

**Additional file 1: Scheme S1, Table S1–S9, and Figures S1–S8**

**Integrative metagenomic and biochemical studies on rifamycin  
ADP-ribosyltransferases discovered in the sediment microbiome**

Jae Hong Shin<sup>1,†</sup>, Hyunuk Eom<sup>2,†</sup>, Woon Ju Song<sup>2,\*</sup>, Mina Rho<sup>1,3,\*</sup>

<sup>1</sup>Department of Computer Science and Engineering, Hanyang University, Seoul, Korea

<sup>2</sup>Department of Chemistry, Seoul National University, 1 Gwanak-ro, Gwanak-gu, Seoul, 08826, Korea

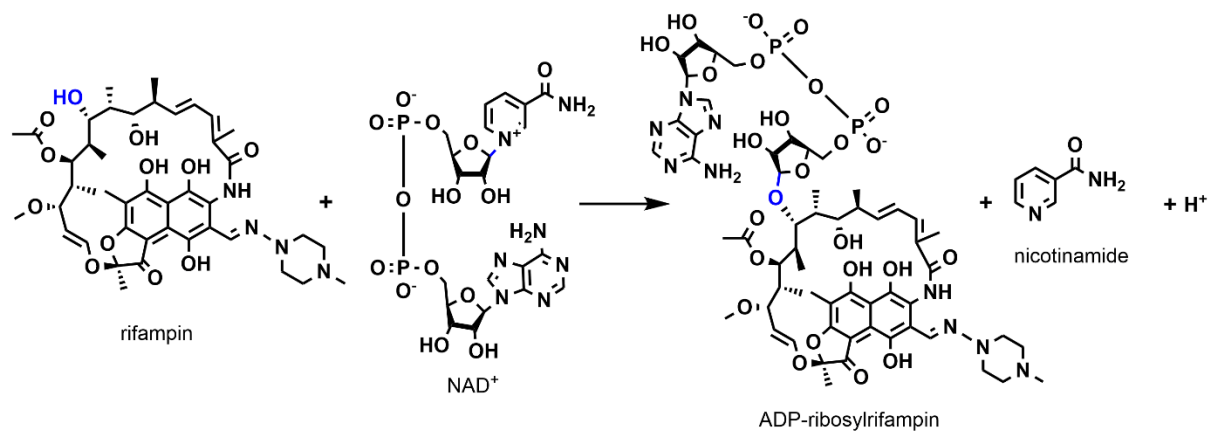
<sup>3</sup>Department of Biomedical Informatics, Hanyang University, Seoul, Korea

<sup>†</sup>These authors contributed equally to this work.

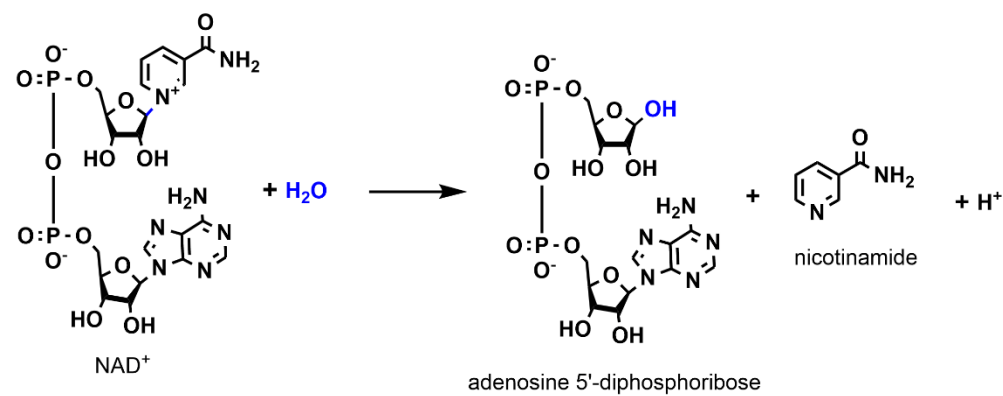
\*Corresponding authors: [woonjusong@snu.ac.kr](mailto:woonjusong@snu.ac.kr), [minarho@hanyang.ac.kr](mailto:minarho@hanyang.ac.kr)

**Scheme S1.** Reaction of mono-ADP-ribosyltransferase (a) ADP-ribosylation of rifampin. (b) Uncoupled reaction of NAD<sup>+</sup> consumption.

(a)



(b)



**Table S1.** A total of 45 rifamycin ADP-ribosyl transferase (*arr*) homologous genes obtained from the sediment microbiome

**Table S2.** Protein sequence similarity between 45 newly identified *arr* genes and the genes in the complete genomes

**Table S3.** The list of nucleotide and amino acid sequences of four representative Arr-wd proteins involved in rifamycin-resistance

**Table S4.** Primer sequences used for the current work.

**Table S5.** ADP-ribosylated product analysis by ESI-MS.

**Table S6.** Steady-state kinetic parameters of Arr-wd proteins in ADP-ribosylation of rifampin and its analogues.

**Table S7.** Substrate dissociation constants ( $K_D$ ) measured from the intrinsic fluorescence quenching.

**Table S8.** Steady-state kinetic parameters of Arr-wd3 variants in ADP-ribosylation of rifampin analogues.

**Table S9.** Determination of minimum inhibitory concentration (MIC) of rifampin and its analogues.

**Table S1.** A total of 45 rifamycin ADP-ribosyl transferase (arr) homologous genes obtained from the sediment microbiome.

>W2\_1488688  
DGIVGPFHGHADLSPGDLIAPGYPSHYGARKLASWVYMTRIVAGAVLAAEIAAGEGRARVYLVEPTGPFEDDPNVTDKKFP  
GNPTQSYRTKDPLRVLEEVLWDWAPTPAERLAQLRDFVERTREGIEAID

>W2\_1852679  
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FPALYFNIAECYEDLGDENAKKNYELADSFHKHPSDKGPFYHGTRADLQVGDLLTAGGKSNYKAELTMNHIFTALVNGAGLA  
AALVGGDGRERVYIVEPTGSFENDPNVTDKKFPGNPTRSYRSEAPLEIVGEVTDWVKITPEELQKWREKLANNKGEIIN

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>W2\_1316525  
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PGNPTKSYRTRDPLRVTVGEVADWPGHPPEQLQAMRDHLDRLESGIEAIED  
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DDPNLTDKKFPGNPTKSYRSKHPFKVVEITIWQGHSPQVKAMNEGLAKLKEQGIEAIED  
>W2\_921462  
TDGPKPFVHESGALLHGTADLAIGDMLVPGYLANYGERRANHVYVETLDAAAWAAELAVGPGRGRIYIIVEPTGPLEDDP  
NLTDKKFPGNPTSYRTRAPVKVVGELVDWVGHSSEEQQAMRDGLADLERGLAIVEDD

