

## Supplementary Materials for

### **Regulated nuclear accumulation of a histone methyltransferase times the onset of heterochromatin formation in *C. elegans* embryos**

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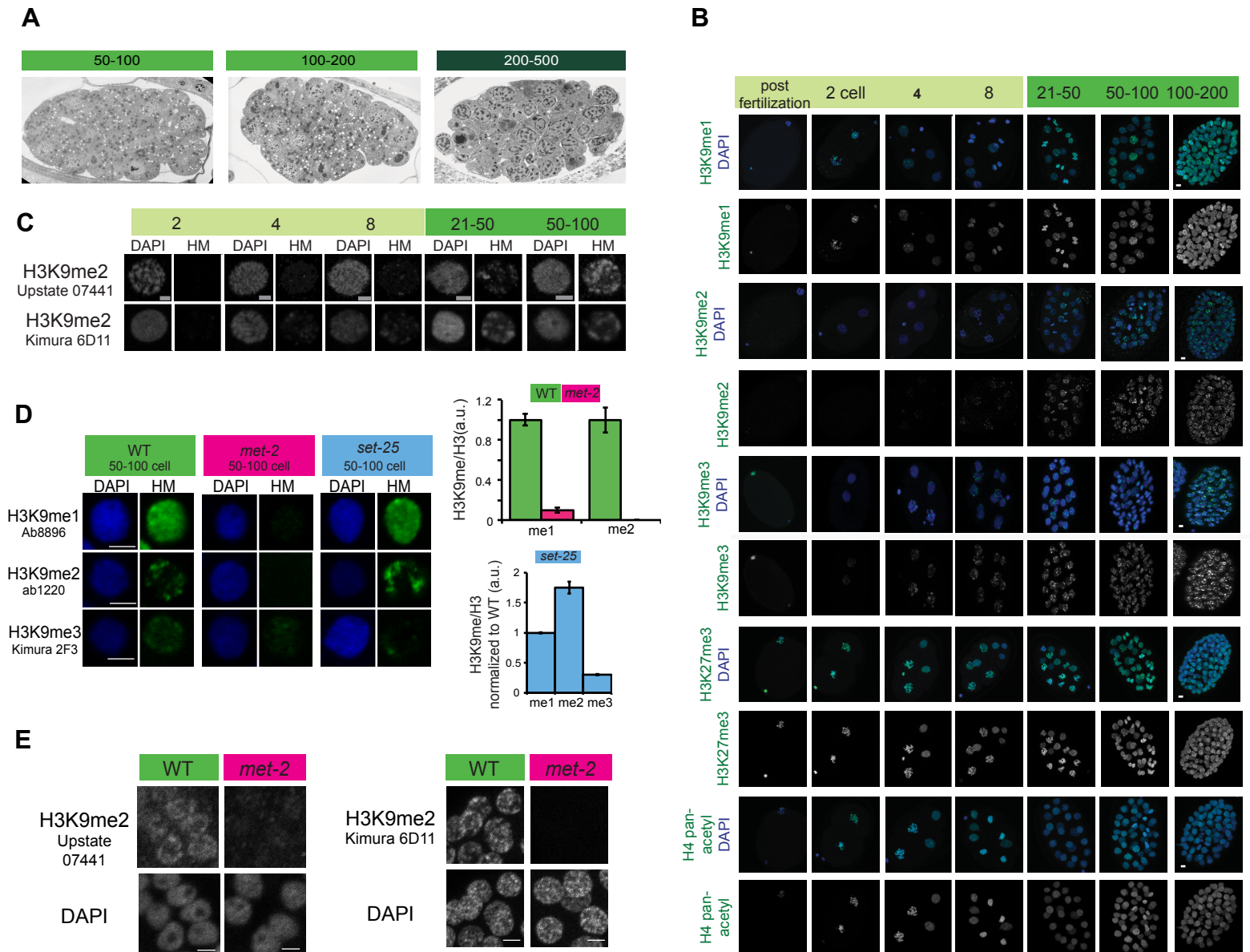
#### **The PDF file includes:**

- Fig. S1. Heterochromatin formation and establishment of H3K9me in WT embryos.
- Fig. S2. Control for TEM conditions in WT versus *met-2* mutants.
- Fig. S3. MET-2 localization with additional reagents and during cell cycle.
- Fig. S4. Specificity controls for MET-2 coimmunoprecipitation and PLA.
- Fig. S5. Further analysis of LIN-65 and dosage compensation for MET-2 protein.

