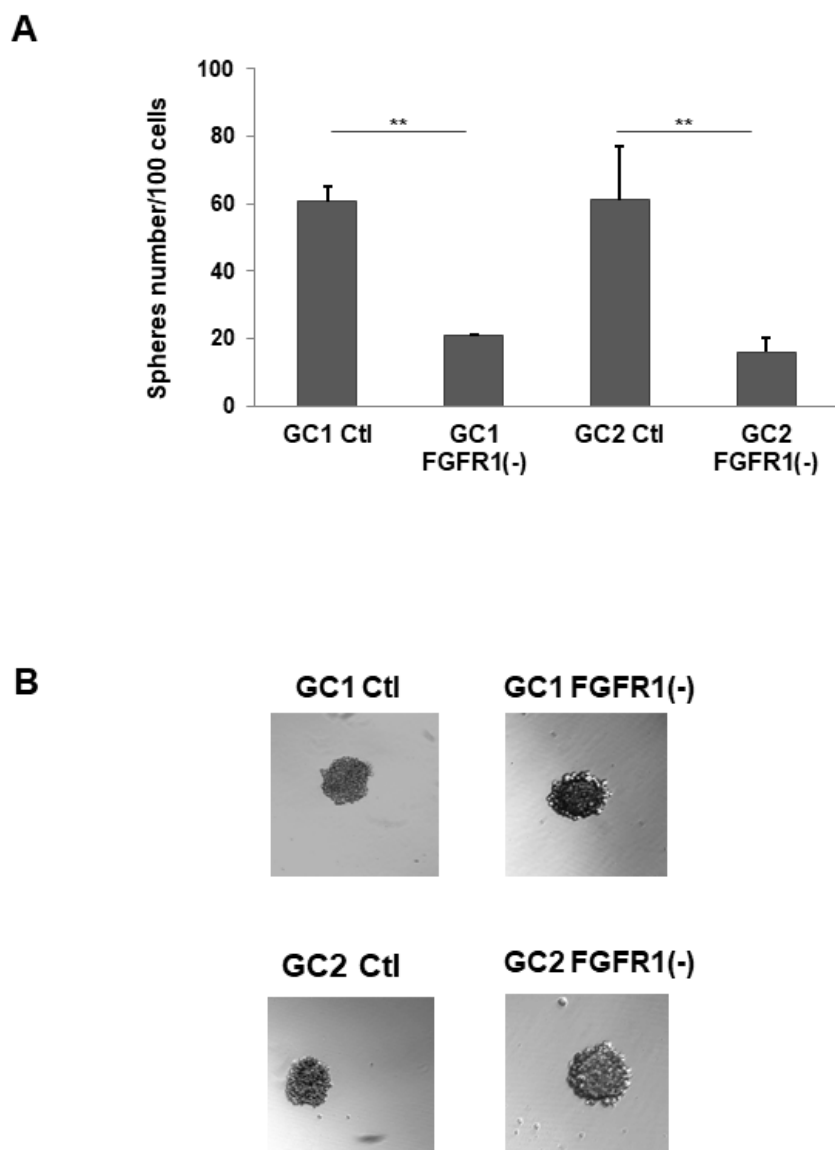
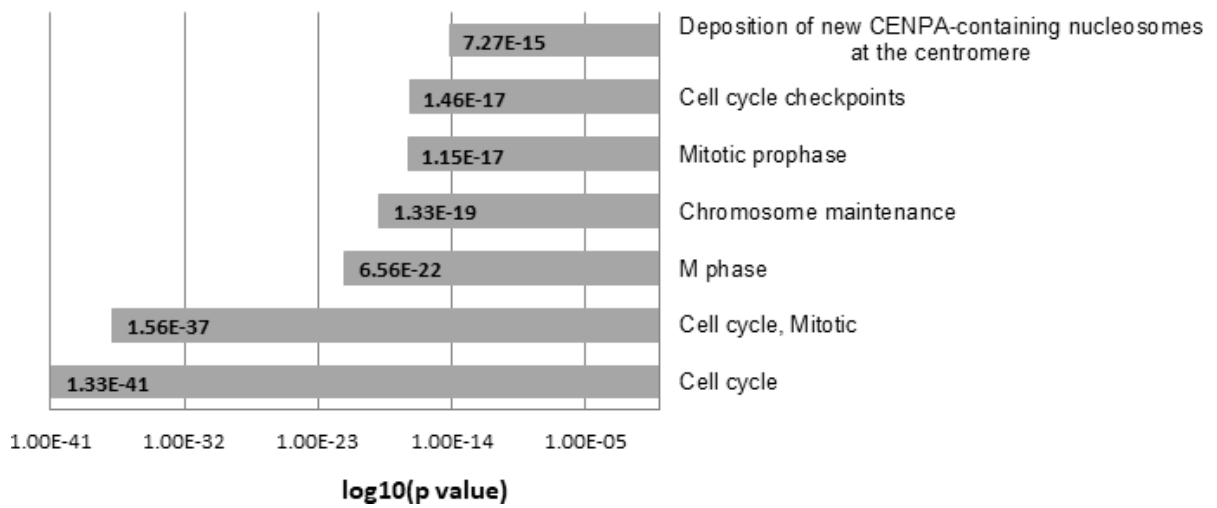


