Imaging Drug Uptake by Bioorthogonal Stimulated Raman Scattering Microscopy

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Supplementary Figures, Supplementary Materials and

Synthetic Procedures

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Figure S1 *N*-pyrrolidine labelled anisomycin derivatives (**2b-i**) used in the experimental determination of vibrational shifts and Raman scattering intensities. (**2h** = **PhDY-ANS**; **2i** = **BADY-ANS**) Dimethylamine counterparts (**3b-i**) used to as truncated models to enable rapid DFT calculations of vibrational shifts and Raman scattering activities (I_{Ram}).

Raman Label	R	Compound	Raman shift (cm ⁻¹) ^a
N=	N-labelled anisomycin	2b	2234
Ř	Me ₂ N	3b	-
	N-labelled anisomycin	2c	2136
Ř	Me ₂ N	3c	-
	N-labelled anisomycin	2d	2249
Ř	Me ₂ N	3d	-
	N-labelled anisomycin	2e	2239
Ř	Me ₂ N	Зе	-
D ₃ C	N-labelled anisomycin	2f	2260 ^b
Ř	Me ₂ N	3f	-
	N-labelled anisomycin	2g	N.D. ^c
	Me ₂ N	Зg	-
	N-labelled anisomycin	2h (PhDY_ANS)	2236 ^d
	Me ₂ N	3h	-
	N-labelled anisomycin	2i	2219
PhR	Me ₂ N	(BADY-ANS) 3i	-

 Table S1 Anisomycin derivatives and their dimethylamine counterparts used in DFT studies.

^{*a*} Values presented for the solid material in the bioorthogonal region $1800 - 2800 \text{ cm}^{-1}$. ^{*b*} The bioorthogonal Raman shift due to the CD₃ group is of much lower intensity and was not used in this study. ^{*c*} Compound **2g** was found to be unstable to laser irradiation. ^{*d*} Intracellular shift 2219 cm⁻¹ determined by SRS imaging.



Figure S2 Raman activities anisomycin analogues and their dimethylamine counterparts. A Predicted Raman activity of dimethylamine adducts **3b-i**, and EdU **1**. DFT predicted Raman scattering activity (I_{ram}) of the peak maximum between 2100 – 2250 cm⁻¹ [gas phase intensities calculated at B3LYP/6-31G(d,p)]. EdU is highlighted in red for clarity. **B** Comparison of the DFT-predicted Raman activities and experimental spontaneous Raman scattering intensities for peaks in the region 2100 – 2250 cm⁻¹. DFT gas phase activities (I_{Ram}) for dimethylamine adducts **3b-i** calculated at the B3LYP/6-31G(d,p) level. Relative spontaneous Raman scattering intensities for the solid anisomycin analogues **2b-i** normalised to the peak area_(819 cm-1) for each analogue and expressed as a percentage of the BADY-ANS peak; mean of ten replicates with error bars ± S.D. (**2h = PhDY-ANS**; **2i = BADY-ANS**)



Figure S3 Spontaneous Raman spectroscopy of anisomycin ANS and analogues 2b-f, 2h and 2i as solid material. Spectra normalised to the anisomycin peak at 819 cm⁻¹ present in each sample, and offset for clarity. (2h = PhDY-ANS; 2i = BADY-ANS)

Note: compound 2g is unstable upon laser irradiation, and therefore no Raman spectrum is presented.



Figure S4 Correlation of the predicted Raman scattering activities calculated by Gaussian (I_{ram}), with experimental Raman intensities measured by spontaneous Raman scattering (I_{exp}). Due to the large dynamic ranges, these are plotted as their logarithms.



Figure S5 Effect of anisomycin **ANS** and analogues **2b-i** on the phosphorylation of JNK1/2 isoforms in SKBR3 cells. (**A**) Cells were exposed to either DMSO or anisomycin ANS (10 μ M) at selected timepoints; (**B**) Cells were exposed to DMSO (lane 1), and 5 μ M concentrations of anisomycin **ANS** (lane 2) and analogues **2b-i** (lanes 3-10) for 30 min. Western blot analysis was carried out with antibodies to phosphorylated JNK (pJNK1/2) and JNK1/2. β -actin was used as a loading control. (**2h** = **PhDY-ANS**; **2i** = **BADY-ANS**)



Figure S6 Correlation between SRS intensity measured at 2219 cm⁻¹ (**BADY-ANS** on-resonance) and **BADY-ANS** concentration as DMSO stock solutions. Average pixel intensities were calculated using ImageJ software. Images were acquired at 512×512 pixels with a 20 µs pixel dwell time. Data represented as mean of 6 replicates with error bars ± SD.



Figure S7 Time-lapse imaging of anisomycin **ANS** uptake into live SKBR3 cells. SKBR3 cells were treated with **ANS** (10 μ M at t = 0 min) and imaged at 2219 cm⁻¹ (C=C, **BADY-ANS**) every minute for 60 mins. Images were acquired at 1024 × 1024 pixels, 20 μ s pixel dwell time. Scale bars: 10 μ m.



Figure S8. Fast-acquisition SRS images of fixed SKBR3 cells treated with **BADY-ANS**. SKBR3 cells treated with **BADY-ANS** (100 μ M, 20 mins) and images acquired at (i) 2953 cm⁻¹ (CH₃, proteins); (ii) 2844 cm⁻¹ (CH₂, lipids); (iii) 2219 cm⁻¹ (C≡C, **BADY-ANS**); and (iv) 2202 cm⁻¹ (cell silent region). Images acquired at 512 × 512 pixels, 2 μ s pixel dwell time, false colour for images applied to different detection wavenumbers. Scale bars: 10 μ m.



Figure S9 Multi-colour SRS imaging of fixed SKBR3 cells treated with **BADY-ANS**. SKBR3 cells were treated with **BADY-ANS** (10 μ M, 30 min) and SRS images acquired at (i) 2844 cm⁻¹ (CH₂, lipids) and (ii) 2219 cm⁻¹ (C=C, **BADY-ANS**) and (iii) overlay of (i) and (ii) showing that **BADY-ANS** is initially concentrated in lipid droplets in some of the cells. Images acquired at 1024 × 1024 pixels, 20 µs pixel dwell time, false colour for images applied to different detection wavenumbers. Scale bars: 10 µm.



Typical field of view



Figure S10 Quantification of the number of lipid droplets present in SKBR3 cells following treatment with (i) DMSO; (ii) **ANS** (10 μ M, 30 min) and (iii) **BADY-ANS** (10 μ M, 30 min). SRS images were acquired at 2844 cm⁻¹ (CH₂, lipids) across a typical field of view (~40 cells, 20× objective lens, n = 9 repeats), and the number of lipid droplets >1 μ m were counted using ImageJ. Inset: maximum intensity Z-projection for a typical field of view following acquisition of a Z-stack of SRS images at 2844 cm⁻¹. Scale bar: 10 μ m. See Materials and Methods for further details. The average number of lipid droplets under each treatment is reported across n = 9 repeats.



Figure S11 Dual colour alkyne-label imaging by spontaneous Raman spectrosocpy. Spontaneous Raman spectrum of SKBR3 cells treated with EdU (100 μ M, 18 h) and BADY-ANS (100 μ M, 20 min). Peak at 2120 cm⁻¹ indicative of EdU (red) and peak at 2219 cm⁻¹ indicative of BADY-ANS (black). Raman spectrum acquired at $\lambda_{ex} = 785$ nm for 60 s using a 50× objective.

Movie BADY-ANS

Live SKBR3 cells were treated with **BADY-ANS** (10 μ M) at t=0 min and SRS images were acquired at 2219 cm⁻¹ (C=C, **BADY-ANS**) every minute for 60 minutes. The individual frames were compiled on ImageJ as an image stack to generate the movie. Scale bars: 50 μ m.

Movie ANS

Live SKBR3 cells were treated with ANS (10 μ M) at t=0 min and SRS images were acquired at 2219 cm⁻¹ (C=C, **BADY-ANS**) every minute for 60 minutes. The individual frames were compiled on ImageJ as an image stack to generate the movie. Scale bars: 50 μ m.

