis TE, Schmeler KM, Ferris DG, Savage AH et al. Clinical Evaluation of the Cartridge-Based GeneXpert Human Papillomavirus Assay in Women Referred for Colposcopy. <i>J Clin Microbiol</i> 2014.
16 17 18 19 20	559 560 561 562	(43)	Toliman P, Badman SG, Gabuzzi J, Silim S, Forereme L, Kumbia A et al. Field Evaluation of Xpert HPV Point-of-Care Test for Detection of Human Papillomavirus Infection by Use of Self-Collected Vaginal and Clinician-Collected Cervical Specimens. <i>J Clin Microbiol</i> 2016; 54(7):1734-1737.
21 22 23 24 25	563 564 565 566	(44)	Cuschieri K, Geraets D, Cuzick J, Cadman L, Moore C, Vanden Broeck D et al. Performance of a Cartridge-Based Assay for Detection of Clinically Significant Human Papillomavirus (HPV) Infection: Lessons from VALGENT (Validation of HPV Genotyping Tests). <i>J Clin Microbiol</i> 2016; 54(9):2337-2342.
26 27 28 29	567 568 569	(45)	Kunckler M, Schumacher F, Kenfack B, Catarino R, Viviano M, Tincho E et al. Cervical cancer screening in a low-resource setting: a pilot study on an HPV-based screen-and-treat approach. <i>Cancer Med</i> 2017;10.
30 31 32 33	570 571 572	(46)	Guimaraes MD, Grinsztejn B, Melo VH, Rocha GM, Campos LN, Pilotto JH et al. Anal HPV prevalence and associated factors among HIV-seropositive men under antiretroviral treatment in Brazil. <i>J Acquir Immune Defic Syndr</i> 2011; 57 Suppl 3:S217-S224.
34 35 36 37	573 574	(47)	Castle PE, Schiffman M, Gravitt PE, Kendall H, Fishman S, Dong H et al. Comparisons of HPV DNA detection by MY09/11 PCR methods. <i>J Med Virol</i> 2002; 68(3):417-423.
38 39 40	575 576	(48)	Hayatsu H, Shiraishi M, Negishi K. Bisulfite modification for analysis of DNA methylation. <i>Curr Protoc Nucleic Acid Chem</i> 2008; Chapter 6:Unit.
41 42 43 44	577 578 579	(49)	Smith BC, McAndrew T, Chen Z, Harari A, Barris DM, Viswanathan S et al. The cervical microbiome over 7 years and a comparison of methodologies for its characterization. <i>PLoS One</i> 2012; 7(7):e40425.
45 46 47	580 581	(50)	Langmead B, Trapnell C, Pop M, Salzberg SL. Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. <i>Genome Biol</i> 2009; 10(3):R25.
48 49 50	582 583	(51)	Krueger F, Andrews SR. Bismark: a flexible aligner and methylation caller for Bisulfite-Seq applications. <i>Bioinformatics</i> 2011; 27(11):1571-1572.
51 52 53 54 55 56 57 58	584 585	(52)	Qiao YL, Jeronimo J, Zhao FH, Schweizer J, Chen W, Valdez M et al. Lower cost strategies for triage of human papillomavirus DNA-positive women. <i>Int J Cancer</i> 2013.
59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

1 2			
3 4 5 6	586 587 588	(53)	Zhao FH, Jeronimo J, Qiao YL, Schweizer J, Chen W, Valdez M et al. An evaluation of novel, lower-cost molecular screening tests for human papillomavirus in rural China. <i>Cancer Prev Res (Phila)</i> 2013; 6(9):938-948.
7 8 9 10 11 12	589 590 591 592	(54)	Leisenring W, Alonzo T, Pepe MS. Comparisons of predictive values of binary medical diagnostic tests for paired designs. <i>Biometrics</i> 2000; 56(2):345-351.
$\begin{array}{c} 12\\ 13\\ 14\\ 15\\ 16\\ 17\\ 18\\ 19\\ 20\\ 21\\ 22\\ 32\\ 42\\ 52\\ 62\\ 7\\ 28\\ 29\\ 30\\ 31\\ 32\\ 33\\ 34\\ 35\\ 36\\ 37\\ 38\\ 39\\ 40\\ 41\\ 43\\ 44\\ 56\\ 47\\ 48\\ 49\\ 50\\ 51\\ 53\\ 54\\ 55\end{array}$	593		
56 57 58 59 60			For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml
50			

Table 1: Recruitment sites and estimated eligible population.

