

## Expanded View Figures

### Figure EV1. Prolonged NF- $\kappa$ B activation in astrocytes induces microglia differentiation.

- A, B Co-immunostaining of CD11b, TMEM119, DAPI, and quantification of CD11b<sup>+</sup>/TMEM119<sup>+</sup> and CD11b<sup>+</sup>/TMEM119<sup>-</sup> cells in SOD1/IKK mice; more than 90% of CD11b<sup>+</sup> cells are TMEM119<sup>+</sup>, confirming the microglial identity.
- C, D Immunostaining for microglia (IBA1), macrophages (F4/80), DAPI, and quantification of F4/80<sup>+</sup>/IBA1<sup>+</sup> cells. F4/80<sup>+</sup> cells represent only a very small percentage of IBA1<sup>+</sup> cells.
- E, F Co-immunostaining of TMEM119, CD45, DAPI, and quantification of TMEM119<sup>+</sup>/CD45<sup>+</sup>; CD45 identifies a subset of TMEM119<sup>+</sup> microglia.
- G, H Immunostaining for MAC-2, CD45, DAPI, and the relative quantification of CD45<sup>+</sup>/MAC-2<sup>+</sup> and CD45<sup>+</sup>/MAC-2<sup>-</sup> fraction; only a subset of CD45<sup>+</sup> cells are also MAC-2<sup>+</sup>.
- I, J Immunostaining of CD11b, MAC-2, DAPI, and quantification of CD11b<sup>+</sup>/MAC-2<sup>+</sup> fraction, confirming that MAC-2<sup>+</sup> cells represent a subset of microglia.
- K Schematic summary of cell identity according to immunohistochemistry in Figs 2 and EV1.
- L, M Co-immunostaining of CD11b, CD11c, DAPI, and quantification of CD11b<sup>+</sup>/CD11c<sup>+</sup> (DAM) fraction; CD11c<sup>+</sup> cells represent about 25% of myeloid cells.
- N, O Immunostaining of TMEM119, CD11c, DAPI, and quantification of CD11c<sup>+</sup>/TMEM119<sup>+</sup>, showing that nearly all CD11c<sup>+</sup> are microglia and not infiltrating cells.
- P–R Immunostaining of TMEM119, CD11c, CD169, and quantification of TMEM119<sup>+</sup>/CD11c<sup>+</sup> fraction expressing the monocyte marker CD169; almost no CD11c<sup>+</sup> cell is CD169<sup>+</sup>, confirming that CD11c<sup>+</sup> are not infiltrating monocytes; nevertheless, a distinct population of TMEM119<sup>+</sup>/CD169<sup>+</sup> early-activated microglia can be identified (R).
- S Time course of CD11c<sup>+</sup> cell fraction in the overall CD11b<sup>+</sup> population; the DAM population does not expand in the progression phase.

Data information: Representative images are shown only for SOD1/IKK. Quantification is shown for both SOD1/IKK ( $n = 3$ ) and IKK ( $n = 3$ ) at P50. In (A, C, E, G, I, L and N), the bottom panels represent the high-magnified views of the upper panels. Data are presented as means  $\pm$  SD. Scale bars, 10  $\mu$ m.