General Methods

All non-aqueous reactions were carried out under an atmosphere of nitrogen or argon using ovendried glassware that was cooled in a desiccator prior to use. Unless otherwise noted, starting materials and reagents were obtained from commercial suppliers and were used without further purification. Toluene, THF, CH₂Cl₂, and Et₂O were dried and purified by passage through activated alumina columns using a Glass Contour Solvent Purification System. Triethylamine was distilled from calcium hydride and stored over activated 4 Å molecular sieves under an argon atmosphere. Anhydrous DMF was purchased from Acros Organics. Saturated aqueous solutions of inorganic salts are represented as (volume, sat aq.). Nuclear magnetic resonance (NMR) spectra were recorded at ambient temperature (298 K, unless otherwise stated) on a Bruker AVA400, AVA500 or AVA600 spectrometer running at 400, 500, or 600 MHz (¹H spectra) or 101, 126, 151 Hz (¹³C spectra), respectively. Chemical shifts (δ values) are reported in parts-per-million (ppm) relative to tetramethylsilane (¹H and ¹³C spectra; $\delta_{\text{TMS}} = 0$) and are calibrated to the residual solvent peak. ¹H NMR data are reported as follows: chemical shift, relative intensity, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, qn = quintet, m = multiplet, br = broad), coupling constants (J value, Hz), and interpretation. ¹³C NMR data are reported as follows: chemical shift, relative intensity and assignment (Q = quaternary, CH = methane, CH_2 = methylene, CH_3 = methyl). Infra-red spectra (IR) were recorded neat on Shimadzu IRAffinity-1. The value of peaks at maximum absorbance (v_{max}) are quoted in wavenumbers (cm^{-1}) . Mass spectra were obtained by electrospray (ESI) on a Bruker microTOF II mass spectrometer, or by electron ionisation (EI) on a Kratos MS50TC mass spectrometer. Mass-to-charge ratios (m/z) of all parent (molecular) ions ($[M]^{+/-}$) and their intensities are reported, followed by (major) fragment or adduct ions and their intensities. Melting points (mp) were determined on a Gallenkamp Electrothermal Melting Point apparatus and are uncorrected the temperature range. R_f values (R_f) were recorded using Merck Silicagel 60 F254 aluminium backed plates. Flash chromatography was carried out using Merck Kieselgel 60 (Merck 9385) under positive pressure. Eluent compositions are quoted as v/v ratios. Rt values were recorded by analytical reverse phase HPLC analysis using a Waters 600E (100 µL) gradient pump using a 717plus autosampler and a Waters 996 PDA equipped with a Phenomenex Luna C18(2), 5 µm, 250 x 4.6 mm column at a flow rate of 1 mL min⁻¹. Preparative reverse phase HPLC was performed using a Waters 600 (225 µL) system using a Waters 486 tuneable absorbance detector recording at 254 nm equipped with a Phenomonex Luna C18(2), 5 μ m, 250 x 21.2 mm column at a flow rate of 21.2 mL min⁻¹.

Flow Rate: 21.2 mL min ⁻¹		λ: 275 nm	
Time (min)	% H ₂ O + 0.1% T	FA	% MeCN + 0.1% TFA
0.0	80		20
20	60		40
50	60		40
55	5		95
65	5		95

Preparative RP HPLC Method A

Analytical RP HPLC Method B

Flow Rate: 1.0 mL min ⁻¹	λ: 275 nm	
Time (min)	% H ₂ O + 0.1% TFA	% MeCN + 0.1% TFA
0	80	20
20	60	40
25	5	95
35	5	95
40	80	20
50	80	20

Analytical RP HPLC Method C

Flow Rate: 1.0 mL min ⁻¹	λ: 2	275 nm
Time (min)	% H ₂ O + 0.1% TFA	% MeCN + 0.1% TFA
0.0	95	5
30	5	95
35	5	95
40	95	5
50	95	5



Scheme S1 Synthesis of anisomycin analogues. Reagents and Conditions: ANS (60 μ mol), 4b-i (60 μ mol), K₂CO₃, DMF, 9 h, 71-95%.³⁷ Labels 4e and 4g were reacted as their alkyne protected-counterparts 4e-TMS and 4g-TMS which were deprotected *in situ*.

Synthesis of labels 4e-i

5-Trimethylsilyl-2,4-pentadiyn-1-ol, S1

TMS



A solution of 1,4-bis(trimethylsilyl)butadiyne (486 mg, 2.50 mmol) and LiBr (271 mg, 3.13 mmol) in anhydrous Et_2O (25 mL) at -10 °C, was stirred for 10 min in darkness. MeLi (1.95 mL, 3.13 mmol; 1.6 M in Et_2O) was added dropwise and the mixture was stirred at -10 °C for 15 min,

warmed to rt and stirred for a further 2 h in darkness. The mixture was cooled to 0 °C, and a suspension of paraformaldehyde (225 mg, 7.50 mmol) in dry Et₂O (30 mL) was added slowly, the mixture was warmed to rt and stirred for 18 h in darkness. The mixture was washed with NH₄Cl (50 mL; sat. aq.), NaHCO₃ (50 mL; sat. aq.), and brine (50 mL). The combined aqueous extracts were extracted with Et₂O (3 × 50 mL). The combined organic extracts were dried (MgSO₄), filtered, and concentrated *in vacuo*. The crude material was purified using flash column chromatography (Hexane:EtOAc, 5:1) to afford the product as a pale golden oil (199 mg, 52%). **R**_f (Hexane:EtOAc, 5:1) = 0.19; **IR** (neat, cm⁻¹) 3323 (OH), 2224 (C≡C), 2108 (C≡C); ¹**H** NMR (CD₃OD, 500 MHz) δ 4.26 (2H, s, CH₂); 0.20 (9H, s, 3CH₃); ¹³C NMR (CD₃OD, 126 MHz) 87.2 (1C, Q), 85.5 (1C, Q), 76.5 (1C, Q), 68.7 (1C, Q), 49.6 (1C, CH₂), -1.9 (3C, 3CH₃); *m*/z (EI) 152.1 ([M]⁺, 22%), 137.0 (100), 109.0 (17), 77.0 (10), 75.0 (31), 63 (3); **HRMS** (EI) calcd. for C₈H₁₂OSi [M]⁺ 152.0652, found 152.0647. ¹H and ¹³C NMR spectroscopic data were in good agreement with the literature.¹

5-Bromo-1-trimethylsilyl-1,3-pentadiyne, 4e-TMS



A solution of 5-trimethylsilyl-2,4-pentadiyn-1-ol **S1** (59.8 mg, 400 μ mol) in anhydrous Et₂O (1.0 mL) at 0 °C, was treated successively with pyridine (2.5 μ L, 32 μ mol) and PBr₃ (15.2 μ L, 160 μ mol). The reaction mixture was warmed to rt and stirred for 18 h in darkness. The mixture was diluted with

Et₂O (30 mL) and washed with Na₂CO₃ (15 mL; sat. aq.). The aqueous layer was separated and extracted with Et₂O (3 × 30 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified using flash column chromatography (Hexane) to afford the product as a pale yellow oil (68 mg, 79%). **R**_f (Hexane) = 0.74; **IR** (neat, cm⁻¹) 2358 (C=C), 2112 (C=C); ¹H NMR (CD₃OD, 500 MHz) δ 4.15 (2H, s, CH₂); 0.22 (9H, s, 3CH₃); ¹³C NMR (CD₃OD, 126 MHz) δ 87.8 (1C, Q), 86.6 (1C, Q), 72.6 (1C, Q), 70.1 (1C, Q), 13.0 (1C, CH₂), -2.0 (3C, 3CH₃); *m/z* (EI) 216.0 ([⁸¹BrM]⁺, 42%), 214.0 ([⁷⁹BrM]⁺, 40), 201.0 (100), 199.0 (97), 172.9 (34), 170.9 (26), 135.1 (44), 64.1 (78); **HRMS** (EI) calcd. for C₈H₁₁BrSi [⁷⁹BrM]⁺ 213.9808, found 213.9807. ¹H and ¹³C NMR spectroscopic data are in good agreement with the literature.²

[5-²H₃]-1-Trimethylsilyl-1,3-pentadiyne, S2

 CD_3



A solution of 1,4-bis(trimethylsilyl)butadiyne (972 mg, 5.00 mmol) in anhydrous THF (10 mL) was treated with MeLi•LiBr complex (3.78 mL, 5.67 mmol, 1.5 N in Et₂O), and the resultant mixture stirred at rt for 5 h in darkness. The reaction mixture was cooled to -78 °C, and CD₃I (335 μ L,

5.38 mmol) was added dropwise. The mixture was stirred at -78 °C for 1 h, and slowly warmed to rt over 12 h. The reaction mixture was cooled to 0 °C, and slowly quenched with NH₄Cl (20 mL; sat. aq.). The aqueous mixture was then extracted with Et₂O (3×15 mL), and the combined organic

extracts were washed with water (3 × 15 mL) and brine (15 mL). The resultant organic mixture was dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified using flash column chromatography (Hexane) to give the product as a pale yellow oil (400 mg, 57%). \mathbf{R}_f (Hexane) = 0.82; **IR** (neat, cm⁻¹) 2266 (C≡C), 2230 (C≡C), 2110 (CD); ¹H NMR (CD₃OD, 500 MHz) δ 0.18 (9H, s, 3CH₃); ¹³C NMR (CD₃OD, 126 MHz) δ 88.4 (1C, Q), 80.8 (1C, Q), 75.1 (1C, Q), 64.0 (1C, Q), 1.6 (1C, sept, *J* 20.9 Hz, *C*D₃), -1.7 (3C, 3 × CH₃); *m*/z (EI) 139.0 ([M]⁺, 39%), 124.0 (100); **HRMS** (EI) calcd. for C₈H₉D₃Si [M]⁺ 139.0891, found 139.0889.