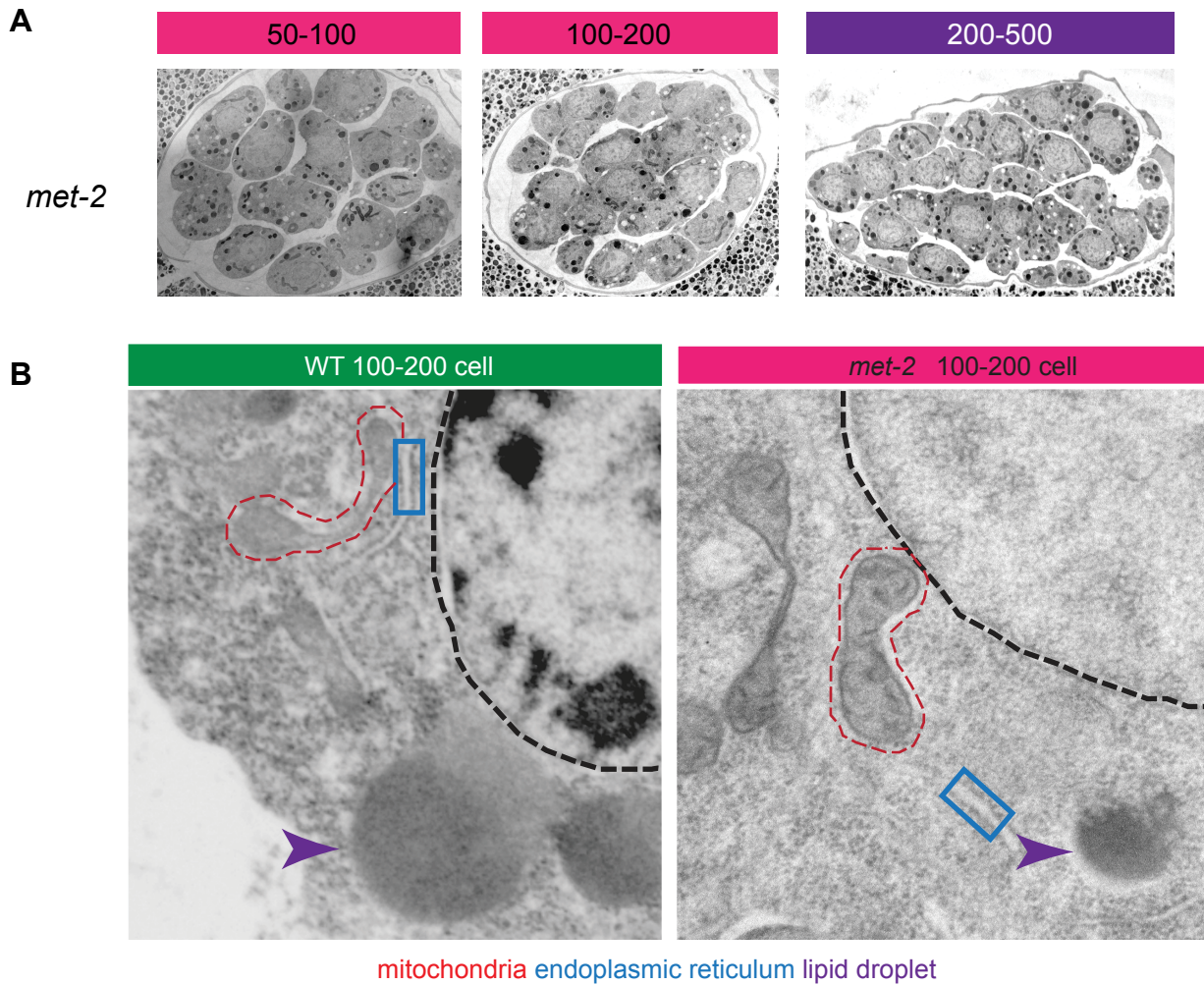
#### **Other Supplementary Material for this manuscript includes the following:**

(available at [advances.sciencemag.org/cgi/content/full/4/8/eaat6224/DC1](https://advances.sciencemag.org/cgi/content/full/4/8/eaat6224/DC1))

