

Supplemental material

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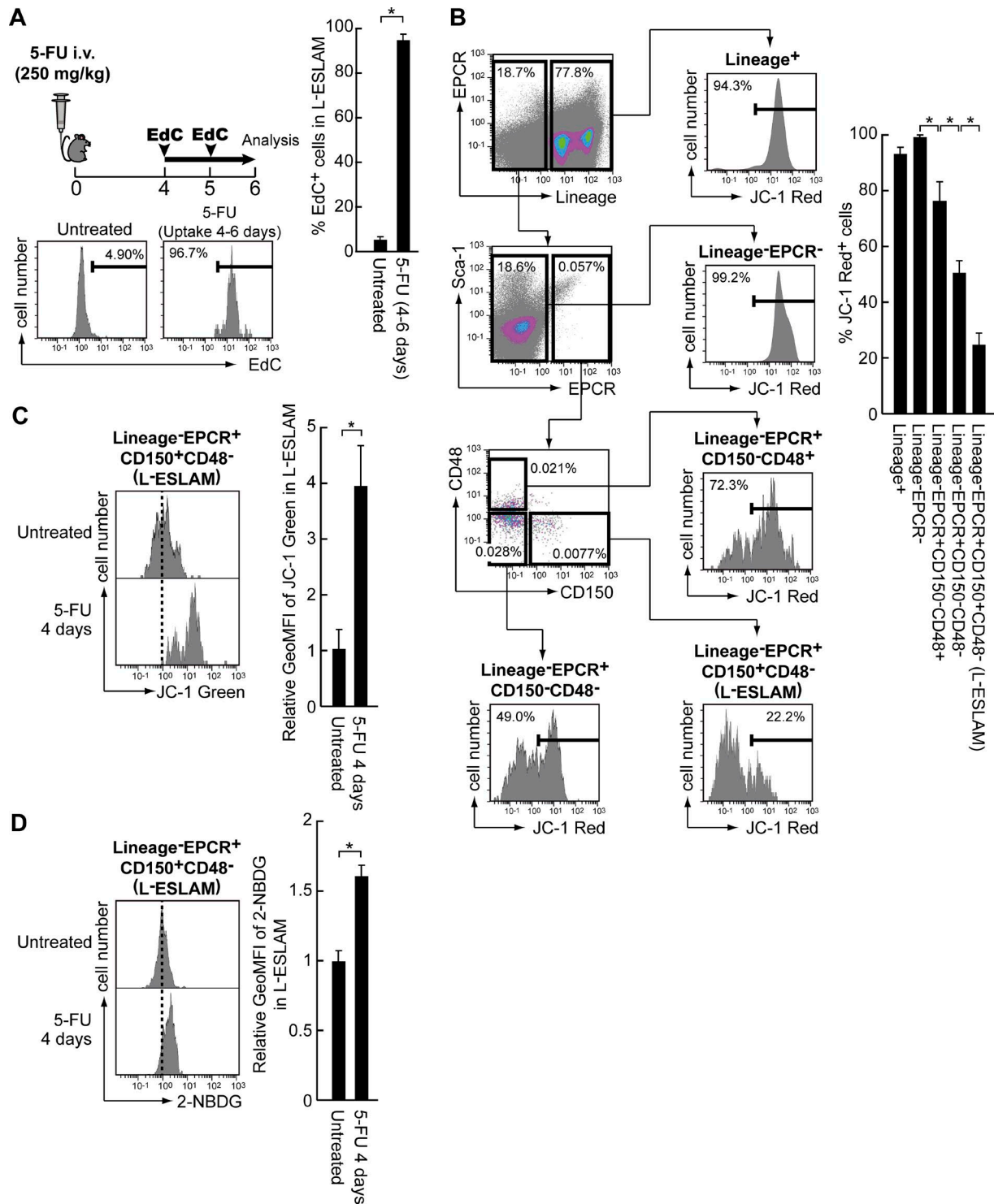