[6-²H₃]-2,4-Hexadiyn-1-ol, S3

 CD_3



A solution of $[5-{}^{2}H_{3}]$ -1-trimethylsilyl-1,3-pentadiyne **S2** (139 mg, 1.00 mmol) in anhydrous THF (10 mL) was cooled to -10 °C, and treated with MeLi•LiBr complex (830 µL, 1.25 mmol, 1.5 N in Et₂O). The mixture was stirred at -10 °C for 15 min, warmed to rt and stirred for an additional

2 h. After complete desilylation, the mixture was cooled to -10 °C, and a suspension of paraformaldehyde (60.9 mg, 2.03 mmol) in anhydrous THF (3.0 mL) was added. The mixture was warmed to rt and stirred for 18 h. The reaction was quenched by the slow addition of NH₄Cl (20 mL; sat. aq.), and the organic layer was washed with NaHCO₃ (20 mL; sat. aq.) and brine (20 mL). The combined aqueous layers were extracted with Et₂O (3 × 30 mL), and the combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane:EtOAc, 10:1 → Hexane:EtOAc, 4:1) to afford the product as pale yellow needles (95 mg, 98%). **R**_f (Hexane:EtOAc, 4:1) = 0.39; **IR** (neat, cm⁻¹) 3329 (OH), 2259 (C≡C); ¹**H** NMR (CD₃OD, 500 MHz) δ 4.21 (2H, s, CH₂); ¹³C NMR (CD₃OD, 126 MHz) δ 75.8 (1C, Q), 73.1 (1C, Q), 69.1 (1C, Q), 63.2 (1C, Q), 49.6 (1C, CH₂), 1.5 (1C, sept., *J* 20.7 Hz, CD₃); *m*/z (EI) 97.0 ([M]⁺, 100%), 80.0 (60), 77.9 (73), 69.0 (72), 68.0 (46), 62.9 (60), 52.9 (39); **HRMS** (EI) calcd. for C₆H₃D₃O [M]⁺ 97.0602, found 97.0598.

[6-²H₃]-1-Bromo-2,4-hexadiyne, 4f



A solution of $[6^{-2}H_3]$ -2,4-hexadiyn-1-ol **S3** (27.2 mg, 280 µmol) in anhydrous Et₂O (1.0 mL) at 0 °C, was treated successively with pyridine (1.9 µL, 24 µmol) and PBr₃ (10.5 µL, 110 µmol). The reaction mixture was warmed to rt and stirred for 18 h in darkness. The mixture was diluted with

Et₂O (20 mL) and washed with Na₂CO₃ (10 mL; sat. aq.). The aqueous layer was separated and extracted with Et₂O (3 × 25 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane) to afford the product as a colourless oil (36 mg, 80%). **R**_f(Hexane) = 0.85; **IR** (neat, cm⁻¹) 2257 (C=C); ¹**H NMR** (CD₃OD, 600 MHz) δ 4.10 (2H, s, CH₂Br); ¹³**C NMR** (CD₃OD, 151 MHz) δ 78.0 (1C, Q), 70.8 (1C, Q), 69.4 (1C, Q), 62.9 (1C, Q), 13.6 (1C, CH₂), 1.7 (1C, sept. *J* 20.1 Hz, CD₃); *m/z* (EI) 161.0 ([⁸¹BrM]⁺, 27%), 159.0 ([⁷⁹BrM]⁺, 28), 80.1 (100), 78.1 (58), 63.0 (62) **HRMS** (EI) calcd. for C₆H₂D₃Br [⁷⁹BrM]⁺ 158.9757, found 158.9762.

4-Iodobenzyl alcohol, S4

1-Trimethylsilyl-1,3-butadiyne, S5

A solution of 1,4-bis(trimethylsilyl)butadiyne (972 mg, 5.00 mmol) in anhydrous Et₂O (10 mL) at 0 °C was treated with MeLi•LiBr complex (5.0 mL, 7.50 mmol, 1.5 N in Et₂O). The reaction mixture was warmed to rt, and stirred for 5 h. The reaction was quenched by the addition of NH₄Cl (10 mL, sat. aq.), and the resulting mixture extracted with Et₂O (3 × 20 mL). The combined organic extracts were washed with water (20 mL) and brine (20 mL), dried (MgSO₄), filtered and concentrated at ambient conditions overnight. The product was found to be volatile and unstable when concentrating to dryness; approximate concentration was determined by ¹H NMR (0.16 N in Et₂O). **R**_f (Hexane) = 0.98; ¹H NMR (CDCl₃, 500 MHz) δ 2.13 (1H, s, *H*C=CC=C), 0.23 (9H, s, 3 × CH₃); ¹³C NMR (CDCl₃, 126 MHz) δ 88.0 (1C, Q), 84.7 (1C, Q), 68.3 (1C, CH), 66.6 (1C, Q), -0.6 (3C, 3CH₃). ¹H and ¹³C NMR spectroscopic data are in good agreement with the literature.⁴

4-(4-Trimethylsilyl-1,3-butadiyn-1-yl)benzyl alcohol, S6

A mixture of 4-iodobenzyl alcohol **S4** (748 mg, 3.20 mmol), Et₃N (890 μ L, 6.40 mmol), PdCl₂(PPh₃)₂ (45.5 mg, 65.0 μ mol, 2 mol%) and CuI (34.2 mg, 180 μ mol, 5.6 mol%) were dissolved

in anhydrous Et₂O (4 mL). The reaction mixture was stirred for 10 minutes, after which 1trimethylsilyl-1,3-butadiyne **S5** (20 mL, 3.2 mmol, 0.16 N in Et₂O) was added, and the mixture stirred at rt for 18 h. The resulting mixture was concentrated *in vacuo* and re-dissolved in CH₂Cl₂ (50 mL). The organic mixture was washed with HCl (2 × 40 mL, 1 N aq.) and water (40 mL). The combined aqueous washings were extracted CH₂Cl₂ (3 × 100 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (CH₂Cl₂) to afford the product as a light brown solid (272 mg, 37%). **R**_f(CH₂Cl₂) = 0.36; **mp** 88 - 90 °C; **IR** (neat, cm⁻¹) 3256 (OH), 2203 (C=C), 2102 (C=C), 1508 (C=C); ¹**H** NMR (CDCl₃, 500 MHz) δ 7.51 (2H, d, *J* 8.4 Hz, 2 × Ar*H*), 7.35 (2H, d, *J* 8.4 Hz, 2 × Ar*H*), 4.73 (2H, d, *J* 5.9 Hz, CH₂OH), 1.72 (1H, t, *J* 5.9 Hz, CH₂OH), 0.26 (9H, s, 3 × CH₃); ¹³**C** NMR (CDCl₃, 126 MHz) δ 142.2 (1C, Q), 132.9 (2C, CH), 126.8 (2C, CH), 120.6 (1C, Q), 90.8 (1C, Q), 87.8 (1C, Q), 76.6 (1C, Q), 74.2 (1C, Q), 64.8 (1C, CH₂), -0.4 (3C, CH₃); m/z (EI) 228.1 ([M]⁺, 39%), 213.1 (100); **HRMS** (EI) calcd. for C₁₄H₁₆OSi [M]⁺ 228.0965, found 228.0978. ¹H NMR spectroscopic data is in good agreement with the literature.⁵

4-(4-Trimethylsilyl-1,3-butadiyn-1-yl)benzyl bromide, 4g-TMS

A solution of 4-(4-trimethylsilyl-1,3-butadiyn-1-yl)benzyl alcohol **S6** (91 mg, 400 μ mol) in anhydrous Et₂O (1 mL) at 0 °C, was treated with pyridine (2.5 μ L, 32 μ mol) then PBr₃(15.2 μ L,

160 μmol). The reaction mixture was warmed to rt and stirred for 18 h in darkness. The reaction mixture was diluted with Et₂O (30 mL) and washed with Na₂CO₃ (15 mL; sat. aq.). The aqueous layer was separated and extracted with Et₂O (3 × 30 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane) to afford the product as a pale yellow solid (90 mg, 77%). **R**_f (Hexane) = 0.48; **mp** 65 – 67 °C; **IR** (neat, cm⁻¹) 2203 (C≡C), 2099 (C≡C); ¹**H** NMR (CD₃OD, 500 MHz) δ 7.50 (2H, d, *J* 8.4 Hz, 2 × Ar*H*), 7.44 (2H, d, *J* 8.4 Hz, 2 × Ar*H*), 4.59 (2H, s, CH₂Br), 0.24 (9H, s, 3 × CH₃); ¹³C NMR (CD₃OD, 126 MHz) δ 140.0 (1C, Q), 132.6 (2C, CH), 129.1 (2C, CH), 121.0 (1C, Q), 90.0 (1C, Q), 87.2 (1C, Q), 75.6 (1C, Q), 74.0 (1C, Q), 31.7 (1C, CH₂), -1.9 (3C, CH₃); *m/z* (EI) 292.0 ([⁸¹BrM]⁺, 14%), 290.0 ([⁷⁹BrM]⁺, 14), 211.1 (100), 196.0 (27), 183.0 (24); **HRMS** (EI) calcd. for C₁₄H₁₅BrSi [⁷⁹BrM]⁺ 290.0121, found 290.0132.

