

FIGURE S1. Phenotype and sorting purity of intestinal CX₃CR1⁺CD11c⁺ macrophages.

(A) CD11c⁺ cells isolated from small intestinal lamina propria of CX₃CR1^(GFP/+) mice were analyzed for expression of Siglec F/CD169 in the intestinal CX₃CR1⁺CD11c⁺ macrophages. (B) CD11c⁺ cells were isolated from lamina propria of small intestine of CX₃CR1^(GFP/+) mice and sorted to CX₃CR1⁺ and CX₃CR1⁻ subsets. Purity of sorted subsets from intestinal CD11c⁺ cells were about 90%. (C) Total IgA and IgG level from culture supernatants of CD11c⁺CX₃CR1⁺ macrophages co-cultured with B cells or each cell alone in presence of anti-IgM (10 μg/ml). (D-F) The level of total IgA and IgG were determined from culture supernatants of B cells co-cultured with CD11c⁺CX₃CR1⁺ macrophages (D) in presence of 10 μg/ml anti-mouse IL-10 or (E and F) CpG (1 μM) and MyD88 inhibitor (100 Mm, IMG2005; Novus Biologicals, Littleton, CO). Data are representative of three independent experiments. Graphs show mean ± SEM. ns; not significant, *p < 0.05, **p < 0.01 and ***p < 0.001 using one-way ANOVA.

