Supplementary Materials

Bioengineering of bacterial pathogens for noninvasive imaging and *in vivo* evaluation of therapeutics

Authors: Sathish Rajamani¹*, Kyle Kuszpit¹, Jennifer M. Scarff¹, Linnea Lundh¹, Maisha Khan¹, Jennifer Brown¹, Robert Stafford¹, Lisa H. Cazares¹, Rekha G. Panchal¹ and Thomas Bocan¹*

Affiliations:

¹Molecular & Translational Sciences Division, US Army Medical Research Institute of Infectious Diseases, 1425 Porter Street, Frederick, MD 21702

Supplementary Information Data 1. DNA sequences of Promoter – thymidine kinase fusion in *P. aeruginosa*

P. aeruginosa Promoter_lasR (428bp, GenBank: M59425.1) + *E. coli* TK (GenBank: DQ384607.1) fusion for pUC18T-mini-Tn7T-Gm (Genbank: AY599232.2) cloning for *P. aeruginosa* chromosomal integration (BamHI and HindIII)

Sequentially cloned into pBBR1MCS5 following PCR with below primers and cloning with desired restriction sites.

5'ProLasR w/KpnI GAAAC<u>GGTACC</u>CGCTGTGCCTTTGCGCGTG

3' ProLasR w/ClaI GAAAC<u>ATCGAT</u>AGCGCTACGTTCTTCTTAAACTATTAACCAATCAGCC

5'E.coli_TK w/ClaI CAAAC<u>ATCGAT</u>ATGGCACAGCTATATTTCTACTATTCCGCAATGAATGC

3'E.coli_TK w/XbaI CAAAC<u>TCTAGA</u>TTAATCGTGGCGATGCCTTTCCTGAATAGC

Primers for PCR amplification and subcloning of PromoterLasR-TK into pUC18T-mini-Tn7T-Gm vector using pBBR1MCS5-PlasR-TK plasmid template

5'ProLasR w/BamHI GAAAC<u>GGATCC</u>CGCTGTGCCTTTGCGCGTG

3'E.coli_TK w/HindIII CAAAC<u>AAGCTT</u>TTAATCGTGGCGATGCCTTTCCTGAATAGC Supplementary Information Data 2. DNA sequences of Promoter – thymidine kinase fusion in *A. baumannii*

A. baumannii **Promoter_16srDNA** (A1S_r01) **promoter** (327bp, GenBank: CP000521.1) + pantoate--beta-alanine ligase (vitB5 pathway) Ribosome Binding Site and *E. coli* TK

Primers for Prrn_RBS-TK cloning into pUC18T-mini-Tn7T-Gm vector using pUC57-Prrn_RBS-TK

5'Pro_16S_AB w/BamHI GAAAC<u>GGATCC</u>GATTAGATTGGTTGCTTTAAGTGATGAATTTGATGATG

3'E.coli_TK w/HindIII CAAAC<u>AAGCTT</u>TTAATCGTGGCGATGCCTTTCCTGAATAGC

PTn7R (Chromosomal integration verification) GTAAACTGAAATCAGTCCAGTTATGCTGTGA

PglmSF1 (Chromosomal integration verification) TTCGTTTTTGCTGATGAAAATAGCGGTG Supplementary Information Data 3. DNA sequences of Promoter – thymidine kinase fusion in Bp82

B. pseudomallei S12 gene promoter (205bp, GenBank: CP004379.1) + *B. pseudomallei* codonoptimized *E. coli* TK gene fusions for gene synthesis and cloning into pUC18TminiTn7-Km::FRT (GenBank: EU223384.2)

BpS12 promoter fusion and with protein secretory signal sequences for bacterial TK secretion into host cell. Codon optimized of *E. coli* TK for Bp engineering was carried out using http://www.jcat.de/

Primers for PCR amplification of ProS12-TK promoter fusions into pUC18TminiTn7-Km::FRT using pUC57 clones (Genewiz DNA synthesis) of above inserts

