

**Transcriptional dynamics during lens fiber cell differentiation and novel insights into the denucleation process**

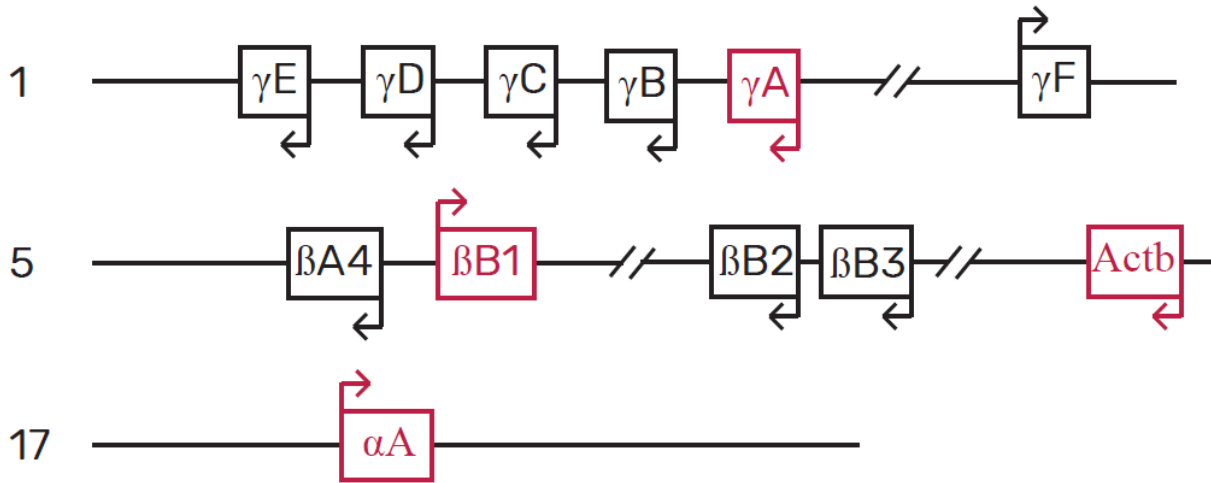
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**Materials Included:**

Figures S1 to S7 and Tables S1-S2

**Fig. S1. Chromosomal localization, probe design, and stages of lens fiber cell differentiation.** A) Chromosomal localization of genes on three mouse chromosomes: 1 ( $\gamma$ A-Crystallin), 5 ( $\beta$ B1-Crystallin and  $\beta$ -actin), and 17 ( $\alpha$ A-Crystallin). Genes examined and their neighbors are shown in red and black boxes, respectively. B) A summary of probes and their dyes used in this study.