4-Phenyl-1-trimethylsilyl-1,3-butadiyne, S7

A mixture of iodobenzene (180 μ L, 1.6 mmol), Et₃N (450 μ L, TMS 3.2 mmol), PdCl₂(PPh₃)₂ (11.5 mg, 16.4 µmol, 1 mol%) and CuI (8.8 mg, 46.2 µmol, 3 mol%) were dissolved in anhydrous Et₂O (2 mL). The mixture was stirred for 10 min, after which 1-trimethylsilyl-1,3-butadiyne S5 (10 mL, 1.6 mmol, 0.16 N in Et₂O) was added, and the mixture stirred at rt for 18 h. The mixture was concentrated in vacuo and re-dissolved in CH_2Cl_2 (25 mL). The organic mixture was washed with HCl (2 × 20 mL, 1 N aq.) and water (20 mL). The combined aqueous washings were extracted CH_2Cl_2 (3 × 50 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated in vacuo. The crude material was purified by flash column chromatography (CH₂Cl₂) to afford the product as a pale yellow oil (260 mg, 82%). \mathbf{R}_{f} (Hexane) = 0.64; IR (neat, cm⁻¹) 2207 (C=C), 2102 (C=C); ¹H NMR (CD₃OD, 500 MHz) δ 7.53 - 7.49 (2H, m, 2 × ArH), 7.46 - 7.41 (1H, m, ArH), 7.38 (2H, t, J 7.5 Hz, 2 × ArH), 0.24 (9H, s, 3 × CH₃); ¹³C NMR (CD₃OD, 126 MHz) δ 132.2 (2C, CH), 129.3 (1C, CH), 128.3 (2C, CH), 121.1 (1C, Q), 89.5 (1C, Q), 87.4 (1C, Q), 76.0 (1C, Q), 73.3 (1C, Q), -1.8 (3C, CH₃); *m/z* (EI) 198.1 $([M]^+, 32\%)$, 183.2 (100); **HRMS** (EI) *m/z* calcd. for C₁₃H₁₄Si $[M]^+$ 198.0859, found 198.0862. ¹H and ¹³C NMR spectroscopic data are in good agreement with the literature.⁶

5-Phenyl-2,4-pentadiyn-1-ol, S8



A solution of 4-phenyl-1-trimethylsilyl-1,3-butadiyne S7 (198.3 mg, 1.00 mmol) in anhydrous THF (10 mL) was cooled to -10 °C, and treated with MeLi•LiBr complex (1.3 mL, 2.00 mmol, 1.5 N in Et_2O).

The mixture was stirred at -10 °C for 15 min, warmed to rt and stirred for an additional 2 h. After complete desilylation, the mixture was cooled to -10 °C, and a suspension of paraformaldehyde (90.7 mg, 3.02 mmol) in anhydrous THF (3.0 mL) was added. The mixture was warmed to rt and stirred for 18 h. The reaction was quenched by the slow addition of NH₄Cl (20 mL; sat. aq.), and the organic layer was washed with NaHCO₃ (20 mL; sat. aq.) and brine (20 mL). The combined aqueous layers were extracted with Et₂O (3 × 30 mL), and the combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane:EtOAc, 10:1) to afford the product as a pale yellow oil (117.2 mg, 75%). **R**_f (Hexane:EtOAc, 5:2) = 0.54; **IR** (neat, cm⁻¹) 3293 (OH), 2241 (C=C); ¹**H** NMR (CD₃OD, 500 MHz) δ 7.53 – 7.48 (2H, m, 2 × Ar*H*), 7.45 – 7.35 (3H, m, 3 × Ar*H*), 4.34 (2H, s, C*H*₂OH); ¹³C NMR (CD₃OD, 126 MHz) δ 132.1 (2C, CH), 129.1 (1C, CH), 128.3 (2C, CH), 121.4 (1C, Q), 81.2 (1C, Q), 77.1 (1C, Q), 72.7 (1C, Q), 68.3 (1C, Q), 49.8 (1C, CH₂); *m/z* (EI) 156.0 ([M]⁺, 100%), 139.0 (22), 128.1 (65), 102.0 (64); **HRMS** (EI) *m/z* calcd. for C₁₁H₈O [M]⁺ 156.0570, found 156.0565. ¹H and ¹³C NMR spectroscopic data are in good agreement with the literature.⁷

5-Bromo-1-phenyl-1,3-pentadiyne, 4h



A solution of 5-phenyl-2,4-pentadiyn-1-ol **S8** (117.2 mg, 750 μ mol) in anhydrous Et₂O (3 mL) at 0 °C, was treated with pyridine (4.8 μ L, 60 μ mol) then PBr₃ (28.5 μ L, 300 μ mol). The reaction mixture was

warmed to rt and stirred for 18 h in darkness. The mixture was diluted with Et₂O (30 mL) and washed with Na₂CO₃ (15 mL; sat. aq.). The aqueous layer was separated and extracted with Et₂O (3 × 30 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane) to afford the product as a yellow oil (97 mg, 59 %). **R**_f (Hexane) = 0.47; **IR** (neat, cm⁻¹) 2243 (C≡C), 2222 (C≡C); ¹**H** NMR (CD₃OD, 500 MHz) δ 7.55 – 7.51 (2H, m, 2 × Ar*H*), 7.47 – 7.42 (1H, m, Ar*H*), 7.41 – 7.37 (2H, m, 2 × Ar*H*), 4.24 (2H, s, C*H*₂Br); ¹³**C** NMR (CD₃OD, 126 MHz) δ 132.2 (2C, CH), 129.4 (1C, CH), 128.3 (2C, CH), 121.0 (1C, Q), 78.9 (1C, Q), 77.3 (1C, Q), 72.3 (1C, Q), 69.8 (1C, Q), 13.4 (1C, CH₂); *m/z* (EI) 219.9 ([⁸¹BrM]⁺, 11%), 217.9 ([⁷⁹BrM]⁺, 11), 139.0 (100); **HRMS** (EI) calcd. for C₁₁H₇Br [⁷⁹BrM]⁺ 217.9726, found 217.9711.

4-(4-Phenyl-1,3-butadiyn-1-yl)benzyl alcohol, S9

A solution of 4-phenyl-1-trimethylsilyl-1,3-butadiyne S7 (100.1 mg, 500 μ mol) in THF:MeOH (2 mL, 1:1 v/v) was treated with K₂CO₃ (138.2 mg, 1.00 mmol) and the mixture

stirred at rt for 2 h. The reaction was quenched by the addition of NH₄Cl (2.0 mL, sat. aq.) and Et₂O (4.0 mL). The organic phase was separated, washed with brine (2 × 5 mL), dried (MgSO₄) and filtered. To the deprotected alkyne was added Et₃N (4.2 mL, 30.0 mmol), 4-iodobenzyl alcohol **S4** (117.1 mg, 500 µmol), PdCl₂(PPh₃)₂ (3.5 mg, 5.0 µmol, 1 mol%) and CuI (2.7 mg, 14 µmol, 2.8 mol%), and the mixture stirred at rt for 18 h. The reaction was quenched by the addition of NH₄Cl (10 mL, sat. aq.) and the resulting mixture extracted with Et₂O (3 × 20 mL). The combined organic extracts were washed with brine (15 mL), dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane:EtOAc, 5:2) to give the product as a white solid (97.6 mg, 84%). **R**_f (Hexane:EtOAc, 5:2) = 0.35; **mp** 126 – 128 °C;

