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# **Supplemental Data**

# **Identifying Genes Whose Mutant Transcripts**

### **Cause Dominant Disease Traits**

# by Potential Gain-of-Function Alleles

Zeynep Coban-Akdemir, Janson J. White, Xiaofei Song, Shalini N. Jhangiani, Jawid M. Fatih, Tomasz Gambin, Yavuz Bayram, Ivan K. Chinn, Ender Karaca, Jaya Punetha, Cecilia Poli, Baylor-Hopkins Center for Mendelian Genomics, Eric Boerwinkle, Chad A. Shaw, Jordan S. Orange, Richard A. Gibbs, Tuuli Lappalainen, James R. Lupski, and Claudia M.B. Carvalho



Figure S1: Dissection of quality control features for frameshifting indels and stopgain variants in the Baylor-CMG database

Box plots display variant read (vR) to total read (tR) ratio (vR/tR) and quality score values across variants called in the Baylor-CMG database with vR=2,3,4,5 and 6. vR/tR plateaus when vR reaches 5. Therefore, in the extraction of high-quality frameshifting indels and stopgain variants, the criteria that vR should be at least 5 reads was used.



Figure S2: Variant filtering criteria for indels in Baylor-CMG and ARIC database

Variant prioritization workflow for frameshifting indels in the Baylor-CMG and the ARIC database was applied as follows. If an on-target indel has a variant read number (vR) >=5, it was included for further analysis. Then the indels were filtered based on the minor allele frequencies in our internal database (MAF <= 0.01). At this step, in-frame indels were also removed from the analysis.



Figure S3: Sensitivity analysis in terms of the overlap of top 5% depleted genes for NMD<sup>-</sup> variants in ARIC vs. ExAC databases using: synonymous variant count normalization and coding sequence length normalization at different MAF cutoffs.

The sensitivity analysis was performed at different MAF cutoffs (0.1, 0.05, 0.01, 0.005 and 0.001) using synonymous variant count normalization (Left panel) and coding sequence length normalization (Right panel). The overlap between top 5% depleted genes for NMD<sup>-</sup> variants in ARIC vs. ExAC databases is similar using synonymous variant count normalization (29.2%) and coding sequence length normalization (30.3%) at MAF <= 0.01.





The sensitivity analysis was performed at different MAF cutoffs (0.1, 0.05, 0.01, 0.005 and 0.001) using synonymous variant count normalization (Left panel) and coding sequence length normalization (Right panel). The overlap between top 5% depleted genes for NMD<sup>-</sup> variants in either control database and our control OMIM list of genes that cause disease via potential gain-of-function (N=39) drops significantly using coding-sequence length normalization (28.2%) compared to synonymous variant count normalization (33.33%) at MAF <= 0.01.

#### NMDEscPredictor



### Figure S5: An example of NMD prediction of a frameshifting indel in

NMDescPredictor web-based tool (<u>https://nmdpredictions.shinyapps.io/shiny/</u>)



Figure S6: The normalization by the number of synonymous variants to calculate the expected number of NMD<sup>-</sup> variants

Number of expected NMD<sup>-</sup> variants per each canonical transcript was calculated as follows: The total # of variants was multiplied by the ratio of # of rare synonymous variants in NMD<sup>-</sup> region to the total # of rare synonymous variants.



The correlation is -0.27

Total number of variants in each tissue

Figure S7: Correlation between the number of variants in each GTEx tissue and corresponding p-values.



Figure S8: An example of a frameshifting indel in the fourth exon (nearly in the middle of the *BTN2A1* gene) was predicted to lead to NMD<sup>-</sup> by our tool. Allele-specific expression data in GTEx is concordant with this prediction.

The variant chr6:26,465,566\_CAA>C that is located in exon 4 of transcript ENST0000042938 (7 coding exons in total) belonging to the *BTN2A1* gene was predicted to be NMD<sup>-</sup> by NMDEscPredictor based on the location of the boundary PTC. The average ratio of variant read count to total read count for this variant was extracted from GTEx RNA-Seq data and quantified as 0.478. This experimental finding supported the computational prediction of this variant as NMD<sup>-</sup> by NMDEscPredictor.



