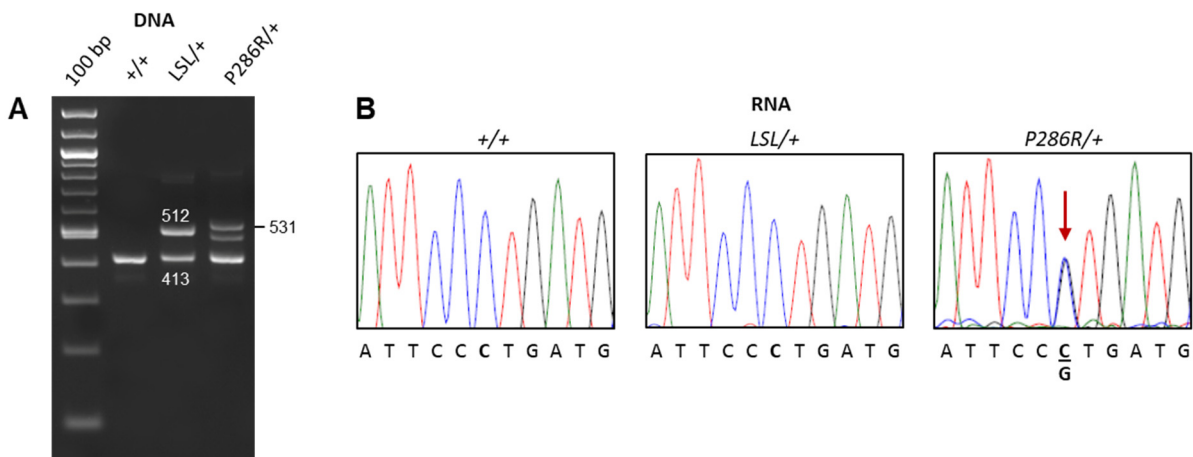


Supplementary Figures 1-5

Li *et al*,

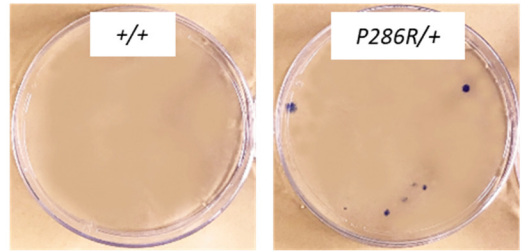
Polymerase-Mediated Ultramutagenesis in Mice Produces Diverse Cancers with High Mutational Load



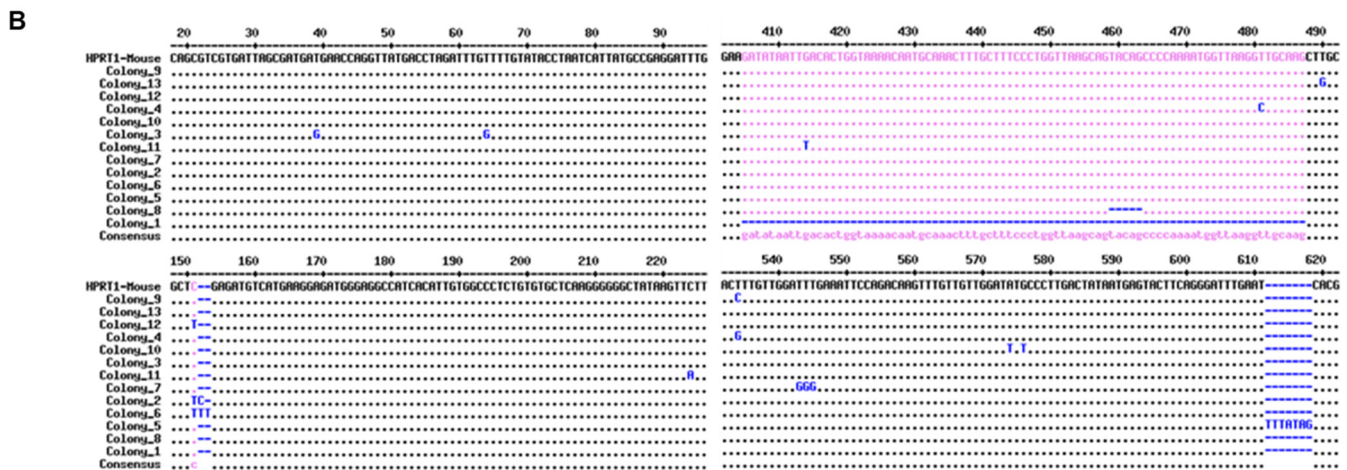
Supplementary Fig. 1. Molecular validation of allele function. A) Multiplex genotyping protocol (tail DNA) distinguished between + (wt), *LSL* (*LSL-Pole^{P286R}*), and *P286R* (*Pole^{P286R}*) alleles. Product sizes (bp) are shown. The *P286R* product typically appears as a doublet but this does not affect interpretation of genotyping. B) RT-PCR using intron-spanning primers to characterize allele expression at RNA level. There is no expression from the *LSL* allele, as expected, whereas the *P286R* and wild-type alleles result in equal expression at that RNA level.