IR (neat, cm⁻¹) 3246 (OH), 2216 (C=C), 2145 (C=C); ¹**H** NMR (CD₃OD, 500 MHz) δ 7.58 – 7.51 (4H, m, 4 × Ar*H*), 7.48 – 7.35 (5H, m, 5 × Ar*H*), 4.65 (2H, s, C*H*₂OH); ¹³C NMR (CD₃OD, 126 MHz) δ 143.3 (1C, Q), 132.1 (2C, CH), 132.0 (2C, CH), 129.1 (1C, CH), 128.3 (2C, CH), 126.6 (2C, CH), 121.6 (1C, Q), 120.2 (1C, Q), 80.9 (1C, Q), 80.7 (1C, Q), 73.0 (1C, Q), 72.8 (1C, Q), 63.2 (1C, CH₂); *m*/*z* (EI) 232.1 ([M]⁺, 100%), 202.1 (44); **HRMS** (EI) calcd. for C₁₇H₁₂O [M]⁺ 232.0883, found 232.0889. ¹H and ¹³C NMR spectroscopic data are in good agreement with the literature.⁸

4-(4-Phenyl-1,3-butadiyn-1-yl)benzyl bromide, 4i



A solution of 4-(4-phenyl-1,3-butadiyn-1-yl)benzyl alcohol **S9** (116.1 mg, 500 μ mol) in anhydrous Et₂O (3 mL) at 0 °C, was treated with pyridine (4.8 μ L, 60 μ mol) then PBr₃ (28.5 μ L,

300 µmol). The reaction mixture was warmed to rt and stirred for 18 h in darkness. The reaction mixture was diluted with Et₂O (30 mL) and washed with Na₂CO₃ (15 mL; sat. aq.). The aqueous layer was separated and extracted with Et₂O (3 × 30 mL). The combined organic extracts were dried (MgSO₄), filtered and concentrated *in vacuo*. The crude material was purified by flash column chromatography (Hexane) to afford the product as white solid (91 mg, 62 %). **R**_f (Hexane) = 0.25; **IR** (neat, cm⁻¹) 2216 (C≡C), 2145 (C≡C); ¹**H NMR** (CD₃OD, 500 MHz) δ 7.57 – 7.51 (4H, m, 4 × Ar*H*), 7.49 – 7.38 (5H, m, 5 × Ar*H*), 4.60 (2H, s, C*H*₂Br); ¹³C **NMR** (CD₃OD, 126 MHz) δ 139.8 (1C, Q), 132.4 (2C, CH), 132.1 (2C, CH), 129.2 (1C, CH), 129.1 (2C, CH), 128.3 (2C, CH), 121.4 (1C, Q), 121.4 (1C, Q), 81.2 (1C, Q), 80.4 (1C, Q), 73.7 (1C, Q), 72.9 (1C, Q), 31.7 (1C, CH₂); *m/z* (EI) 295.9 ([⁸¹BrM]⁺, 12%), 293.9 ([⁷⁹BrM]⁺, 12), 215.0 (100), 213.0 (44), 107.4 (10); **HRMS** (EI) calcd. for C₁₇H₁₁Br [⁷⁹BrM]⁺ 294.0039, found 294.0034.

Characterisation of anisomycin analogues 2b-i

General procedure for the synthesis of Raman-labelled anisomycin derivatives

To a solution of **ANS** (60.0 μ mol) in DMF (1.5 mL) was added potassium carbonate (60.0 μ mol) and **4b-i** (60.0 μ mol), the solution was stirred at room temperature for 9 h. The solution was then concentrated *in vacuo*, and the residue was purified by RP HPLC, and freeze dried to afford the product.

N-Cyanomethyl anisomycin, 2b



Purification (Method A); colourless oil (20.9 mg, 87%). **R**_t (Method B) = 19.4 min; **IR** (neat, cm⁻¹) 3410 (OH), 2363 (C=N), 1748 (C=O), 1672 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹**H NMR** (CDCl₃, 500 MHz) δ 7.17 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 6.89 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 4.82 (1H, d, *J* 3.7 Hz, C₃*H*), 4.31 – 4.28 (1H, m C₄*H*), 3.89 – 3.70 (2H, dd, *J* 41.5, 17.5 Hz, NC*H*₂C=N), 3.82 (3H, s, OMe), 3.79 (1H, dd, *J* 11.2, 6.1 Hz, C₅*H*_dH_b), 3.65 (1H, dd, *J*

12.4, 7.4 Hz, C₂*H*), 3.01 – 2.92 (2H, m, C*H*₂Ar), 2.86 (1H, dd, *J* 11.2, 6.1 Hz, C₅H_a*H_b*), 2.19 (3H, s, O*Ac*); ¹³C NMR (CDCl₃, 126 MHz) δ 170.9 (1C, Q), 158.8 (1C, Q), 129.8 (2C, CH), 127.9 (1C, Q), 114.5 (2C, CH), 112.8 (1C, Q), 80.1 (1C, CH), 73.5 (1C, CH), 65.8 (1C, CH), 59.1 (1C, CH₂), 55.3

(1C, CH₃), 40.6 (1C, CH₂), 31.4 (1C, CH₂), 20.7 (1C, CH₃); *m/z* (ESI) 305.2 ($[M + H]^+$, 100%), 263.1 (11), 179.0 (41), 117.0 (22); **HRMS** (ESI) calcd. for C₁₆H₂₀O₄N₂Na $[M + Na]^+$ 327.1315, found 327.1315.

N-Propargyl anisomycin, 2c



Purification (Method A); colourless oil (21.6 mg, 90%). **R**_t (Method B) = 16.6 min; **IR** (neat, cm⁻¹) 3277 (OH), 2131 (C=C), 1749 (C=O), 1672 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹**H** NMR (CDCl₃, 500 MHz) δ 7.14 (1H, d, *J* 8.6 Hz, 2 × ArH), 6.88 (1H, d, *J* 8.6 Hz, 2 × ArH), 4.82 (1H, d, *J* 4.0 Hz, C₃H), 4.40 – 4.36 (1H, m, C₄H), 4.22 – 4.15 (1H, m, C₅H_aH_b), 4.03 – 4.00 (1H, m,

C₂H), 3.95 (2H, ddd, *J* 38.2, 17.2, 2.1 Hz, NC*H*₂C=CH), 3.82 (3H, s, O*Me*), 3.21 – 3.15 (1H, m, C₅H_a*H*_b), 3.15 – 3.07 (1H, m, C*H*₂Ar), 2.62 (1H, t, *J* 2.1 Hz, NCH₂C=C*H*), 2.20 (3H, s, O*Ac*); ¹³C NMR (CDCl₃, 126 MHz) δ 170.4 (1C, Q), 158.9 (1C, Q), 129.8 (2C, CH), 127.2 (1C, Q), 114.5 (2C, CH), 78.74 (1C, Q), 78.68 (1C, CH), 77.2 (1C, CH),* 72.3 (1C, CH), 63.3 (1C, CH), 57.8 (1C, CH₂), 55.3 (1C, CH₃), 41.2 (1C, CH₂), 29.6 (1C, CH₂), 20.6 (1C, CH₃); *m/z* (ESI) 326.1 ([M + Na]⁺, 100%), 304.2 ([M + H]⁺, 8), 284.1 (18), 262.1 (15), 244.1 (6); **HRMS** (ESI) calcd. for C₁₇H₂₂O₄N [M + H]⁺ 304.1549, found 304.1547. *Obscured by solvent peak – observed in HSQC.

N-2-Butyn-1-yl anisomycin, 2d



Purification (Method A); colourless oil (17.6 mg, 71%). **R**_t (Method B) = 23.1 min; **IR** (neat, cm⁻¹) 3271 (OH), 2247 (C=C), 1751 (C=O), 1670 (C=O, TFA), 1612 (C=C), 1585 (C=C), 1514 (C=C); ¹H NMR (CDCl₃, 500 MHz) δ 7.11 (2H, d, *J* 8.2 Hz, 2 × Ar*H*), 6.87 (2H, d, *J* 8.2 Hz, 2 × Ar*H*), 4.87 (1H, s, C₃*H*), 4.34 (1H, s, C₄*H*), 4.18 (1H, br. s, C₅*H*_aH_b), 4.08 (1H, br. s, C₂H), 3.91 (2H, dd, *J* 31.0, 17.0 Hz, NCH₂C=CMe), 3.81 (3H, s, OMe), 3.24 (1H, d, *J* 11.6 Hz,

C₅H_a*H_b*), 3.17 − 3.06 (2H, m, C*H*₂Ar), 2.17 (3H, s, O*Ac*), 1.94 (3H, s, NCH₂C≡C*Me*); ¹³C NMR (CDCl₃, 126 MHz) δ 170.2 (1C, Q), 158.9 (1C, Q), 129.8 (2C, CH), 127.1 (1C, Q), 114.5 (2C, CH), 87.8 (1C, Q), 78.0 (1C, CH), 71.7 (1C, CH), 66.7 (1C, Q), 65.7 (1C, CH), 58.0 (1C, CH₂), 55.3 (1C, CH₃), 42.5 (1C, CH₂), 29.4 (1C, CH₂), 20.6 (1C, CH₃), 3.6 (1C, CH₃); *m*/*z* (ESI) 318.2 ([M + H]⁺, 100%); **HRMS** (ESI) calcd. for C₁₈H₂₄O₄N [M + H]⁺ 318.1700, found 318.1705.

