

Figure S1: Q-PCR analysis of transcript depletion in transgenic RNAi manipulations

A) - E) Quantitative PCR (Q-PCR) from retinal tissue of genotypes indicated. Detected transcript, normalized to the house keeping transcript RP49, is indicated on y-axis. n=75retinae per sample(** - $p < 0.01$, two-tailed unpaired t-test).The experiment has been repeated two times, one of the trials is shown here.

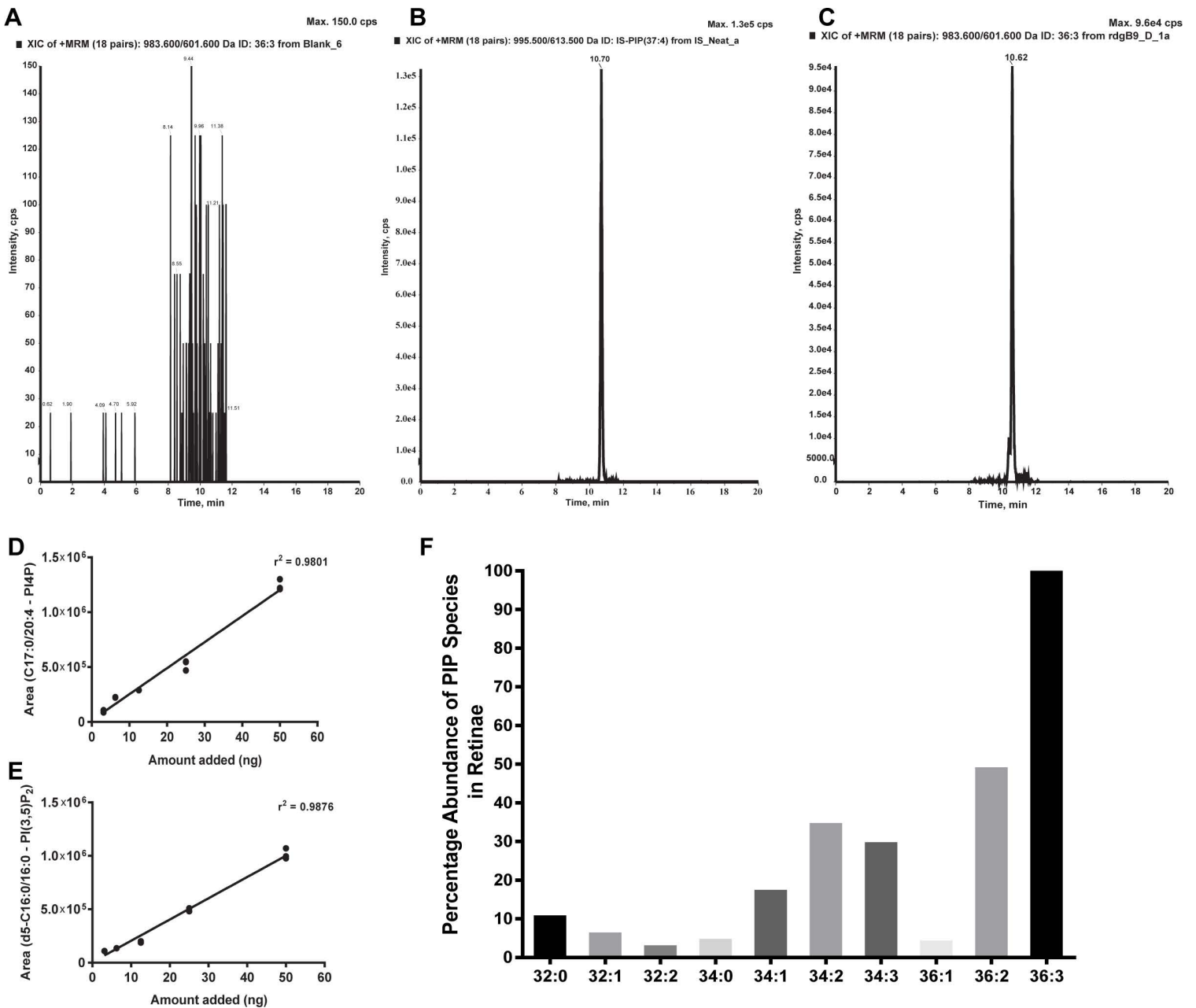


Figure S2: LC-MS analysis of PIP and PIP₂ levels in retinal lipid extracts

A) - C) Representative chromatograms of blank, synthetic internal standard (C_{17:0/20:4}-PI₄P) and retinal lipid extract (36:3-PIP) respectively. The 36:3 PIP elutes at 10.62 mins, which is not observed in the chromatograms from blank injections. X-axis is elution time; y-axis is signal in counts per second.

D) and E) Response linearity curves for extracted internal standards C_{17:0/20:4}-PI₄P and d₅-C_{16:0/16:0}-PI(3,5)P₂, respectively. X-axis represents amount added before extraction; y-axis represents area under curve for the peak being quantified.

F) Representation of the acyl chain length and desaturation of PIP species extracted and detected from *Drosophila* retinae. X-axis represents the species detected from wild type *Drosophila* retinae in the format a:b where a is the combined acyl chain length of *sn-1* and *sn-2* and b is the number of double bonds in both acyl chains. Y-axis represents the relative abundance of each species. The PIP species 36:3 is most abundant with 36:2, the next most abundant species, being the about 50% of the levels of 36:3.