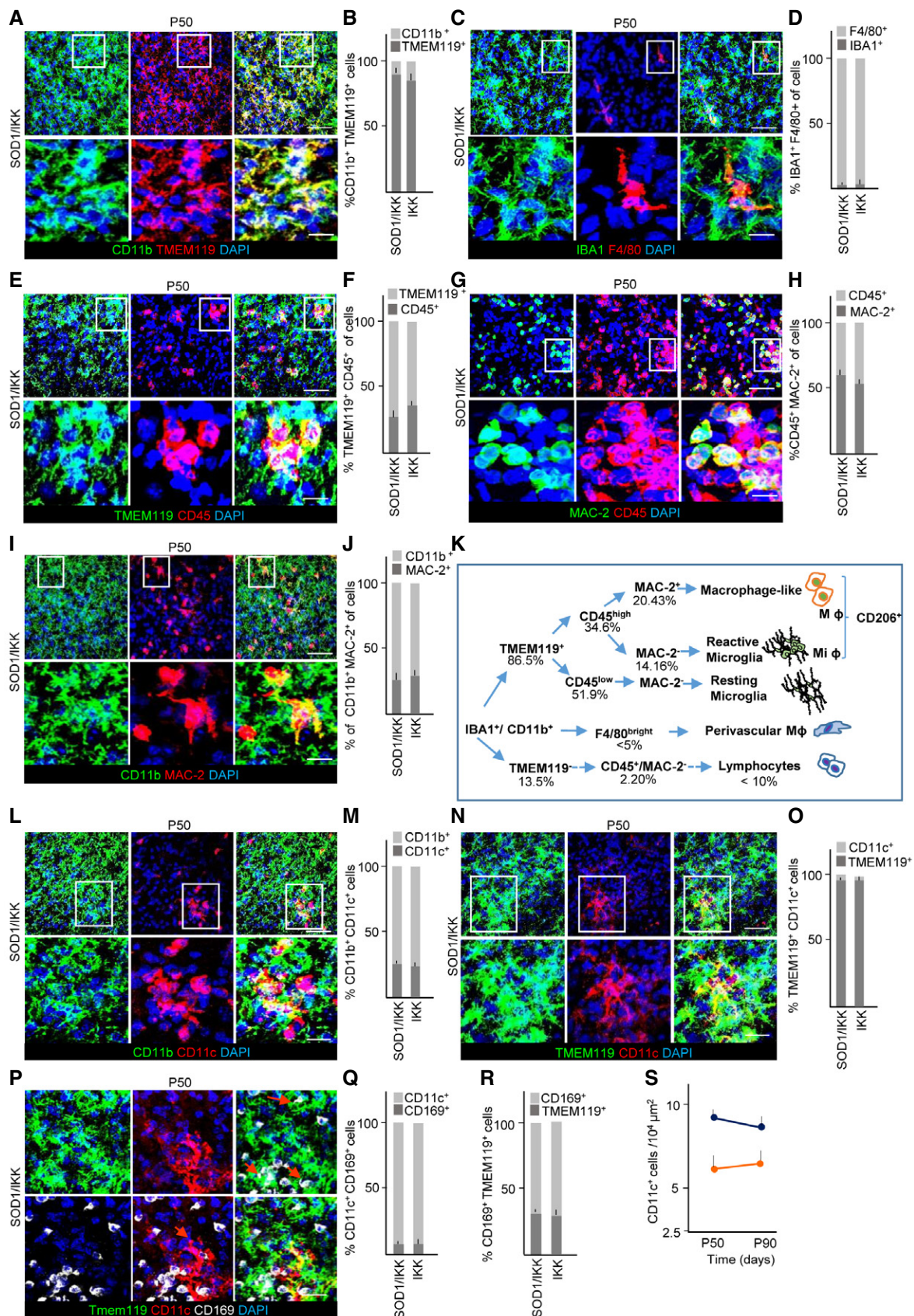
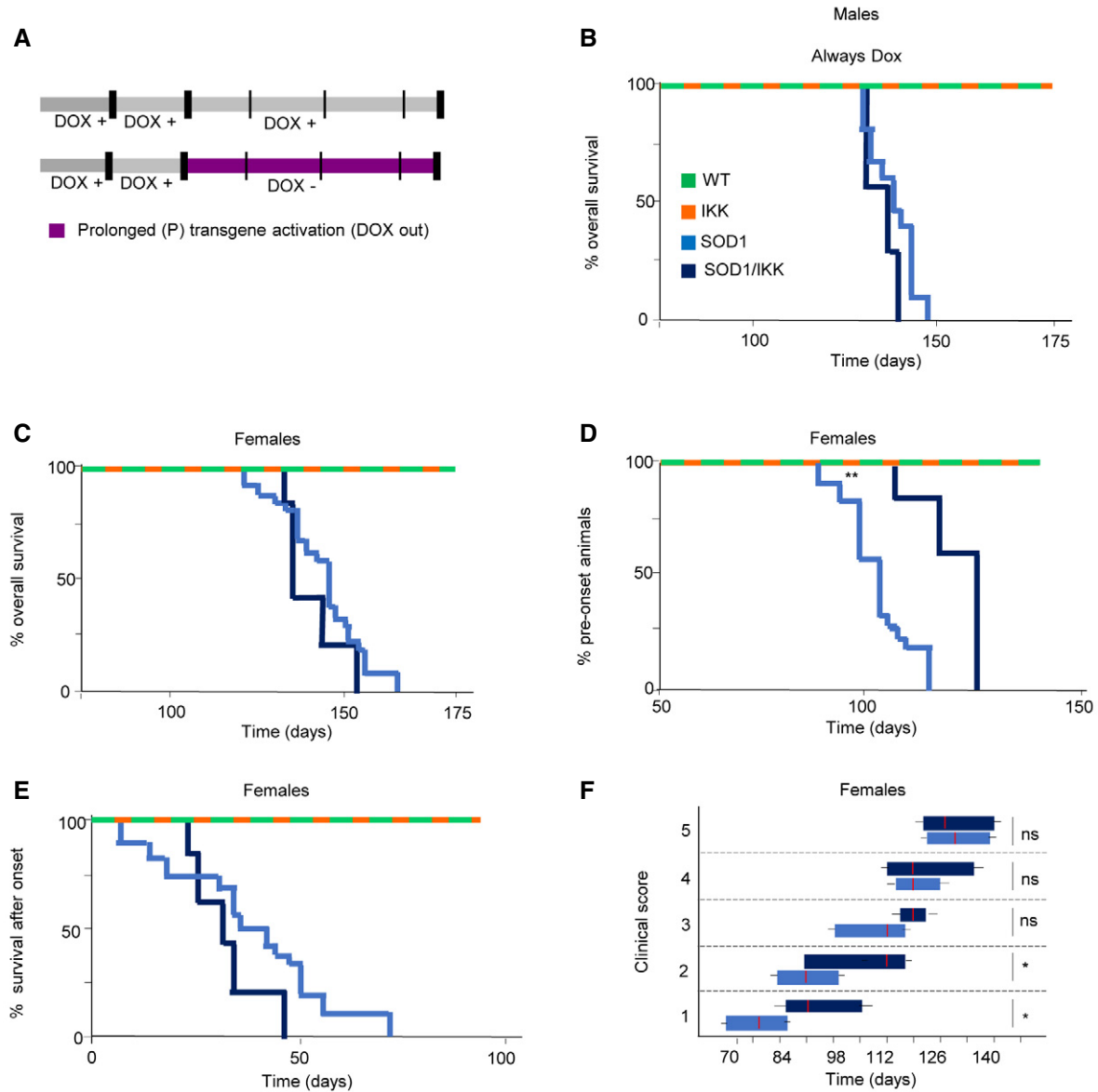


