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Supplemental Information

Immunological Synapse Predicts Effectiveness

of Chimeric Antigen Receptor Cells

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Supplementary Figures

Figure S1



Figure S1: Quantification of IS quality. CAR T cells were added to the lipid bilayer containing Alexa Fluor 647 labeled Kappa IgG1. Cells were stained with Abs against perforin, pZeta, and phalloidin (F-actin staining). The IS under the lipid bilayer was quantified by measuring the mean fluorescence intensities of F-actin, pZeta, and Kappa cluster, as well as the percentage of perforin-positive cells on the glass-supported planar lipid bilayer containing Kappa IgG1. Error bars show ± standard deviation (s.d.). NS, not significant.







Figure S3: Specific interactions between Kappa-CAR T cells and the lipid bilayer carrying Kappa antigen. (a and b) Representative confocal images of 4-1BB- and CD28-CAR T cells. CAR T cells on the lipid bilayer carrying Alexa Fluor 647-labled Lambda IgG1 (a, as a control for kappa antigen) or Alexa Fluor 647- labeled Kappa IgG1 (b, tumor antigen). Fixed CAR T cells were stained for perforin, pZap70, LCK, and F-actin, as indicated. Scale bars represent 25.0 μ m. (c) Quantification of the mean fluorescence intensities of F-actin, pZAP70, and cluster of Kappa. Error bars show \pm s.d.



Figure S4: Superior IS quality in CD19 specific 4-1BB-CAR T cells. (a) Representative confocal microscopy of CD19-CAR T cells with 4-1BB or CD28

activated on lipid bilayer carrying CD19-Alexa Fluor 568 (red). Fixed and permeabilized CD19-CAR T cells were stained for perforin and pZeta. Then, these cells were incubated with phalloidin, Alexa Fluor 532 (magenta), Alexa Fluor 647-(green), and Alexa Fluor 488-(cyan) conjugated secondary Abs, respectively. Scale bars represent 25.0 μ m. (**b**) Quantification of the IS quality under the lipid bilayer obtained by measuring the mean fluorescence intensities of F-actin, pZeta, and CD19 cluster, as well as the percentage of perforin-positive cells on the lipid bilayer containing CD19. Error bars show \pm s.d.



Figure S5: Standard ⁵¹Cr release assay cannot distinguish the difference between 4-1BB-CAR and CD28-CAR from cytotoxicity of CAR T cell to multiple tumor cell lines. The cytotoxicity of Kappa-CAR T (a) and CD19-CAR (b) cells from two

individuals (Donor #1 and Donor #2) was measured using a standard 4-h 51 Cr-release assay. Three Kappa positive B-cell lymphoma cell lines (Daudi, JEKO1, and BJAB) were used as Kappa-CAR T cell's target cells. The two CD19 positive B- cell lymphoma cell lines (Daudi and Raji) were used as the target cells. Error bars show \pm s. d. PBMCs from individuals were transduced with 4-1BB construct (red dots) or CD28 construct (black dots) retrovirus, as described in Figure 1.



Figure S6: Comparable IL-2 expression between CD28-CAR T and 4-1BB-CAR T cells.

(a) Representative flow cytometry analysis of IL-2 expression in CD8 positive population from CD28- and 4-1BB-CAR T cells (CD19-CAR and Kappa-CAR). (b) PBMCs from five donors transduced with 4-1BB-CAR (red dots) or CD28-CAR (black dots), respectively. The ratio (%) and expression (MFI) of IL-2 in CD8 subset were calculated. This data is pooled from at least two independent experiments. *P* value is for paired t-test.



Figure S7: 4-1BB-CAR T cells have enhanced antitumor activity and proliferation measured by long-term killing assays.

(a) Kappa-CAR T cells were isolated from four different individuals and transduced with TM, 4-1BB, and CD28 constructs. The target Daudi cells expressing fluorescent protein mCherry were mixed with CAR T cells for 7 days. The number of both target cells and CAR T cells were measured by flow cytometry, as described in Figure 4. (b) CD19-CAR T cells were isolated from four different individuals and transduced with TM, 4-1BB, and CD28 constructs. The Raji-GFP target cells were mixed with CAR T cells for 7 days. The number of both target cells and CAR T cells and CAR T cells for 7 days. The number of both target cells and CAR T cells was measured by flow cytometry. This data is pooled from at least three independent experiments.



Figure S8: 4-1BB-CAR T cells show higher IS quality from five individuals. PBMCs from five individuals were transduced with Kappa-4-1BB-CAR (red dots) or Kappa-CD28-CAR construct (black dots) retrovirus. The MFIs of F-actin, pZeta, antigen cluster, and percentage of perforin polarization at the IS from Kappa-CAR T cells were calculated. The transduced CAR T cells were activated by lipid bilayers carrying Kappa-Alexa Fluor 647 to quantify the MFI on the plasma membrane to evaluate the IS quality. This data is pooled from at least two independent experiments. *P* value is for paired t-test.



Figure S9: Superior anti-tumor activity from 4-1BB-Kappa-CAR T cells from five individuals. PBMCs from five individuals were transduced with 4-1BB Kappa-CAR (red dots) or CD28 Kappa-CAR (black dots) retrovirus. The reciprocal of the area under the curve of target cell numbers (killing efficiency, left) and area under the curve of effector cell numbers from Kappa-CAR T cells (proliferation efficiency, right) were calculated, respectively. The transduced CAR T cells were activated by co-culturing with a Kappa-positive Daudi cell line to quantify the anti-tumor activity. This data is pooled from at least two independent experiments. *P* value is for paired t-test.



Figure S10: 4-1BB-CAR-Kappa NK-92 cells show superior anti-solid tumor activity *in vivo*. (a) NSG mice (n=6) were subcutaneously injected with 2×10^6 luciferase-expressing Daudi (FFLuc-Daudi) cells mixed with Matrigel (Day -7) to mimic solid tumor xenograft animal model. At day 1, mice were injected (i.v.) with one-dose 5×10^6 effector CAR NK-92 cells (CD28 and 4-1BB). As a control, no treatment (NT) group was infused with PBS. On day 18 and 42, all the mice were subjected to intra-tumor injection of the 5×10^6 effector CAR NK92 cells and PBS (as a control). IVIS imaging was recorded using the bioluminescence value (photons/second) at the indicated time points. (b) Tumor growth inhibition (TGI) was calculated, as described previously at indicated time points. Error bars show \pm s.d.