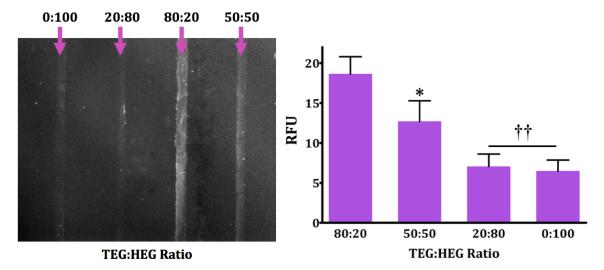
## Supporting Information

## Aptamer Recognition of Multiplexed Small-Molecule-Functionalized Substrates

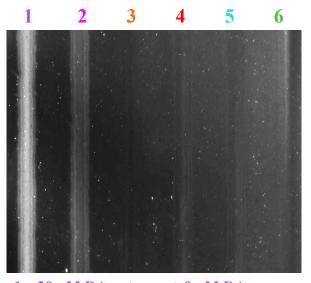
Nako Nakatsuka,<sup>1,2</sup> Huan H. Cao,<sup>1,2</sup> Stephanie Deshayes,<sup>3</sup> Arin L. Melkonian,<sup>3</sup> Andrea M. Kasko,<sup>2,3</sup> Paul S. Weiss,<sup>1,2,4</sup> and Anne M. Andrews<sup>1,2,5\*</sup>

<sup>1</sup>Department of Chemistry and Biochemistry, <sup>2</sup>California NanoSystems Institute, <sup>3</sup>Department of Bioengineering, <sup>4</sup>Department of Materials Science and Engineering, <sup>5</sup>Department of Psychiatry and Biobehavioral Sciences, Semel Institute for Neuroscience & Human Behavior, and Hatos Center for Neuropharmacology, University of California, Los Angeles, Los Angeles, CA 90095, United States

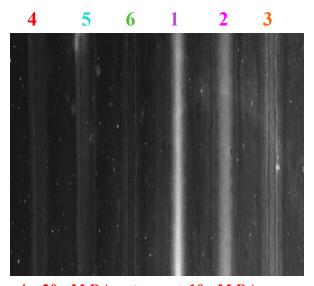
\*To whom correspondence should be addressed: <u>aandrews@mednet.ucla.edu</u>



**Figure S1.** Optimizing surface-tethered small molecule densities. **(Left)** Different ratios of hydroxyl-terminated alkanethiols (background molecules; hydroxyl triethylene glycol (undecanethiol) (TEG)) *vs.* carboxyl-terminated alkanethiols (small-molecule tethers; carboxyl hexa(ethylene glycol) undecanethiol (HEG)) were incubated in each channel. Representative fluorescence image for dopamine aptamer recognition of dopamine functionalized channels. **(Right)** Maximal relative fluorescence intensities were observed for 80% TEG:20% HEG. This ratio was used for all subsequent experiments. Error bars are standard errors of the means for *N*=3 substrates. Group means were significantly different \**P*<0.05 and ++*P*<0.01 *vs.* the 80:20 TEG:HEG ratio.

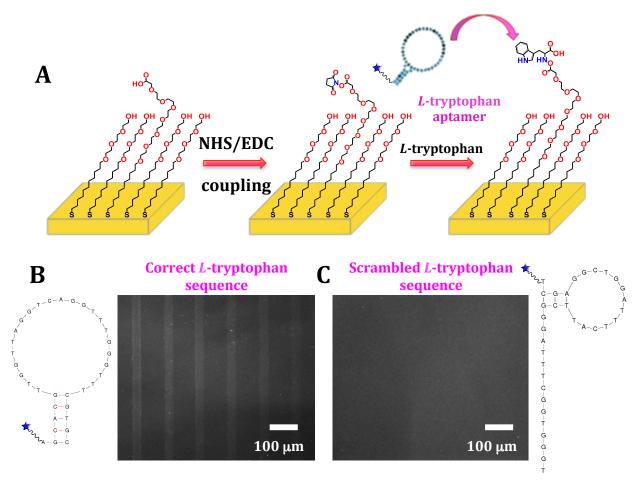




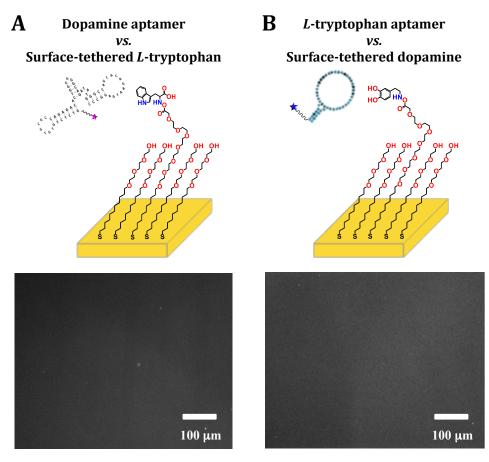


 $4 - 20 \mu M DA aptamer + 10 \mu M DA$  $5 - 20 \mu M DA aptamer + 20 \mu M DA$  $6 - 20 \mu M DA aptamer + 50 \mu M DA$ 

**Figure S2.** Representative fluorescence images for the competitive displacement experiment to investigate reversible binding of the dopamine (DA) aptamer to surface-tethered dopamine. A different concentration of free dopamine (0-50  $\mu$ M) was added to each channel in different orders on different substrates. Binding was quantified for *N*=4 substrates in Figure 3A.



**Figure S3.** *L*-Tryptophan aptamer capture on patterned substrates. **(A)** Schematic (not to scale) of patterning and functionalization of *L*-tryptophan. **(B)** Substrates were incubated with the 34-base *L*-tryptophan-specific aptamer sequence or **(C)** a scrambled sequence with the same numbers of each nucleotide as the correct sequence but randomized to generate a different secondary structure. The secondary structure of the correct and scrambled sequences were generated using *Mfold*. Substrates were imaged at an emission wavelength of 525 nm for AlexaFluor<sup>®</sup> 488 (excitation at 490 nm).



**Figure S4.** Selectivity of dopamine and *L*-tryptophan aptamers. **(A)** Patterned *L*-tryptophan-functionalized substrates imaged at an emission wavelength of 605 nm for AlexaFluor® 546 (excitation at 556 nm) to visualize bound dopamine aptamers. **(B)** Patterned dopamine-functionalized substrates imaged at an emission wavelength of 525 nm for AlexaFluor® 488 (excitation at 490 nm) to visualize bound *L*-tryptophan aptamers. In both cases, no observable patterns were detected indicating minimal cross-reactivity for the incorrect targets.