Figure EV1.



**Figure EV2. Time-dependent NF- $\kappa$ B activation in astrocytes differentially affects disease onset and progression.**

A Experimental design illustrating no transgene activation (male mice in panel B) or prolonged transgene activation (female mice in panels C–F) induced by DOX withdrawal (DOX out, purple).

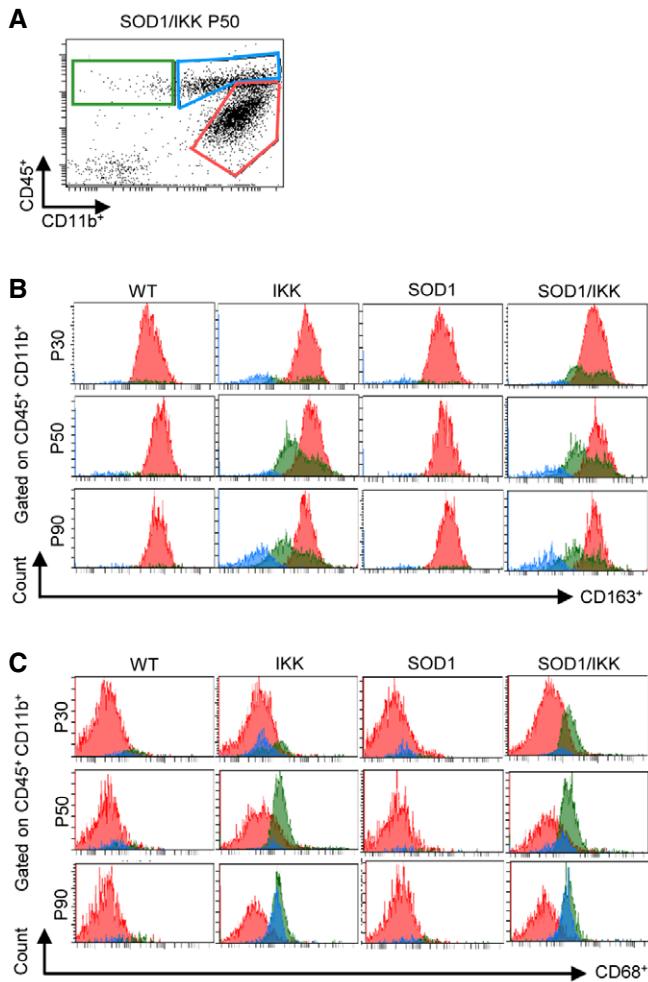
B Kaplan–Meier curves of overall survival (% of alive mice over time) of male mice treated with DOX at all time, resulting in inhibition of IKK2-CA transgene expression. No difference between the SOD1 and the SOD1/IKK mice was observed (Mantel–Cox  $P = 0.0612$ ; SOD1  $n = 25$ ; SOD1/IKK  $n = 3$ ).

