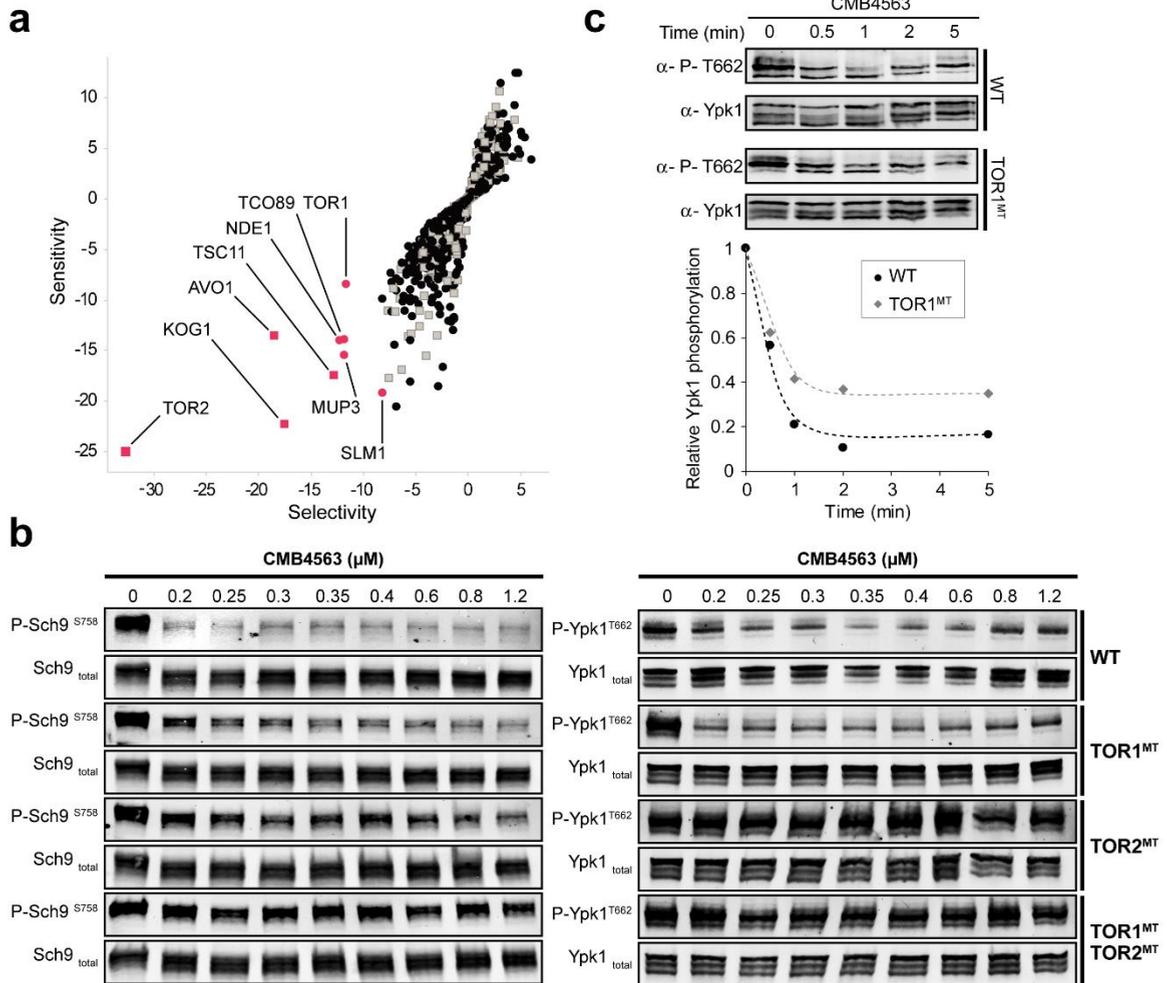


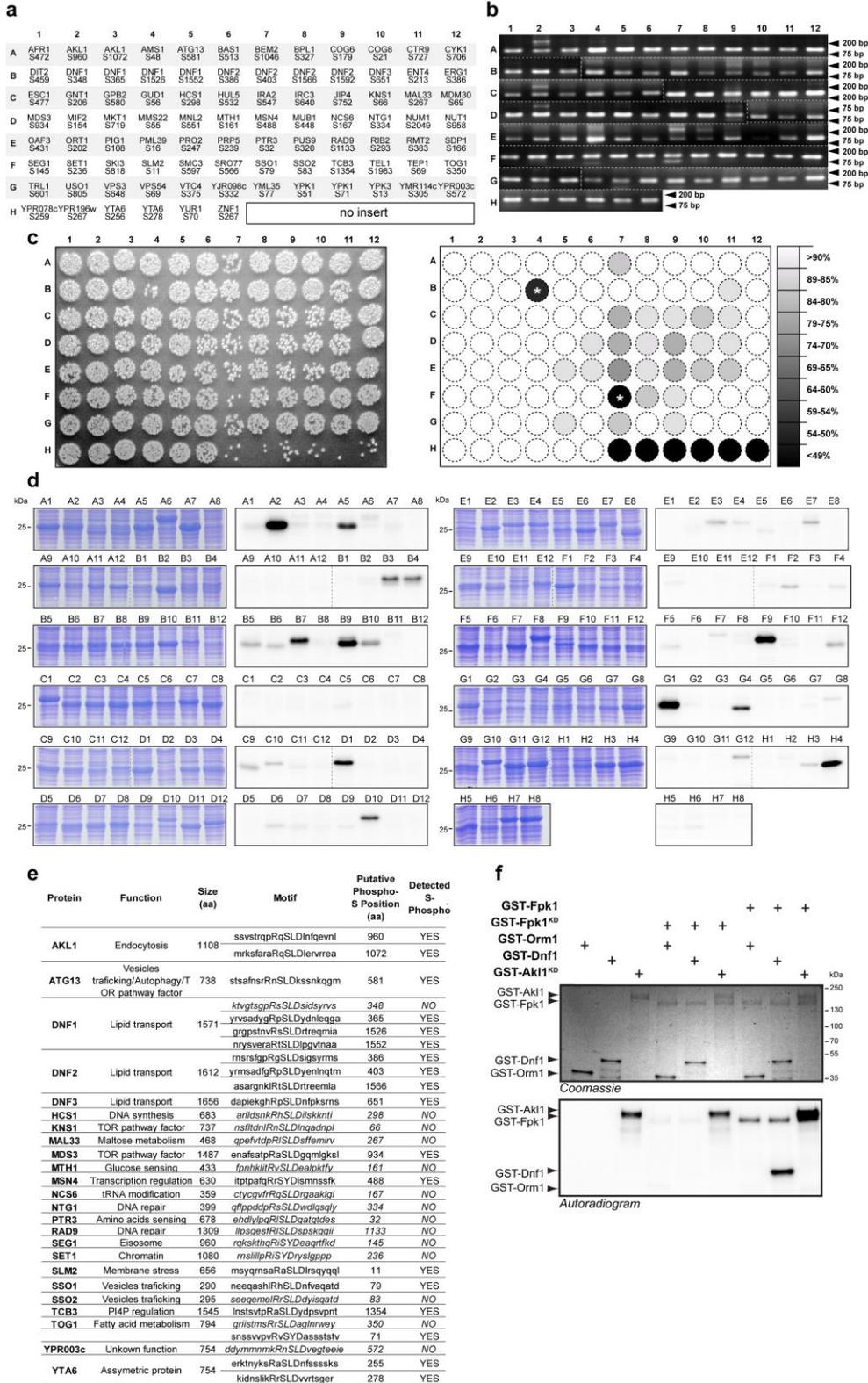
SUPPORTING INFORMATION

Supplemental figure 1



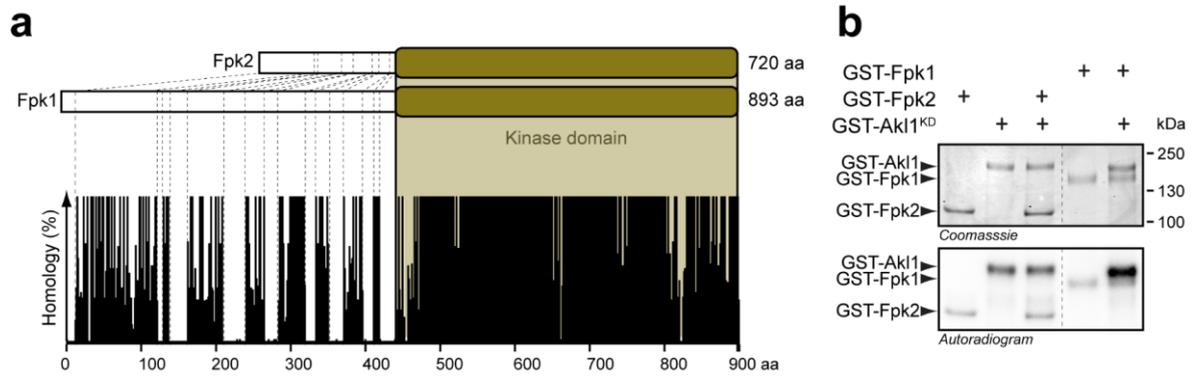
Supplemental figure 1: CMB4563 is a potent and fast inhibitor of TOR1 and TORC2. **a**, Heterozygous single deletion mutant strains covering 6200 genes (Open Biosystems) were grown as a pool in the presence or absence of 0.5 μM CMB4563 for 20 generations. The relative abundances of the individual strains were then compared. Sensitivity of individual deletion mutants are observed on the vertical axis and associated with a selectivity on the horizontal axis which was determined by statistical analysis to correct for how frequently particular mutants scored in multiple assays. The most selective and sensitive strains are found in the bottom left corner (1). **b**, Western blots assessing the respective activity of TORC1 and TORC2 after 12-minute CMB4563 