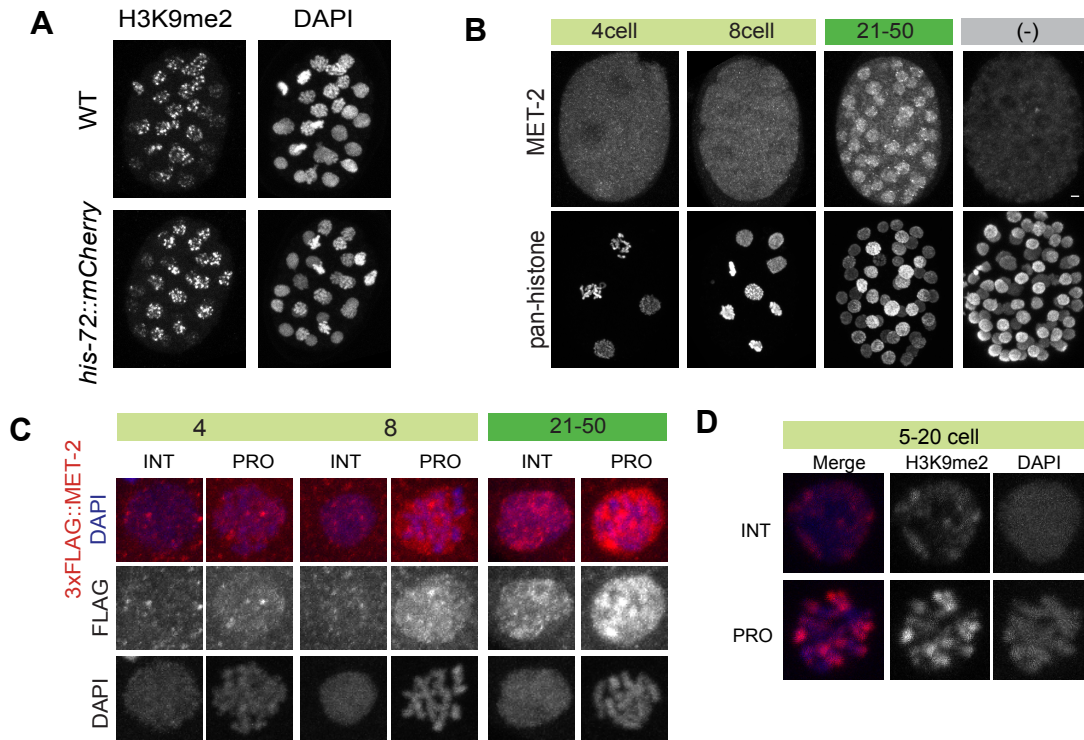
Table S1 (Microsoft Excel format). Spectral counts and sequence coverage for proteins identified in the GFP immunoprecipitation and mass spectrometry experiment. Strains that lack GFP (N2), and contain MET-2::GFP or ZEN-4::GFP were processed in parallel.



**Fig. S1. Heterochromatin formation and establishment of H3K9me in WT embryos.** (A) TEM single sections of whole embryos at designated stages. Each embryo is approximately 50 $\mu$ m long. (B) Whole embryos stained with H3K9me1/2/3, HK27me3 or H4 pan-acetylation at different stages of development (Scale bar 2  $\mu$ m). (C) Representative single nuclei at designated stages showing H3K9me2 staining with two additional H3K9me2 antibodies: Upstate 07-441 and Kimura 6D11. (D) Antibody specificity for H3K9me2(ab1222) and H3K9me3 (Kimura 2F3): representative single nuclei at designated stages showing H3K9me staining in wild-type (green) vs. *met-2* (magenta) and *set-25* (blue) mutant embryos, Histone Modification (HM, green), DAPI (blue). H3K9me/Histone levels normalized to wild-type. Error bars denote standard error of the mean. (E) Antibody specificity for H3K9me2 (Upstate 07-441 and Kimura 6D11).



**Fig. S2. Control for TEM conditions in WT versus *met-2* mutants.** (A) TEM single sections of whole *met-2* embryos. Note the electron dense droplets in the cytoplasm vs. the electron lucent nuclei. (B) Cytosolic components in WT vs. *met-2* mutants by TEM. Mitochondria (red circle), endoplasmic reticulum (blue box), lipid droplet (purple arrowhead). Note the similar appearance of the cytosol vs. the different morphologies of the nuclei for wild-type vs *met-2*.

