

advances.sciencemag.org/cgi/content/full/4/8/eaar4008/DC1

## Supplementary Materials for

## Synthetic matrix enhances transplanted satellite cell engraftment in dystrophic and aged skeletal muscle with comorbid trauma

Woojin M. Han, Shannon E. Anderson, Mahir Mohiuddin, Daniela Barros, Shadi A. Nakhai, Eunjung Shin, Isabel Freitas Amaral, Ana Paula Pêgo, Andrés J. García\*, Young C. Jang\*

\*Corresponding author. Email: andres.garcia@me.gatech.edu (A.J.G.); young.jang@gatech.edu (Y.C.J.)

Published 15 August 2018, *Sci. Adv.* **4**, eaar4008 (2018) DOI: 10.1126/sciadv.aar4008

## The PDF file includes:

- Fig. S1. Isolated primary MuSCs are Pax7<sup>+</sup>.
- Fig. S2. RGD-presenting hydrogels promote MuSC activation and proliferation.
- Fig. S3. RGD-presenting hydrogels promote MuSC differentiation.
- Fig. S4. Pax7/MyoD expression of MuSCs in 4 and 6% 20-kDa PEG-4MAL hydrogels.
- Fig. S5. 1D diffusion assay in PEG-4MAL hydrogels.

Fig. S6. Synthetic matrix supports higher MuSC proliferation potential than collagen gel.

Table S1. List of cell-adhesive synthetic peptides and their targets.

## Other Supplementary Material for this manuscript includes the following:

(available at advances.sciencemag.org/cgi/content/full/4/8/eaar4008/DC1)

Movie S1 (.mp4 format). Differentiated myotubes in RGD hydrogels contract in vitro.



**Fig. S1. Isolated primary MuSCs are Pax7<sup>+</sup>.** Freshly isolated cells were encapsulated in PEG-4MAL hydrogel and immediately fixed and stained. Top row scale bar: 100 μm. Bottom row scale bar: 10 μm.



**Fig. S2. RGD-presenting hydrogels promote MuSC activation and proliferation.** (**A**) Representative z-projections myogenic colonies formed in hydrogels presenting synthetic cell adhesive peptides. Day 3. Scale bars: 100  $\mu$ m. (**B**) MuSCs encapsulated and cultured in PEG-4MAL hydrogels become activated (Pax7<sup>+</sup>/MyoD<sup>+</sup>) by 72 hours postencapsulation in culture. Scale bars: 50  $\mu$ m. (**C**) Quantification of Pax7<sup>+</sup> and MyoD<sup>+</sup> cells. n=9-18. *p*=0.28 (Pax7) and *p*=0.69 (MyoD) Kruskal-Wallis test.



**Fig. S3. RGD-presenting hydrogels promote MuSC differentiation.** (**A**) Freshly isolated GFP<sup>+</sup> and TdTomato<sup>+</sup> MuSCs were seeded (1:1) collagen/laminin-coated tissue culture plastic to demonstrate fusion through co-expression of GFP and TdTomato. Scale bar: 200  $\mu$ m. (**B**) Freshly isolated GFP<sup>+</sup> and TdTomato<sup>+</sup> MuSCs were seeded (1:1) in PEG-4MAL hydrogels functionalized with RGD or RDG. Cells cultured in RGD-functionalized hydrogels fuse (arrow) and become more elongated compared to the cells cultured in RDG-functionalized hydrogels. Scale bar: 100  $\mu$ m. (**C**) Quantification of fusion index (%). n=8. *p*<0.0001 via unpaired two-tailed *t*-test. (**D**) Cells cultured in both RGD- and RDG-functionalized hydrogels stain positive for desmin and myosin heavy chain (MyHC). Cells were cultured in growth media for 6 days, then in differentiation media for 4 days. Scale bars: 100  $\mu$ m (Desmin), 500  $\mu$ m (MyHC). (**E**) Quantification of myogenin and desmin gene expression. n=4-5. n.s. *p*>0.05. Mean ± SEM.



**Fig. S4. Pax7/MyoD expression of MuSCs in 4 and 6% 20-kDa PEG-4MAL hydrogels.** (**A**) MuSCs encapsulated and cultured in PEG-4MAL hydrogels become activated ( $Pax7^+/MyoD^+$ ) by 76 hours post-encapsulation in culture. Scale bars: 50 µm. (**B**) Quantification of  $Pax7^+$  and  $MyoD^+$  cells. n=13-17. \* *p*<0.05 via unpaired *t*-test.



**Fig. S5. 1D diffusion assay in PEG-4MAL hydrogels.** (**A**) Representative micrographs of Alexa-555-labeled  $\alpha$ bungartoxin (8 kDa) diffusing into PEG-4MAL gels over time. (**B**) Quantification of intensity. The solution of Fick's second law was fitted through the data to determine the diffusion coefficients. n=5. Mean ± SEM. (**C**) Diffusion coefficients determined from curve fitting. n=5. Mean ± SEM. (**D-F**) Rheological assessments of 4% 20 kDa and 2.5% 10 kDa PEG-4MAL hydrogels. n=5. Mean ± SD. # *p*<0.05 via unpaired two-tailed *t*-test.



Fig. S6. Synthetic matrix supports higher MuSC proliferation potential than collagen gel. (A) Representative z-projections of GFP<sup>+</sup> MuSCs 1-day post-encapsulation in 4% PEG-4MAL hydrogel and 2.7 mg·ml<sup>-1</sup> collagen gel. Scale bars: 100  $\mu$ m. (B) Storage modulus of 2.8% PEG-4MAL hydrogel and 2.7 mg·ml<sup>-1</sup> collagen gel. n=4-5. *p*=0.14 via unpaired *t*-test. (C) Quantification of GFP<sup>+</sup> myogenic colony area in 4% PEG-4MAL hydrogel and 2.7 mg·ml<sup>-1</sup> collagen gel. n=10. \*\*\* *p*<0.001 via unpaired two-tailed *t*-test. (D) Representative z-projections of GFP<sup>+</sup> MuSCs 3-days post-encapsulation in 2.8% PEG-4MAL hydrogel and 2.7 mg·ml<sup>-1</sup> collagen gel. Scale bars: 100  $\mu$ m.

 Table S1. List of cell-adhesive synthetic peptides and their targets.

Common Name	Sequence	Targets
RGD	GRGDSPC	$\alpha_{v}\beta_{1}, \alpha_{v}\beta_{3}, \alpha_{v}\beta_{5}, \alpha_{v}\beta_{6}, \alpha_{v}\beta_{8}, \alpha_{5}\beta_{1}, \alpha_{8}\beta_{1}, \alpha_{11b}\beta_{3}$
RDG (scrambled control)	GRDGSPC	N/A (inactive)
YIGSR	CGGEGYGEGYIGSR	67 kDa laminin receptor
C16	CGGKAFDITYVRLKF	Syndecan, $\alpha_{v}\beta_{3}$ , $\alpha_{5}\beta_{1}$ , $\beta_{1}$