

Appendix: P2X4 receptor controls microglia activation and favours remyelination in autoimmune encephalitis

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Appendix Supplementary Methods

T cell proliferation and cytokine production

To measure proliferation, splenocytes were labeled with eFluor670 proliferation dye (5 μ M, eBioscience), washed and stimulated with anti-CD3/CD28 (1 μ g/ml). On day 3 cells were stained with anti-CD4 (#100509, 1:100, Biolegend) and anti-CD8 (#100732, 1:100, Biolegend). Proliferation was measured by dilution of eFluor670 and quantified as division index using FlowJo Software (Treestar).

To measure cytokine production, splenocytes were stimulated for 4h with phorbol-12-myristat-13-acetat (PMA, 50 ng/ml, Sigma) and ionomycin (1 μ M, Sigma). Cells were washed and stained with anti-CD4 (#100509, 1:100, Biolegend), anti-CD8 (#100732, 1:100, Biolegend), anti- $\gamma\delta$ TCR (#118119, 1:100, Biolegend). Cells were fixed with 4 % paraformaldehyde (PFA, Sigma), permeabilized with 0.5% saponin buffer and stained for anti-IFN γ (#54413, 1:100, BD Bioscience) and anti-IL-17A (#12-7177-81, 1:100, eBioscience) and intracellular cytokines were measured by FACS (LSRFortessa, BD).

Immunocytochemistry

Primary antibodies were used as follow to: iNOS (#610329, 1:500, BD Bioscience), MRC1 (#ab64693, 1:1000, Abcam), NG2 (#AB5320, 1:500, Abcam), Iba1 (#019-19741, 1:1000; Wako), MBP (#SMI-99P, 1:500, Covance), Olig2 (#MABN50, 1:400, Millipore), P2X7R (#APR-004, 1:100, Alomone), P2X4R (#APR-002, 1:400, Alomone) and extracellular P2X4R (#APR-024, 1:400, Alomone). We used the following secondary antibodies goat anti-rabbit Alexa Fluor 488 (A11008, 1:200, Invitrogen) and goat anti-mouse Alexa Fluor 594 (A11032, 1:200, Invitrogen). For isolectin B4 (IB4) staining, live cultures were incubated for 30 min with IB4 (#FL 1201, 1:100, Vector),

washed with PBS, fixed in 4% PFA in PBS for 20 min and processed for conventional immunocytochemistry.

FACS

CD4-Bv605 (#100547, 1:100, Biolegend), CD8-perCP (#100732, 1:100, Biolegend), $\gamma\delta$ TCR-Bv650 (#118129, 1:100, Biolegend), CD3e-Bv421 (#100228, 1:100, Biolegend), CD45APC-eFluor780 (#47-0451-82, 1:100, eBioscience), Ly6G-AF700 (#127622, 1:100, Biolegend), CD11b-FITC (#101205, 1:100, Biolegend), CD11c-APC (#17-0114-82, 1:100, eBioscience).