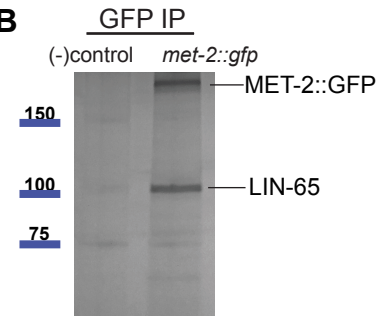


**Fig. S3. MET-2 localization with additional reagents and during cell cycle. (A)** H3K9me2 staining in wild-type vs. *his-72::mCherry* embryos showing that the mCherry tag doesn't interfere with H3K9me2. **(B)** Whole embryos stained with an antibody against endogenous MET-2 (raised against the first 17 amino acids of MET-2 protein) and pan-histone. (-) represents *met-2* mutants. Scale bar, 2  $\mu$ m. **(C)** Representative single nuclei showing Crispr reporter 3xFLAG::MET-2 during interphase (INT) and prophase (PRO, Scale bar, 2  $\mu$ m). **(D)** H3K9me2 levels in interphase (INT) and prophase (PRO) nuclei from the same embryo at the 15 cell stage.

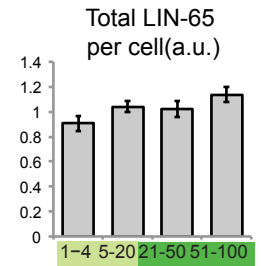
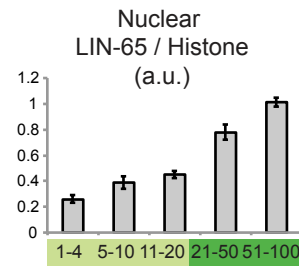
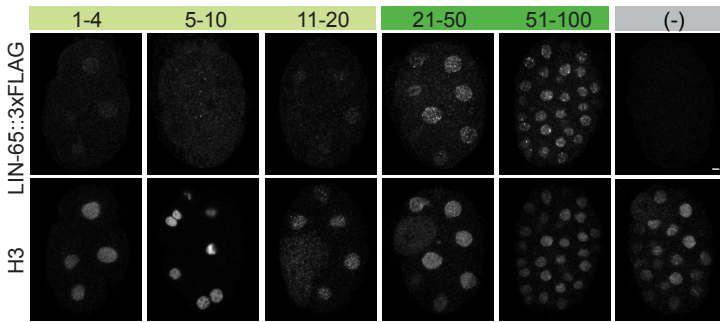
A

	MET-2::GFP		(-)control	GFP control
	Spectral counts	Coverage	Spectral counts	Spectral counts
<b>MET-2</b>	895	36%	0	7
<b>LIN-65</b>	171	21%	0	0
<b>B0336.5</b>	8	21%	0	0
<b>Histone H3</b>	291	7%	0	0

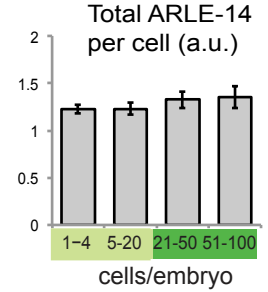
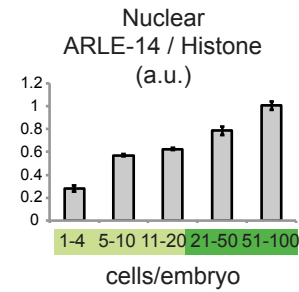
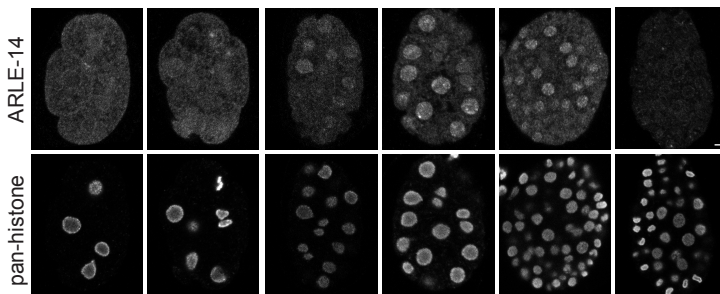
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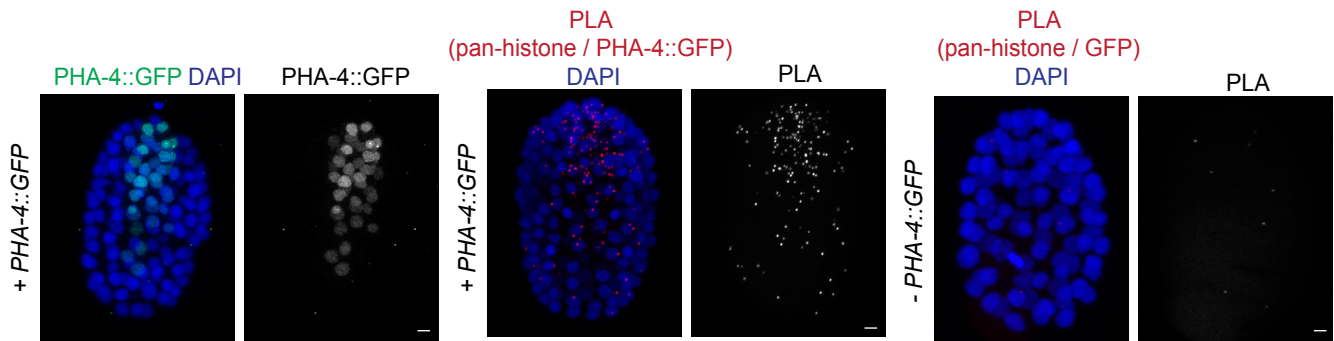
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D