A

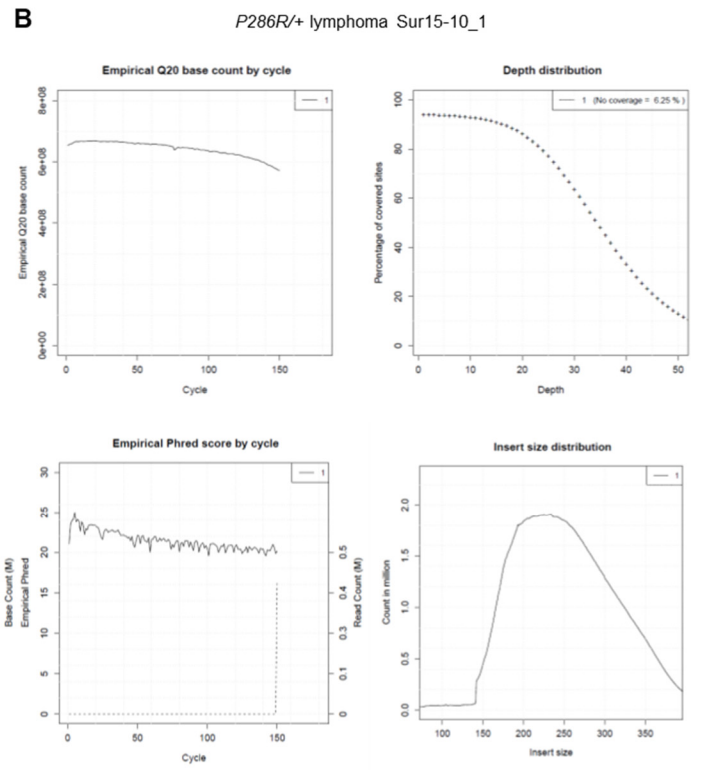
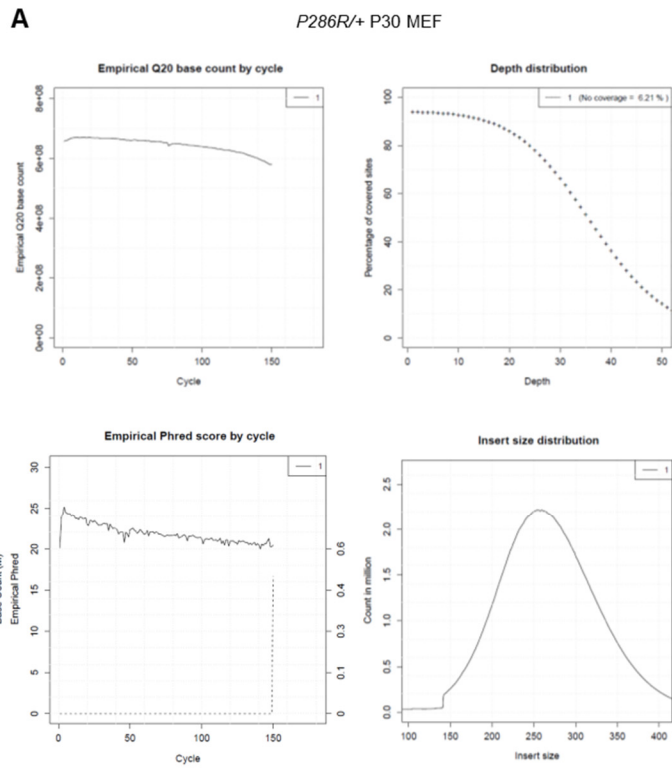
	500 cells regular media ↓		10 ⁶ cells, 60μM 6-TG ↓	
	plating efficiency		6-TG resistant colonies	
	<i>P286R/+</i>	<i>+/+</i>	<i>P286R/+</i>	<i>+/+</i>
	140	124	7	0
	131	129	10	0
	142	115	9	0
	138	130	5	0
	120	126	8	0
mean	134	125	7.8	0