qPCR primers

Sequences for mouse primers used for qPCR

| Target gene | Forward (5'->3') | Reverse (5'->3') |
|--------------------|--------------------------------|----------------------------|
| <i>Arg1</i> | GGATTGGCAAGGTGATGGAA | CGACATCAAAGCTCAGGTGAA |
| <i>Bdnf</i> | TCCAAAGGCCAACTGAAGCA | CTGCAGCCTTCCTTGGTGTA |
| <i>Ccl2</i> | AGCAGCAGGTGTCCCAA | TTCTTGGGGTCAGCACAGAC |
| <i>Ccr7</i> | GTGGTGGCTCTCCTTGTC | GGTATTCTCGCCGATGTAGTCA |
| <i>Chi3l3</i> | GCCCACCAGGAAAGTACACA | CCTCAGTGGCTCCTTCATTCA |
| <i>Clec7a</i> | ACCACAAGCCCACAGAATCA | AGGAAGGCAAGGCTGAGAAA |
| <i>Foxp3</i> | ACCACACTTCATGCATCAGCTC | GGCTGGGTTGTCCAGTGGAC |
| <i>Ifny</i> | TAACTATTTAACTCAAGTGGCATAGATGTG | GCCAGTTCCTCCAGATATCCAAG |
| <i>Il10</i> | AAAGGACCAGCTGGACAACA | TAAGGCTTGGCAACCCAAGTA |
| <i>Il12a</i> | AAACCAGCACATTGAAGACC | GGAAGAAGTCTCTCTAGTAGCC |
| <i>Il1b</i> | TGGCAACTGTTCTGAACTCA | GGGTCCGTCAACTTCAAAGAAC |
| <i>Il4</i> | ACGGAGATGGATGTGCCAAA | GAAGCACCTTGAAGCCCTA |
| <i>Il6</i> | CGATGATGCACTTGCAGAAA | ACTCCAGAAGACCAGAGGAA |
| <i>Irf5</i> | TGATGTCAAACCCCGAGAGAA | GAACATCTCCAGCAGCAACC |
| <i>Irf8</i> | GATATGCCGCCTATGACACA | CCCGTAGTAGAAGCTGATGAC |
| <i>Jak3</i> | CATAGAGGACGTGGACACTCAA | TGACATGTCTCCAGCCCAA |
| <i>Marco</i> | TTCTGTGCGATGCTCGGTTA | TTGTCCAGCCAGATGTTCCC |
| <i>Mbp</i> | CCCTCACAGCGATCCAAGTA | CTCTGTGCCTTGGGAGGAA |
| <i>Mr1</i> | GCTCGCTGTATTCTTGGTGAA | ACCAGGATCGGAAACAGCTA |
| <i>Mrc1</i> | CACAAAGCCATGCTGTAGTACC | GTAAAACCCATGCCGTTTCCA |
| <i>Msx3</i> | CTCCAGTCGCGCACTCTT | CCGTGGTTTGCGATTGGTTT |
| <i>Nos2</i> | GAGGAGCAGGTGGAAGACTA | GGAAAAGACTGCACCGAAGATA |

| | | |
|---------------|-------------------------|---------------------------|
| <i>P2rx4</i> | TTTGCGATTGACAGCGCCAAC | ATGGAACACACCTTCCAGTCC |
| <i>Ptgs2</i> | CTTCTCCCTGAAGCCGTACA | TGCTACTGTAGAGGGCTTTCAA |
| <i>Retnla</i> | ATCCCTCCACTGTAACGAAGAC | ACAAGCACACCCAGTAGCA |
| <i>Ror</i> | ACTGAAAGCAGGAGCAATGGAAG | TTCAAAAAAGACTGTGTGGTTGTTG |
| <i>Stat1</i> | GCAGGTGTTGTCAGATCGAAC | ATGCACGGCTGTCGTTCTA |
| <i>Stat3</i> | TGGGCATCAATCCTGTGGTA | CCAATTGGCGGCTTAGTGAA |
| <i>Stat6</i> | TGACTTTCCACAACGCCTAC | CATCTGAACCGACCAGGAAC |
| <i>Tgfb1</i> | GCTGCGCTTGACAGAGATTA | GTAACGCCAGGAATTGTTGCTA |
| <i>Tnf</i> | GGGTGATCGGTCCCAAA | TGAGGGTCTGGGCCATAGAA |