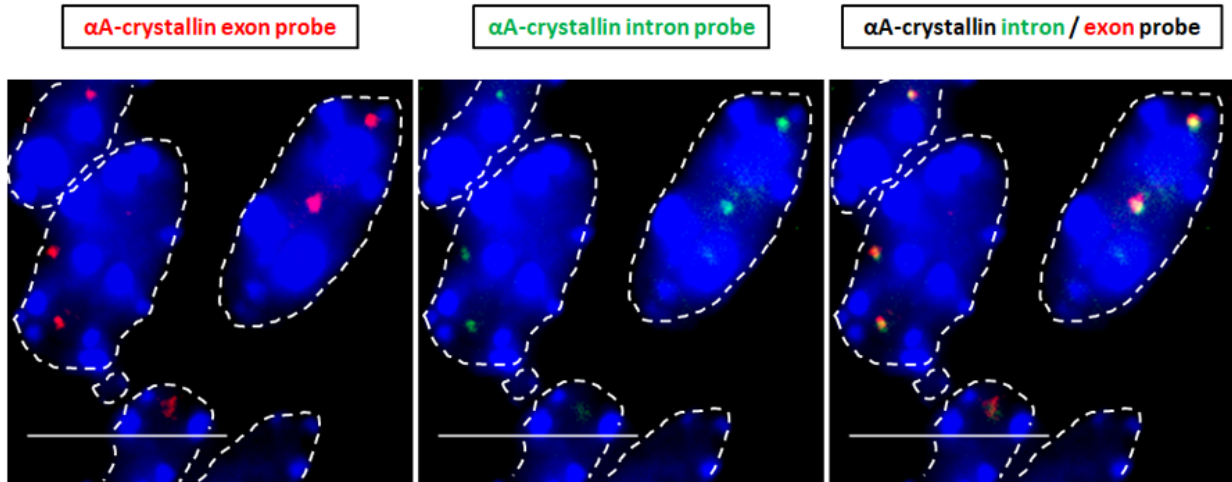
**Fig. S1A**



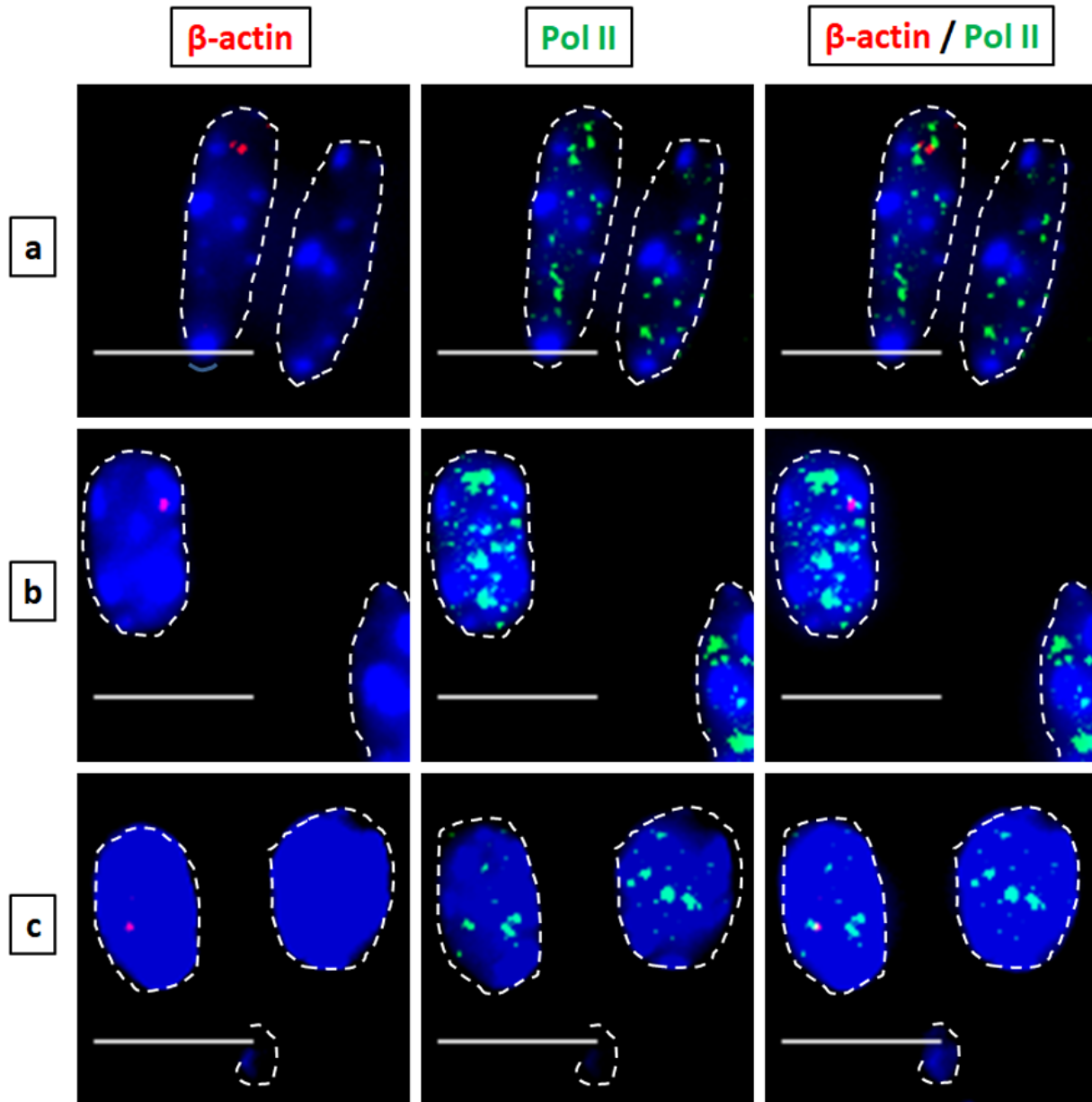
**Fig. S1B**

Gene	Probe length	Exon or Intron	Dye	# of probes
$\alpha$ A-Crystallin	20	Exon	Quasar 570	25
$\alpha$ A-Crystallin	20	Intron	Quasar 670	21
$\gamma$ A-Crystallin	20	Exon	Quasar 670	16
$\beta$ B1-Crystallin	20	Exon	CAL Fluore Red 610	12