Observed *HPRT1* mutation frequency:
+/+ : 0
P286R/+ : 2.9×10^{-5}
 P value, unpaired T test: $p < 0.0001$



Supplementary Fig. 2. 6-TG selection of passage 20 MEFs. A) Plating efficiency and resistant colonies. Each row represents a separate plate (selection of 10⁶ cells). B) cDNA sequencing of *HPRT1* locus reveals mutations in each of 13 independent 6-TG resistant colonies; most are nonsynonymous substitutions. Only regions of the cDNA with mutations are shown.



Supplementary Fig. 3. General quality control metrics of WGS data. Representative analyses are provided for two of the samples; metrics were comparable across all samples.

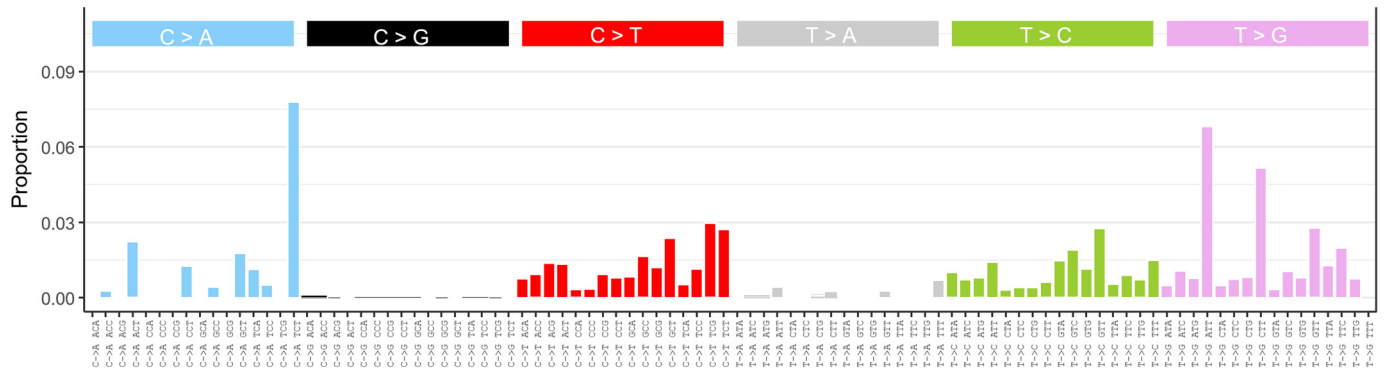


Supplementary Fig. 4. Copy number plots from WGS data (individual samples). For each sample the y-axis represents the normalized depth ($\log_2(\text{depth}/\text{median depth across genome})$). The x-axis represents the position among the 19 autosomes as illustrated in the legend. The analysis shows that *Pole*^{P286R} does not result in large-scale chromosomal copy-number instability. A) Parental mouse strains. B) Representative passage 30 MEF line clones. Some rearrangements occur during passaging of MEFs but these are qualitatively the same among wild-type and mutant MEFs. C) Tumors from *Pole*^{P286R/+} mice. D) Tumors from *Pole*^{P286R/LSL} (hemizygous) mice.