**Table S2.** Protein sequence similarity between 45 newly identified *arr* genes and the genes in the complete genomes

Arr gene name	Accession number	Species name	Sequence similarity (%)
W2_1488688	WP_069949724.1	<i>Arthrobacter sp. U41</i>	57.81
W2_1852679	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	74.39
W2_2666971	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	76.83
W2_399810	WP_077133846.1	<i>Spirosoma montaniterrae</i>	80.14
W2_931432	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	75.20
W2_1076727	WP_053195305.1	<i>Herbaspirillum hiltneri</i>	82.35
W2_1252150	WP_096357565.1	<i>Mucilaginibacter gotjawali</i>	76.22
W2_493312	WP_015132086.1	<i>Calothrix sp. PCC 7507</i>	66.93
wd4	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	75.61
W2_1524164	WP_061991201.1	<i>Flammeovirgaceae bacterium 311</i>	56.59
W2_2157429	WP_013239403.1	<i>Clostridium ljungdahlii</i>	98.54
W2_1786964	WP_048919688.1	<i>Rufibacter sp. DG31D</i>	71.97
W2_1931402	WP_077133846.1	<i>Spirosoma montaniterrae</i>	73.79
W2_611506	WP_061991201.1	<i>Flammeovirgaceae bacterium 311</i>	57.36
W2_2027090	WP_011585690.1	<i>Cytophaga hutchinsonii</i>	77.78
W2_1316525	WP_077202317.1	<i>Vibrio</i>	78.83
W2_1614168	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	78.46
W2_705194	WP_077133846.1	<i>Spirosoma montaniterrae</i>	74.45
W2_691914	WP_066405829.1	<i>Flavisolibacter tropicus</i>	81.89
W2_1580944	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	71.66
W2_456471	WP_069243308.1	<i>Burkholderia latens</i>	81.25
W2_1294492	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	67.07
W2_2321962	WP_077133846.1	<i>Spirosoma montaniterrae</i>	77.24
W2_1976254	WP_080238844.1	<i>Spirosoma rigui</i>	78.03
W2_2509215	WP_013930076.1	<i>Runella slithyformis</i>	66.89
W2_436021	WP_075153047.1	<i>Sphingomonas koreensis</i>	70.77
W2_982741	WP_080238844.1	<i>Spirosoma rigui</i>	79.70
W2_2284918	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	72.06
W2_928841	WP_084490216.1	<i>Niabella ginsenosidivorans</i>	74.80
W2_1056966	WP_015132086.1	<i>Calothrix sp. PCC 7507</i>	73.72
W2_759404	WP_080238844.1	<i>Spirosoma rigui</i>	71.94
W2_505284	WP_015175623.1	<i>Oscillatoria nigro-viridis</i>	81.42
W2_2302663	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	80.88
W2_1051501	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	81.62
W2_1844246	WP_015175623.1	<i>Oscillatoria nigro-viridis</i>	82.27
W2_1355083	WP_015175623.1	<i>Oscillatoria nigro-viridis</i>	85.62
W2_1008891	WP_015132086.1	<i>Calothrix sp. PCC 7507</i>	84.29
Wd3	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	80.88
W2_498816	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	85.07
W2_1851401	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	82.84
W2_1182683	WP_015171984.1	<i>Geitlerinema sp. PCC 7407</i>	84.78
Wd1	WP_015175623.1	<i>Oscillatoria nigro-viridis</i>	74.80
W2_770591	WP_015132086.1	<i>Calothrix sp. PCC 7507</i>	84.56
Wd2	WP_096357565.1	<i>Mucilaginibacter gotjawali</i>	78.72
W2_921462	WP_025433386.1	<i>Rhodococcus</i>	81.02

**Table S3.** The list of nucleotide and amino acid sequences of four representative Arr-wd proteins involved in rifamycin-resistance

	nucleotide and amino acid sequence
arr-wd1	AGCGATACGATGCCGCCGAGGGCGCCCGCGCTTGCCGCCGTTCTACACGGCACAAAGGCCGACCTG AAGCCGGGAGACCTCATCGCGCCCGCTTCCGCTCCAACCTCGGCAGCGGACGCGCGGCCGCTACGTC TACCTGACCGCTACGCTCGACGCGGCCGCTGCGGGCGCCGAACCTAGCCCTGGGGACGCGCCGAGCG CATCTACGTCGTGGAGCCGACCGGGCCTATCGAGGACGACCCTAACCTGACCAACGCCAAGTATCCCGGC AACCCGACGCGAGTCGTACCGCACGCGGACCCGCTCCGGGTCTCCGGCGAGGTGACTTCGTGGCAGGG GCACACGCCCGAGCAGCTTCGGGAGATGCGGGAGATGGTCGCGCGGATGCGGCGGCGCGCATCGAG GCCATCGAGGAC
Arr-wd1	SDTMPPEGAPRLPPFYHGTKADLKPGLIAPGFRSNFGSGRAAAVYVLTATLDAAVWGAELALGDGRERIYV VEPTGPIEDDPNLNNAKYPGNPTQSYRTRDPLRVSGEVTSWQGHTEQLREMREVMRRRGIEAIEDLE HHHHHH (149 aa, 16.6 kDa, $\epsilon_{280} = 19940 \text{ cm}^{-1}\text{M}^{-1}$ )
arr-wd2	CAAATAACAAAAGTGAACAATAAATGAACCAAGTCAAAGCCCTTTGTTCAACTTTCTCCACG GCACAAAAGGCTGATCTAAAGATTGGAGACTTTATTGAAGTTGGTCTTAACTCAAACACGGTCAAAGAAA AAATGCGAAATATCTACCTGACAGCAACTTTGGATGCAGCTATTGGGGCGCTGAGCTTGCTCTGGGG GAAGGACGCGAAAGAATCTATTTAGTAGAACCGACAGGGCAATTGAAGACGCCCAATTTAACTGATA AAAAATCCCGGTAAATCAAACAAAATCTTATCGTTCAAACATCCGTTTAAAGTTGTCGCGGAGATTACT ATTTGGCAAGGACACTCACCCGAACAAGTCAAAGCAATGAATGAGGGATTAGCAAACTTAAAGAACAA GGTATTGAGGCAATAGAGGAT
Arr-wd2	QTNKTENKSNEPSQSPFVQTFHGTAKDLKIGDFIEVGLNSNYGQRKNAKYIYLTATLDAAIWGAELALGEGR ERILVEPTGPIEDDPNLTKKFPNPTKSYRSKHPFKVVEITIWQGHSPHEQVKAMNEGLAKLKEQIEAIED LEHHHHHH (155 aa, 17.5 kDa, $18450 \text{ cm}^{-1}\text{M}^{-1}$ )
arr-wd3	GACAAGACCGCGATGATGCCGAACCGCGCAGCCGTTCTACACGGCACGAAGGCCGACCTGAAGCC GGGCGACCTCATCGCGCCGGGCTACGCTCCAACCTACGGCGCGCGAAGAAAGCAGCCTTCGTCTACCT CACGGCCACGCTCGACGCCGCCACCTGGGGCGCCGAGCTGGCCGCGGCGAGGGCCGCGCCGGATC TACTTGGTGGAGCCGACGGGCCCATCGAGGACGACCCCAACCTCACGGACAAGCGGTTCCCGGGCAA CCCCACGAAGTCGTATCGCTCGCGGGACCCGCTGCGGGTACGGGCGAGGTCACGGACTGGCAGGGGC ACCCGACGAGCAGTTGCAGGCCATGCGGGAGCACCTGGAGCGCCTCCGCGCGCAGGGCATCGAGGC GATCGAGGAC
Arr-wd3	DKTADDARTAQPFYHGTKADLKPGLIAPGYASNYGARKKAAFVYLTATLDAATWGAELAAGEGRGRIYV PTGPIEDDPNLTKRFPNPTKSYRSRDLRVTEVTDWQGHPEQLQAMREHLERLRAQIEAIEDLEHH HHHH (147 aa, 16.4 kDa, $19940 \text{ cm}^{-1}\text{M}^{-1}$ )
arr-wd4	GAATTTAGCCGAACAATAATGTTGTCAAACCTGTCTTCAAGGTATGGAGTTGGAAGAGAAAGGAAA GCCTGAAGAAGCGAGCGAAATATTTCTTCAAGCGTGAACGAAGCGGCTGACGATTTGAAAAATTTA CTGCCGTTATTATGTGGCGCGTCATCAAACGATACTGCCGACAAATTAATGGCTGGAACGGTTT TGGAGTTCGGATTAAGATCAATGACGACACGGTTAAGTCGGCTTTTCCCGCTTTGTATCAAAACCTTG CCAAATGCTTTGAGGATTTAGGCGATGCCCGCAACCGGAAAAAGAATTATGAATTAGCAAATTCATTCA AGAAAAACCGTCCGACAAAGTCTTTTTATCACGGAACGCGAGCGGATTTGCAAATCGGCGATTTG CTGACCGCCGAGGCAAGTCTAATTACAAGCCGAACCTCGTAATGAATCACATTTACTTTACGGCATTG GTGAACGGCGCGGGACTTGCCTCGGCTTTGGCTAACGGCGACGGACGCGAGCGGGTTTATATAGTCG AACCGACGGGGAGTTTTGAAAATGATCCGAATGTGACGGACAAAAATTTCCGGGCAATCCGACACGC TCATATCGCAGTCAAGCACCTTTGAAAATCGTCGGCGAAGTAACAGATTGGGTGAGAATAACGCCTGA GGAATCCAAAAATGGCGGGAAAAGTTGGCGAACAGCAAAGGAGAAATTATTAAT
Arr-wd4	EFSPNNNVKLCQMELEEKGPKEEASEIFLQAWNAAADDFEKFAGYVVARHQNDADKLKWLTVLEF GLKINDDTVKSAPFALYQNLAKCFEDLGDARNKKNYELANSFKKPSDKGPFYHGTRADLQIGDLLTAGGKS NYKAEVLMNHIFTALVNGAGLAAALANGDGRERVYVEPTGSFENDPNVTDKFKPNPTRSYRSQAPLKIV GEVTDWVRITPEELQKWREKLANSKGEIINLEHHHHHH (254 aa, 28.6 kDa, $35410 \text{ cm}^{-1}\text{M}^{-1}$ )