$\beta$ -actin	20	Exon	Quasar 670	35

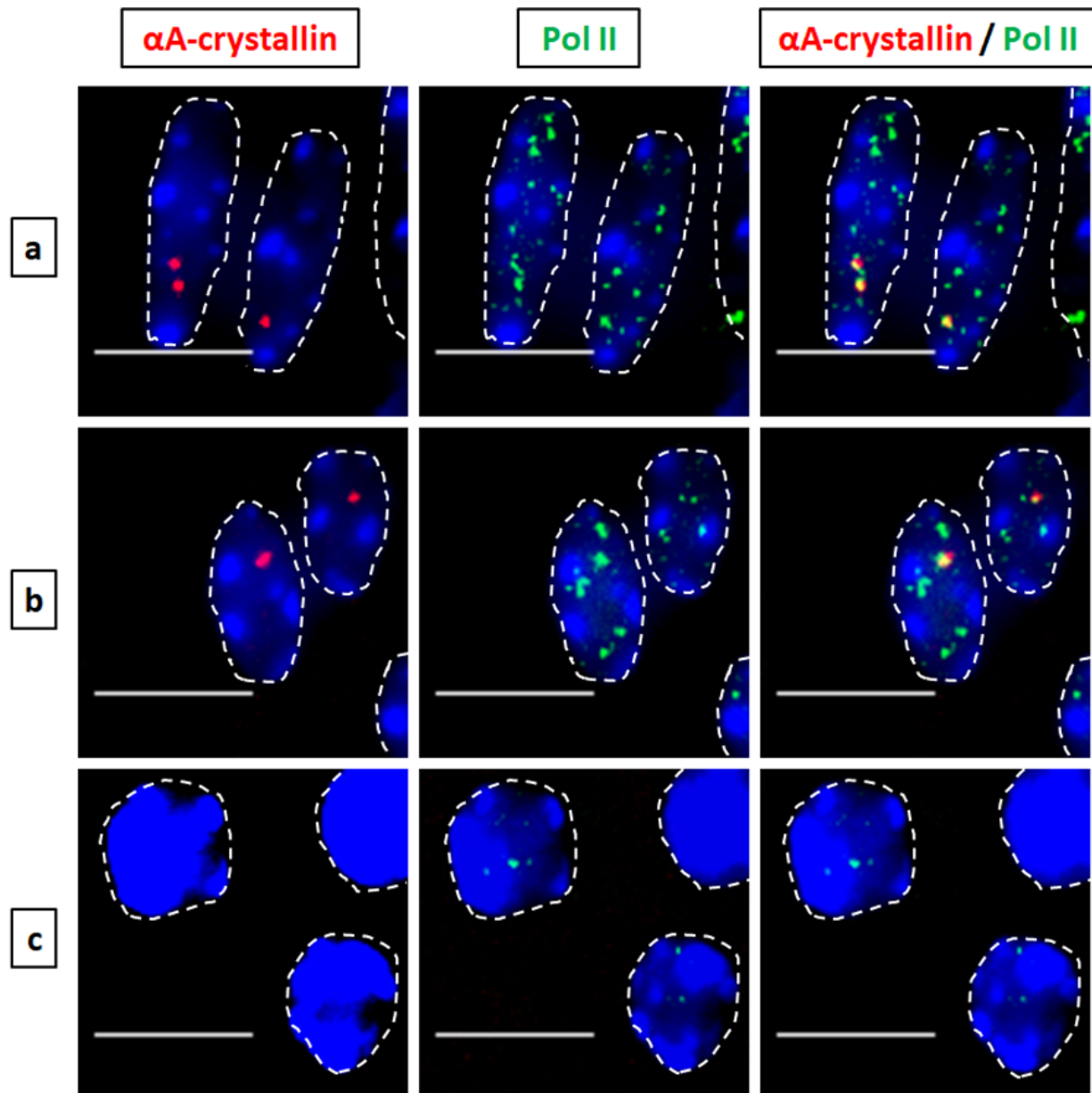
**Fig. S2. RNA FISH of  $\alpha$ A-crystallin introns and exons to identify and confirm nascent transcription sites in cell nuclei of newborn mouse lens tissue. Exons and introns were detected by Quasar 570- and Quasar 670-labeled probes (see Fig. S1B), respectively.**