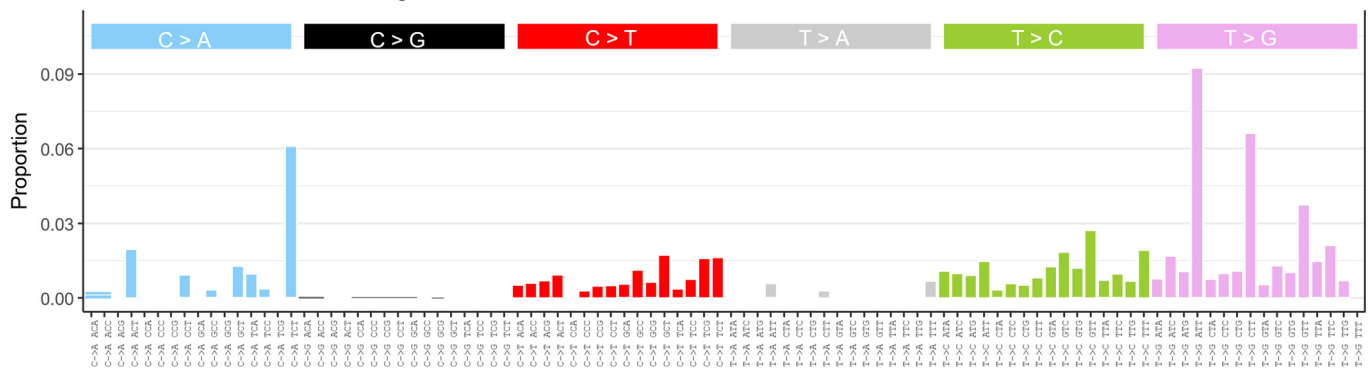
Supplementary Figure 5.