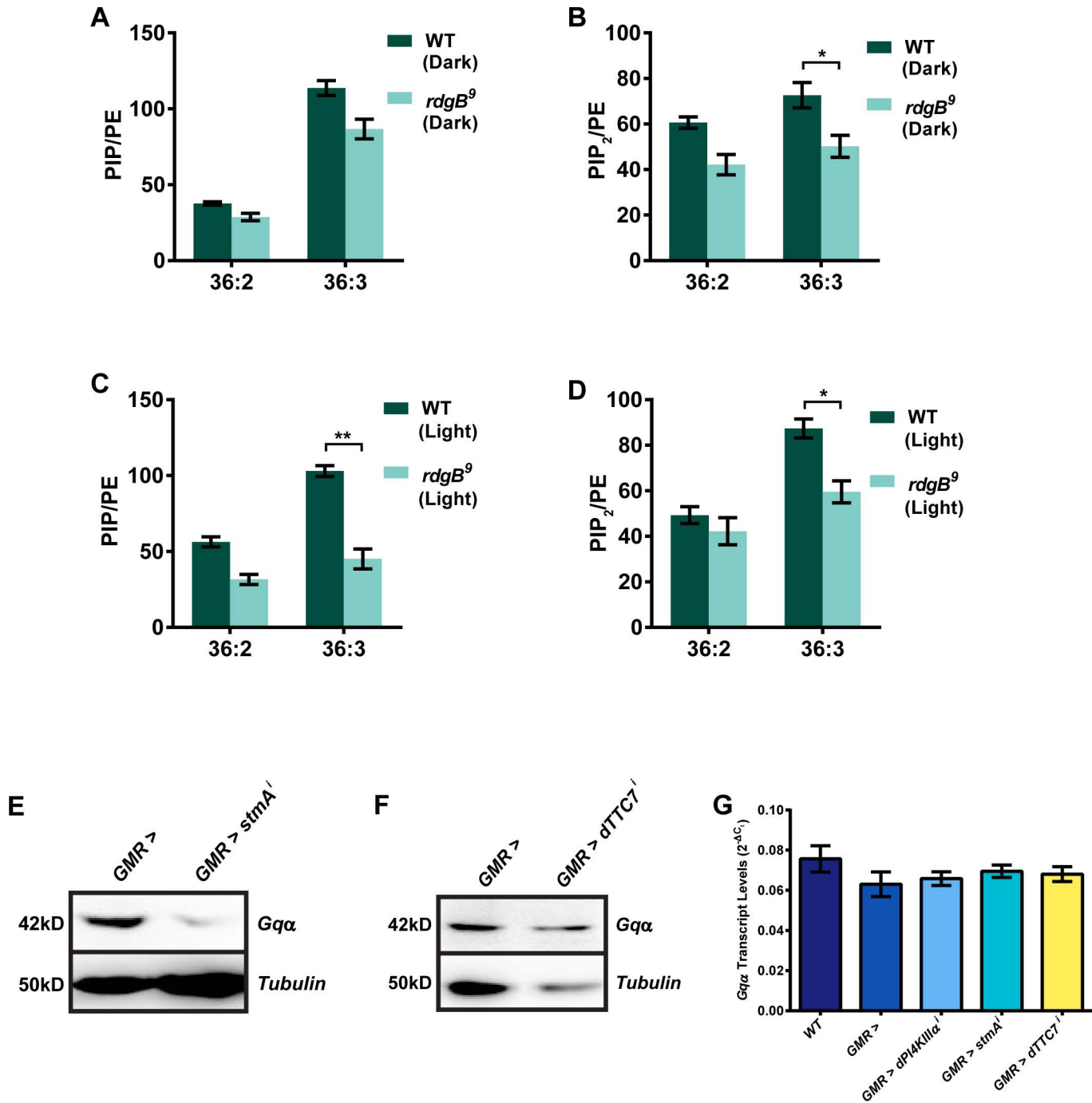


Figure S3: LC-MS measurements of lipids from *rdgB*⁹ flies

A) and B) LC-MS measurement of total PIP and PIP₂ levels, respectively, from retinæ of one day old WT and *rdgB*⁹ flies. Flies were reared and processed completely in dark. Levels for the two most abundant molecular species of PIP and PIP₂ [36:2 and 36:3] are shown. Y-axis represents PIP and PIP₂ levels normalized to phosphatidylethanolamine (PE). Values are mean ± s.e.m., n=25 retinæ per sample (* - p < 0.05, ANOVA followed by Tukey's multiple comparison test). The experiment has been repeated three times, one of the trials is shown here.

- C) and D) LC-MS measurement of total PIP and PIP₂ levels, respectively, from retinae of one day old WT and *rdgB* flies. Flies were reared in dark and exposed to one minute of bright illumination before processing. Y-axis represents PIP and PIP₂ levels normalized to phosphatidylethanolamine (PE). Levels for the two most abundant molecular species of PIP and PIP₂ [36:2 and 36:3] are shown. Values are mean \pm s.e.m., n=25 retinae per sample (* - p < 0.05, ** - p < 0.01, ANOVA followed by Tukey's multiple comparison test). The experiment has been repeated three times, one of the trials is shown here.
- E) and F) Western blot of head extracts made from flies of the indicated genotypes. Tubulin is used as a loading control, genotypes are indicated above.
- G) Quantitative PCR (Q-PCR) from retinal tissue of genotypes indicated. Detected transcript normalized to the house keeping transcript RP49 is indicated on y-axis. n=75 retinae per sample.