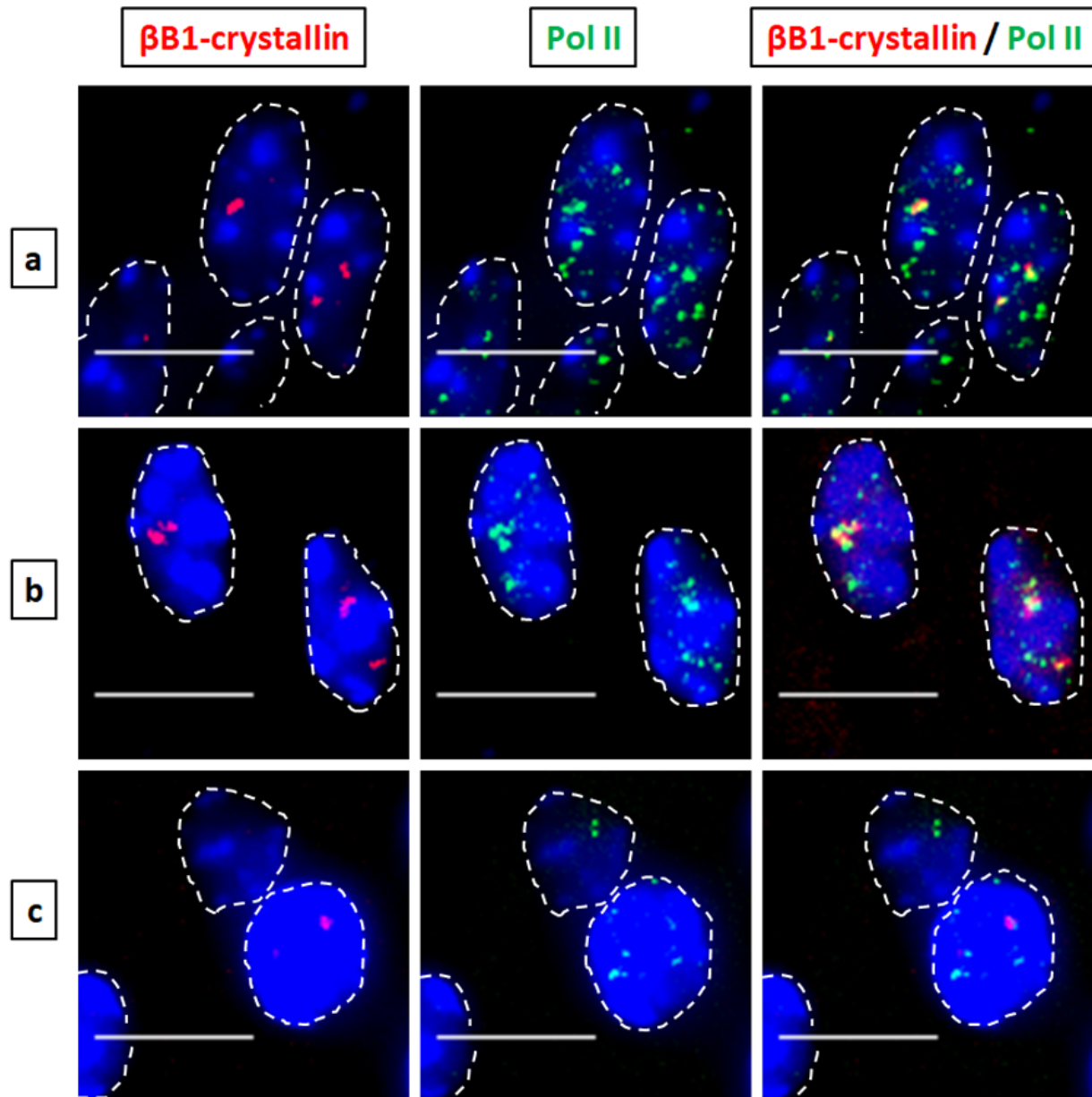
**Fig. S3. Co-localization of active RNA PolII and the transcription sites of  $\beta$ -actin gene to confirm that RNA FISH technique detects nascent transcription sites.** The co-localization shows  $\beta$ -actin by RNA FISH (red) and RNA polymerase II (green) by immunofluorescence in newborn mouse lens fiber cell nuclei. RNA FISH signal were detected by various Quasar or CAL Fluor dyes indicated in Fig. S1B and RNA polymerase II signal was detected by Alexa-Fluor 488 conjugated secondary antibody against an anti-Pol II primary antibody.