P286R/+ MEF line clones

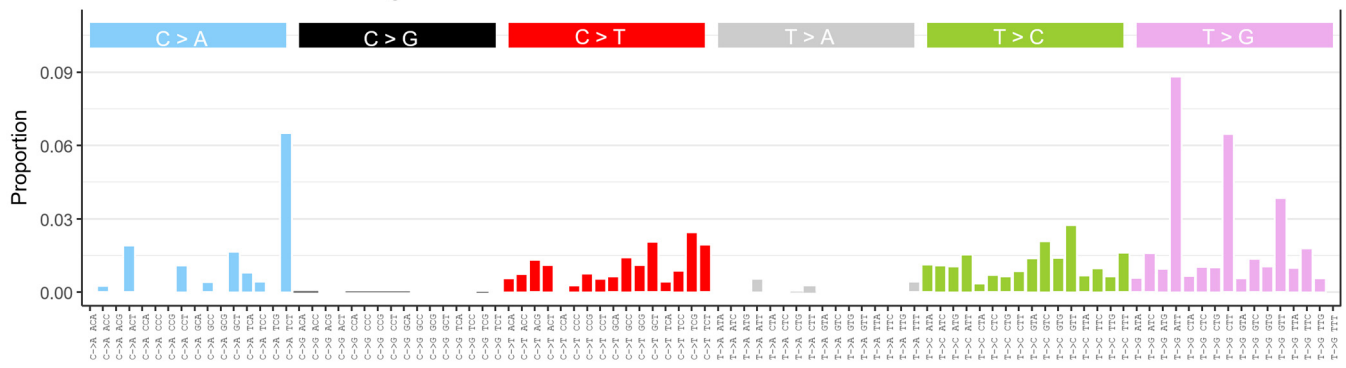
P286R/+ P15 MEF A3-3 single cell clone



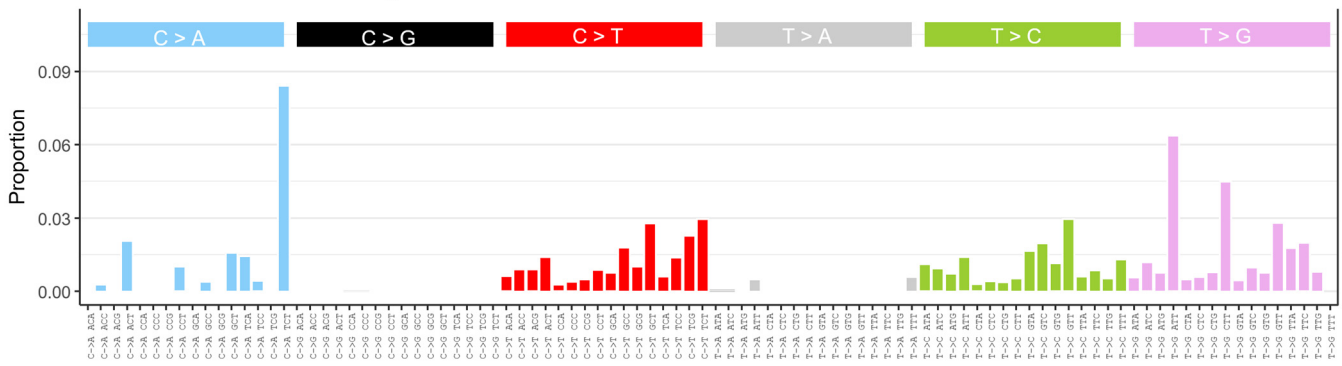
P286R/+ P30 MEF A3-3 single cell clone



P286R/+ P15 MEF A3-6 single cell clone



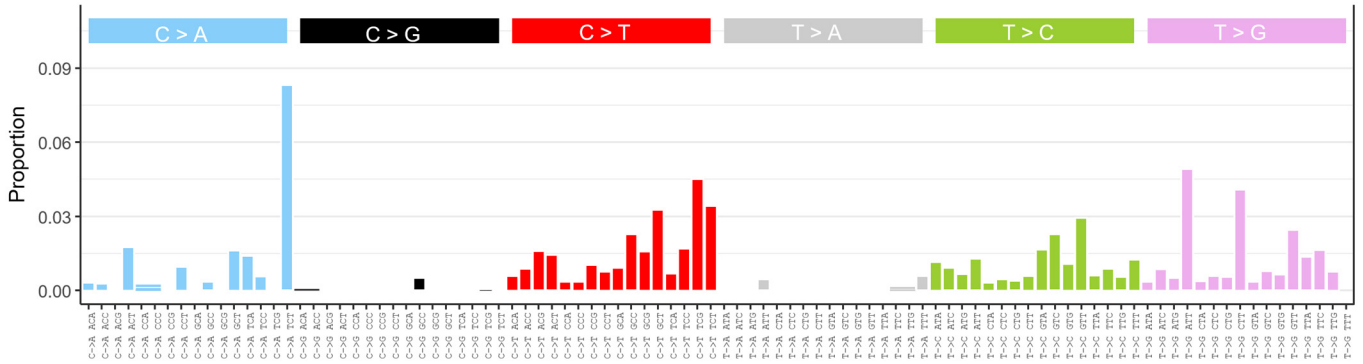
P286R/+ P30 MEF A3-6 single cell clone



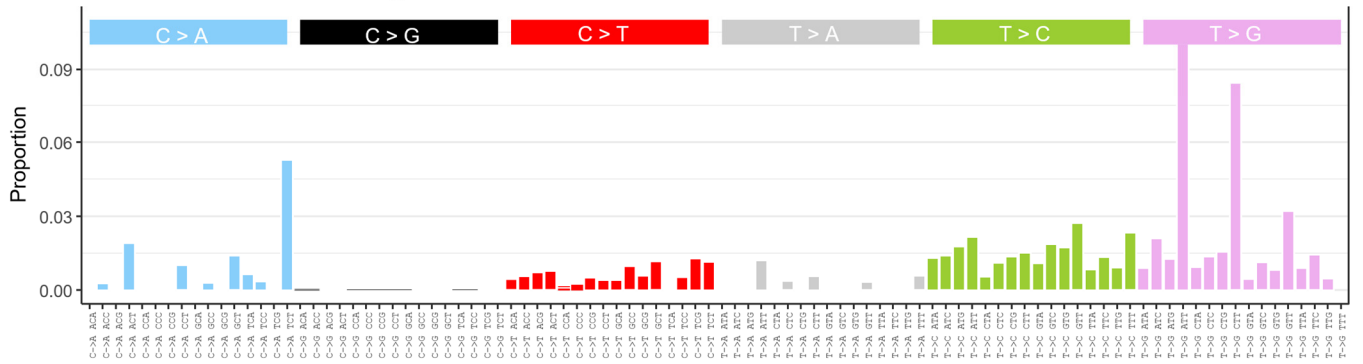
