

## Supplementary Information

Magnetosomes extracted from *Magnetospirillum gryphiswaldense* as theranostic agents in an experimental model of glioblastoma

Silvia Mannucci<sup>1\*</sup>, Stefano Tambalo<sup>2\*</sup>, Giamaica Conti<sup>1</sup>, Leonardo Ghin<sup>3</sup>, Alessio Milanese<sup>2</sup>, Anna Carboncino<sup>1</sup>, Elena Nicolato<sup>1</sup>, Maria Rosaria Marinozzi<sup>1</sup>, Donatella Benati<sup>1</sup>, Roberto Bassi<sup>3</sup>, Pasquina Marzola<sup>4</sup>, Andrea Sbarbati<sup>1,3</sup>

<sup>1</sup>Department of Neuroscience, Biomedicine and Movement Sciences, University of Verona, Strada Le Grazie 8, I-37134, Verona, Italy; <sup>2</sup>Consorzio INSTM, Via G. Giusti 9, I-50121 Firenze, Italy;

<sup>3</sup>Department of Biotechnology, University of Verona, Strada Le Grazie 15, I-37134 Verona, Italy;

<sup>4</sup>Department of Computer Science, University of Verona, Strada Le Grazie 15, I-37134 Verona, Italy.

\*These authors contributed equally to the paper

#present address: Center for Neuroscience and Cognitive Systems @UniTn, Istituto Italiano di Tecnologia, Corso Bettini 31, 38068 Rovereto, Italy;

### **Information S1: Cancer cells culture**

U87MG cell line (purchased by ATCC Manassas, VA), was cultured in Eagle's Minimum Essential Medium (EMEM) with 10% of Fetal Bovine Serum (FBS), 1% of a mix of penicillin/streptomycin 1:1 and 1% of L-glutamine 200 mM, in 25 cm<sup>2</sup> plates and incubated at 37°C in humidified air with 5% CO<sub>2</sub>. Medium and L-glutamine were purchased by Sigma-Aldrich (Italy),

while serum and antibiotic mix were acquired by GIBCO Life Technologies (USA). When at confluence, cells were treated with trypsin-EDTA 1% (GIBCO Life Technologies, USA), harvested and centrifuged at 1200 rpm for 5 min. The supernatant was discarded and cells pellet was resuspended in 1 ml of complete medium, placed in 75 cm<sup>2</sup> plates and incubated at 37°C and 5% of CO<sub>2</sub> until 80% confluence was detectable.

***Information S2: MNs-uptake in cancer cells***

Batches of 2X10<sup>5</sup> cells were seeded in specific glasses multiwell for optical microscopy (Falcon BD, Italy) with 1 ml of growth medium. Cells were incubated at 37°C in humidified air with 5% CO<sub>2</sub> for 24 h. Different concentrations of MNs (1 mg/ml, 0.5 mg/ml, 0.2 mg/ml) were added to the wells and incubated for 6 h, 12 h and 24 h. At the selected time points, the growth medium was discarded and cells were washed with 1 ml of 1X sterile phosphate buffer saline (PBS, GIBCO, Life Technologies, USA). Cells were fixed with 1 ml of 4% buffered formalin (Biotica, Italy) for 30 min at room temperature. Once formalin was discarded, samples were double stained with Prussian Blue and Nuclear Fast Red (Biotica, Italy).

***Information S3: Transmission Electron Microscopy (TEM) of cells incubated with MNs***

Whole mount bacteria of different samples of MNs, extracted and purified, were fixed with glutaraldehyde 2% in Sorensen buffer pH 7.4 for 2 h, post-fixed in 1% osmium tetroxide in aqueous solution for 2 h, dehydrated in graded concentrations of acetone. At the end of dehydrating process, samples were positioned in a multi-well grid for electron microscopy and observed using Philips Morgagni TEM and equipped with a Megaview II camera for digital image acquisition.

U87MG cancer cells, plated on a 2.4 cm culture glass, were positioned on the bottom of 3.5 cm Petri dishes and incubated at 37°C in humidified air with 5% of CO<sub>2</sub> in 3 ml of growth medium. After 24h, the growth medium was then discarded and replaced with fresh medium added with MNs at different concentrations: 1 mg/ml, 0.5 mg/ml, 0.2 mg/ml. Cells were incubated as previously described for 6 h, 12 h and 24 h. At each time point, the growth medium was discarded and cells were washed with 1 ml of 1X sterile phosphate buffer saline (PBS, GIBCO, Life Technologies,

USA). Cells were fixed with glutaraldehyde 2% in Sorensen buffer pH 7.4 for 2 h, then post-fixed in 1% osmium tetroxide ( $\text{OsO}_4$ ) in aqueous solution for 2 h, and finally dehydrated in graded concentrations of acetone. At the end of dehydrating process, glasses were stained with lead citrate.

**Information S4: Magnetic Resonance Imaging**

MRI was performed using a Bruker tomograph operating at 4.7 T, equipped with an actively shielded gradient insert (Bruker, Germany) having a maximum gradient strength of 40 G/cm.  $T_2$  and  $T_2^*$  weighted images were acquired with the following parameters:  $T_2$  w RARE 3D sequence with  $\text{TR}=1200$  ms,  $\text{TE}_{\text{eff}}=47.5$  ms,  $\text{FOV}=5 \times 2.5 \times 2.5$  cm<sup>3</sup>,  $\text{NEX}=1$ ,  $\text{MTX}=256/128/32$ , Slice Thickness=0.78 mm;  $T_2^*$  weighted FLASH gradient echo sequence with  $\text{TR}=400$  ms,  $\text{TE}=4.4$  ms, flip angle=10°,  $\text{FOV}=5 \times 2.5$  cm,  $\text{NEX}=2$ ,  $\text{MTX}=256/128$ ,  $\text{NSLICES}=8$ , Slice Thickness=2 mm.

**Information S5: Histology**

At the end of the last imaging session, tumors were excised, washed with PBS 0.1 M and fixed with 10% buffered formalin for 4 h. Afterwards, samples were dehydrated with increasing concentration gradient of alcohol from 70% to 100% and then with xylene for final processing. Tissues were embedded in paraffin and sections of 5  $\mu\text{m}$  were obtained and dried at 37°C for 24 h. Section were then stained with Prussian blue and Nuclear Fast Red (Biotica, Italy).