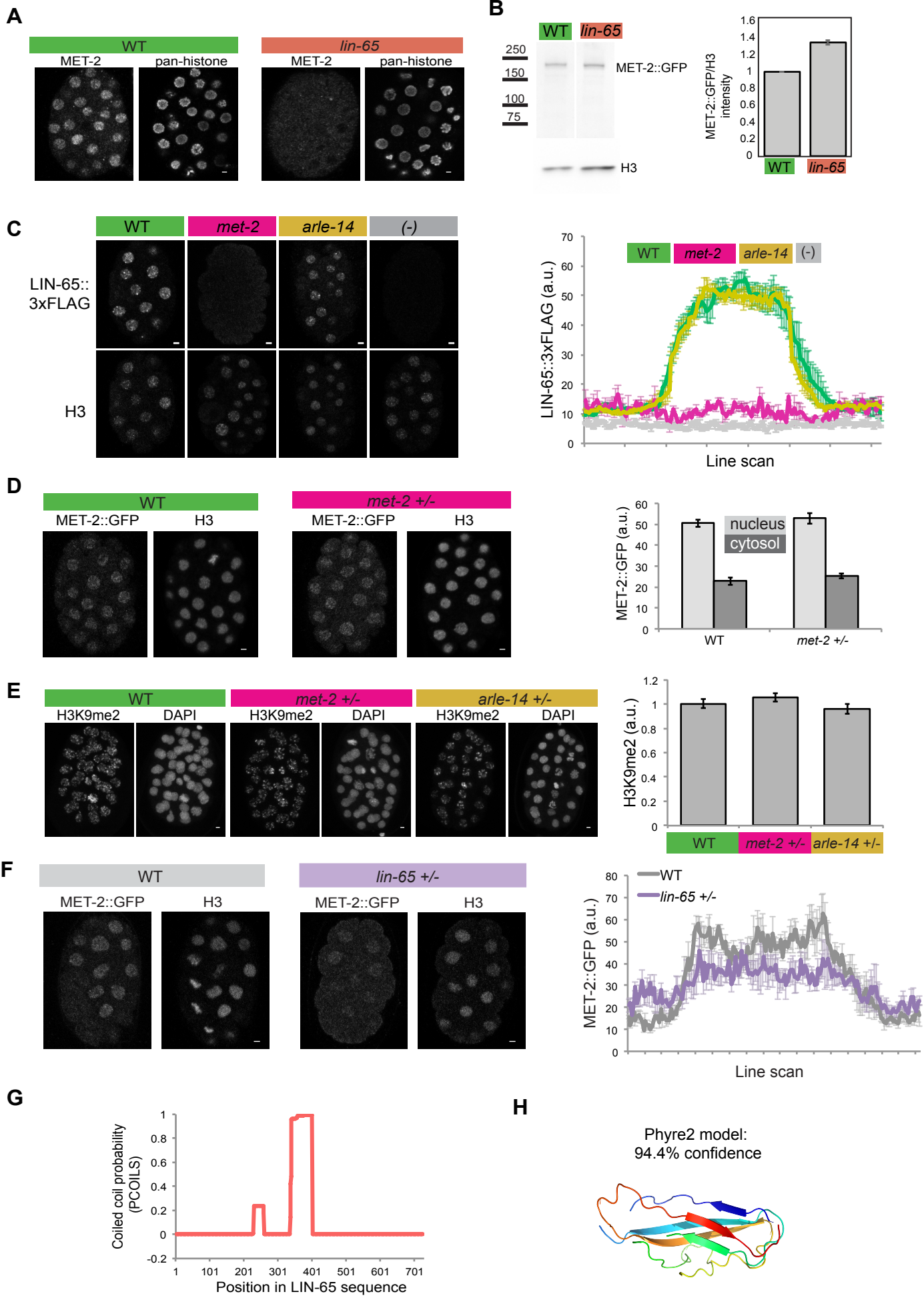
E



F



**Fig. S4. Specificity controls for MET-2 coimmunoprecipitation and PLA.** (A) Table showing spectral counts and peptide coverage after GFP IP-MudPIT MS for the following strains: *met-2::gfp*, wild-type (no GFP) and *zen-4::gfp* (GFP control). (B) Silver stain showing the 100 kDa LIN-65 band after MET-2::GFP IP, identified by cutting out the 100kDa band and Mass Spec. (C and D) Whole embryos showing LIN-65::3xFLAG ( $\alpha$ FLAG antibody; C) and endogenous ARLE-14 (D) at different stages of embryonic development with a H3 or pan-histone co-stain (Scale bar, 2  $\mu$ m). Quantitation of nuclear and total protein during embryogenesis, normalized to histone. (E) Left panel shows the staining pattern of transcription factor PHA-4::GFP (green) in whole embryos, with DNA (DAPI). The two panels on the right show PLA signal between GFP and pan-histone (red) with DNA (DAPI) in the following strains: *pha-4::gfp* or wild-type *N2* (no GFP -PHA-4::GFP) (Scalebar 2  $\mu$ m). (F) Specificity controls for Fig. 4K. PLA for LIN-65::3xFLAG/MET-2::GFP (left panel) and MET-2::GFP/ARLE-14 (right panel) under conditions where individual proteins are depleted. (Scale bar 2  $\mu$ m)



**Fig. S5. Further analysis of LIN-65 and dosage compensation for MET-2 protein.