**Table S4.** Primer sequences used for the current work

template protein	mutations	sequences
Arr-wd3	N34H (Arr-wd3+N34H)	5'-G GGC TAT GCA AGC <u>CAT</u> TAT GGT GCA CG-3' 5'-C GTG CAC CATA <u>ATG</u> GCT TGC ATA GCC C-3'
Arr-wd3	H125M (Arr-wd3+H125M)	5'-GCA ATG CGC GAA <u>ATG</u> CTG GAA CGC CTG-3' 5'-CAG GCG TTC CAG <u>CAT</u> TTC GCG CAT TGC-3'
Arr-wd3	H125G (Arr-wd3+H125G)	5'-CAA TGC GCG AA <u>GGC</u> CTG GAA CGC CTG-3' 5'-CA GGC GTT CCA <u>GGC</u> CTT CGC GCA TTG-3'
Arr-wd3	H125K (Arr-wd3+H125K)	5'-GCA ATG CGC GAA <u>AAA</u> CTG GAA CGC CTG-3' 5'-CAG GCG TTC CAG <u>TTT</u> TTC GCG CAT TGC-3'
Arr-wd3	H142 to stop codon (Arr-wd3+Δ6His)	5'-GAA GCA ATT GAA GAT <u>TAA</u> GAG CAC CAC CAC-3' 5'-GTG GTG GTG CTC <u>TTA</u> ATC TTC AAT TGC TTC-3'

**Table S5.** ADP-ribosylated product analysis by ESI-MS in negative mode

substrate	Product*	calculated m/z	detected m/z
rifampin	ADP-ribosylrifampin	$[M+H]^- = 1362.5$	1362.8
		$[M+Na]^- = 1384.5$	1385.0
rifapentine	ADP-ribosylrifapentine	$[M+H]^- = 1416.5$	1416.8
rifaximin	ADP-ribosylrifaximin	$[M+H]^- = 1325.4$	1325.8
rifabutin	ADP-ribosylrifabutin	$[M+H]^- = 1386.5$	1386.8
		$[M+Na]^- = 1408.5$	1408.9

\*The ADP-ribosylated product includes a phosphate group, which makes the molecule to exhibit -2 charge by itself.

**Table S6.** Steady-state kinetic parameters of Arr-wd proteins in ADP-ribosylation of rifampin and its analogues

(a) NAD<sup>+</sup>

	Arr-wd1	Arr-wd2	Arr-wd3	Arr-wd4
$k_{\text{cat}}$ (s <sup>-1</sup> )	0.023(3)	0.14(2)	0.027(2)	0.108(9)
$K_M$ (mM)	0.4(2)	2.3(6)	0.11(4)	0.8 (2)
$k_{\text{cat}}/K_M$ (s <sup>-1</sup> M <sup>-1</sup> )	65(9)	61(11)	246(19)	139(12)

(b) rifampin

	Arr-wd1	Arr-wd2	Arr-wd3	Arr-wd4
$k_{\text{cat}}$ (s <sup>-1</sup> )	0.010(2)	0.056(3)	0.0206(2)	0.060(3)
$K_M$ (mM)	0.23(19)	0.48(7)	0.18(6)	0.17(8)
$k_{\text{cat}}/K_M$ (s <sup>-1</sup> M <sup>-1</sup> )	45(8)	118(6)	113(8)	357(37)

(c) rifapentine

	Arr-wd1	Arr-wd2	Arr-wd3	Arr-wd4
$k_{\text{cat}}$ (s <sup>-1</sup> )	0.014(1)	0.031(1)	0.010(1)	0.045(3)
$K_M$ (mM)	0.11(5)	0.13(3)	0.07(5)	0.11(4)
$k_{\text{cat}}/K_M$ (s <sup>-1</sup> M <sup>-1</sup> )	128(12)	245(10)	147(17)	419(30)