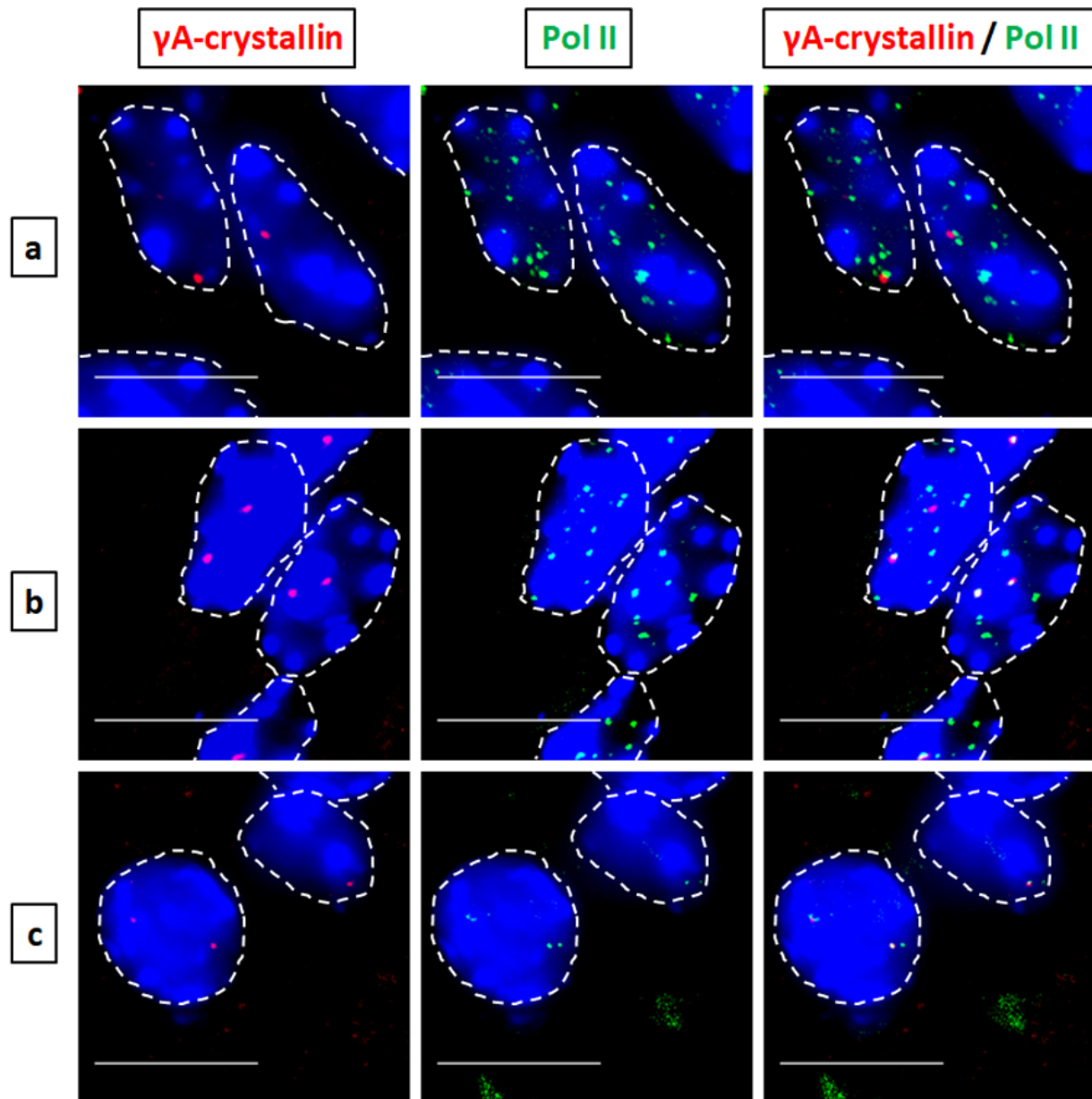
**Fig. S4. Co-localization of active RNA PolII and the transcription sites of  $\alpha$ A-crystallin gene to confirm that RNA FISH technique detects nascent transcription sites.** The co-localization shows  $\alpha$ A-crystallin by RNA FISH (red) and RNA polymerase II (green) by immunofluorescence in newborn mouse lens fiber cell nuclei. RNA FISH signal were detected by various Quasar or CAL Fluor dyes indicated in Fig. S1B and RNA polymerase II signal was detected by Alexa-Fluor 488 conjugated secondary antibody against an anti-Pol II primary antibody.



