

Supplementary Material

A gene-based positive selection detection approach to identify vaccine candidates using *Toxoplasma gondii* as a test case protozoan pathogen

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Supplementary Table S1: Command-line syntax for programs used in study

Program^a	Command-line syntax with parameters^b
BLASTP	<code>blastp -query prot_Toxoplasma_ME49.fasta -subject prot_Toxoplasma_FOU.fasta -outfmt "10 evaluate qseqid qlen sseqid slen bitscore score pident nident mismatches positive" -max_target_seqs 5 -out blastp_Toxoplasma_ME49</code>
Clustal Omega	<code>clustalo -i mrna_sequences.txt -o clustal_output --force</code>
PAL2NAL	<code>pal2nal.pl clustal_output mRNA_sequences.fasta -output paml -nogap > pal2nal_output</code>
RAxML	<code>raxmlHPC-PTHREADS -f a -x 12345 -p 12345 -# 100 -m GTRGAMMA -s pal2nal_output -n raxml_output -w out_dir</code>
CODEML	<code>codeml control_file.ctl > output_file</code>
predict_binding.py	<code>predict_binding.py IEDB_recommended HLA-A*23:01 9 input.fasta</code>
MKtest	<code>GENOME.class <- set.populations(GENOME.class,list(within_pop,between_pop)) GENOME.class <- MKT(GENOME.class,do.fisher.test=TRUE) results <- get.MKT(GENOME.class) neutrality.index <- results[1,"neutrality.index"]</code>
Tajima's D	<code>GENOME.class <- neutrality.stats(GENOME.class) tajima_result <- GENOME.class@Tajima.D</code>
F _{ST}	<code>GENOME.class <- F_ST.stats(GENOME.class) get.F_ST (GENOME.class,mode="nucleotide") Results <- get.diversity (GENOME.class) fst_nucleotide <- results[1,"nucleotide.F_ST"]</code>

^aMKtest = the McDonald–Kreitman test, F_{ST} = Fixation index. MKtest, Tajima's D and F_{ST} methods are performed using R functions from PopGenome (<https://cran.r-project.org/web/packages/PopGenome/index.html>).

^b-f a = selects the algorithm to be 'rapid Bootstrap analysis and search for best scoring ML tree in one program run'; -x = rapid Bootstrap random number seed; -p = parsimony random seed; -# = number of runs; -m GTRGAMMA = generalised time reversible (GTR) GAMMA substitution model.