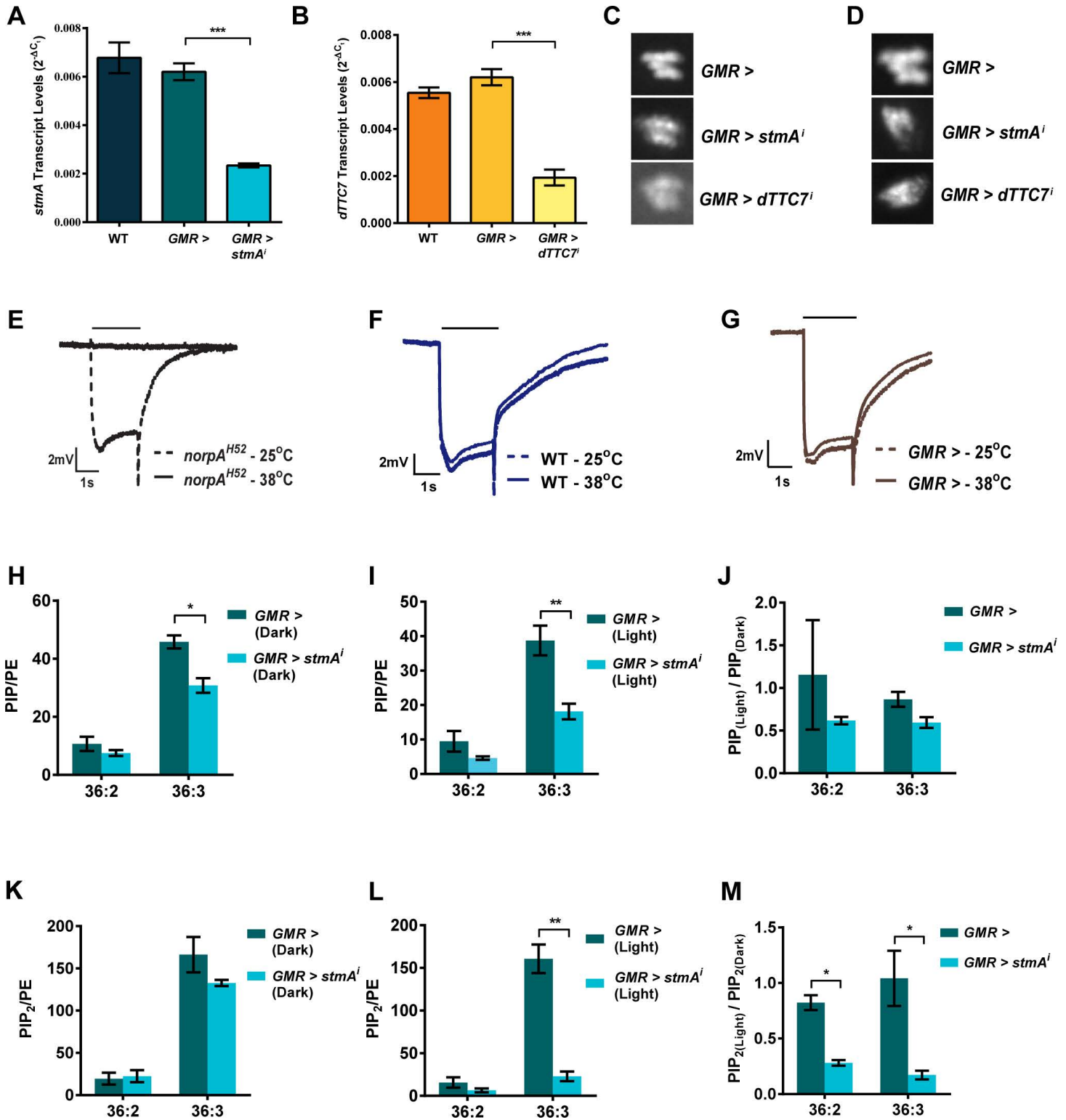


Figure S4: *stmA* is required for maintenance of PIP and PIP₂ levels in photoreceptors

A) - B) Quantitative PCR (Q-PCR) from retinal tissue of genotypes indicated. Detected transcript, normalized to the house keeping transcript RP49, is indicated on y-axis. n=75 retinae per sample (***) - $p < 0.001$, two-tailed unpaired t-test).