C Overall survival depicted in a Kaplan–Meier curve of survival for female mice with prolonged activation. (Mantel–Cox  $P = 0.5397$ ; SOD1  $n = 18$ ; SOD1/IKK  $n = 5$ ).

D Kaplan–Meier curves representing percentage of female animals in the presymptomatic phase of ALS disease (based on peak weight); (Mantel–Cox  $**P = 0.0048$ ).

E Kaplan–Meier curves representing duration of the progression phase in female mice (Mantel–Cox  $P = 0.2266$ ; SOD1  $n = 18$ ; SOD1/IKK  $n = 5$ ).

F Time course of the neurological score progression, showing delayed symptoms onset in female SOD1/IKK mice. Data are shown as box and whiskers plot: The horizontal red lines in the middle represent median, box limits represent 10% to 90%, while left and right whisker represent min. and max. value range, respectively. Values were calculated by nonparametric Mann–Whitney test.  $*P \leq 0.05$ ; ns = not significant  $P > 0.05$ . (Mann–Whitney score 1  $*P = 0.0333$ ; score 2  $*P = 0.0433$ ).

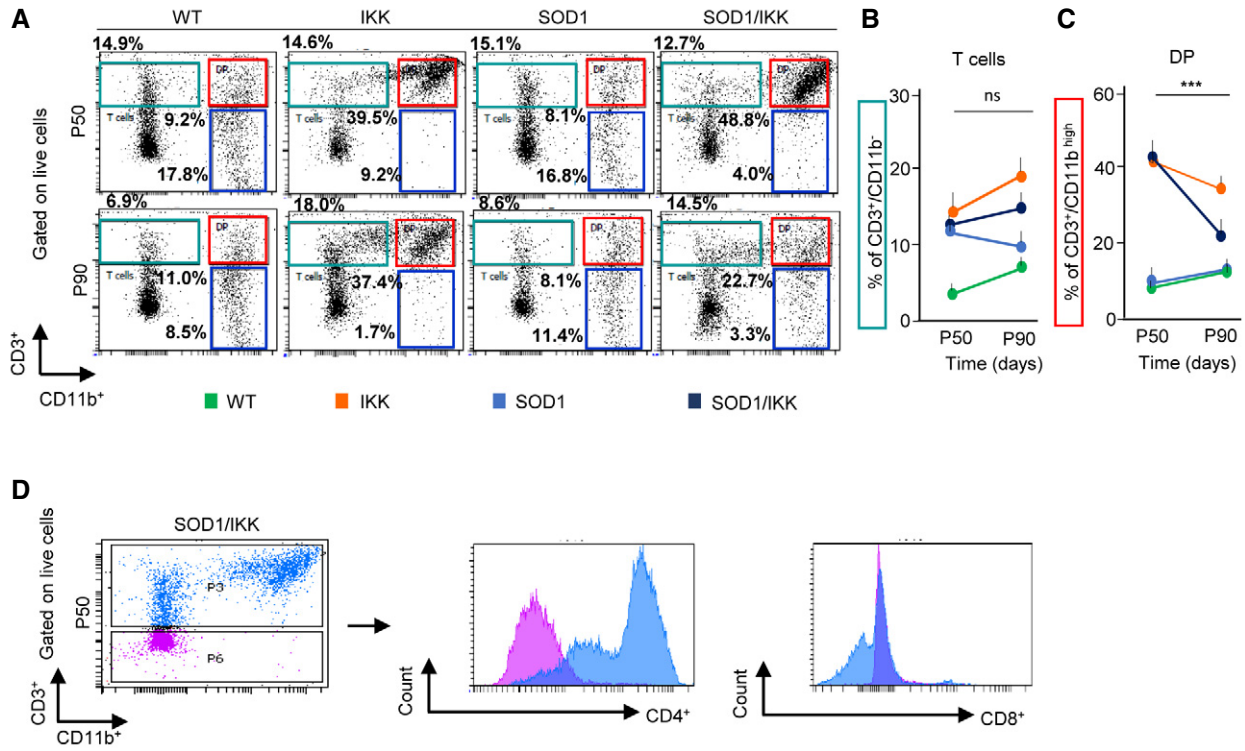


**Figure EV3. Microglia immunophenotype characterization after different time points of IKK/NF- $\kappa$ B activation in astrocytes.**

A Representative FACS dot-plot of CD11b<sup>+</sup> and CD45<sup>+</sup> immune cells at P50 on which histograms in (B and C) is gated.