(d) rifaximin

	Arr-wd1	Arr-wd2	Arr-wd3	Arr-wd4
$k_{\text{cat}}$ (s <sup>-1</sup> )	0.0047(4)	0.0165(2)	0.0028(4)	0.086(7)
$K_M$ (mM)	0.11(6)	0.03(0)	0.13(10)	0.28(8)
$k_{\text{cat}}/K_M$ (s <sup>-1</sup> M <sup>-1</sup> )	41(4)	533(8)	22(3)	304(23)

(e) rifabutin

	Arr-wd1	Arr-wd2	Arr-wd3	Arr-wd4
$k_{\text{cat}}$ (s <sup>-1</sup> )	0.010(1)	0.033(1)	0.013(2)	0.063(6)
$K_M$ (mM)	0.17(10)	0.24(3)	0.22(10)	0.27(9)
$k_{\text{cat}}/K_M$ (s <sup>-1</sup> M <sup>-1</sup> )	60(8)	142(6)	58(7)	230(21)

**Table S7.** Substrate dissociation constants ( $K_D$ ) measured from the intrinsic fluorescence quenching(a) NAD<sup>+</sup> and NADP<sup>+</sup>

$K_D$ (mM)	NAD <sup>+</sup>	NADP <sup>+</sup>
Arr-wd1	1.23(4)	2.3(1)
Arr-wd2	2.14(5)	2.33(7)
Arr-wd3	1.67(6)	1.89(8)
Arr-wd4	1.75(7)	1.90(6)
Arr-wd3+Δ6His	1.53(4)	2.03(8)
Arr-wd3+N34H	2.08(3)	2.07(5)
Arr-wd3+H125M	1.39(1)	2.5(1)
Arr-wd3+H125G	1.48(5)	2.1 (1)
Arr-wd3+H125K	1.23(4)	2.54(1)

(b) rifampin analogues

$K_D$ (μM)	rifampin	rifapentine	rifaximin	rifabutin
Arr-wd1	12.8(4)	22.5(8)	8.6(2)	7.8(1)
Arr-wd2	3.5(1)	5.4(2)	5.1(2)	5.1(2)
Arr-wd3	25.4(4)	30.6(4)	36.2(9)	34.0(14)
Arr-wd4	1.2(1)	1.7(1)	1.4(1)	1.9(1)
Arr-wd3+Δ6His	12.7(1)	19.4(4)	20.5(6)	20.1(4)
Arr-wd3+N34H	7.1(1)	10.1(1)	5.1(1)	7.2(1)
Arr-wd3+H125M	8.9(3)	13.1(1)	7.0(2)	6.9(2)
Arr-wd3+H125G	5.9(2)	9.5(1)	6.3(2)	8.0(2)
Arr-wd3+H125K	12.8(5)	20.6(6)	11.1(1)	18.1(5)

**Table S8.** Steady-state kinetic parameters of Arr-wd3 variants in ADP-ribosylation of rifampin analogues

(a) NAD<sup>+</sup>

variants	$k_{\text{cat}}$ (s <sup>-1</sup> )	$K_{\text{M}}$ (mM)	$k_{\text{cat}}/K_{\text{M}}$ (s <sup>-1</sup> M <sup>-1</sup> )
Arr-wd3	0.027(2)	0.11(4)	246(19)
+ N34H	0.012(1)	0.3(1)	39(4)
+ H125M	0.016(2)	0.20(12)	81(12)
+ H125G	0.018(3)	0.84(30)	21(3)
+ H125K	0.024(2)	0.41(10)	57(5)

(b) rifampin

variants	$k_{\text{cat}}$ (s <sup>-1</sup> )	$K_{\text{M}}$ (mM)	$k_{\text{cat}}/K_{\text{M}}$ (s <sup>-1</sup> M <sup>-1</sup> )
Arr-wd3	0.0206(2)	0.18(6)	113(8)
+ N34H	0.0077(2)	0.011(5)	702(15)
+ H125M	0.026(1)	0.06(2)	417(21)
+ H125G	0.018(1)	0.09(4)	207(16)
+ H125K	0.0143(5)	0.06(1)	227(7)

(c) rifapentine

variants	$k_{\text{cat}}$ (s <sup>-1</sup> )	$K_{\text{M}}$ (mM)	$k_{\text{cat}}/K_{\text{M}}$ (s <sup>-1</sup> M <sup>-1</sup> )
Arr-wd3	0.010(1)	0.07(5)	147(17)
+ H125M	0.0214(10)	0.05(2)	409(19)
+ H125G	0.018(1)	0.08(3)	229(14)
+ H125K	0.0180(6)	0.06(1)	305(9)

(d) rifaximin

variants	$k_{\text{cat}}$ (s <sup>-1</sup> )	$K_{\text{M}}$ (mM)	$k_{\text{cat}}/K_{\text{M}}$ (s <sup>-1</sup> M <sup>-1</sup> )
Arr-wd3	0.0028(4)	0.13(10)	22(3)
+ H125M	0.0238(9)	0.09(2)	277(11)
+ H125G	0.0240(9)	0.07(2)	354(13)
+ H125K	0.0205(6)	0.10(1)	204(6)

(e) rifabutin

variants	$k_{\text{cat}}$ (s <sup>-1</sup> )	$K_{\text{M}}$ (mM)	$k_{\text{cat}}/K_{\text{M}}$ (s <sup>-1</sup> M <sup>-1</sup> )
Arr-wd3	0.013(2)	0.22(10)	58(7)
+ H125M	0.0138(6)	0.05(2)	288(13)
+ H125G	0.0226(7)	0.09(2)	249(8)
+ H125K	0.0154(2)	0.06(1)	241(3)

**Table S9.** Determination of minimum inhibitory concentration (MIC) of rifampin and its analogues

(a) MIC values from the literatures (1).

plasmids	IPTG	rifampin	rifaximin	rifabutin
pET28a(+)	-	4	8	8
	+	8	16	8
pET28a(+)-arr-2	-	512	>512	>512
	+	>512	>512	>512
pET22b(+)	-	8	16	16
	+	8	16	16
pET22b(+)-arr- <i>ms</i>	-	128	>512	64
	+	>512	>512	>512
pET15b(+)	-	8	16	16
	+	8	16	16
pET15b(+)-arr- <i>sc</i>	-	128	>512	64
	+	512	512	512