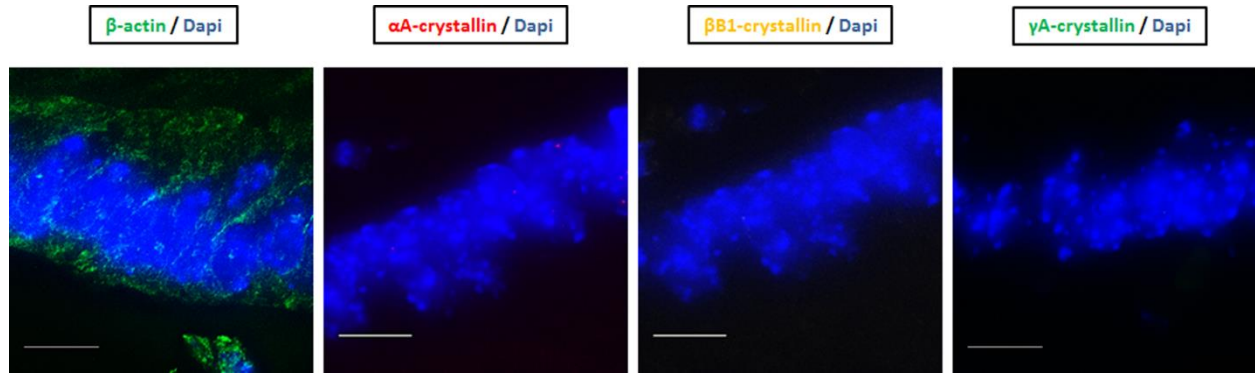
**Fig. S5. Co-localization of active RNA PolII and the transcription sites of  $\beta$ B1-crystallin gene to confirm that RNA FISH technique detects nascent transcription sites.** The co-localization shows  $\beta$ B1-crystallin by RNA FISH (red) and RNA polymerase II (green) by immunofluorescence in newborn mouse lens fiber cell nuclei. RNA FISH signal were detected by various Quasar or CAL Fluor dyes indicated in Fig. S1B and RNA polymerase II signal was detected by Alexa-Fluor 488 conjugated secondary antibody against an anti-Pol II primary antibody.