FGFR1/FOXM1 pathway: a key regulator of glioblastoma stem cells radioresistance and a prognosis biomarker

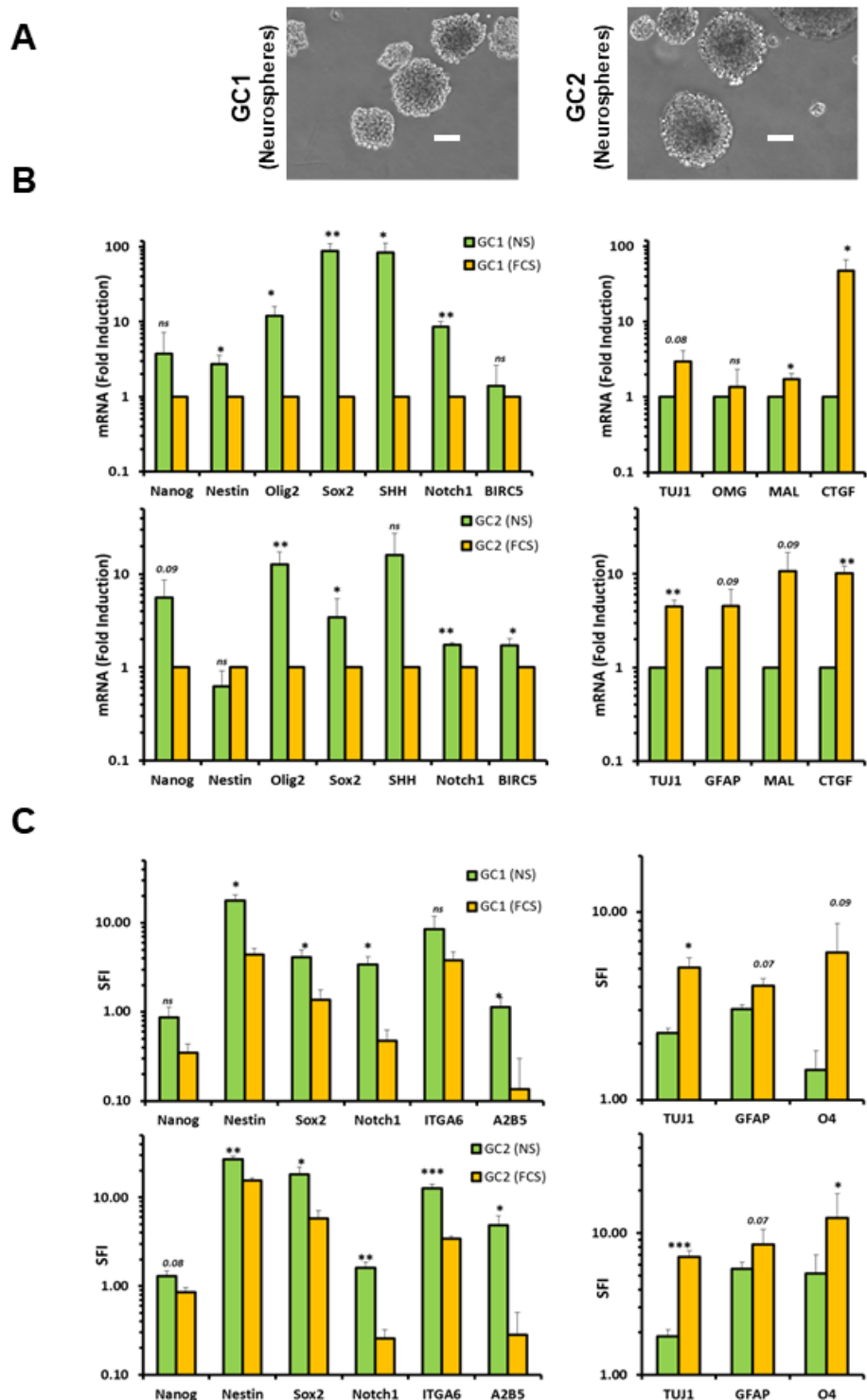
SUPPLEMENTARY MATERIALS



Supplementary Figure 1: Cells derived from human GBM biopsy specimens inhibited for FGFR1 form neurospheres. Cells derived from 2 GBM biopsy specimen (GC1 and GC2) were transfected with siFGFR1(11) (GC1FGFR1(-) or GC2FGFR1(-)) or a scramble control (Ctl). (A) Cells were analyzed for their capacity to form neurospheres as described in “Materials and Methods”. Quantifications of 3 experiments are presented as means \pm SD. *** $p < 0.001$; ** $p < 0.01$; * $0.01 < p < 0.05$. (B) Phase contrast photomicrographs of GSLC-enriched neurosphere cell lines, magnification: $\times 10$, scale bar: 6 μm .



Supplementary Figure 2: FGFR1 inhibition induces a downregulation of cell cycle pathways. The genes list regulated in GCS2FGFR1(-) cells was compared to Reactome database according to “Materials and Methods”.



Supplementary Figure 3: Characterization of the stem and differentiated phenotypes in GSLC-enriched neurospheres and in differentiated GBM cultures (FCS).

(A) Phase contrast photomicrographs of GSLC-enriched neurosphere cell lines isolated from 2 patient tumors (GC1, GC2). Magnification: $\times 10$, scale bar: 6 μm . (B–C) GSLC-enriched neurosphere cell lines (GC1, GC2) were kept in stem cell medium or allowed to differentiate as adherent GBM cells for at least 15 days in FCS medium. (B) Real-time Quantitative PCR analysis of the stem (left panel) and differentiation (right panel) markers in neurospheres or GBM differentiated cells for both GC1 and GC2 cell lines. Shown are the fold inductions expressed as means \pm SEM of at least 3 independent experiments. * $p < 0.05$, ** $p < 0.01$ compared with the related control. ns: not significant. (C) Immunofluorescence FACS analysis of the stem (left panel) and differentiation (right panel) markers in neurospheres or GBM differentiated cells. The results, expressed as SFI (Specific Fluorescence Index), were representative of at least 3 independent experiments. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ compared with the related control.

Supplementary Table 1: Forward and reverse primer sequences used for quantitative PCR detection