(b) Measured MIC values in this study

plasmids	IPTG	rifampin	rifapentine	rifaximin	rifabutin
pET22b(+)	-	4	4	4	4
	+	2	4	4	4
pET22b(+)-arr-wd1	-	4	4	4	4
	+	32	64	512	64
pET22b(+)-arr-wd2	-	32	64	256	32
	+	512	>512	>512	256
pET22b(+)-arr-wd3	-	8	8	16	16
	+	64	128	512	128
pET22b(+)-arr-wd4	-	8	16	32	8
	+	128	256	>512	256
pET22b(+)-arr-wd3+N43H	-	8	8	16	8
	+	32	64	>512	128
pET22b(+)-arr-wd3+H125M	-	8	8	16	8
	+	32	64	512	128
pET22b(+)-arr-wd3+H125G	-	16	8	64	16
	+	64	128	>512	128
pET22b(+)-arr-wd3+H125K	-	8	16	16	8
	+	16	32	512	64

**Figure S1.** Structural analysis of Arr-*ms* protein complexed with rifampin (2HW2.pdb). (a) Annotated secondary structure information and rifampin binding loop (b) 2D diagram of rifampin-Arr-*ms* interaction (color scheme: pink and purple: hydrophobic interaction, green: van der Waals contact, lightblue: hydrogen bond, orange: cation-pi interaction) (c) Water mediated hydrogen bonds between rifampin and Arr-*ms*

**Figure S2.** Autodock simulation of NAD<sup>+</sup>-binding site in Arr-*ms*

**Figure S3.** Protein purification and characterization of Arr-wd proteins. (a) A representative FPLC trace of Arr-wd1 protein (b) 15% SDS-PAGE of Arr-wd proteins and the single variants (c) A representative MADLI-TOF spectra of Arr-wd1 protein

**Figure S4.** A representative HPLC analysis of steady-state activity assay by the (a) previously reported HPLC method and by the (b) modified quenching method (c) The standard curve of nicotinamide (d-e) Michaelis-Menten kinetic analysis under various concentrations of (d) NAD<sup>+</sup> or (e) rifampin

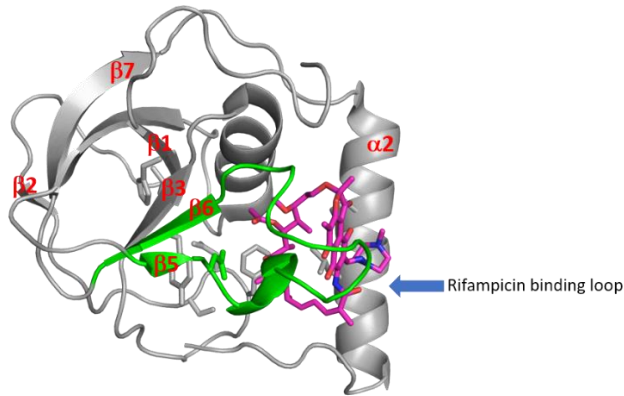
**Figure S5.** Intrinsic fluorescence changes upon the addition of either NAD<sup>+</sup> or rifampin. (a) Relative location of the tryptophan residues in Arr-*ms* (PDB 2HW2) (b) UV-Vis spectra of rifampin and NAD<sup>+</sup>. Rifampin exhibits strong absorption at 295 and 325 nm. (c) Representative intrinsic fluorescence quenching upon the addition of rifampin. The fluorescence intensity was corrected for the secondary filter effect with rifampin as shown in (b). (d) Plots of  $F_0/F$  vs the concentration of rifampin. (e) Dissociation constant ( $K_D$ ) determined by the fluorescence changes **Figure S6.** Rifampin analogues used for the current study

**Figure S7.** Representative HPLC traces of the reactions with rifampin analogues. (a) rifapentine (b) rifaximin (c) rifabutin

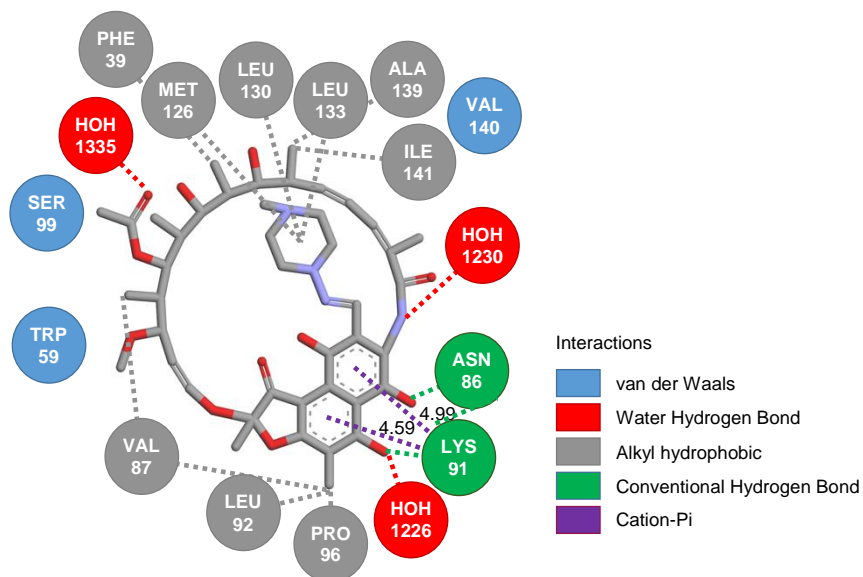
**Figure S8.** MIC values of Arr-wd proteins. (a) Arr-wd 1-4 proteins (b) Arr-wd3 variants

**Figure S1.** Structural analysis of Arr-*ms* protein complexed with rifampin (2HW2.pdb). (a) Annotated secondary structure information and rifampin binding loop (b) 2D diagram of rifampin-Arr-*ms* interaction (color scheme: pink and purple: hydrophobic interaction, green: van der Waals contact, lightblue: hydrogen bond, orange: cation-pi interaction) (c) Water mediated hydrogen bonds between rifampin and Arr-*ms*.

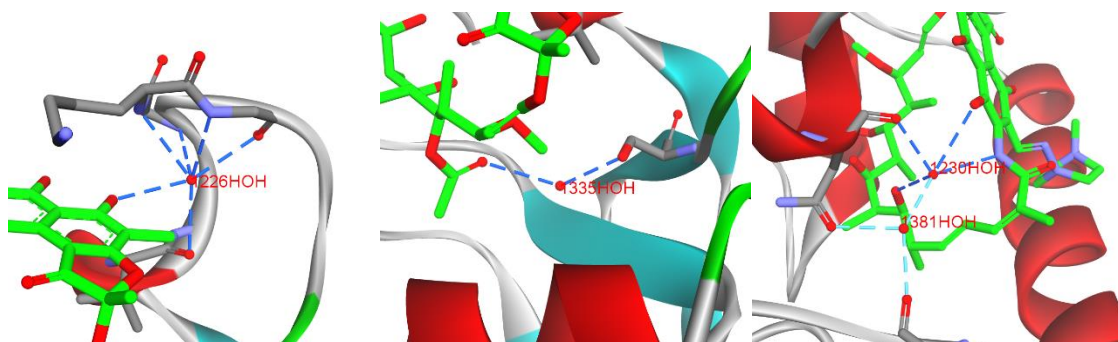
(a)



(b)