Figure S1. Cell division, mitochondrial statuses and the potential for glucose uptake of HSCs before and after 5-FU administration. **(A)** 5-FU-treated mice (250 mg/kg, i.v.) were subjected to EdC administration twice (150 mg/kg/shot i.p.) at indicated schedule. After 48 h from first EdC administration, EdC⁺ cells within Lineage⁻EPCR⁺CD150⁺CD48⁻ (L-ESLAM) HSCs were detected by flow cytometry. Numbers in histograms represent the frequency of gated cells within L-ESLAM cells. The graph depicts the frequency of EdC⁺ cells. ($n = 4$, two independent experiments). **(B)** $\Delta\Psi_m$ in indicated fractions was examined by JC-1 staining. Numbers in dot plots or histograms represent the frequency of gated cells within total BM cells or indicated fractions, respectively. The graph depicts the frequency of JC-1 Red⁺ cells within indicated fractions. ($n = 4$, two independent experiments). **(C)** After 5-FU treatment, mitochondrial mass in L-ESLAM HSCs was examined using JC-1 Green fluorescence, which reflects the height of mitochondrial mass. Histograms show the fluorescent intensity of JC-1 Green. The graph depicts geometric mean fluorescence intensity (GeoMFI) relative to the value of HSCs derived from untreated mice. ($n = 4$, two independent experiments). **(D)** L-ESLAM HSCs derived from untreated or 5-FU-treated mice were cultured with 2-NBDG, and subsequently the uptake of 2-NBDG was determined by a flow cytometry. The graph depicts geometric mean fluorescence intensity (GeoMFI) relative to the value of HSCs derived from untreated mice. Data are presented as means \pm SD ($n = 4$, two independent experiments; *, $P < 0.01$).

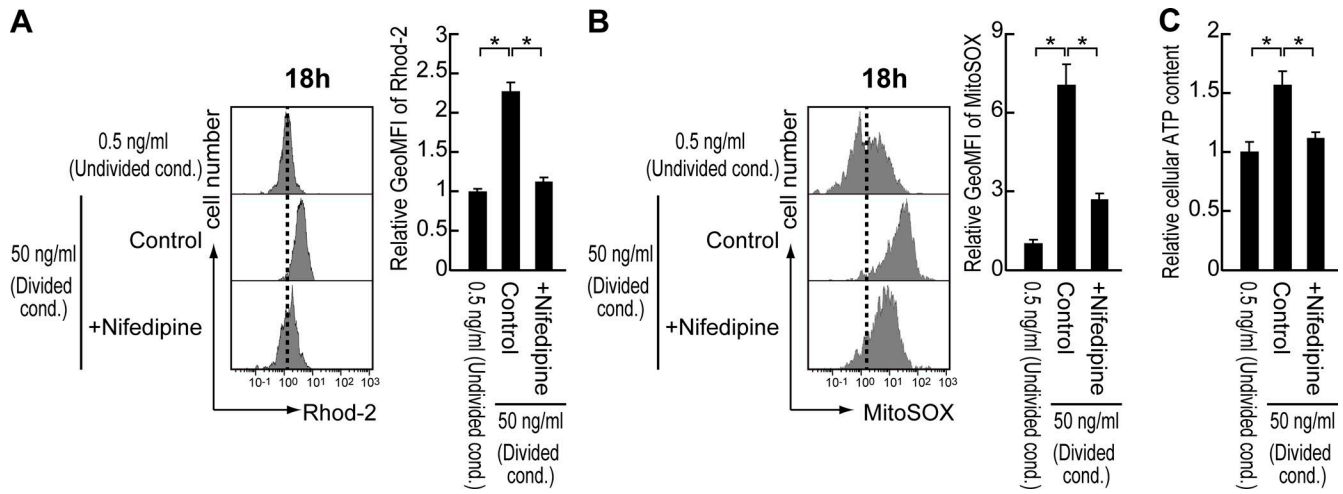


Figure S2. **Nifedipine suppressed mitochondrial functions. (A and B)** After 18 h culture under divided conditions (50 ng/ml SCF and TPO) in the absence (Control) or presence of 60 μ M Nifedipine (+Nifedipine), mitochondrial Ca^{2+} or superoxide level of culture HSCs were examined using Rhod-2 or MitoSOX. Graphs depict geometric mean fluorescence intensity (GeoMFI) relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions; $n = 4$, two independent experiments). **(C)** After the culture under divided conditions (50 ng/ml SCF and TPO) for 18 h in the absence (Control) or presence of 60 μ M Nifedipine (+Nifedipine), ATP content in L⁻ESLAM fractions of BM was examined by luminescence-based assay. The graph depicts intracellular ATP content relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions). Data are expressed as the mean \pm SD ($n = 4$, two independent experiments; *, $P < 0.01$; **, $P < 0.05$).