5'NewBpProS12 w/KpnI CAAAT<u>GGTACC</u>GGA TCC CGC CTT TGC ATT CCG G

3'TKBPcodw/HindIII CAAAT<u>AAGCTT</u>TTAGTCGTGGCGGTGGCG Supplementary Information Figure 1. Growth inhibition studies with WT and *tk* engineered *Acinetobacter baumannii* ATCC 17978 in the presence of Zidovudine and FIAU





Supplementary Information Figure 2. Growth inhibition studies with WT and *tk* engineered *B. pseudomallei* Bp82 in the presence of Zidovudine and FIAU





Supplementary Information Figure 3. Dynamic PET/CT scans with PAO1 and PAO1TK infected mice

In vivo images of mice (m03, upper panel, and m04, lower panel) injected with 10⁹ CFU PAO1TK (left upper and lower quadrants) and PAO1 (right upper and lower quadrants) and imaged at different time points after [¹⁸F]FIAU injection. Mouse m03 images are threshold adjusted to focus on the upper limbs injected with bacteria, while m04 is threshold adjusted to show lower limbs with bacteria. Table below shows the volume-of-interest (VOI) and signal intensities (UL-upper right, LL-lower right UR-upper right, LR- lower right, and BG-background). The signal intensities were used to tabulate the corresponding signal-to-noise ratios.

To quantify the PET data, a sphere equivalent to a 50 μ l volume was used so that the sample size was the same for each treatment. The 50 μ l sphere or VOI was applied to the images and the amount of radioactivity quantified. The background signal was defined as the amount of PET signal present in the VOI applied to the muscle in the non-infected limb, e.g., in mouse 03, background (BG) equals 4.61 %ID/g at 1 h10min. For the determination of the bacterial-tk signal, the 50 μ l VOI was applied to the limb containing the bacteria expressing thymidine kinase, e.g., mouse 03 signal in the upper right limb (UL) was 6.06 %ID/gm. The signal-to-noise was the ratio of the activity in the UL divided by BG or 6.06/4.61 which equals 1.32.

In vivo images of mouse (m03) with 10⁹ PAO1 (WT) and PA01TK (TK)

~1h10m ~2h30m ~3h50m ~4h55m



~50µL VOIs used for analysis

m03	Mean (nCi/cc)			Sig to Noise	
Scan @	UL	LL	BG	UL-BG	LL- <mark>B</mark> G
~1h10m	12219.90	11734.20	9287.46	1.32	1.26
~2h30m	7558.64	7490.90	5017.08	1.51	1.49
~3h50m	4569.47	4656.05	2934.45	1.56	1.59
~4h55m	3157.26	2922.62	2134.91	1.48	1.37

	Decay Corrected				
m03	M	lean (%ID/	Sig to Noise		
Scan @	UL	LL	BG	UL-BG	LL-BG
~1h10m	6.06	5.82	4.61	1.32	1.26
~2h30m	6.23	6.18	4.14	1.51	1.49
~3h50m	6.36	6.48	4.08	1.56	1.59
~4h55m	6.49	6.01	4.39	1.48	1.37

In vivo images of mouse (m04) with 10⁹ PAO1 (WT) and PA01TK (TK)

~1h10m ~2h30m ~3h50m ~4h55m



~50µL VOIs used for analysis

101.000				1	
m04	N	lean (nCi/o	Sig to Noise		
Scan @	UL	LL	BG	UL-BG	LL-BG
~1h10m	11556.00	11203.40	7802.61	1.48	1.44
~2h30m	7567.50	7022.72	3951.73	1.91	1.78
~3h50m	4683.13	4512.75	2399.43	1.95	1.88
~4h55m	3064.61	3006.66	2024.68	1.51	1.49
	Decay Corrected				
m04	Mean (%ID/g)			Sig to Noise	
Scan @	UL	LL	BG	UL-BG	LL-BG
~1h10m	5.72	5.55	3.86	1.48	1.44
~2h30m	6.23	5.78	3.25	1.91	1.78
~3h50m	6.50	6.26	3.33	1.95	1.88
~4h55m	6.29	6.17	4.15	1.51	1.49

Note: Animals m03 and m04 both died just prior to scanning at ~4h55m

Supplementary Information Figure 4: Bacterial imaging in BALB/c after infection with 10⁷ CFU PAO1 (WT) or PAO1TK (TK) detected with [¹⁸F]FIAU