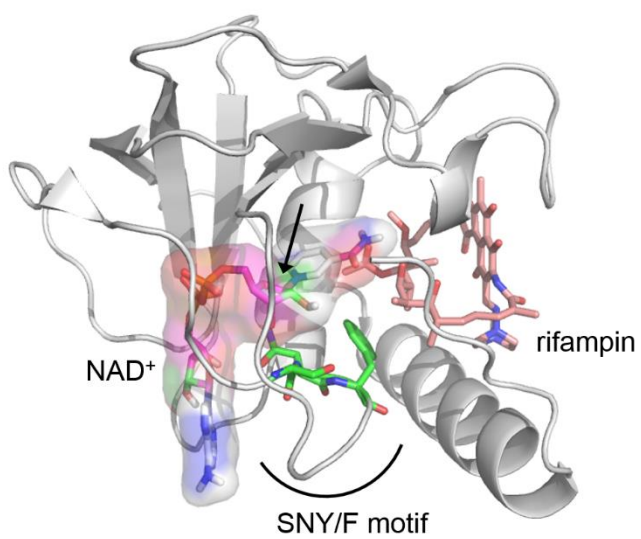


(c)



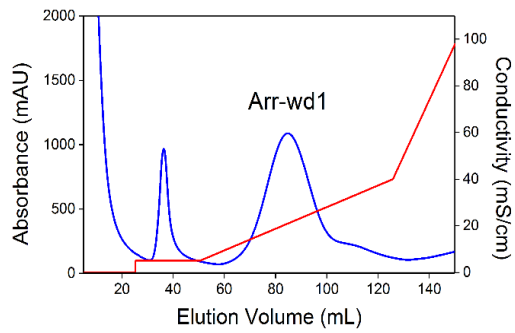


**Figure S2.** Autodock simulation of NAD<sup>+</sup>-binding site in Arr-*ms*. The black arrows indicate the scissile C-N bond in NAD<sup>+</sup> that generates an oxocarbenium ion in accordance with the release of nicotinamide.

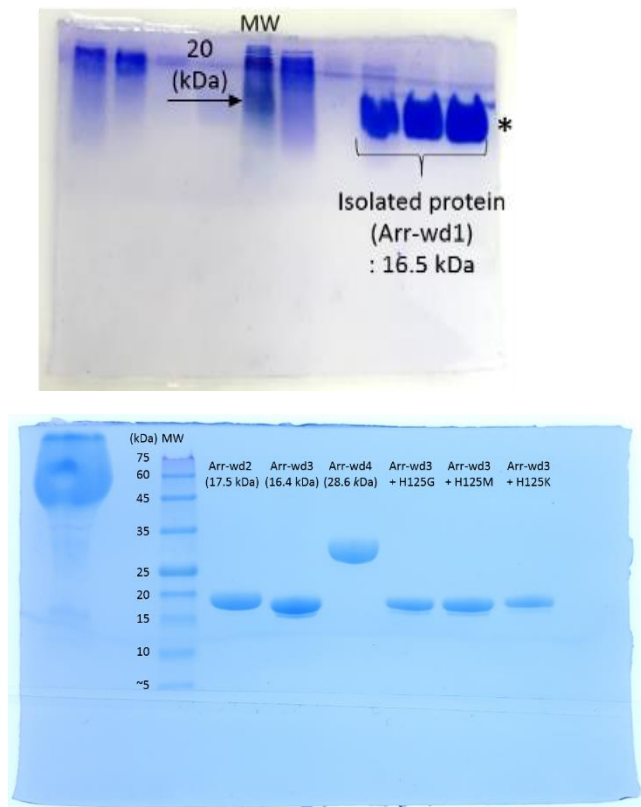


**Figure S3.** Protein purification and characterization of Arr-wd proteins. (a) A representative FPLC trace of Arr-wd1 protein (b) 15% SDS-PAGE of Arr-wd proteins and the single variants (c) A representative MADLI-TOF spectra of Arr-wd1 protein.

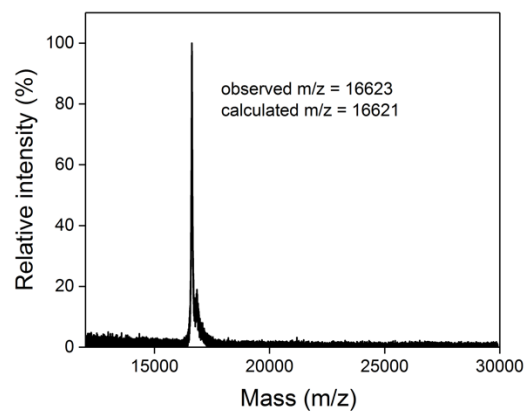
(a)