#### Figure S9: Pipeline workflow for our algorithm

At the first step of the algorithm, as a quality control (QC) step, high-quality and rare (MAF <=0.01) frameshifting indels and stopgain variants were extracted from the Baylor-CMG (disease database, 6,109 exomes) and the ExAC and the ARIC control databases (60,706 and 10,940 exomes, respectively). Then, using the NMDEscPredictor algorithm, frameshifting indels and stopgain variants in each database were categorized into three categories as NMD escaping (NMD<sup>-</sup>), NMD triggering (NMD<sup>+</sup>). We then removed the variants that could not be annotated to any canonical transcript in Ensembl version 19 as well as variants mapped to transcripts without a predicted PTC, without a boundary PTC or mapped to singleexon canonical transcripts. Next, each gene in the genome is ranked based on the depletion of NMD<sup>-</sup> relative to NMD<sup>+</sup> variants in control databases (NMD escape intolerance score metric). This analysis revealed a total of 1, 996 genes as the most depleted in either database (i.e. ranked in the top 5%). Those genes were further investigated for NMD<sup>-</sup> variants in the Baylor-CMG database (disease database). A subset of significantly depleted genes has NMD<sup>-</sup> variants in multiple unrelated individuals with similar clinical phenotypes (based on the phenotypic similarity scoring) in the Baylor-CMG database. Some of those genes were found to be causative for human disease through escape from NMD and include novel and known disease genes.



Figure S10: The overlapping number of genes depleted for NMD<sup>-</sup> variants (i.e. ranked in the top 5%) in each category of truncating variants within ARIC and ExAC database

Venn Diagrams display the number of genes depleted for NMD<sup>-</sup> variants (i.e. ranked in the top 5%) in -1 frame, +1 frame and stopgain categories in the ARIC and ExAC control databases.



ARIC and ExAC 
Neither ARIC or ExAC • Only ARIC • Only ExAC

Figure S11: Expected and observed number of NMD<sup>-</sup> variants in each category of truncating variants in the ARIC and ExAC control databases

The plots show the expected # of escape (NMD<sup>-</sup>) variants (x axis) compared to the observed # of escape (NMD<sup>-</sup>) variants (y axis) per gene in the -1 frame, +1 frame and stopgain categories in the ARIC and ExAC control databases. The genes for which the observed # of variants/gene relative to expected # of variants/gene were depleted in both databases were colored in orange, and those depleted only in ARIC database were colored in purple and those not depleted in either control database were colored in dark green.

#### **Permutation test**



The number of overlapping genes in each permutation

# **Figure S12:** Permutation test results to quantify how often overlap between top 5% depleted genes for NMD<sup>-</sup> variants in ARIC vs. ExAC control database might occur by chance

10,000 permutations were done by generating random subsets of all of the genes considered in the analysis (N=16,411) at the size of ExAC gene set (N=1,385). In each permutation, the number of genes overlapping between ARIC and random set of genes were calculated. The red dashed line shows the observed value (N=252) of the number of genes overlapping between top 5% depleted genes for NMD<sup>-</sup> variants in ARIC (N=863) vs. ExAC database (N=1,385).



# Figure S13: The percentage of overlapping frameshift variants between the ExAC and ARIC databases at different MAF cutoffs

The number of overlapping frameshift variants (light blue boxes) between the ExAC and ARIC databases were shown at different MAF intervals including 0-0.0019, 0.0019-0.001, 0.001-0.01 and 0.01-0.1.



# Figure S14: The percentage of overlapping stopgain variants between the ExAC and ARIC databases at different MAF cutoffs

The number of overlapping stopgain variants (light blue boxes) between ExAC and ARIC databases were shown at different MAF intervals including 0-0.0019, 0.0019-0.001, 0.001-0.01 and 0.01-0.1.