**

(A) Whole embryos stained with antibodies against endogenous MET-2 protein in wild-type (green) vs. *lin-65* (red) mutants, and co-stained for pan-histone. (B) MET-2::GFP IP and Western blot with GFP and histone antibodies in wild-type vs. *lin-65* embryonic extracts on the same gel. The lane between wild-type vs. *lin-65* is not shown. The graph shows pixel counts for the intensity of MET-2::GFP normalized to histone H3. Error bars denote standard error of the mean for N=2 experiments. (C) LIN-65::3xFLAG ( $\alpha$ FLAG antibody) staining in wild-type vs. *met-2* and *arle-14* mutants. Line scan quantitation. Average of line scans across multiple nuclei are shown and error bars denote standard error of the mean. (D) MET-2::GFP levels in progeny of *met-2*(+/-) moms. Quantitation of MET-2::GFP signal intensity in the cytosol (dark grey) and the nucleus (light grey). Error bars denote standard error of the mean. (E) H3K9me2 levels in progeny of *met-2*(+/-) and *arle-14*(+/-) moms and quantified. Error bars denote standard error of the mean. (F) MET-2::GFP levels in WT vs. progeny of *lin-65*(+/-) heterozygous moms. Line scan quantitation showing mean accumulation of MET-2::GFP in wild-type (grey) or *lin-65*(+/-) (purple) embryos at the 21-50 cell stage. Average of line scans across multiple nuclei are shown and error bars denote standard error of the mean. (G) Coiled-coil probability across LIN-65 amino acid sequence predicted by PCOILS (<https://toolkit.tuebingen.mpg.de/#/tools/pcoils>). (H) Model structure of LIN-65 C-terminus predicted by Phyre2 (<http://www.sbg.bio.ic.ac.uk/phyre2/html/page.cgi?id=index>). Full LIN-65 a.a. sequence was used as input for Phyre2.