Province	Site	Type of site	Potential participants p
			site (approximate)
Kigali	Cor-unum HC	Public Health Center	1,405
Kigali	Kimironko HC	Public Health Center	1,227
Kigali	Rwampara HC	Public Referral Hospital	1,098
Kigali	Kicukiro HC	Public Health Center	1,054
Kigali	Kacyiru HC	Public Health Center	905
Kigali	Gikondo HC	Public Health Center	925
Kigali	Rwanda Military Hospital	Public Referral Hospital	300
Kigali	WEACTx for Hope	Private HIV Clinic	500
Kigali	Busanza HC	Public Health Center	100
Kigali	Nyarugunga HC	Public Health Center	100
West	Gisenyi DH	Public District Hospital	810
Total			8,424
			1



BMJ Open	Pa
Appendix I. Study Questionnaire	
A. <u>Socio-demographics</u>	
1. What is your date of birth? <u>D</u> <u>D</u> / <u>M</u> <u>M</u> / <u>Y</u> <u>Y</u> <u>Y</u> (enter all 0 if not remembered) (If date of birth or year of birth is given, skip to Question 3)	
2. What is your age?(Years)	
3. What is your marital status?	
 Married/Cohabiting Divorced Widowed Separated Single Choose not to answer 	
4. Do you live in Kigali?	
 Yes No {Skip to Question A5} Choose not to answer {Skip to Question A5} 	
5. In what area of Kigali City Province do you live?	
 Nyarugenge Kicukiro Gasabo urban (e.g., Kacyiru, Kimironko, Kinyinya, etc.) Gasabo rural (e.g., Nduba, Jabana, Gikomero, etc.) Do not know Choose not to answer 	
6. What is your household income per month in FRW?	
$ < 5,000 \\ 5,000 - 9,999 \\ 10,000 - 34,999 \\ 35,000 - 59,999 $	

1 2	
2 3	
4	60,000 - 84,999
5	85,000 - 109,999
6 7	110,000 – 134,999
8	135,000 - 269,999
9 10	270,000 or more
11	
12 13	7. How many people live in your household? {Enter 00 if refuse to answer}
14	8. What is your occupation?
15 16	
17	Employed by government, another institution, or company
18 19	Self-employed (Small and medium enterprises)
20	Self-employed (High income earnings)
21	Farming (peasants)
22 23	Unemployed/Does not work
24	Other (specify)
25 26	
27	Choose not to answer {Skip to Question B1}
28	{The following questions are sensitive and personal in nature. Your answers will be kept
29 30	confidential. You may choose not to answer certain questions. Answering any question is
31	voluntary}
32 33	B. <u>Sexual Behaviors</u>
34	
35	1. How old were you when you first had sex? (Years) {Enter 00 if refuse to
36 37	answer}
38	2. How many sexual partners have you had in your lifetime?
39 40	
41	No Partners (Ineligible)
42	1 Partner
43 44	2-3 Partners
45	4-5 Partners
46 47	6-9 Partners
48	
49 50	10 or more partners
51	Choose not to answer
52	3. How many sexual partners have you had in the last 6 months?
53 54	
55	No Partners
56 57	
57 58	
59	
60	For peer review only - http://bmjopen.bmj.com/site/about/guidelines.xhtml

2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
12	
13 14	
14	
15	
16	
17	
18	
19	
20	
21	
22	
23	
24	
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
55 54	
55	
56	
57	
58	
59	