N-2,4-Pentadiyn-1-yl anisomycin, 2e



Purification (Method A); pale yellow oil (18.8 mg, 74%). **R**_t (Method B) = 24.2 min; **IR** (neat, cm⁻¹) 3271 (OH), 2236 (C=C), 1751 (C=O), 1670 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹**H NMR** (CDCl₃, 500 MHz) δ 7.15 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 6.89 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 4.90 – 4.85 (1H, m, C₃*H*), 4.39 – 4.35 (1H, m, C₄*H*), 4.12 (1H, dd, *J* 12.1, 5.7 Hz, C₅*H*_aH_b), 4.00 – 3.94 (1H, m, C₂*H*), 3.97 (2H, dd, *J* 59.7, 18.0 Hz, NC*H*₂C=C-), 3.83 (3H, s, OMe),

3.17 – 3.06 (3H, m, C₅H_a*H_b* + C*H*₂Ar), 2.29 (1H, s, -C=C*H*), 2.20 (3H, s, O*Ac*); ¹³C NMR (CDCl₃, 126 MHz) δ 170.2 (1C, Q), 158.9 (1C, Q), 129.8 (2C, CH), 127.1 (1C, Q), 114.6 (2C, CH), 78.3 (1C, CH), 74.1 (1C, Q), 72.0 (1C, CH), 69.6 (1C, Q), 66.5 (1C, Q), 66.3 (1C, CH), 64.9 (1C, Q), 58.6 (1C, CH₂), 55.3 (1C, CH₃), 42.6 (1C, CH₂), 29.9 (1C, CH₂), 20.6 (1C, CH₃); *m*/*z* (ESI) 328.2 ([M + H]⁺, 100%), 179.0 (7); **HRMS** (EI) calcd. for C₁₉H₂₂O₄N [M + H]⁺ 328.1543, found 328.1544.

N-[6-²H₃]-2,4-Hexadiyn-1-yl anisomycin, 2f



Purification (Method A); colourless oil (20.4 mg, 77%). **R**_t (Method C) = 21.1 min; **IR** (neat, cm⁻¹) 3289 (OH), 2264 (C=C), 1751 (C=O), 1670 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹**H NMR** (CDCl₃, 500 MHz) δ 7.15 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 6.89 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 4.90 (1H, d, *J* 3.7 Hz, C₃*H*), 4.41 – 4.38 (1H, m, C₄*H*), 4.23 (1H, dd, *J* 12.5, 5.6 Hz, C₅*H*_aH_b), 4.12 – 4.07 (1H, m, C₂*H*) 4.04 (2H, dd, *J* 51.5, 17.8 Hz, NC*H*₂C≡C-), 3.83 (3H, s, O*Me*), 3.24 (1H, d, *J* 12.5 Hz, C₅H_aH_b), 3.19 – 3.09 (2H, m, C*H*₂Ar), 2.20 (3H, s, O*Ac*); ¹³C **NMR** (CDCl₃, 126 MHz) δ 170.2 (1C, Q),

159.0 (1C, Q), 129.8 (2C, CH), 126.8 (1C, Q), 114.6 (2C, CH), 79.1 (1C, Q), 78.1 (1C, CH), 75.8 (1C, Q), 71.9 (1C, CH), 66.0 (1C, CH), 63.0 (1C, Q), 62.0 (1C, Q), 58.2 (1C, CH₂), 55.3 (1C, CH₃), 42.6 (1C, CH₂), 29.5 (1C, CH₂), 20.5 (1C, CH₃) 3.73 (1C, m, CD₃); m/z (ESI) 345.2 ([M + H]⁺, 100%), 179.0 (6); **HRMS** (ESI) calcd. for C₂₀H₂₁D₃O₄N [M + H]⁺ 345.1888, found 345.1888.

N-4-(1,3-Butadiyn-1-yl)benzyl anisomycin, 2g



Purification (Method A); light brown oil (28.5 mg, 95%). **R**_t (Method B) = 28.2 min; **IR** (neat, cm⁻¹) 3273 (OH), 1751 (C=O), 1670 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹H NMR (CDCl₃, 500 MHz) δ 7.58 (2H, d, *J* 8.2 Hz, 2 × Ar*H*), 7.49 (2H, d, *J* 8.2 Hz, 2 × Ar*H*), 6.99 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 6.84 (2H, d, *J* 8.6 Hz, 2 × Ar*H*), 4.93 (1H, d, *J* 3.2 Hz, C₃*H*), 4.32 – 4.23 (3H, m, C₄*H* + NC*H*₂Ar), 4.03 – 3.89 (2H, m, C₂*H* + C₅*H*_a*H*_b), 3.79 (3H, s, O*Me*), 3.15 – 3.08 (2H, m, C₅H_a*H*_b + C*H*_XH_YAr), 2.91 – 2.84 (1H, m, CH_X*H*_YAr), 2.55 (1H, s, -C=C*H*), 2.16 (3H, s, O*Ac*); ¹³C NMR (CDCl₃, 126 MHz) δ 169.9 (1C, Q), 158.9 (1C, Q), 133.5 (2C, CH), 130.9 (2C, CH), 129.7

(2C, CH), 127.2 (1C, Q), 123.0 (1C, Q), 117.9 (1C, Q), 114.5 (2C, CH), 77.8 (1C, Q), 77.2 (1C, CH), 75.3 (1C, Q), 74.0 (1C, Q), 72.3 (1C, CH), 72.3 (1C, CH), 67.8 (1C, CH), 59.9 (1C, CH₂), 58.8 (1C, CH₂), 55.3 (1C, CH₃), 30.4 (1C, CH₂), 20.6 (1C, CH₃); *m/z* (ESI) 404.2 ($[M + H]^+$, 100%), 179.0 (17); **HRMS** (ESI) calcd. for C₂₅H₂₆O₄N $[M + H]^+$ 404.1856, found 404.1869. * Obscured by solvent peak – observed in HSQC.

N-5-Phenyl-2,4-pentadiyn-1-yl anisomycin, 2h (PhDY-ANS)



Purification (isocratic 60:40 H₂O: MeCN 0.1% TFA), light brown oil (21.0 mg, 73%). **R**_t (Method C) = 31.3 min; **IR** (neat, cm⁻¹) 3284 (OH), 2253 (C=C), 1751 (C=O), 1672 (C=O, TFA), 1612 (C=C), 1585 (C=C), 1514 (C=C); ¹H NMR (CDCl₃, 500 MHz) δ 7.58 – 7.55 (2H, m, $2 \times \text{Ar}H$), 7.47 – 7.43 (1H, m, ArH), 7.41 – 7.34 (2H, m, $2 \times \text{Ar}H$), 7.18 (2H, d, *J* 8.6 Hz, $2 \times \text{Ar}H$), 6.90 (2H, d, *J* 8.6 Hz, $2 \times \text{Ar}H$), 4.90 (1H, d, *J* 3.8 Hz, C₃H), 4.41 (1H, br s, C₄H), 4.23 – 4.18 (1H, m, C₅H_aH_b), 4.15 – 4.02 (3H, m, C₂H + NCH₂C=C-), 3.83 (3H, s, OMe), 3.25 – 3.22 (1H, m, C₅H_aH_b), 3.19 – 3.10 (2H, m, CH₂Ar), 2.21 (3H, s, OAc); ¹³C NMR (CDCl₃, 126 MHz) δ 170.3 (1C, Q), 159.0 (1C, Q),

132.9 (2C, CH), 130.1 (1C, CH), 129.8 (2C, CH), 128.6 (2C, CH), 127.1 (1C, Q), 120.5 (1C, Q), 114.6 (2C, CH), 79.5 (1C, Q), 78.6 (1C, CH),* 74.9 (1C, Q), 72.3 (1C, CH), 72.3 (1C, Q), 65.7 (1C, CH), 58.1 (1C, CH₂), 55.3 (1C, CH₃), 42.5 (1C, CH₂), 29.7 (1C, CH₂), 20.6 (1C, CH₃); *m/z* (ESI) 404.2 ($[M + H]^+$, 100%), 324.9 (4), 246.9 (5), 179.0 (9); **HRMS** (ESI) calcd. for C₂₅H₂₆O₄N [M + H]⁺ 404.1856, found 404.1861. *Q peak expected at 77.2 ppm obscured by solvent peak.