B Histograms of CD163 expression gated on CD11b<sup>+</sup>/CD45<sup>low-intermediate</sup> (red), CD11b<sup>+</sup>/CD45<sup>+</sup> (blue), and CD11b<sup>-</sup>/CD45<sup>+</sup> (green) cells at P30, P50, and P90. Distinctive CD163 expression is seen only in CD11b<sup>+</sup>/CD45<sup>low-intermediate</sup> population and decreases from P30 and P50 to P90.

C Histograms of CD68 expression in CD11b<sup>+</sup>/CD45<sup>low-intermediate</sup> (red), CD11b<sup>+</sup>/CD45<sup>high</sup> (blue), and CD11b<sup>-</sup>/CD45<sup>+</sup> (green) subpopulations. Whereas CD11b<sup>+</sup>/CD45<sup>low-intermediate</sup> express constant low levels of CD68, a strong increase in CD68 expression is detected in CD11b<sup>+</sup>/CD45<sup>high</sup> from P30, P50 to P90.



**Figure EV4. Characterization of lymphocyte dynamics in lumbar spinal cord in response to NF-κB activation in astrocytes.**

A Representative FACS dot-plot showing overall CD3<sup>+</sup>/CD11b<sup>+</sup> population at P50 and P90 in WT, IKK, SOD1, and SOD1/IKK mice, gated on live cells.  
 B, C Quantification of (B) CD3<sup>+</sup>CD11b<sup>-</sup> cells (expressed as %; green in panel A) and (C) CD3<sup>+</sup>/CD11b<sup>high</sup> double-positive populations (DP) (expressed as %; red in panel A) at P50 and P90. P50: WT *n* = 4, IKK *n* = 6, SOD1 *n* = 4, SOD1/IKK *n* = 4; P90: WT *n* = 3, IKK *n* = 6, SOD1 *n* = 4, SOD1/IKK *n* = 5. Two-way ANOVA. Data are shown as means ± SD, \*\*\**P* ≤ 0.001; ns = not significant, *P* > 0.05.  
 D FACS dot-plot of CD3<sup>+</sup>/CD11b<sup>+</sup> population at P50 in SOD1/IKK animal gated on live cells and histograms of CD4<sup>+</sup> and CD8<sup>+</sup> expression gated on CD3<sup>high</sup> (blue) and CD3<sup>low</sup> (purple).

**Figure EV5. Wnt signaling is involved in astrocyte-driven microglia expansion after IKK/NF-κB activation in astrocytes.**

A Expression of *Wnt1* on mRNA level in spinal cord of P50 mice (*n* = 3–4; relative to HPRT).  
 B Co-immunostaining of WNT7a (green), GFAP (white), and DAPI (blue) in P50 and P90 animals of all four genotypes.  
 C Immunostaining for Wnt5a and GFAP in spinal cord from SOD1/IKK mice treated with vehicle or with the PORCN inhibitor C59; Wnt5a immunoreactivity is reduced to < 10% in C59-treated compared to vehicle-treated SOD1/IKK mice (at P38).  
 D High-magnified representative pictures of IBA1<sup>+</sup> staining in IKK and SOD1/IKK animals with either C-59 or vehicle (Veh) treatment from P26 to P38 (early administration).  
 E Quantification of IBA1<sup>+</sup> area in WT, IKK, SOD1, and SOD1/IKK animals with C-59 or Veh treatment from P26 to P38 (% of total area).  
 F High-magnified representative pictures of IBA1<sup>+</sup> staining of IKK and SOD1/IKK, either C-59- or Veh-treated, in late administration between P90 and P102.  
 G Quantification of IBA1<sup>+</sup> cells of C-59- or Veh-treated mice from P90 to P102.  
 H Quantification of CD45<sup>+</sup> cells of IKK and SOD1/IKK animals treated with Veh or C-59 from P90 to P102.  
 Data information: (E) C59 early treatment: WT *n* = 4, IKK *n* = 3, SOD1 *n* = 2, SOD1/IKK *n* = 4; vehicle-treated: WT *n* = 2, IKK *n* = 2, SOD1 *n* = 2, SOD1/IKK *n* = 7. (G–H) C59 late treatment: WT *n* = 2, IKK *n* = 3, SOD1 *n* = 2, SOD1/IKK *n* = 3; vehicle-treated: WT *n* = 3, IKK *n* = 2, SOD1 *n* = 2, SOD1/IKK *n* = 2. Two-way ANOVA. Data are shown as means ± SD, \*\**P* ≤ 0.01; \*\*\**P* ≤ 0.0001. Scale bars, 40 μm.