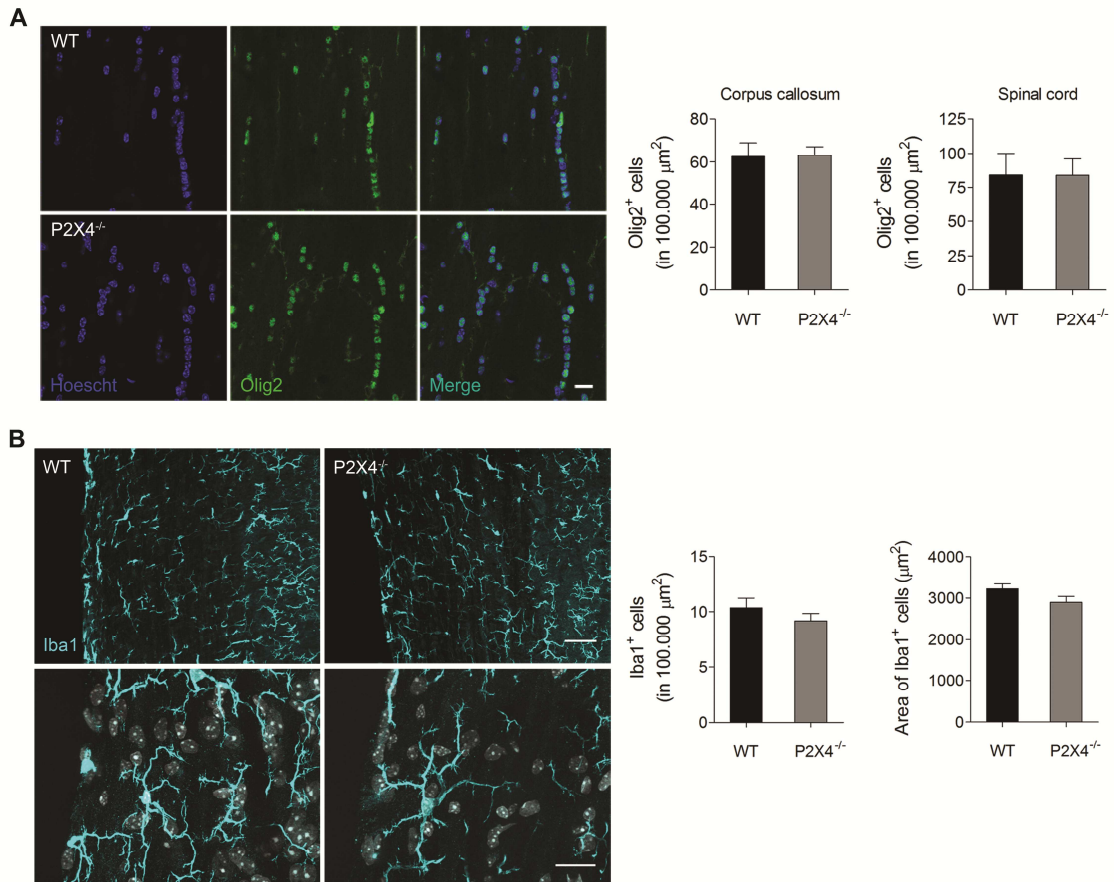
| Housekeeping gene | Forward (5'->3') | Reverse (5'->3') |
|--------------------------|----------------------------|----------------------------|
| <i>B2m</i> | ACTGACCGGCCTGTATGCTA | ATGTTTCGGCTTCCCATTCTCC |
| <i>Gapdh</i> | AGACGGCCGCATCTTCTT | TTCACACCGACCTTACCAT |
| <i>Hprt</i> | CAGTACAGCCCCAAAATGGTTA | AGTCTGGCCTGTATCCAACA |
| <i>Ppia</i> | AGGGTTCCTCCTTTCACAGAA | TGCCGCCAGTGCCATTA |

Sequences for rat primers used for qPCR

| Target gene | Forward (5'->3') | Reverse (5'->3') |
|--------------------|----------------------------|----------------------------|
| <i>Arg1</i> | GTGAAGAACCACGGTCTGTG | GAGATGCTTCCAATTGCCATACTG |
| <i>Ccl2</i> | GTGCTGTCTCAGCCAGATGCA | GCTGCTGGTATTCTTGTAGTT |
| <i>Mrc1</i> | AAGTTTAAGCACTGGCTGGCA | CAGGTTCTGATGATGGACTTCCTG |
| <i>Nos2</i> | GAGATTTTTTACGACACCCCTTAC | CATGCATAATTTGGACTTGCAAG |

| Housekeeping gene | Forward (5'->3') | Reverse (5'->3') |
|--------------------------|----------------------------|----------------------------|
| <i>Cyclophilin A</i> | CAAAGTTCAAAGACAGCAGAAAA | CCACCCTGGCACATGAATC |
| <i>Gapdh</i> | GAAGGTCGGTGTCAACGGATTT | CAATGTCCACTTTGTCACAAGAGA |
| <i>Hprt</i> | ATGGACTGATTATGGACAGGACTGA | ACACAGAGGGCCACAATGTG |