N-4-(4-Phenyl-1,3-butadiyn-1-yl)benzyl anisomycin, 2i (BADY-ANS)



Purification (isocratic 50:50 H₂O:MeCN 0.1% TFA), pale yellow oil (28.0 mg, 81%). **R**_t (Method C) = 29.9 min; **IR** (neat, cm⁻¹) 3284 (OH), 2218 (C=C), 1751 (C=O), 1670 (C=O, TFA), 1613 (C=C), 1585 (C=C), 1514 (C=C); ¹H NMR (CDCl₃, 500 MHz) δ 7.61 – 7.54 (4H, m, 4 × ArH), 7.50 (2H, d, *J* 7.9 Hz, 2 × ArH), 7.43 – 7.34 (3H, m, 3 × ArH), 6.99 (2H, d, *J* 8.6 Hz, 2 × ArH), 6.84 (2H, d, *J* 8.6 Hz, 2 × ArH), 4.95 (1H, s, C₃H), 4.32 – 4.25 (3H, m, C₄H + NCH₂Ar), 4.05 – 3.87 (2H, m, C₂H + C₅H_aH_b), 3.79 (3H, s, OMe), 3.17 – 3.06 (2H, m, CH_XH_YAr + C₅H_aH_b), 2.91 – 2.81 (1H, m, CH_XH_YAr), 2.15 (3H, s, OAc); ¹³C NMR (CDCl₃, 126 MHz) δ 169.9 (1C, Q), 158.9 (1C, Q), 133.2 (2C, CH), 132.6 (2C, CH), 131.0 (2C, CH), 129.7 (2C, CH), 129.5 (1C, CH), 128.5 (2C, CH), 127.3 (1C, Q), 123.8

(1C, Q), 121.5 (2C, Q), 114.5 (2C, CH), 82.6 (1C, Q), 80.2 (1C, Q), 77.2 (1C, CH),* 75.0 (1C, Q), 73.6 (1C, Q), 72.3 (1C, CH), 69.2 (1C, CH), 60.2 (1C, CH₂), 58.9 (1C, CH₂), 55.3 (1C, CH₃), 30.5 (1C, CH₂), 20.6 (1C, CH₃); *m/z* (ESI) 480.2 ($[M + H]^+$, 100%), 324.9 (4), 246.9 (6); **HRMS** (ESI) calcd. for C₃₁H₃₀O₄N $[M + H]^+$ 480.2169, found 480.2179. *Obscured by solvent peak – observed in HSQC.

References

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-S21-







¹H NMR CD₃OD, 500 MHz



— 4.21



110 100 f1 (ppm) . 170



f1 (ppm) , 70







100 90 f1 (ppm) 0 200 190 . 180 170 160 . 150 140 . 130 . 120 110 80 . 70 60 50 . 40 . 30 20 10









-S33-











70°C	5	
N·TFA		- 10
SS'02 MeO ACO		20
¹³ C NMR ^{0+'62 —} CDCl ₃ , 126 MHz		30 -
67.24		- 4
	KARAA MAKANGA KANA M	20
62°55 — 00°85 —		- 09
12 59 ~ 12'99 ~ 82'12 —		70
Þ0 82 —		- 80
08.78 —		- 06
	e de vergen andere de la companya d	100 100 11 (ppm)
70'LTT	u na	110
		120
92'5ZT — – — — — — — — — — — — — — — — — — —		130
		140
	o-baylo retrokandi Verene	150
ī6'8Sī —		160
∠τ·0ζτ —		170
		180
		190
	and the second	200



		$\lceil \circ \rceil$
		- 10
09'0Z MeO AcO		50
^{58'6Z} — ¹³ C NMR CDCl ₃ , 126 MHz		30 -
65'ZÞ —		- 4
— 22°35		- 20
- 28'64		- 09
		- 70
26.87 ~~		- 8
		- 6
		f1 (ppm)
		110
09 #11		120
28.271 —		130
	the state of the	140
	o'hord' a shekek ka	150
₩ ^{128.} 94		160
81.071 —		170
		180
		190
		200

















Optimised geometries for compounds $\mathbf{3b} - \mathbf{i}$ and EdU 1 reported in standard orientation.

Compound **3b**

Cent	ter	Atomic	Atomic	Coordinates (Angstroms)		
Num	ber	Number	Туре	х	Y	Z
	1	7	0	-0.734458	0.019048	-0.315890
	2	6	0	0.313091	-0.749856	0.346684
	3	6	0	1.659521	-0.258487	0.012313
	4	7	0	2.725570	0.128507	-0.232204
	5	1	0	0.221497	-0.755436	1.451740
	6	1	0	0.248247	-1.793650	0.018611
	7	6	0	-0.767810	1.407920	0.133985
	8	6	0	-2.028421	-0.633888	-0.155670
	9	1	0	-2.358687	-0.702430	0.898480
	10	1	0	-1.989625	-1.646617	-0.569082
	11	1	0	-2.784963	-0.071902	-0.710056
	12	1	0	-1.525026	1.951952	-0.437096
	13	1	0	-1.007331	1.508945	1.209557
	14	1	0	0.199817	1.882118	-0.049365

Compound 3c

Cente	er	Atomic	Atomic	Coor	dinates (Ang	gstroms)
Numbe	er	Number	Туре	х	Y	Z
	1	7	0	-0.767768	0.026340	-0.327453
	2	6	0	0.277283	-0.765840	0.326502
	3	6	0	1.632731	-0.291730	0.032283
	4	6	0	2.758672	0.085260	-0.186153
	5	1	0	3.750362	0.415941	-0.391754
	6	1	0	0.144504	-0.799805	1.428299
	7	1	0	0.183575	-1.801230	-0.024993
	8	6	0	-0.781121	1.408745	0.136043
	9	6	0	-2.068516	-0.605670	-0.157562
1	10	1	0	-2.388366	-0.678099	0.900767
1	11	1	0	-2.050200	-1.617216	-0.576164
1	12	1	0	-2.825977	-0.030151	-0.697916
1	13	1	0	-1.528160	1.973716	-0.429597
1	14	1	0	-1.022327	1.503315	1.213072
1	15	1	0	0.196675	1.864557	-0.036221

Compound 3d

Cent	ter Atomic Atomic Coordinates (Angstroms)					
Numk	ber	Number	Туре	x	Ŷ	Z
	1	7	0	-1.392028	0.05737	B -0.334655
	2	6	0	-0.447850	-0.83708	3 0.344648
	3	6	0	0.955656	-0.46345	5 0.146229
	4	6	0	2.120160	-0.16843	0.006758
	5	6	0	3.525513	0.18315	B -0.175691
	6	1	0	-0.652795	-0.89753	3 1.434543
	7	1	0	-0.607190	-1.84858	1 -0.050995
	8	6	0	-1.320897	1.42100	5 0.174688
	9	6	0	-2.745833	-0.47115	9 -0.252143
	10	1	0	-3.126560	-0.54853	3 0.785792
	11	1	0	-2.785498	-1.46865	6 -0.702101
	12	1	0	-3.426196	0.17887	5 -0.810630
	13	1	0	-1.988051	2.06260	7 -0.409240
	14	1	0	-1.612802	1.503112	2 1.240510
	15	1	0	-0.301068	1.79858	4 0.070852
	16	1	0	3.657016	0.84837	9 -1.036357
	17	1	0	4.137220	-0.70878	1 -0.351827
	18	1	0	3.929622	0.69467	6 0.705101

Compound 3e

Cent	ter	Atomic	Atomic	C001	rdinates (An	gstroms)
Num	ber	Number	Туре	х	Y	Z
	1	7	0	1.743369	0.088101	-0.341381
	2	6	0	0.851697	-0.851133	0.344060
	3	6	0	-0.565912	-0.531043	0.175870
	4	6	0	-1.748950	-0.277082	0.060645
	5	6	0	-3.081257	0.004226	-0.071134
	6	6	0	-4.262749	0.254405	-0.186994
	7	6	0	1.639722	1.439034	0.198637
	8	6	0	3.116762	-0.396923	-0.310218
	9	1	0	2.272599	2.112049	-0.387209
	10	1	0	0.607299	1.788793	0.123557
	11	1	0	1.952840	1.508386	1.258658
	12	1	0	3.175704	-1.382550	-0.783182
	13	1	0	3.530632	-0.481379	0.713691
	14	1	0	3.755788	0.287765	-0.875389
	15	1	0	1.070778	-0.919510	1.430810
	16	1	0	1.035101	-1.852197	-0.067659
	17	1	0	-5.300200	0.473039	-0.288809

Center	Atomic	Atomic	.c Coordinates (Angstroms)		
Number	Number	Туре	х	Y	Z
1	7	0	2.448883	0.130785	-0.355908
2	6	0	1.610169	-0.865899	0.317498
3	6	0	0.176990	-0.601446	0.191101
4	6	0	-1.019035	-0.395767	0.112814
5	6	0	-2.364893	-0.169503	0.021800
6	6	0	-3.561139	0.031825	-0.058714
7	6	0	2.294290	1.460356	0.222479
8	6	0	3.842372	-0.292795	-0.356710
9	1	0	2.885420	2.177552	-0.354801
10	1	0	1.245621	1.763349	0.174825
11	1	0	2.623024	1.515620	1.278831
12	1	0	3.937830	-1.262356	-0.856363
13	1	0	4.274909	-0.385997	0.658998
14	1	0	4.442802	0.434073	-0.911695
15	1	0	1.859959	-0.957681	1.396078
16	1	0	1.828900	-1.844584	-0.129685
17	6	0	-4.993769	0.271936	-0.154556
18	1	0	-5.543572	-0.317460	0.588139
19	1	0	-5.376746	0.001618	-1.145239
20	1	0	-5.230234	1.328138	0.017994

Compound 3f (note the CD₃ group in compound 3f was modelled as a CH₃ group).