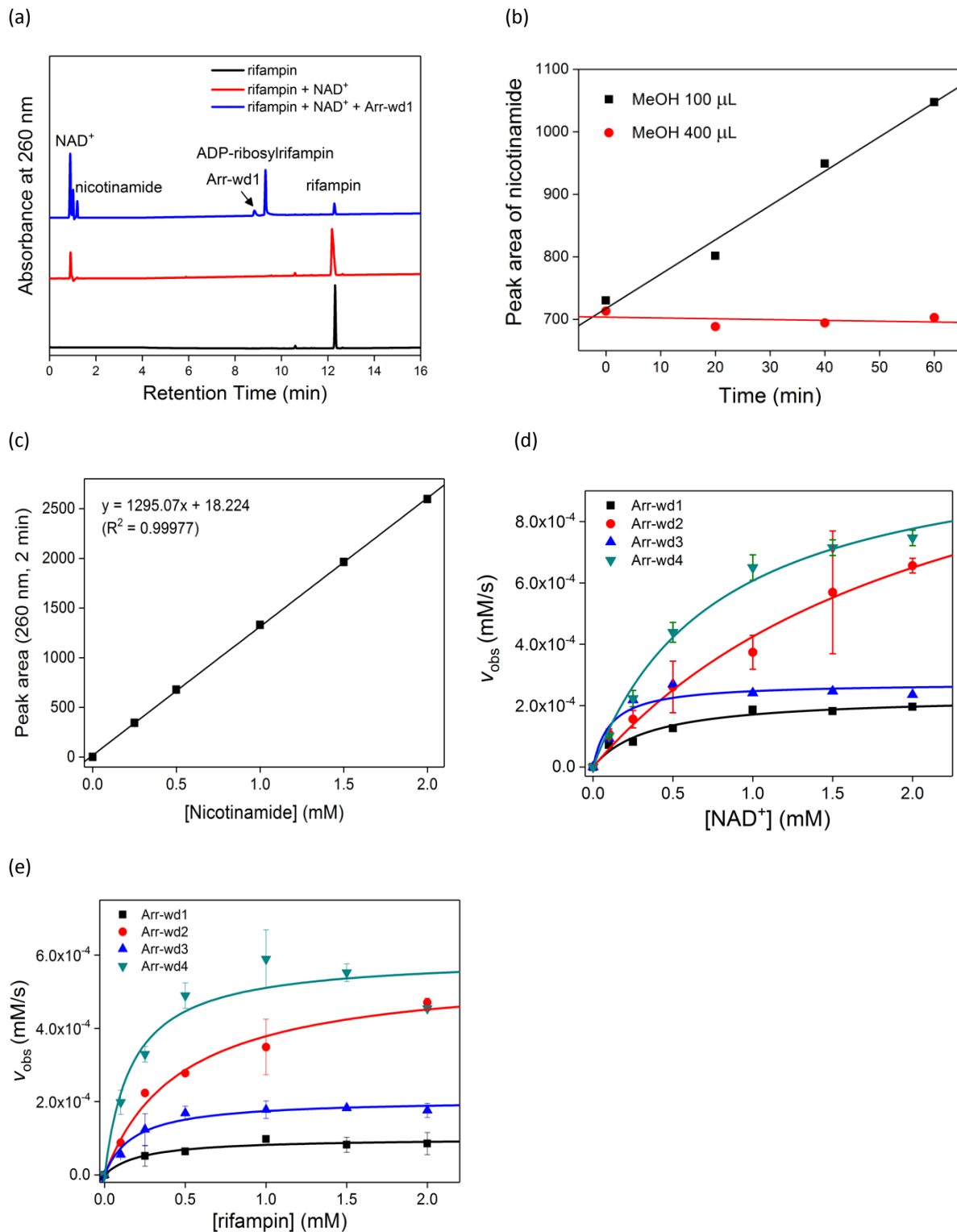
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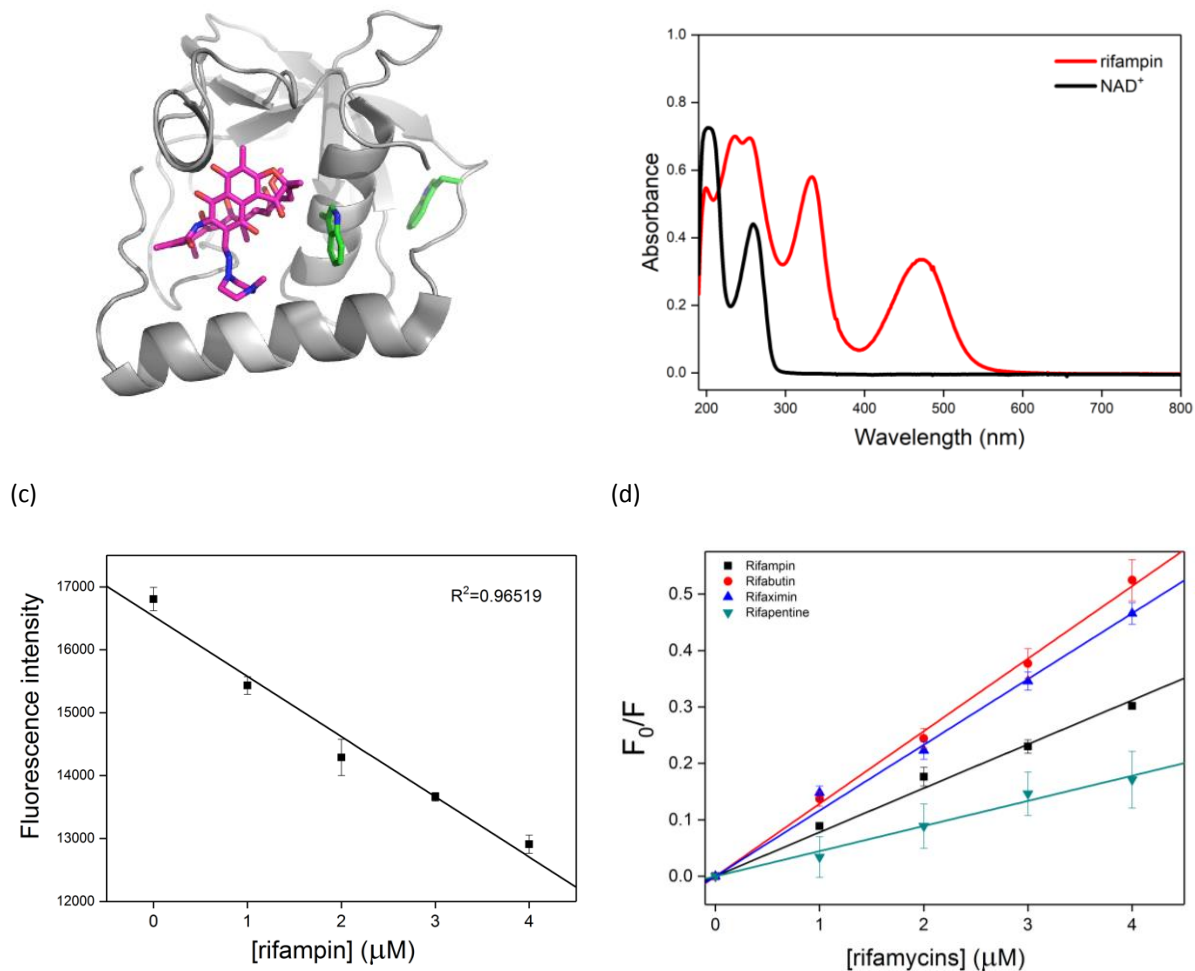
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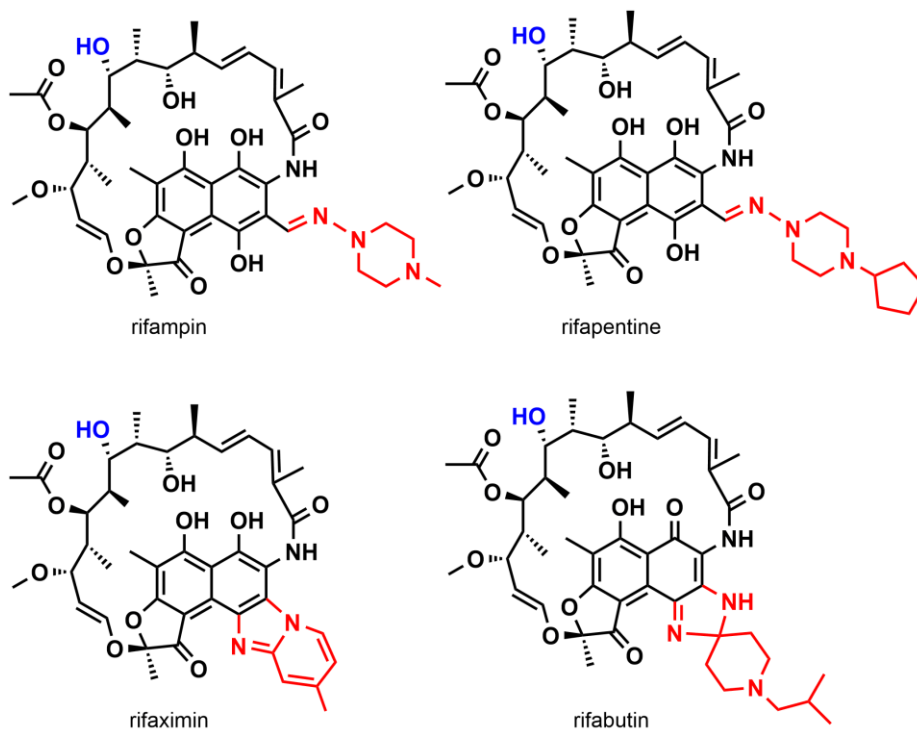
**Figure S4.** A representative HPLC analysis of steady-state activity assay. (a) Previously reported HPLC method (b) Modified quenching method by increasing the volume of methanol from 100  $\mu\text{L}$  to 400  $\mu\text{L}$ . (c) The standard curve of nicotinamide by the modified HPLC method (d-e) Michaelis-Menten kinetic analysis under various concentrations of (d)  $\text{NAD}^+$  or (e) rifampin.