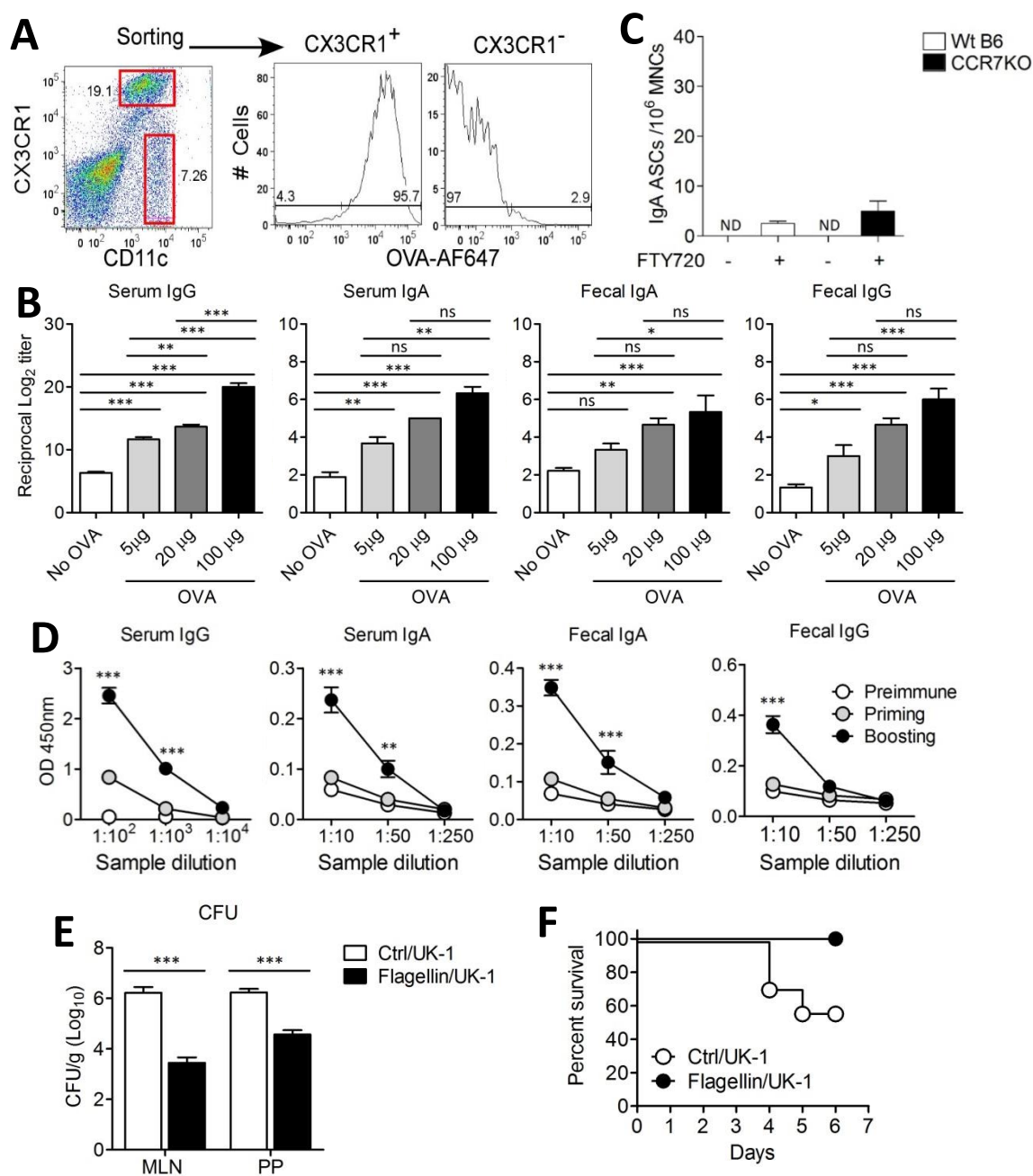


FIGURE S2. Preferential uptake of circulatory Ag by intestinal CX₃CR1⁺ macrophages.

(A) CX₃CR1^(GFP/+) mice *i.v.* injected with 100 μg OVA-AF647. At 1hr following injection, CD11c⁺ cells were isolated from small intestinal lamina propria of CX₃CR1^(GFP/+) mice and sorted to CX₃CR1⁺ and CX₃CR1⁻ subsets. Fluorescence signal from OVA-AF647 was analyzed to investigate the preferential uptake of circulatory antigen by CD11c⁺CX₃CR1⁺ macrophages. (B) WT B6 mice were *i.v.* injected with three different doses of OVA and boosted 2 weeks later. Serum and feces were collected 1 week after boosting (n=3). The titers of OVA specific serum IgG, serum IgA, fecal IgA and fecal IgG were analyzed. (C) The generation of circulatory Ag specific IgA ASCs was not induced in the MLN. WT B6 and CCR7 KO mice were *i.v.* immunized with 100 μg OVA and boosted 2 weeks later. Mice were treated with FTY720 and then OVA specific IgA ASCs were determined in the MLN. ND, not detected. (D-F) WT mice were *i.v.* injected with 2 μg flagellin isolated from *Salmonella Typhimurium* and boosted 2 weeks later. At 10 days following final boosting, mice were challenged orally with 10⁷ CFU of wild-type invasive *Salmonella Typhimurium*. (D) Serum and feces were collected 1 week after boosting (n=10). The level of flagellin specific serum IgG, serum IgA, fecal IgA and fecal IgG were analyzed. (E) CFU from PP and MLN were determined at day 6 following challenge and (F) survival rates were monitored.