treatments to cells of the indicated genotypes. The corresponding merge channels version is in Fig. 1d. TORC1 activity is followed by Sch9^{P-S758} and TORC2 by Ypk1^{P-T662} phosphorylation. **c**, Western blot and corresponding quantification assessing the kinetics of Ypk1^{T662} dephosphorylation in *TOR1^{WT}* and *TOR1^{MT}* strains with 0.8 μM CMB4563 treatment at indicated time points (above, minutes).

Supplemental figure 2



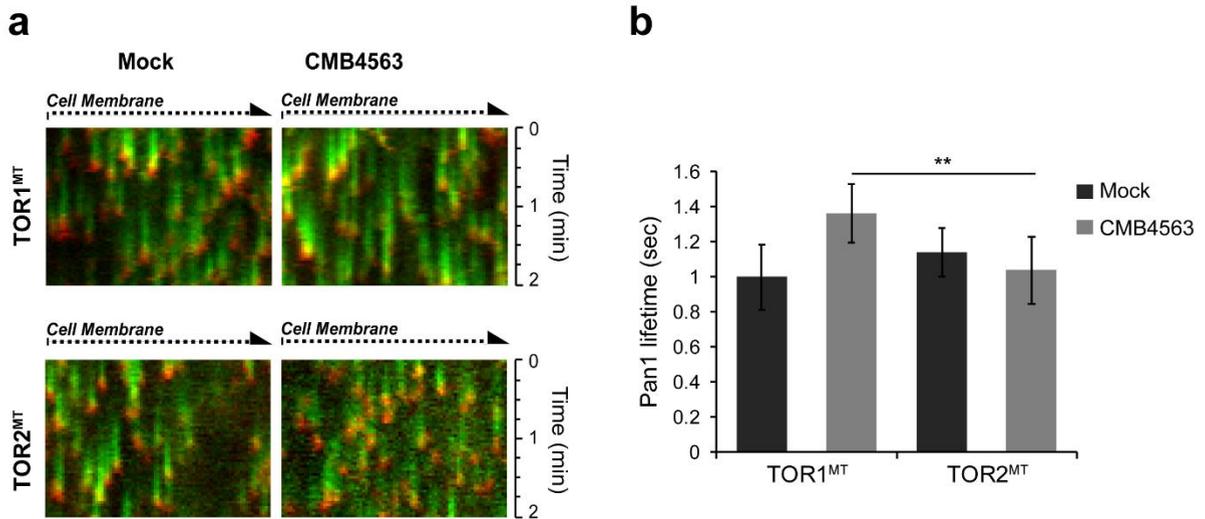
Supplemental figure 2: Fpk1 substrate screen. **a**, Organization of the 96-well-plate containing negative controls and the primer dimers corresponding to the 90 candidate peptide substrates. **b**, PCR products of the primer dimers analysed on 2% agarose gels **c**, Left Panel: Gibson assembly reactions were transformed into bacteria. 1/10 of the transformed cells were spotted onto LB plates containing kanamycin, including multiple negative controls corresponding to linearized vector without insert. Right Panel: Heat map reporting the cloning efficiency assessed with respect to the negative controls. The (*) indicated failed wells in which the cloning/transformation initially failed (these were subsequently repeated, successfully). **d**, Results of the initial *in vitro* kinase assays screen. Dashed lines define splicing of the gels. Bacterial lysates containing one of the 90 candidate peptides were individually incubated with GST-Fpk1 and ³²P-ATP for 30 min at 30°C. Reactants were subsequently resolved by SDS-PAGE, analysed by Coomassie stain (left panel) and autoradiography (right panel). **e**, Table recapitulating the peptides identified in both the initial and secondary screens (left) **f**, *In vitro* kinase assay assessing the ability of GST-Fpk1^{WT/KD} to phosphorylate GST-Ak11^{KD}, GST-Orm1 (aa 1-85; negative control) or GST-Dnf1 (aa 1404-1571; positive control).