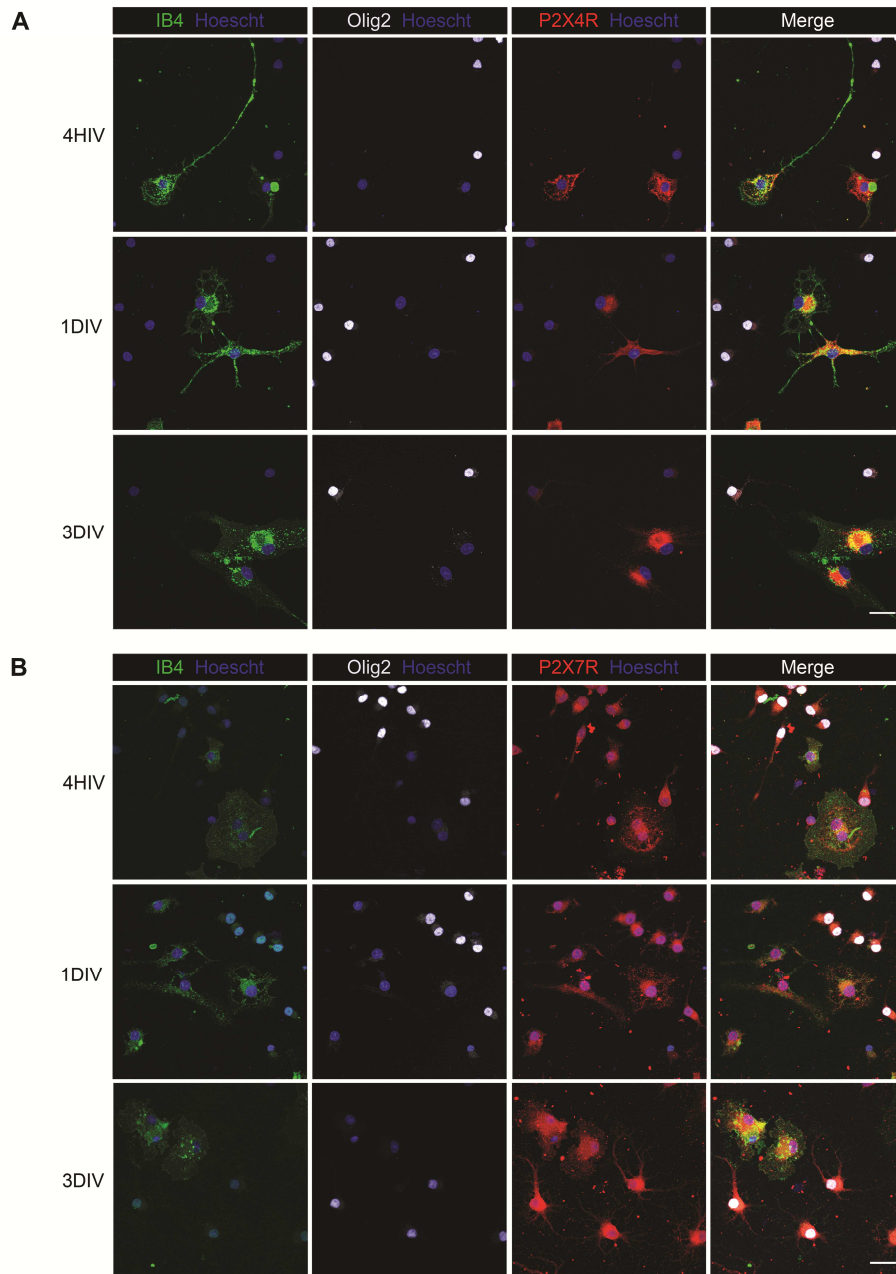
Appendix Figures



Appendix Figure S1. Microglia and oligodendroglial characterization of WT and P2X4^{-/-} mice.