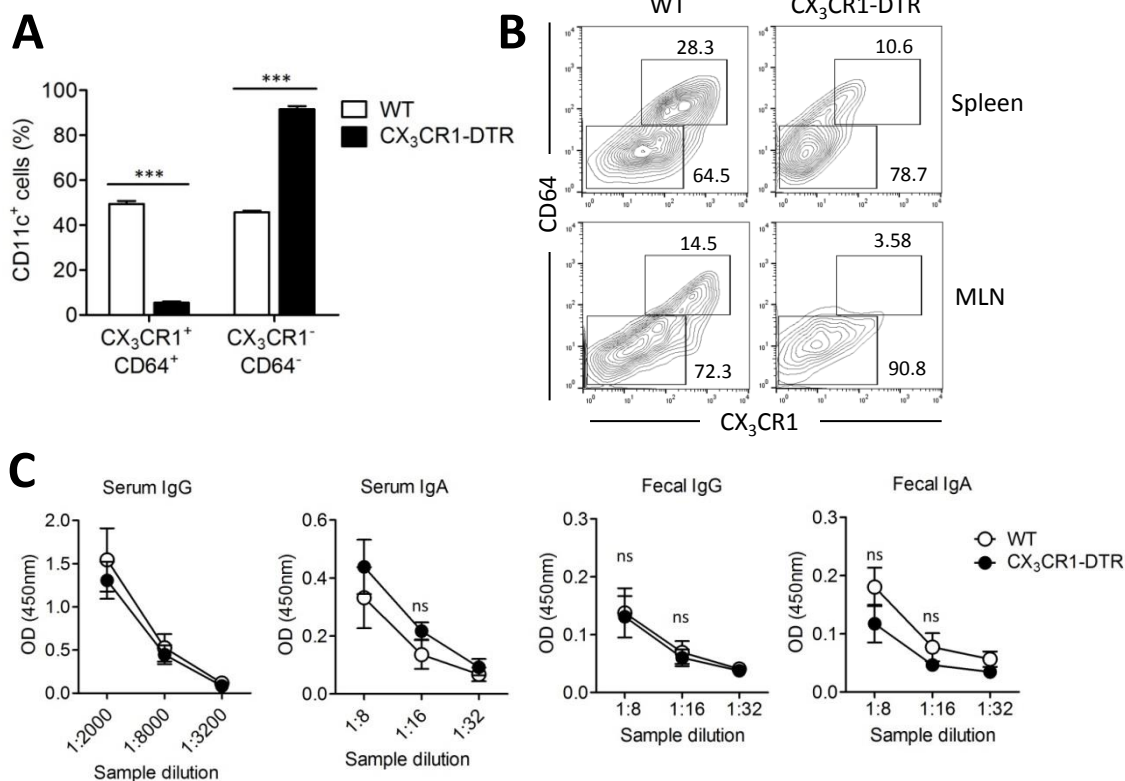


FIGURE S3. Generation of antigen specific IgA antibodies in the gut upon delivery of antigen by oral gavage partially reduced in absence of CX₃CR1⁺ macrophages.

For depletion of CX₃CR1⁺ macrophages, mice were *i.p.* injected with 200 ng diphtheria toxin before 1 day and after 2 days of OVA immunization. **(A-B)** To confirm the depletion of CD11c⁺CX₃CR1⁺ macrophages, CD11c⁺ cells were isolated from the small intestinal lamina propria, spleen, and MLN. CD64⁺CX₃CR1⁺ cells were analyzed from the CD11c⁺ population. **(A)** The percentages of CD64⁺CX₃CR1⁺ population in the lamina propria among CD11c⁺ cells were summarized. **(B)** Depletion of CD64⁺CX₃CR1⁺ cells in the spleen and MLN. **(C-F)** WT B6 and CX₃CR1-DTR mice were treated with 10 mg OVA with 10 μg cholera toxin and boosted 2 weeks later. Serum and feces were collected 1 week after 2nd immunization (n=5). **(C)** OVA specific serum IgG, **(D)** serum IgA, **(E)** fecal IgG and **(F)** fecal IgA were analyzed. Graphs show mean ± SEM. ns; not significant, using Student's *t*-test.