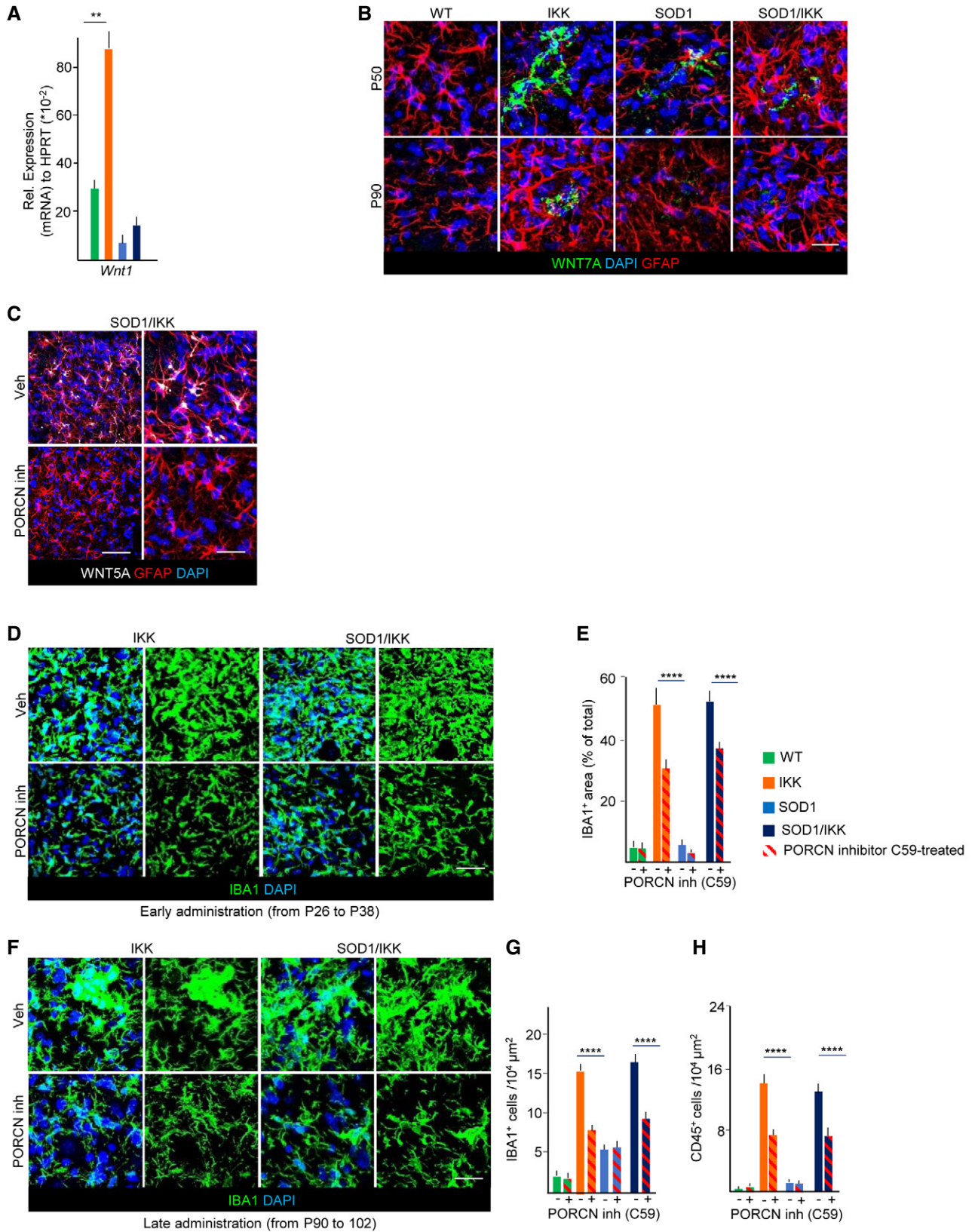


Figure EV5.