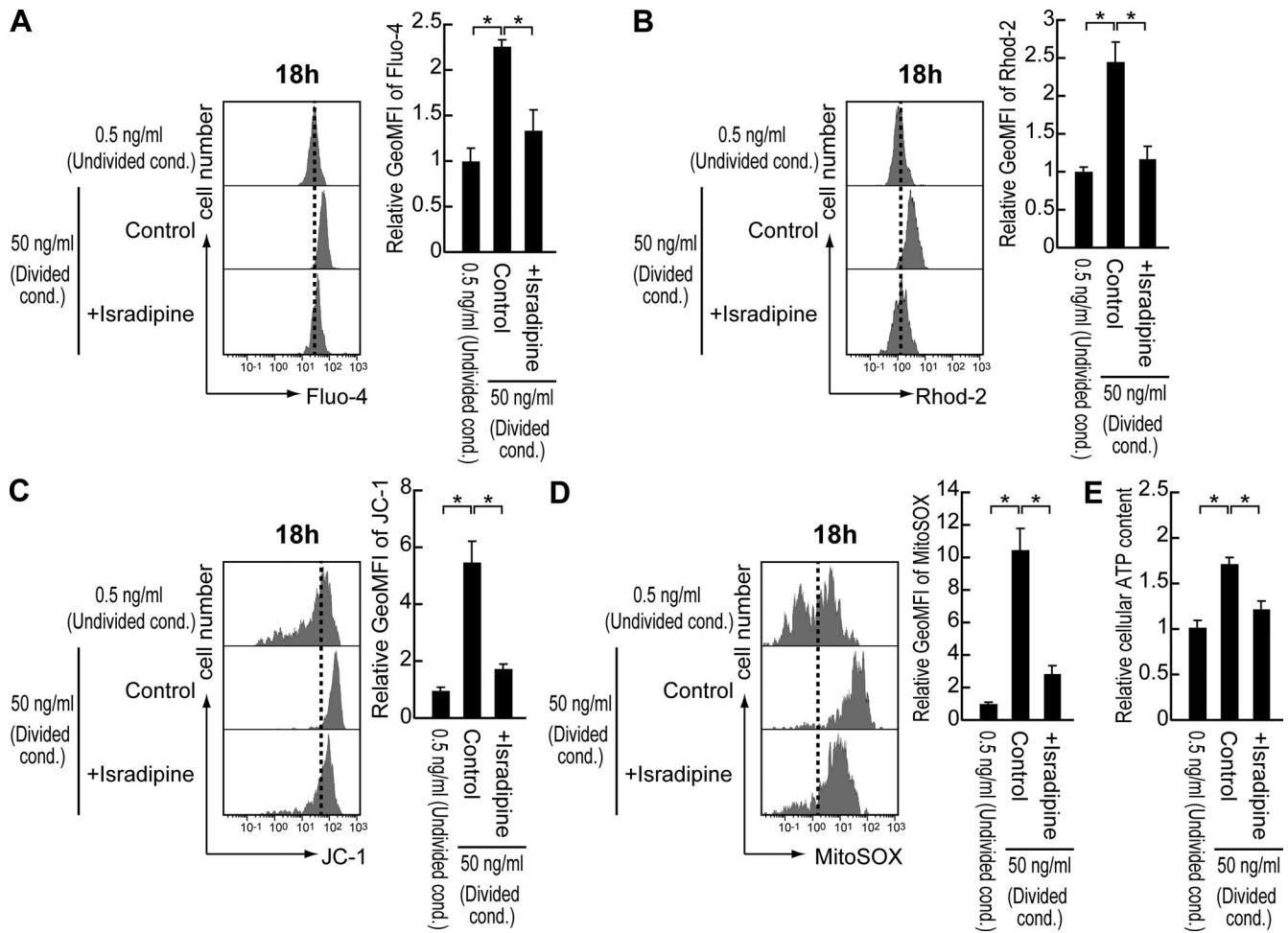


Figure S3. **Isradipine, alternative blocker for LTCCs, also suppressed Ca^{2+} -mitochondria pathway.** (A–D) After the culture under divided conditions (50 ng/ml SCF and TPO) for 18 h in the absence (Control) or presence of 50 μM Isradipine (+Isradipine), $\Delta\Psi_m$ (A), intracellular Ca^{2+} (B), mitochondrial Ca^{2+} (C), or mitochondrial superoxide level (D) of culture HSCs were examined by JC-1, Fluo-4, Rhod-2, or MitoSOX staining. Graphs depict geometric mean fluorescence intensity (GeoMFI) relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions; $n = 4$, two independent experiments). (E) After 18 h culture under divided conditions (50 ng/ml SCF and TPO) in the absence (Control) or presence of 50 μM Isradipine (+Isradipine), ATP content in cultured