**Fig. S6. Co-localization of active RNA PolII and the transcription sites of  $\gamma$ A-crystallin gene to confirm that RNA FISH technique detects nascent transcription sites.** The co-localization shows  $\gamma$ A-crystallin by RNA FISH (red) and RNA polymerase II (green) by immunofluorescence in newborn mouse lens fiber cell nuclei. RNA FISH signal were detected by various Quasar or CAL Fluor dyes indicated in Fig. S1B and RNA polymerase II signal was detected by Alexa-Fluor 488 conjugated secondary antibody against an anti-Pol II primary antibody.



**Fig. S7. RNA FISH in newborn mouse lens epithelium shows that  $\beta$ B1- and  $\gamma$ A-crystallin shows no expression within the lens epithelium.** The various indicated genes were detected by Quasar and CAL Fluor dyes indicated in figure S1B.





**Table. S1.** Number of cells counted at each of the four developmental stages, E12.5, E14.5, E16.5 and P1, from each of the four areas ‘a’, ‘b’, ‘c’, ‘d’, for each for the four genes studied,  $\beta$ -actin,  $\alpha$ A-,  $\beta$ B1- and  $\gamma$ A-crystallin.

<b>E12.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	145	132	152	138
<b><math>\alpha</math>A-crystallin</b>	293	267	301	276
<b><math>\beta</math>B1-crystallin</b>	109	116	147	162
<b><math>\gamma</math>A-crystallin</b>	148	135	149	138

<b>E14.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	195	92	82	185
<b><math>\alpha</math>A-crystallin</b>	306	222	159	259
<b><math>\beta</math>B1-crystallin</b>	194	113	91	70
<b><math>\gamma</math>A-crystallin</b>	112	109	68	189

<b>E16.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	229	140	107	111
<b><math>\alpha</math>A-crystallin</b>	340	219	177	108
<b><math>\beta</math>B1-crystallin</b>	207	158	116	80
<b><math>\gamma</math>A-crystallin</b>	272	133	106	125

<b>P1</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	559	329	211	0
<b><math>\alpha</math>A-crystallin</b>	750	434	214	0
<b><math>\beta</math>B1-crystallin</b>	381	211	114	0
<b><math>\gamma</math>A-crystallin</b>	326	246	162	0

	<b>WT P1</b>	<b>Snf2h null P1</b>
<b><math>\beta</math>-actin</b>	901	540
<b><math>\alpha</math>A-crystallin</b>	1319	717
<b><math>\beta</math>B1-crystallin</b>	794	534
<b><math>\gamma</math>A-crystallin</b>	805	504

**Table. S2.** Number of transcription sites counted at each of the four developmental stages, E12.5, E14.5, E16.5 and P1, from each of the four areas ‘a’, ‘b’, ‘c’, ‘d’, for each for the four genes studied,  $\beta$ -actin,  $\alpha$ A-,  $\beta$ B1- and  $\gamma$ A-crystallin.

<b>E12.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	50	50	80	73
<b><math>\alpha</math>A-crystallin</b>	100	198	285	203
<b><math>\beta</math>B1-crystallin</b>	23	67	86	58
<b><math>\gamma</math>A-crystallin</b>	4	8	11	8

<b>E14.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	71	73	68	97
<b><math>\alpha</math>A-crystallin</b>	226	289	173	372
<b><math>\beta</math>B1-crystallin</b>	112	108	84	58
<b><math>\gamma</math>A-crystallin</b>	26	110	95	241

<b>E16.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	203	110	68	51
<b><math>\alpha</math>A-crystallin</b>	448	272	139	13
<b><math>\beta</math>B1-crystallin</b>	128	157	93	41
<b><math>\gamma</math>A-crystallin</b>	76	115	83	112

<b>P1</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	439	249	121	0
<b><math>\alpha</math>A-crystallin</b>	791	306	11	0
<b><math>\beta</math>B1-crystallin</b>	275	151	41	0
<b><math>\gamma</math>A-crystallin</b>	146	272	182	0

	<b>WT P1</b>	<b>Snf2h null P1</b>
<b><math>\beta</math>-actin</b>	651	240
<b><math>\alpha</math>A-crystallin</b>	909	315
<b><math>\beta</math>B1-crystallin</b>	543	268
<b><math>\gamma</math>A-crystallin</b>	555	235