Supplementary Fig. 5

P286R/+ MEF line clones *continued*

P286R/+ P15 MEF A3-8 single cell clone

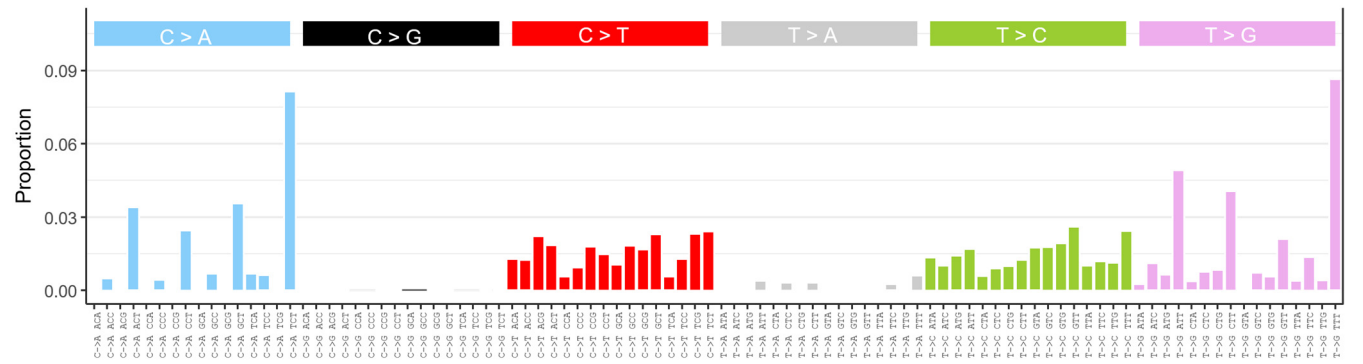


P286R/+ P30 MEF A3-8 single cell clone

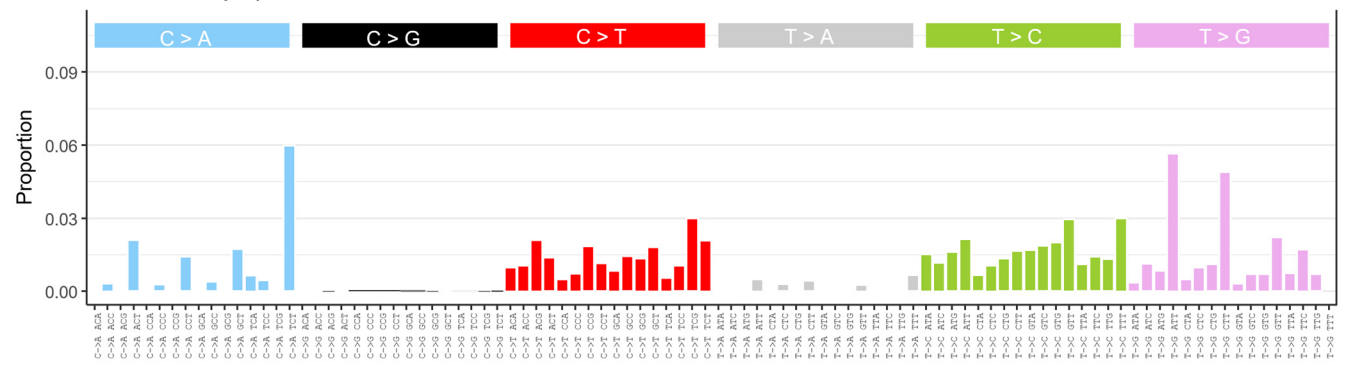


P286R/LSL (hemizygous) tumors

P286R/LSL lung adenocarcinoma 5R

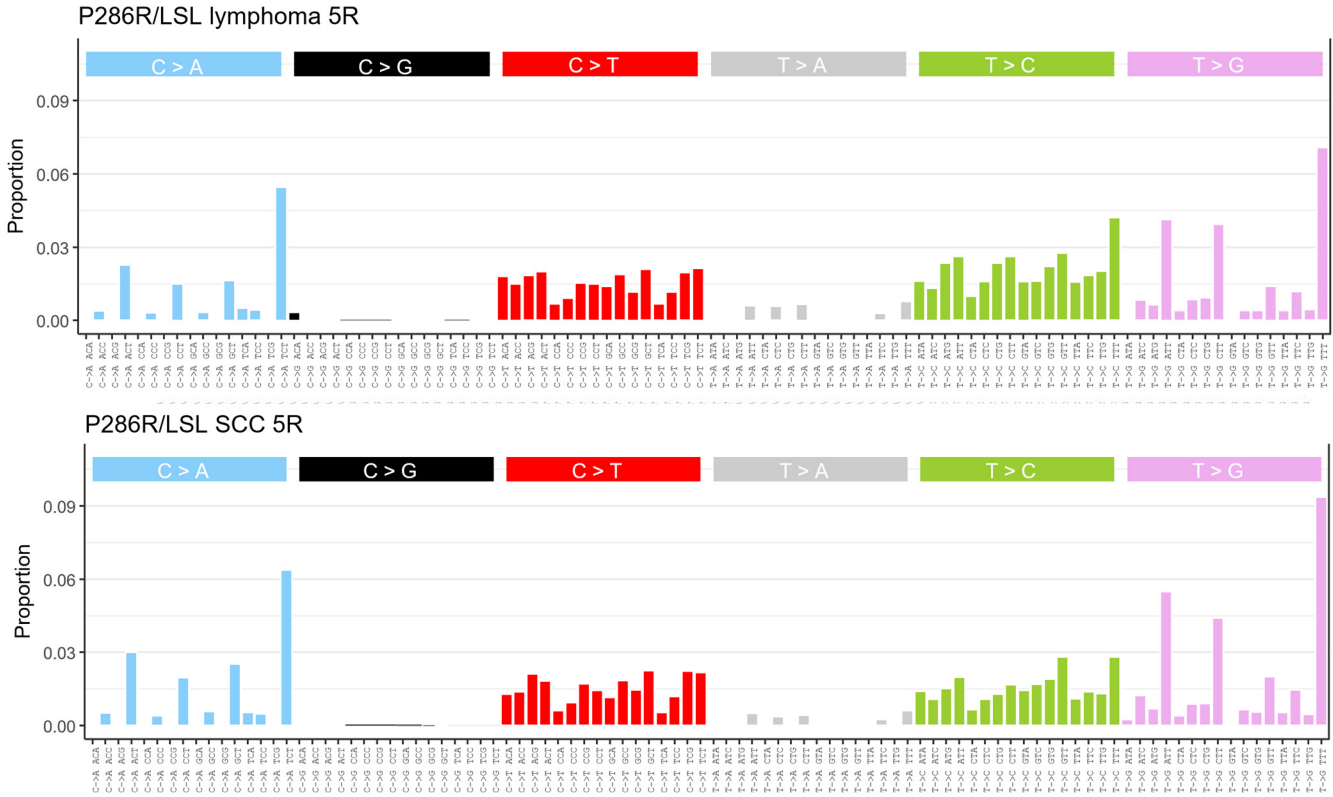