1

1 Partner	
-----------	--

2 or more Partners

Choose not to answer

C. Parity

1. What age did you have your first child? ____ (Years) {Enter 00 if refuse to answer, Enter 99 if never pregnant} (if 00 or 99, skip to Question D1)

2. How many live births have you had in your lifetime? _____

3. Have you given birth in the last year?

Yes

No

Choose not to answer

D. <u>Tobacco Use</u>

1. Have you ever smoked cigarettes?

Yes

No (skip to Question D3)

Choose not to answer (skip to Question D3)

2. Do you currently smoke cigarettes?

Yes

No

Choose not to answer

3. Have you ever chewed/used tobacco orally (Ubugoro)?

Yes

No (skip to Question D5)

Choose not to answer (skip to Question D5)

4. Are you currently chewing/using tobacco orally (Ubugoro)?

Yes

] No

Choose not to answer

5.	Have you ever chewed/used tobacco orally (Tobacco leaves-Igikamba)?
Y	es
	o (skip to Question E1)
Cł	noose not to answer (skip to Question E1)
6.	Are you currently chewing/using tobacco orally (Tobacco leaves-Igikamba)?
Y	es
No)
	noose not to answer
E.	Contraceptive Use
1.	Have you ever used oral contraceptives?
	es
	o (Skip to Question E3)
	noose not to answer (Skip to Question E3)
2.	Do you currently use oral contraceptives?
Y	es
N)
Cł	noose not to answer
3.	Have you ever used Depo Provera (contraceptive)?
ΠY	es
	o (Skip to Question E5)
	noose not to answer (Skip to Question E5)
4.	Do you currently use Depo Provera (contraceptive)?
ΠY	es
N	
Cł	noose not to answer
5.	Have you ever used Jadell (contraceptive)?
∏ Ye	

1	
2	
3	
4	
5	
6	
7	
8	
9	
10	
11	
12	
13	
14	
15	
16 17	
17	
18	
19	
20	
21	
22 23	
23	
24	
25	
26	
27	
28	
29 30	
20 21	
31 32	
32 33	
33 34	
35	
36	
37	
38	
39	
40	
40 41	
42	
43	
44	
45	
46	
47	
48	
49	
50	
51	
52	
53	
54	
55	
56	
57	
58	
59	
60	

Choose not to answer (Skip to Question E7)
6. Do you currently use Jadell (contraceptive)?
Yes
No
Choose not to answer
7. Have you ever used IUD (contraceptive)?
Yes
No (Skip to Question E9)
Choose not to answer (Skip to Question E9)
8. Do you currently use IUD (contraceptive)?
☐ Yes
□ No
Choose not to answer
9. Do use condoms as a contraceptive?
Yes
□ No (skip to Question F1)
Choose not to answer (skip to Question F1)
10. Have you used condoms as a contraceptive in the last 6 months?
☐ Yes
□ No
Choose not to answer
F. <u>Infections</u>
1. Have you ever had Malaria
Yes
No (skip to Question F8)
Choose not to answer (skip to Question F8)
2. How many times have you had Malaria?

4.	Have you had your Malaria treated using drugs?
Yes	
🗌 No	
Cho	ose not to answer
5.	If Yes, how was it treated?
Trea	ated only with traditional medicine
Trea	ated only with drugs (e.g. Coartem)
Trea	ated with both (traditional and modern)
Not	treated
Cho	ose not to answer
6.	Was your last episode of Malaria treated using drugs?
Yes	
🗌 No	
Cho	ose not to answer
7.	If Yes, how was it treated?
Trea	ated only with traditional medicine
Trea	ated only with drugs (e.g. Coartem)
Trea	ated with both (traditional and modern)
🗌 Not	treated
Cho	ose not to answer
8.	Have you ever had Tuberculosis (TB)?
Yes	
No	(skip to end)
Cho	ose not to answer (skip to end)
9.	If Yes, how many times have you had TB in your lifetime?
1(). What year did you last have Tuberculosis (TB)? <u>Y</u> Y <u>Y</u>
	END OF QUESTIONNAIRE