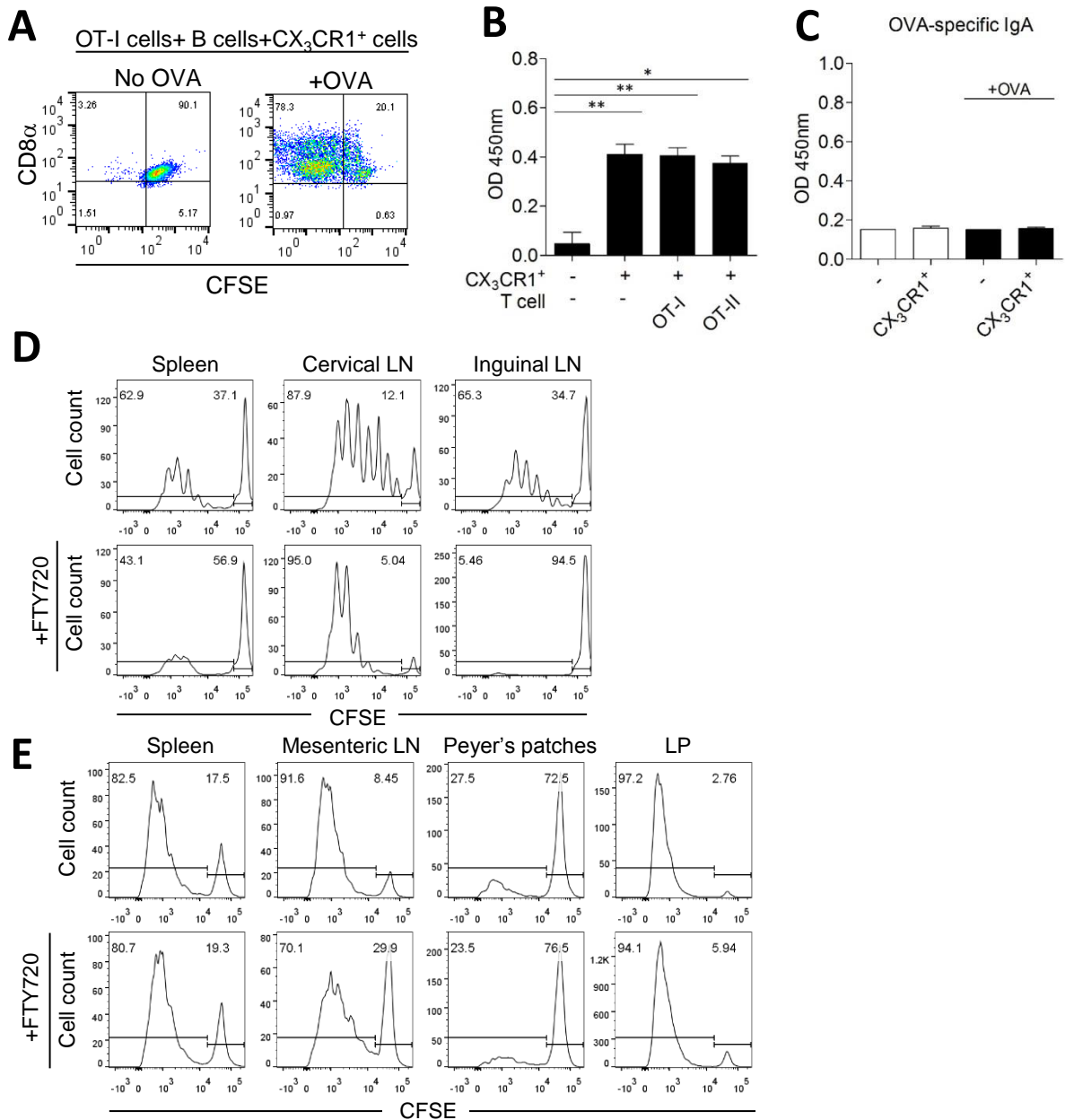


FIGURE S4. CD8⁺ T cells could not enhance IgA production without cognate Ag.

(A-C) CD11c⁺ cells of SI LP from CX₃CR1^(GFP/+) mice were isolated and sorted to CX₃CR1⁺ cells. Splenic B cells stimulated with anti-IgM were co-cultured with OVA pulsed CD11c⁺CX₃CR1⁺ macrophages and OT-I or OT-II cells. (A) CFSE dilution of co-cultured OT-I cells were analyzed. (B) Total IgA was measured from culture supernatant of B cells co-cultured with CX₃CR1⁺ macrophages with OT-I or OT-II cells in absence of OVA. (C) OVA-specific IgA was measured from culture supernatant of B cells co-cultured with CX₃CR1⁺ macrophages in the presence of OVA. Graphs show mean ± SEM. *p < 0.05 and **p < 0.01 using one-way ANOVA with Tukey's post hoc test. (D-E) OT-I CD8⁺ T cells labeled with 9 μM CFSE were adoptively transferred to CD45.1⁺ WT B6 mice. (D) Next day these mice were *s.c.* immunized with 20 μg OVA plus 1 μg cholera toxin and *i.p.* treated with 20 μg FTY720. At 2 days later, FTY720 was treated again. After 3 days, CFSE dilution of CD45.2⁺CD8α⁺ cells were analyzed from the spleen, cervical LN, and inguinal LN. (E) Next day following OT-I cell transfer, OVA was *i.v.* injected and CFSE dilution of CD45.2⁺CD8α⁺ cells were analyzed from the spleen, MLN, Peyer's patches (PP), and small intestine lamina propria (LP) at day 5 following OVA injection.