Supplemental figure 3:



Supplemental figure 3: Ak11 is not a target of Fpk2 *in vitro*. **a**, ClustalX alignments of Fpk1 and Fpk2 protein sequences. The green box represents the conserved kinase domain and the dashed lines delimit sequence insertions. **b**, Coomassie blue and autoradiogram SDS-PAGE images after *in vitro* kinase assay assessing the ability of Fpk1 and Fpk2 to phosphorylate Ak11. Dashed lines define splicing of the gel.

Supplemental figure 4



Supplemental figure 4: The inhibition of TORC1 does not cause delay the endocytosis process. a, Pan1-GFP and Abp1-mCherry kymographs were generated from *TOR1^{MT}* or *TOR2^{MT}* strains treated or not with CMB4563 for 30 min. **b,** Quantification of Pan1-GFP and Abp1-mCherry patch lifetimes determined from *TOR1^{MT}* or *TOR2^{MT}* cells as assessed in a. Bar graphs of Pan1-GFP lifetimes means, +/- standard deviation. Statistical parameters were extracted (Student's test: $P < 0.005$ (**)) based on five patches/kymograph. Five kymographs were analysed.

Supplemental table 2

| Protein | Function | Size (aa) | Motif | Putative Phospho-S Position | Detected S-Phospho | Protein | Function | Size (aa) | Motif | Putative Phospho-S Position | Detected S-Phospho |
|---------|---|-----------|---|-----------------------------|--------------------|---------|------------------------------|-----------|------------------------|-----------------------------|--------------------|
| AFR1 | Shmoo formation | 620 | appprrsskRpSLDnnesaryf | 472 | YES | MTH1 | Glucose sensing | 433 | fophklllRvSLDealpktfy | 161 | NO |
| AKL1 | Endocytosis | 1108 | ssvstrqpRgSLDlnfgevnl mrksfaraRgSLDlervrea | 960 1072 | YES YES | MSN4 | Transcription regulation | 630 | ittpbafqRrSYDismnssfk | 488 | YES |
| AMS1 | Mannose metabolism | 1083 | lpkfydkkrSLDdhdkvkvwv | 48 | NO | MUB1 | Ubiquitin/Proteasome | 620 | kisdnferRvSYDkmkkitn | 448 | NO |
| ATG13 | Vesicles trafficking/Autophagy/TOR pathway factor | 738 | atsafnrRnSLDksnkqgm | 581 | YES | NCS6 | tRNA modification | 359 | ctycgvfrRgSLDrgaakigi | 167 | NO |
| BAS1 | Purine/histidine metabolism | 811 | sdtnpeyRtSLDnmsdfis | 513 | NO | NTG1 | DNA repair | 399 | qflppddpRnSLDwdlqsgiy | 334 | NO |
| BEM2 | Cytoskeleton organisation | 2167 | patpngkmRdSLDttgrlakt | 1046 | YES | NUM1 | Organel migration | 2748 | seirdqidRpSLDvikekaa1 | 2049 | NO |
| BPL1 | Biotinylation | 690 | efufyryyRaSYDaessilh | 327 | NO | NUT1 | Transcription regulation | 1132 | gvpgsenkRgSLDsehvdyf | 958 | NO |
| COG6 | Vesicles trafficking | 839 | xvlecnkRiSLDgkimppe | 179 | NO | OAF3 | Oleate metabolism | 863 | sfkinrhtRpSYDimpdsdyi | 431 | NO |