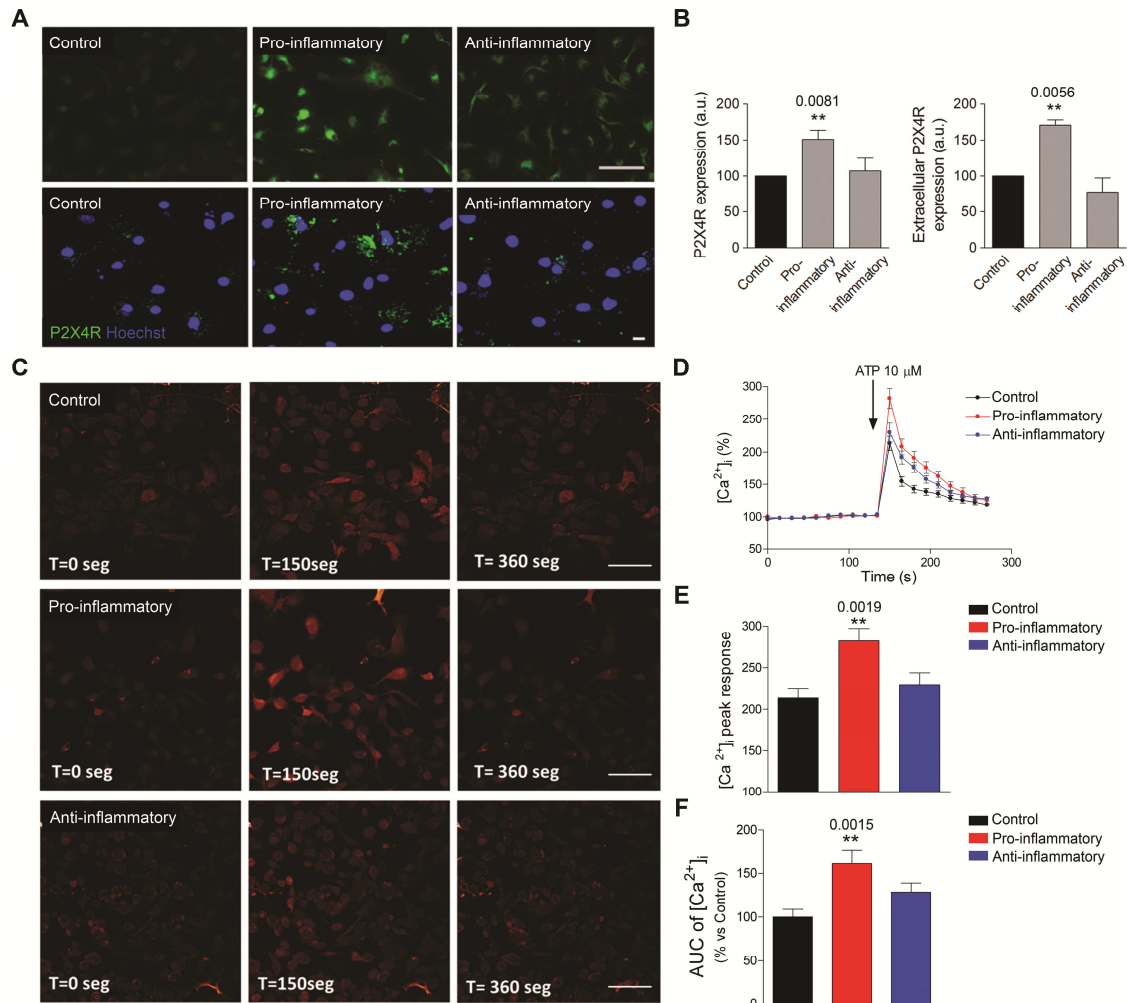
A Immunohistochemistry analysis of Olig2, a marker of oligodendrocytes in corpus callosum and spinal cord of WT and P2X4^{-/-} mice. Images show representative fields in corpus callosum (n = 3). Scale bar = 20 μm.

B Immunohistochemistry analysis of Iba1, a marker of microglia in the spinal cord of WT and P2X4^{-/-} mice (n = 3). Scale bar = 50 (top) and 20 (bottom) μm.



Appendix Figure S2. P2X4R is expressed predominantly in microglia.

A-B) Representative images of microglia-OPCs coculture stained for P2X4R (A; red) or P2X7R (B; red), Olig2 (white), isolectin B4 (IB4; green) and Hoechst (blue) at different stages of oligodendrocyte development, 4 hours in vitro (HIV), 1 day in vitro (DIV) and 3 DIV. Scale bar = 20 μ m



Appendix Figure S3. P2X4R expression and function in differentially polarized microglia.

A, B) Total P2X4R (green, top) (n = 5) or extracellular P2X4R (green, bottom) (n = 3) expression in control, pro-inflammatory and anti-inflammatory microglia phenotypes. Scale Bar= 80 μ m (top) and 10 μ m (bottom).

C-F) Calcium responses to ATP (10 μ M) in control, pro-inflammatory and anti-inflammatory microglia: Representative images at 0, 150 and 360 seconds. ATP was applied at t=150s (n = 150 cells from 3 different cultures). Scale Bar= 80 μ m.