P286R/LSL lymphoma 1R



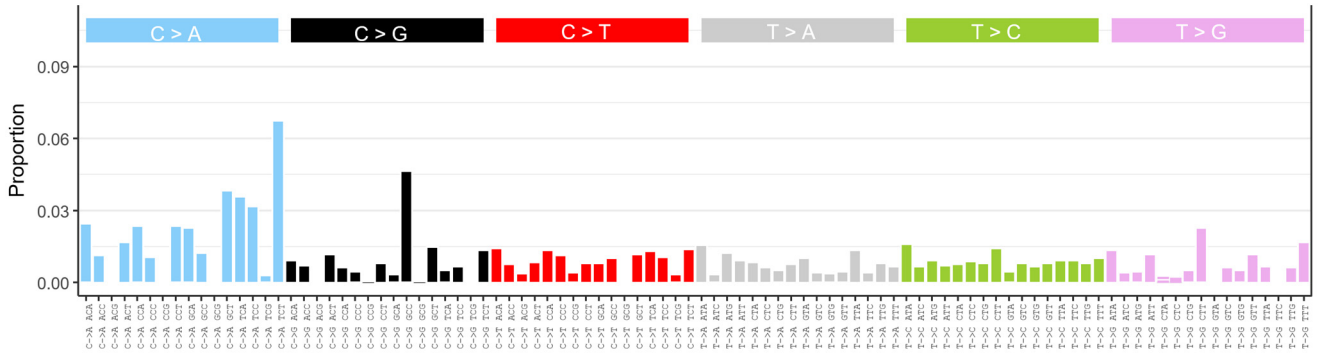
Supplementary Fig. 5

P286R/LSL (hemizygous) tumors *continued*

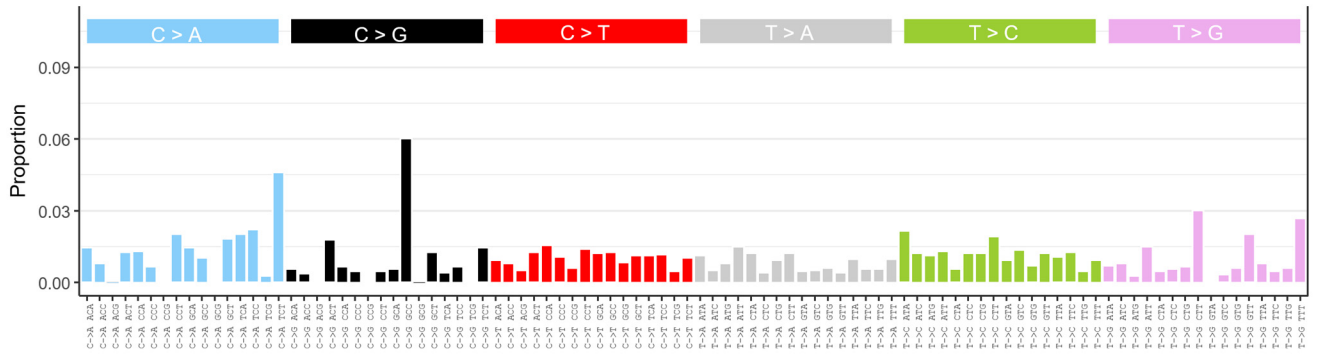


+/+ MEF line clones

+/+ P30 MEF C3-4 single cell clone

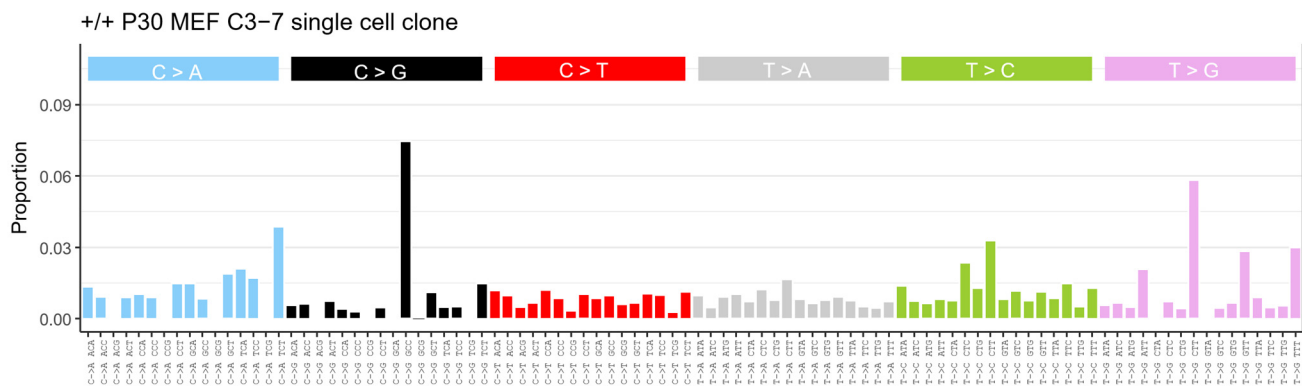


+/+ P30 MEF C3-5 single cell clone



Supplementary Fig. 5

+/+ MEF line clones *continued*



Supplementary Fig. 5. Whole genome sequencing-derived mutational signatures in *Pole*^{P286R/+} (heterozygous) MEFs and tumors, and (hemizygous) *Pole*^{P286R/LSL} tumors, and wild-type (*+/+*) MEFs. The 96 possible mutation types based on trinucleotide context are shown for each sample that underwent WGS in this study.