**Figure S5.** Intrinsic fluorescence changes upon the addition of either NAD<sup>+</sup> or rifampin. (a) Relative location of the tryptophan residues in Arr-ms (PDB 2HW2) (b) UV-Vis spectra of rifampin and NAD<sup>+</sup>. Rifampin exhibits strong absorption at 295 and 325 nm. (c) Representative intrinsic fluorescence quenching upon the addition of rifampin. The fluorescence intensity was corrected for the secondary filter effect with rifampin as shown in (b). (d) Plots of  $F_0/F$  vs the concentration of rifampin. (e) Dissociation constant ( $K_D$ ) determined by the fluorescence changes (a)

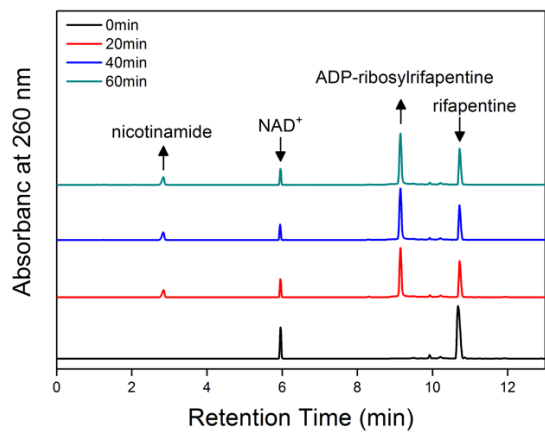


**Figure S6.** Rifampin analogues used for the current study. Altered substructures of the rifampin derivatives are colored in red, and the hydroxyl group that reacts with an oxocarbenium ion is colored in blue.

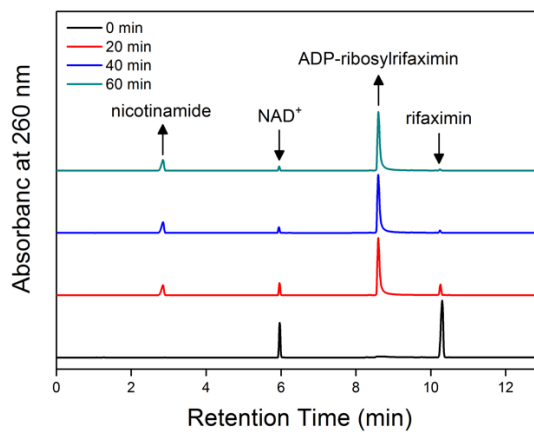


**Figure S7.** Representative HPLC traces of the reactions with rifampin analogues. (a) rifapentine (b) rifaximin (c) rifabutin.

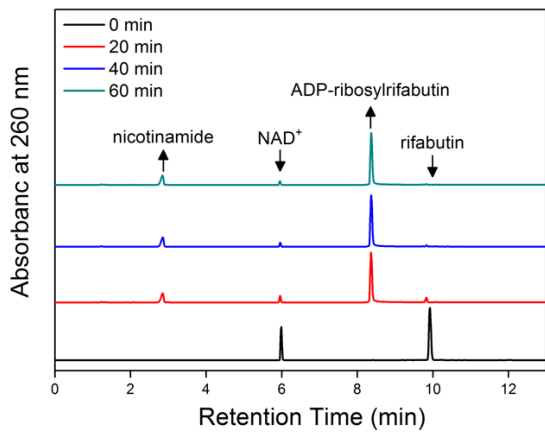
(a)



(b)

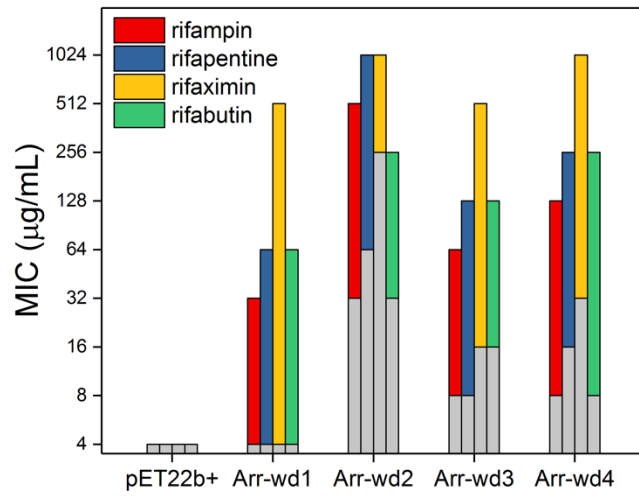


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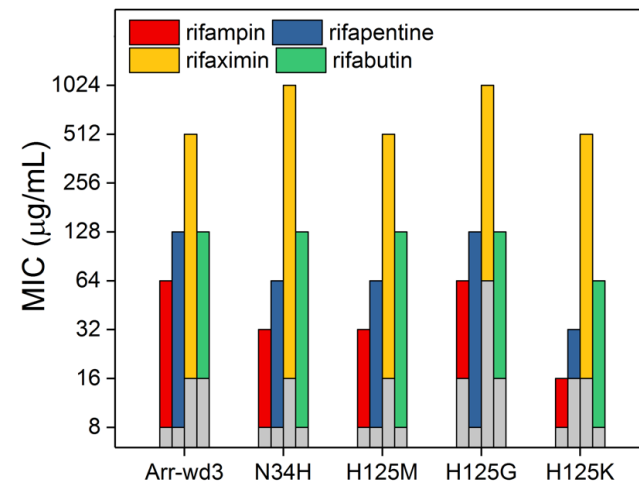


**Figure S8.** MIC values of Arr-wd proteins. (a) Arr-wd 1-4 proteins (b) Arr-wd3 variants.

(a)



(b)





## References

1. Baysarowich, J., Koteva, K., Hughes, D.W., Ejim, L., Griffiths, E., Zhang, K., Junop, M. and Wright, G.D. (2008) Rifamycin antibiotic resistance by ADP-ribosylation: Structure and diversity of Arr. *Proc. Natl. Acad. Sci. U.S.A.*, **105**, 4886-4891.