Compound 3g

Center		Atomic	Atomic	Coor	Coordinates (Angstroms)	
Num	ber	Number	Туре	Х	Y	Z
	1	6	0	1.484532	0.153765	-0.128905
	2	6	0	2.895098	-0.005048	-0.036238
	3	6	0	4.104613	-0.141372	0.043617
	4	6	0	5.457705	-0.293958	0.132962
	5	6	0	6.661833	-0.429750	0.212668
	6	6	0	0.660936	-0.930802	-0.496260
	7	6	0	-0.716072	-0.770711	-0.585674
	8	6	0	-1.316815	0.466110	-0.309944
	9	6	0	-0.499488	1.539897	0.061943
	10	6	0	0.881639	1.395449	0.149696
	11	1	0	7.717864	-0.549104	0.282449
	12	6	0	-2.815067	0.647080	-0.463765
	13	7	0	-3.589291	-0.485779	0.039252
	14	6	0	-4.967655	-0.447127	-0.431861
	15	6	0	-3.530831	-0.587380	1.493502
	16	1	0	-3.991992	0.279904	2.004370
	17	1	0	-2.490991	-0.660805	1.821372
	18	1	0	-4.055172	-1.490887	1.819373
	19	1	0	-5.530120	0.435978	-0.071939
	20	1	0	-5.496614	-1.342109	-0.089781
	21	1	0	-4.987956	-0.438512	-1.526243
	22	1	0	-3.045112	0.748767	-1.532989
	23	1	0	-3.117984	1.603182	0.007751
	24	1	0	-1.350203	-1.608438	-0.857569
	25	1	0	-0.950776	2.502880	0.287326
	26	1	0	1.504697	2.234830	0.440275
	27	1	0	1.116821	-1.892151	-0.709603

Compound 3h

Center		Atomic	Atomic	Coor	rdinates (Ang	gstroms)
Num	ber	Number	Type	Х	Y	Z
	1	7	0	-4.646655	0.153748	-0.404009
	2	6	0	-3.878351	-0.820234	0.377146
	3	6	0	-2.431145	-0.634692	0.275514
	4	6	0	-1.223538	-0.494856	0.217317
	5	6	0	0.130318	-0.344823	0.149261
	6	6	0	1.341520	-0.210944	0.088250
	7	1	0	-4.129916	-1.822433	0.005927
	8	1	0	-4.162651	-0.809512	1.451034
	9	6	0	-4.455154	1.516943	0.077383
	10	6	0	-6.056790	-0.210923	-0.428709
	11	1	0	-6.603158	0.495813	-1.060153
	12	1	0	-6.176816	-1.210831	-0.858182
	13	1	0	-6.530316	-0.208079	0.572801
	14	6	0	2.754731	-0.058537	0.016411
	15	6	0	3.340651	1.222737	0.066347
	16	6	0	4.723058	1.365559	-0.006038
	17	6	0	5.542769	0.240909	-0.127020
	18	6	0	4.971872	-1.033098	-0.176428
	19	6	0	3.590583	-1.187175	-0.106561
	20	1	0	3.142711	-2.174575	-0.145888
	21	1	0	5.605576	-1.909968	-0.270543
	22	1	0	6.621018	0.356682	-0.182624
	23	1	0	5.162793	2.357756	0.032655
	24	1	0	2,700076	2.093286	0.160917
	25	1	0	-3.393172	1.772585	0.049610
	26	- 1	0	-4.821051	1.667160	1.111934
	27	1	0	-4.991663	2.210682	-0.576665
		-				

Compound 3i

Center Atomic Atomic Coordinat			dinates (And	tes (Angstroms)		
Num	ber	Number	Type	х	Y	Z
	1	6	0	-1.315577	0.256791	-0.253756
	2	6	0	0.102430	0.194002	-0.184730
	3	6	0	1.321562	0.136091	-0.129048
	4	6	0	2.678051	0.070319	-0.068597
	5	6	0	3.897006	0.010016	-0.015175
	6	6	0	-2.084959	-0.922580	-0.347723
	7	6	0	-3.470461	-0.857154	-0.419874
	8	6	0	-4.135809	0.377372	-0.397025
	9	6	0	-3.373221	1.546765	-0.294047
	10	6	0	-1.984414	1.496473	-0.226808
	11	6	0	5.315475	-0.063999	0.045713
	12	6	0	6.083557	1.090020	0.305092
	13	6	0	7.471530	1.010723	0.363626
	14	6	0	8.117246	-0.212249	0.165926
	15	6	0	7.365588	-1.361306	-0.091903
	16	6	0	5.977147	-1.293657	-0.152558
	17	6	0	-5.644785	0.442378	-0.535417
	18	7	0	-6.341919	-0.583998	0.237408
	19	6	0	-7.730790	-0.721703	-0.180849
	20	6	0	-6.243209	-0.349965	1.673983
	21	1	0	-6.743122	0.585148	1.992799
	22	1	0	-5.193237	-0.292310	1.971200
	23	1	0	-6.704863	-1.182158	2.214256
	24	1	0	-8.333520	0.191533	-0.011332
	25	1	0	-8.200973	-1.539754	0.373797
	26	1	0	-7.777030	-0.964492	-1.247186
	27	1	0	-5.905045	0.286370	-1.591170
	28	1	0	-5.991810	1.464019	-0.281928
	29	1	0	-4.060407	-1.766112	-0.481429
	30	1	0	-3.874374	2.511101	-0.265512
	31	1	0	-1.404625	2.410082	-0.146821
	32	1	0	-1.579384	-1.882607	-0.365200
	33	1	0	5.389318	-2.183329	-0.352625
	34	1	0	5.577856	2.037610	0.458229
	35	1	0	8.052149	1.906263	0.564220
	36	1	0	9.200600	-0.269551	0.212369
	37	1	0	7.863699	-2.313849	-0.246507

EdU	1
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Center Number		Atomic	Atomic	Coordinates (Angstroms)		gstroms)
		Number	Туре	х	Y	Z
	1	6	0	-3.465456	-0.156665	0.141031
	2	6	0	-2.495357	0.949151	0.069546
	3	6	0	-1.182516	0.663521	-0.174319
	4	7	0	-0.708477	-0.608621	-0.349559
	5	6	0	-1.547545	-1.725838	-0.300667
	6	7	0	-2.872801	-1.418918	-0.067171
	7	8	0	-4.660692	-0.064356	0.349249
	8	8	0	-1.130455	-2.864252	-0.449351
	9	6	0	-2.944260	2.286255	0.242697
	10	6	0	0.730596	-0.893789	-0.597338
	11	8	0	1.377160	0.293538	-1.031385
	12	6	0	2.468053	0.606272	-0.148842
	13	6	0	2.851185	-0.732291	0.502545
	14	6	0	1.478651	-1.388847	0.651891
	15	6	0	3.584842	1.247264	-0.956472
	16	8	0	4.671367	1.409807	-0.043620
	17	8	0	3.498049	-0.593251	1.745262
	18	1	0	3.258898	2.209279	-1.375194
	19	1	0	3.848241	0.581258	-1.791028
	20	1	0	1.006679	-1.000883	1.560312
	21	1	0	1.500216	-2.477110	0.709650
	22	1	0	-0.438225	1.442703	-0.265904
	23	1	0	-3.499896	-2.213420	-0.024743
	24	1	0	0.746835	-1.631428	-1.402974
	25	1	0	3.468511	-1.306020	-0.209650
	26	1	0	2.142207	1.295376	0.644590
	27	6	0	-3.319951	3.426318	0.391604
	28	1	0	-3.665182	4.425265	0.524165
	29	1	0	4.279920	-0.044872	1.576336
	30	1	0	5.427849	1.772633	-0.519746