**Table. S3.** Table of standard deviations for transcription burst fraction measurements.

<b>E12.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	2.7882	2.9289	2.9236	3.2262
<b><math>\alpha</math>A-crystallin</b>	2.4722	2.5925	2.5358	2.4697
<b><math>\beta</math>B1-crystallin</b>	2.6075	3.6279	3.0261	2.5343
<b><math>\gamma</math>A-crystallin</b>	1.121	2.2824	2.7867	2.736

<b>E14.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	2.1847	4.2984	4.629	2.7256
<b><math>\alpha</math>A-crystallin</b>	2.1323	2.1575	2.6829	1.8326
<b><math>\beta</math>B1-crystallin</b>	2.6858	3.8106	4.5014	4.7104
<b><math>\gamma</math>A-crystallin</b>	1.3306	3.1414	3.7352	2.7256

<b>E16.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	2.7003	3.6358	3.8264	3.8217
<b><math>\alpha</math>A-crystallin</b>	2.0324	2.0283	2.2346	0.6192
<b><math>\beta</math>B1-crystallin</b>	2.5718	3.2439	3.7435	4.0658
<b><math>\gamma</math>A-crystallin</b>	1.8998	3.5347	4.1766	3.7829

<b>P1</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	1.7641	2.1979	2.717	NA
<b><math>\alpha</math>A-crystallin</b>	1.4224	1.885	1.097	NA
<b><math>\beta</math>B1-crystallin</b>	2.0262	2.713	2.8809	NA
<b><math>\gamma</math>A-crystallin</b>	1.8854	2.6379	3.1792	NA

	<b>WT P1</b>	<b>Snf2h null P1</b>
<b><math>\beta</math>-actin</b>	1.3676	1.4698
<b><math>\alpha</math>A-crystallin</b>	1.7556	1.8734
<b><math>\beta</math>B1-crystallin</b>	1.5129	1.8816
<b><math>\gamma</math>A-crystallin</b>	2.22	1.5469

**Table. S4.** Table of standard deviations for transcription burst intensity measurements.

<b>E12.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	7.3733	3.6022	3.2183	2.5498
<b><math>\alpha</math>A-crystallin</b>	2.4522	1.6984	1.3836	1.7059
<b><math>\beta</math>B1-crystallin</b>	12.7818	6.5483	6.3446	8.2172
<b><math>\gamma</math>A-crystallin</b>	90.935	8.2466	15.6604	12.0328

<b>E14.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	12.9059	11.2358	10.152	7.4696
<b><math>\alpha</math>A-crystallin</b>	5.5745	7.8456	11.8615	4.0176
<b><math>\beta</math>B1-crystallin</b>	10.0917	12.8424	9.2171	10.4935
<b><math>\gamma</math>A-crystallin</b>	21.931	4.1301	3.9967	2.2957

<b>E16.5</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	2.0217	2.5324	3.8396	3.9512
<b><math>\alpha</math>A-crystallin</b>	1.3673	1.9192	3.4025	7.7329
<b><math>\beta</math>B1-crystallin</b>	3.8434	3.7089	2.9297	6.0089
<b><math>\gamma</math>A-crystallin</b>	6.8814	2.5694	2.4465	7.2049

<b>P1</b>	<b>a</b>	<b>b</b>	<b>c</b>	<b>d</b>
<b><math>\beta</math>-actin</b>	1.2197	2.6165	5.7212	NA
<b><math>\alpha</math>A-crystallin</b>	1.9589	3.6046	20.3828	NA
<b><math>\beta</math>B1-crystallin</b>	1.3478	2.283	6.8795	NA
<b><math>\gamma</math>A-crystallin</b>	4.0081	2.4814	3.5708	NA

	<b>WT P1</b>	<b>Snf2h null P1</b>
<b><math>\beta</math>-actin</b>	1.4502	2.5658
<b><math>\alpha</math>A-crystallin</b>	1.0032	2.858
<b><math>\beta</math>B1-crystallin</b>	1.8053	3.5122
<b><math>\gamma</math>A-crystallin</b>	1.5066	3.506