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Supplemental Information

Efficient Gene Transfer

to the Central Nervous System

by Single-Stranded Anc80L65

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Α

A - DAY 3

DORSAL



Anc80L65-CMV-Luc





4.0

3.0

1.0

(p/sec/cm²/sr Color Scale Min = 3.37e5 Max = 5.26e6





B - DAY 42 AAV9-CMV-Luc

DORSAL

Anc80L65-CMV-Luc



VENTRAL











Ε



Anc80L65

AAV9

scAAV9



AAV9

Microglia

scAAV9

AAV9

Anc80L65





SUPPLEMENTAL FIGURES LEGENDS

Supplementary Figure 1. Kinetics and localization of bioluminescence signal after intravenous injection of BALB/c with Anc80L65-CMV-FLuc or AAV9-CMV-FLuc.

Mice were injected with 2x10¹² gc/kg of vector and imaged at days 3, 7, 21, and 42 post injection. (A) Bioluminescence signal from liver region. A region of interest (ROI) was drawn on each image and photons/sec calculated. Left: graph depicting kinetics of expression. Right: representative images of mice injected with AAV9 or Anc80 at days 3, 7, and 21 post injection. (B) Bioluminescence signal from head region. ROI was drawn on each image and photons/sec calculated. Left: graph depicting and photons/sec calculated. Left: graph depicting kinetics of expression. (B) Bioluminescence signal from head region. ROI was drawn on each image and photons/sec calculated. Left: graph depicting kinetics of expression. Right: representative images of mice injected with AAV9 or Anc80L65 at days 3, 7, and 21 post injection. n=5 mice/group.

Supplementary Figure 2. Whole-body bioluminescence images at days 3 and 42 post intravenous injection of Anc80L65-CMV-FLuc or AAV9-CMV-FLuc

Representative bioluminescence images of BALB/c mice injected systemically with 2x10¹² gc/kg of AAV9-CMV-FLuc or Anc80-CMV-FLuc, at 3 and 42 days after injection in the lateral tail vein.

Supplementary Figure 3. Quantification of AAV genome-copy numbers in peripheral tissues after systemic injection with Anc80L65-CMV-eGFP, AAV9-CMV-eGFP and scAAV9-CMV-eGFP Genomic DNA qPCR analysis was performed to measure the gc/diploid cell in liver (A), heart (B), quadriceps (C) and diaphragm (D), n=3 mice per group; two-way ANOVA followed by Tukey's multiple comparison test; *p<0.05. (E) Representative images of eGFP fluorescence signal detected across the liver after intravenous injection of the same dose of Anc80L65, AAV9 and scAAV9 in wild-type mice $(4x10^{13}gc/kg)$.

Supplementary Figure 4. Anc80L65, AAV9 and scAAV9 transduction of intraparenchymal and vessel-associated astrocytes

Double immunostaining with Lycopersicon Esculentum lectin ("Lectin") and eGFP shows that both astrocytic endfeet and parenchymal astrocytes were equivalently transduced by Anc80L65, AAV9 or scAAV9 after systemic infusion. Scale bar: 50µm

Supplementary Figure 5. Absence of microglial transduction after intravenous delivery of Anc80L65, AAV9 and scAAV9 harboring a self-complementary genome

Double immunostaining with Iba-1 and eGFP was performed in order to determine if Anc80L65, AAV9 or AAV9 containing a self-complementary genome (scAAV9) transduced microglial cells. As shown on those representative images, no colocalization between the two markers was detected. Scale bar: 50µm

Supplementary Figure 6. Vector and experimental design

(A and B) Schematic representation of the expression cassette used to produce the viral vectors. (A) For the bioluminescence experiments a multicistronic vector with a T2A peptide signal was used. A 133 base pairs chimeric intron (I) and the viral SV40 polyA signal (SVPA) were introduced in the cassette. (B) For the systemic and intracerebral injection, the self-complementary (sc) vector 5' ITR contained a deletion of the D-sequence and the terminal resolution site mutation (trs). The transgene design (CMV.eGFP.WPRE.bGH, 2266 base pairs) was identical for the self-complementary and single stranded vectors, and included a 133 bp chimeric intron (I) and the bovine growth hormone polyA signal (bGH). (C) Table including the vector, dose and route of administration used in all the experiments.