HSCs was examined. The graph depicts intracellular ATP content relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions). Data are expressed as the mean \pm SD ($n = 4$, two independent experiments; *, $P < 0.01$).

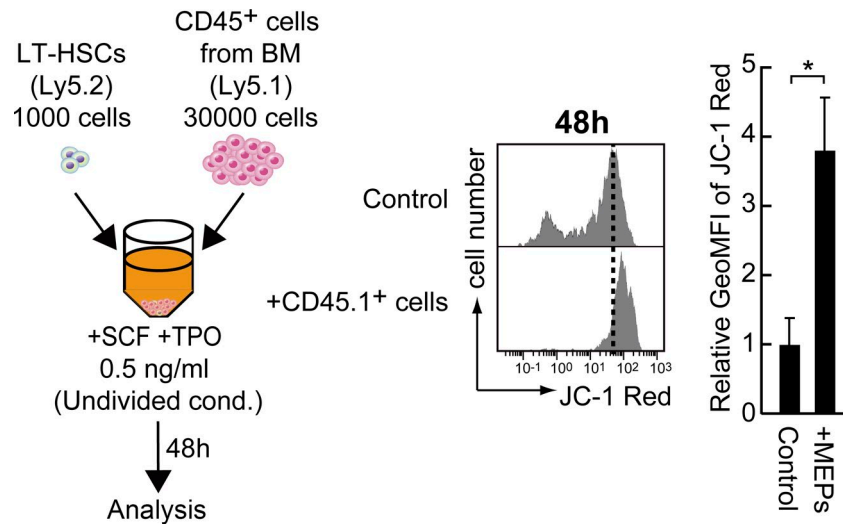


Figure S4. **Mature hematopoietic cells hardly have the potential for the suppression of $\Delta\Psi_m$ in HSCs.** Sorted HSCs (Ly5.2) were cultured with CD45⁺ cells (Ly5.1) derived from total BM for 48 h, and subsequently $\Delta\Psi_m$ of HSCs was examined by using JC-1 staining. Cells cultured without CD45⁺ cells serve as control. The graph depicts geometric mean fluorescence intensity (GeoMFI) relative to the value of cells cultured under control conditions. Data are presented as means \pm SD ($n = 4$, two independent experiments; *, $P < 0.01$).