| COG8 | Vesicles trafficking | 607 | ddlteeqkRiSLDflqdligs | 21 | NO | ORT1 | Arginine metabolism | 292 | ivkkslkdkhSLDpkrdesk | 202 | NO |
| CTR9 | Transcription | 1077 | aloilrkvRdSLDnedvqini | 727 | NO | PIG1 | Glycogen metabolism | 648 | pfllggedhRySLDiydsie | 108 | NO |
| CYK1 | Budding | 1495 | afergalvReSLDgksefykr | 706 | NO | PLM139 | RNA maturation | 334 | leavrksrRiSLDpntklppg | 16 | NO |
| DIT2 | Sporulation | 489 | laemlkqfRvSLDpeweeklt | 459 | NO | PRO2 | Proline metabolism | 456 | daqlkakarSLDaktvnpag | 247 | NO |
| DNF1 | Lipid transport | 1571 | ktvrsqRnSLDaidsvrva | 348 | NO | PRP5 | RNA maturation | 849 | semeveeRiSLDnliklqgt | 239 | NO |
| | | | yrvsadygRpSLDydnlegga | 365 | YES | PTR3 | Amino acids sensing | 678 | ehnoqlpRgSLDgatgdes | 32 | NO |
| | | | grpstvRnSLDrtreemia | 1526 | YES | PUS9 | tRNA modification | 462 | khaktvRnSLDyqktsivkc | 320 | NO |
| DNF2 | Lipid transport | 1612 | nrvsvrRtSLDlpgvtnaa | 1552 | YES | RAD9 | DNA repair | 1309 | lpsgsesRiSLDpsksggii | 1133 | NO |
| | | | rnsrfqRpSLDsigyrcms | 386 | YES | RIB2 | tRNA modification | 591 | khaktvRvSLDyqgtsivkc | 293 | NO |
| | | | yrvsadygRpSLDydnlegga | 303 | YES | RM2 | Translation regulation | 412 | yqmkcdyRySLDdeglwdnd | 383 | NO |
| DNF3 | Lipid transport | 1656 | asargnkRtSLDrtreemia | 1566 | YES | SDP1 | Sit2 phosphatase regulation | 209 | mkyhnlRnSLDnllksradk | 166 | NO |
| | | | trsvsraRaSLDlpginhae | 1592 | YES | SEG1 | Eisosome | 960 | rgkskLhgRiSLDyagrftkd | 145 | NO |
| | | | dapiekgRnSLDnfpksrns | 651 | YES | SET1 | Chromatin | 1080 | rnslllpRiSLDyrslygppp | 236 | NO |
| ENT4 | Trafficking | 247 | rnrthevafSLDplaeedse | 213 | NO | SK13 | RNA degradation | 1432 | lgivevImRcSLDlysqgtii | 818 | NO |
| ERG1 | Ergosterol metabolism | 496 | e1ldyhferkSYDsvinvlsv | 386 | NO | SLM2 | Membrane stress | 656 | msyqznsaRaSLDlrsqqyql | 11 | YES |
| ESC1 | Telomeres | 491 | epekddiRnSLDknfbggnn | 477 | NO | SMC3 | Mitosis | 1230 | rvtlfpRnSLDsdvkiptan | 597 | NO |
| GHT1 | Glycans modifications | 491 | yldndailRnSLDelfrtpny | 206 | NO | SRO77 | Vesicles trafficking | 1010 | dalqlkRiSLDsdktllvd | 566 | NO |
| GPB2 | PKA inhibitor | 880 | whqstlmdRnSLDmrllmkk | 580 | NO | SSO1 | Vesicles trafficking | 290 | neegashlRnSLDnfvagatd | 79 | YES |
| GUID1 | Guanine metabolism | 489 | ngliffvRnSLDpvrkdcldh | 56 | NO | SSO2 | Vesicles trafficking | 295 | seegashlRnSLDdyisqatd | 83 | NO |
| HCS1 | DNA synthesis | 683 | arildsnkRnSLDiskknti | 298 | NO | TCB3 | PI4P regulation | 1545 | lnstsvtRnSLDydpvnt | 1354 | YES |
| HUL5 | Ubiquitin/Proteasome | 910 | efialdkRiSLDdhdnllnm | 532 | NO | TEL1 | Telomeres | 2787 | htlmmvRnSLDnrvkcsky | 1983 | NO |