- C) Representative images of fluorescent deep pseudopupil from one day old flies expressing the P4M::GFP probe, genotype as indicated.
- D) Representative images of fluorescent deep pseudopupil from flies expressing the PLC δ PH::GFP probe, genotype as indicated.
- E) – G) Representative electroretinogram (ERG) trace of one day old flies, genotype as indicated. X-axis is time in (s); y-axis is amplitude in mV. Black bar above traces indicates duration of the light stimulus. The experiment has been repeated two times, one of the trials is shown here.
- H) and K) LC-MS measurement of total PIP and PIP₂ levels, respectively, from retinæ of one day old flies, genotypes as indicated. Flies were reared and processed completely in dark. Levels for the two most abundant molecular species of PIP and PIP₂ [36:2 and 36:3] are shown. Y-axis represents PIP and PIP₂ levels normalized to phosphatidylethanolamine (PE). Values are mean \pm s.e.m., n=25 retinæ per sample (* - p < 0.05, ANOVA followed by Tukey's multiple comparison test).
- I) and L) LC-MS measurement of total PIP and PIP₂ levels, respectively, from retinæ of one day old flies, genotypes as indicated. Flies were reared in dark and exposed to one minute of bright illumination before processing. Y-axis represents PIP and PIP₂ levels normalized to phosphatidylethanolamine (PE). Levels for the two most abundant molecular species of PIP and PIP₂ [36:2 and 36:3] are shown. Values are mean \pm s.e.m., n=25 retinæ per sample (** - p < 0.01, ANOVA followed by Tukey's multiple comparison test).
- J) and M) LC-MS measurement of total PIP and PIP₂ levels, respectively, from retinæ of one day old flies, genotypes as indicated. The y-axis represents a ratio of lipid levels, [PIP_(light)/PIP_(dark)] and [PIP_{2(light)}/PIP_{2(dark)}]. Ratios for the two most abundant molecular species of PIP and PIP₂ [36:2 and 36:3] are shown. A reduction in the ratio indicates a drop in the levels of PIP and PIP₂. Values are mean \pm s.e.m. (* - p < 0.05, ** - p < 0.01, ANOVA followed by Tukey's multiple comparison test).