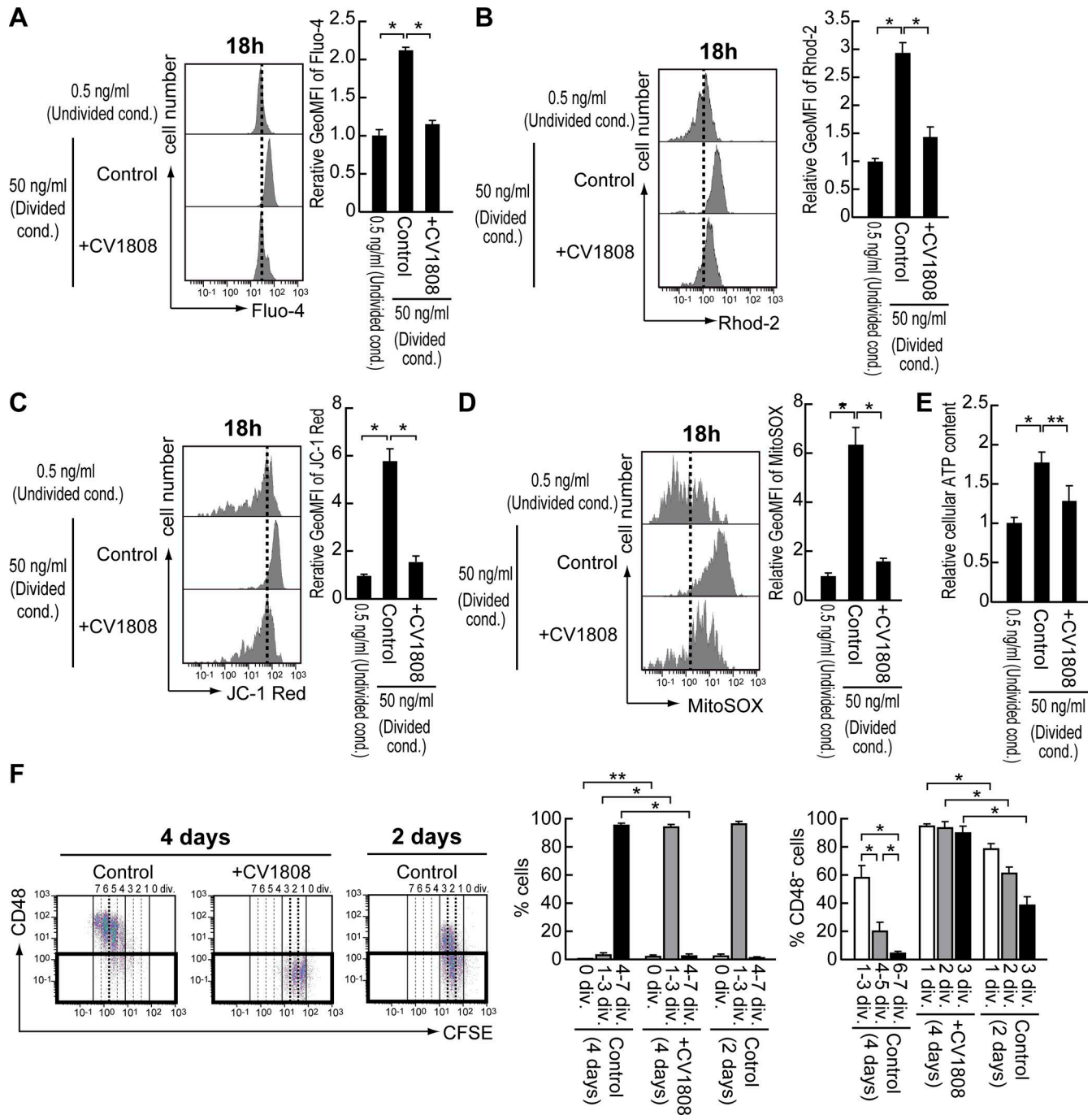


Figure S5. CV1808, an agonist of adenosine A2 receptors, suppressed Ca²⁺-mitochondria pathway and prolongs the interval of cell divisions. (A-D) $\Delta\Psi_m$ (A), intracellular Ca²⁺ (B), mitochondrial Ca²⁺ (C), or mitochondrial superoxide level (D) in HSCs after the culture for 18 h under divided conditions (50 ng/ml SCF and TPO) in the absence (Control) or presence of 50 μ M CV1808 (+CV1808). Graphs depict geometric mean fluorescence intensity (GeoMFI) relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions; $n = 4$, two independent experiments). (E) ATP content of HSCs cultured for 18 h under conditions as described above. The graph depicts intracellular ATP content relative to the value of cells treated with SCF and TPO at 0.5 ng/ml (undivided conditions; $n = 4$, two independent experiments). (F) Cell division number and expressions of CD48 in HSCs cultured under conditions as described above. The gate described by heavy line within each dot plot represents CD48⁺ fractions. Dot lines separated cells that undergoes indicated number of cell division. Graph depicts the frequency of cells undergoing the indicated number of cell divisions in whole cultured cells (left) or CD48⁺ cells within cells undergoing the indicated number of cell divisions (right). Data are presented as means \pm SD ($n = 4$, four independent experiments; *, $P < 0.01$; **, $P < 0.05$).