| IRA2 | cAMP metabolism | 3079 | qyrtaeqqsRaSYDahktgtg | 547 | NO | TEP1 | PI3P Phosphatase | 434 | ntypklyRnSLDdliifltv | 69 | NO |
| IRC3 | DNA synthesis | 689 | qesskdfiRiSLDdvtcyunt | 640 | NO | TOG1 | Fatty acid metabolism | 794 | grlilstnsRnSLDaglnrwey | 350 | NO |
| JIP4 | Unknown function | 876 | aaigaikRnSLDspgdqqqq | 752 | NO | TRL1 | tRNA modification | 827 | ddeietaFRnSLDyktvrvki | 601 | NO |
| KNS1 | TOR pathway factor | 737 | nslftdnRnSLDingadnpi | 66 | NO | USO1 | Vesicles trafficking | 1790 | leteiknvRnSLDdmtglrdv | 805 | NO |
| MAL33 | Maltose metabolism | 468 | qpefvtdpRiSLDsfemirv | 267 | NO | VPS3 | Vesicles trafficking/Vacuole | 1011 | rfrvdmfRnSLDydeliraini | 648 | NO |
| MDM30 | Ubiquitin/Proteasome | 598 | rcydrwinReSLDiltandyd | 69 | NO | VPS54 | Protein recycling | 889 | tpxrarsaRnSLDsltprrsf | 69 | YES |
| MDS3 | TOR pathway factor | 1487 | enafstapRaSLDgqmlgkls1 | 934 | YES | VTCA | Vesicles trafficking/Vacuole | 721 | qfpgdarvRiSLDtelmrvre | 375 | NO |
| MIF2 | Kinetochore | 549 | tykrkystrRySLDtsesppvr | 154 | YES | YJR098c | Unknown function | 656 | vqgfygdwRiSLDlsskhlfi | 332 | NO |
| MKT1 | Transcription regulation | 830 | rlscqfnRiSLDnfgvarki | 719 | NO | YML35 | Translation regulation | 367 | phlidelRnSYDfiegssk | 77 | NO |
| MMS22 | Ubiquitin/Proteasome | 1454 | rsqkivteRiSLDstagesct | 55 | NO | YPK1 | TOR pathway factor | 680 | gehdsasRnSLDdrkgtinps | 51 | YES |
| MNL2 | Mannose metabolism | 849 | tne1dvtirkSYDvlyscr1 | 551 | NO | YPK3 | TOR pathway factor | 525 | snsvvrvRvSYDassststv | 71 | YES |
| | | | | | | YMR114C | Unknown function | 368 | fnqkkslRnSYDglikkneeq | 305 | YES |
| | | | | | | YPR003c | Unknown function | 754 | ddyymmnnRnSLDvsgteie | 572 | NO |
| | | | | | | YPR078c | Unknown function | 372 | tllykryvktSLDmigeekss | 259 | NO |
| | | | | | | YPR196w | Unknown function | 470 | qpeivtdsRiSLDsllevikv | 267 | NO |
| | | | | | | YTA6 | Assymetric protein | 754 | erktnyRnSLDnffssssks | 255 | YES |
| | | | | | | YUR1 | Protein modification | 428 | yfpkvklsRnSYDytlnyt | 70 | NO |
| | | | | | | ZFN1 | Respiratory transition | 465 | qpefvtdpRiSLDslfemirv | 267 | NO |

Supplemental table 2: Table recapitulating all the peptides containing one or more RXSLD/E motif(s) in the *Saccharomyces Cerevisiae* proteome. The table provides (from the left to the right) the name of the protein, its function, its length in amino acids (aa), the sequence used for cloning i.e. the consensus motif surrounded by 5 aa on both sides, the position(s) in the sequence of the serine (S) present in the consensus motif, and whether this motif was phosphorylated *in vitro*.

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