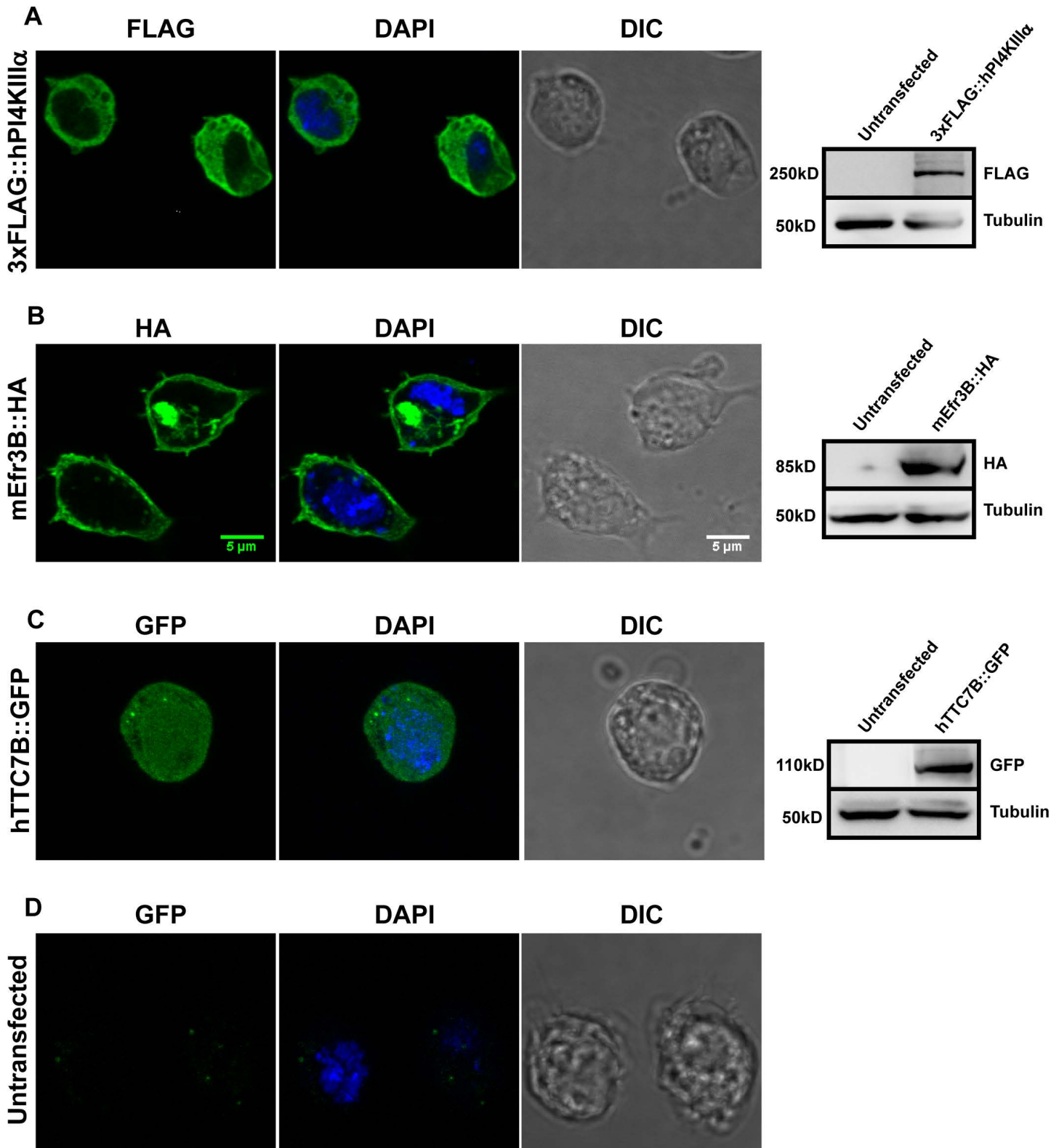


Figure S5: Localization of mammalian PI4KIII α , Efr3 and TTC7 in S2R+ cells

A) Confocal images of S2R+ cells transfected with pUAST-3xFLAG::hPI4KIII α , followed by staining with DAPI and α -FLAG. Blue represents DAPI, which marks the nucleus, and green represents FLAG. Western blot indicates the presence of the FLAG-tagged protein in transfected cells. Scale bar - 5 μ m. The experiment has been repeated two times, one of the trials is shown here.

- B) Confocal images of S2R+ cells transfected with pUAST-mEfr3B::HA followed by staining with DAPI and α -HA. Blue represents DAPI, which marks the nucleus, and green represents HA. Western blot indicates the presence of the HA-tagged protein in transfected cells. Scale bar - 5 μ m. The experiment has been repeated two times, one of the trials is shown here.
- C) Confocal images of S2R+ cells transfected with pUAST-hTTC7B::GFP, followed by staining with DAPI. Blue represents DAPI, which marks the nucleus, and green represents GFP. Western blot indicates the presence of the GFP-tagged protein in transfected cells. Scale bar - 5 μ m. The experiment has been repeated two times, one of the trials is shown here.