| Gene (human) | Primer sequences | Type of marker |
|-------------------------|--|----------------------------|
| β2-microglobulin | forward 5'-ACCCCACTGAAAAAGATGA-3' reverse 5'-ATCTTCAAACCTCCATGATG-3' | endogenous control |
| OMG | forward 5'-TAGGGACTCCATGTTCTACCCA-3' reverse 5'-TCTGCATCCACTTACAGTGA-3' | Oligodendrocytic marker |
| MAL | forward 5'-CGCTGCCCTCTTTTACCTCAG-3' reverse 5'-GAAGCCGTCTTGCATCGTGAT-3' | Oligodendrocytic marker |
| GFAP | forward 5'-GGCAAAGCACCAAAGACGG-3' reverse 5'-GGCGGCGTTCCATTTACAAT-3' | Astrocytic marker |
| TUJ1 | forward 5'-GCTCAGGGGCCCTTTGGACATCTCTT-3' reverse 5'-TTTTCACTCCTTCCGCACCACATC-3' | Neuronal marker |
| CTGF | forward 5'-CATCTCCACCCGGGTTACC-3' reverse 5'-CAGGCGGCTCTGCTTCTCTA-3' | Differentiation marker |
| Nanog | forward 5'-GTCCCGGTCAAGAAACAGAA-3' reverse 5'-TGCGTCACACCATTGCTATT-3' | Stemness-associated marker |
| Nestin | forward 5'-ATCGCTCAGGTCCTGGAAGG-3' reverse 5'-AAGCTGAGGGAAGTCTTGGAG-3' | Stemness-associated marker |
| Sox2 | forward 5'-GCACATGAACGGCTGGAGCAACG-3' reverse 5'-TGCTGCGAGTAGGACATGCTGTAGG-3' | Stemness-associated marker |
| Olig2 | forward 5'-CAGAAGCGCTGATGGTCATA-3' reverse 5'-TCGGCAGTTTTGGGTTATTC-3' | Stemness-associated marker |
| SonicHH | forward 5'-GCGGAAGGTATGAAGGGAAG-3' reverse 5'-GCCAAAGCGTTCAACTTGTC-3' | Stemness-associated marker |
| Notch1 | forward 5'-TGGACCAGATTGGGGAGTTC-3' reverse 5'-GCACACTCGTCTGTGTTGAC-3' | Stemness-associated marker |
| BIRC5/Survivin | forward 5'-CGAGGCTGGCTTCATCCA-3' reverse 5'-AGAAGAAACACTGGGCCAAGTC-3' | Stemness-associated marker |

Supplementary Table 2: Primary antibodies used for flow cytometry

| Marker (human) | Antibodies and fluorochromes | Suppliers | FACS channel | Type of marker |
|-----------------------|---|------------------|---------------------|----------------------------|
| A2B5 | Mouse monoclonal antibody : A2B5 - APC Isotype control : IgM - APC | Miltenyi | FL4-H | Stemness-associated marker |
| ITGA6 | Rat monoclonal antibody : CD49f - PE Isotype control : Igg2A -PE | eBioscience | FL2-H | Stemness-associated marker |
| GFAP | Mouse monoclonal antibody : GFAP - AF647 Isotype control : Igg2B - AF647 | BD Biosciences | FL4-H | Astrocytic marker |
| Nanog | Goat polyclonal antibody : Nanog - PE Isotype control : IgG - PE | R&D Systems | FL2-H | Stemness-associated marker |
| Nestin | Mouse monoclonal antibody : Nestin -Fluorescein Isotype control : Igg1 - Fluorescein | R&D Systems | FL1-H | Stemness-associated marker |
| Notch1 | Mouse monoclonal antibody : Notch1 - APC Isotype control : Igg1 - APC | R&D Systems | FL4-H | Stemness-associated marker |
| O4 | Mouse monoclonal antibody : O4 - PE Isotype control : IgM - PE | Miltenyi | FL2-H | Oligodendrocytic marker |
| TUJ1 | Mouse monoclonal antibody : Tuj1 - AF488 Isotype control : Igg2A - AF488 | BD Biosciences | FL1-H | Neuronal marker |
| Sox2 | Mouse monoclonal antibody : Sox2 - APC Isotype control : Igg2A - APC | R&D Systems | FL4-H | Stemness-associated marker |

Supplementary Table 3: Six-genes set and their corresponding coefficients

| | Coefficient |
|---------------|--------------------|
| FGFR1 | 0.27084 |
| FOXM1 | 0.63132 |
| MELK | -0.45751 |
| GLI2 | -0.60469 |
| TWIST1 | -0.00581 |
| ZEB1 | -0.01317 |

Coefficients determined as described in “Materials and Methods”.