- D) Confocal images of untransfected S2R+ cells followed by staining with DAPI. Blue represents DAPI, which marks the nucleus, and green represents GFP. Scale bar - 5 μ m.

Table S1: Conditions for LC separation Solvent A: Water + 0.1% Formic Acid; Solvent B: Acetonitrile. Gradient:

Time (min)	Flow Rate	%A	%B
0	0.1	55	45
5	0.1	55	45
10	0.1	0	100
15	0.1	0	100
16	0.1	55	45
20	0.1	55	45

Table S2: MRM details

MRMs:

Polarity: Positive; Ion Source: Turbo Spray Ion Drive; Resolution Q1: Unit; Resolution Q3: Unit

Q1 Mass (Da)	Q3 Mass (Da)	Dwell Time (ms)	Species
809.4	535.4	60	IS – PI (17:0/14:1)
823.5	549.5	60	PI - 32:2
849.5	575.5	60	PI - 34:2
851.6	577.6	60	PI - 34:3
877.5	603.6	60	PI - 36:2
879.6	605.6	60	PI - 36:3
938.5	556.4	60	IS - PI4P (17:0/20:4)
957.5	575.5	60	PIP - 34:2
959.5	577.5	60	PIP - 34:3
985.6	603.6	60	PIP - 36:2
987.6	605.6	60	PIP - 36:3
1046.5	556.4	60	IS – d5-PI(3,5)P ₂ (16:0/16:0)
1065.5	575.5	60	PIP ₂ - 34:2
1067.5	577.5	60	PIP ₂ - 34:3
1093.6	603.6	60	PIP ₂ - 36:2
1095.6	605.6	60	PIP ₂ - 36:3
690.5	535.5	60	IS – PE (17:0/14:1)
754.4	601.4	60	PE - 34:2
752.5	603.5	60	PE - 34:3
732.5	577.5	60	PE - 36:2
730.4	575.4	60	PE - 36:3