Supplementary Information

Paatero et al. "Junction-based lamellipodia drive endothelial cell rearrangements in vivo via a VE-cadherin-F-actin based oscillatory cell-cell interactions"





a) Quantification of the fraction of multicellular DLAV segments using Tg(BAC(*cdh5:cdh5-ts*)) embryos during development. n=26 DLAV segments (8 embryos). b) Quantification of the duration from anastomosis until final conversion into a multicellular tube. Black line is median. n=14 segments (8 embryos). c) Quantification of DLAV segments carrying blood flow during development. n=26 segments (8 embryos). hpf, hours post-fertilization.



Supplementary Figure 2. Validation of the rat Ve-cadherin antibody in zebrafish embryo whole-mount immunofluorescence analysis.

a-h) Confocal images of a Tg(*kdrl:EGFP*^{s843}) wild-type sibling (a-d) or a VE-cad null mutant embryo (Tg(*kdrl:EGFP*^{s843});*cdh5*^{ubs8/ubs8}) (e-h), stained for VE-cadherin (rat anti-Cdh5) and ZO1. a and e shows the merged channels, b-d and e-h the individual channels. Arrow points to an endothelial cell-cell junction. Scale bar 20 μ m.



Supplementary Figure 3. Analysis of F-actin oscillations of the remodeling junctions.

a) Kymograph across the junction in dorsal aorta in EGFP-UCHD fish (data generated as in Fig. 3). Solid arrow denotes forward movement. b) Intensity plotting of EGFP-UCHD kymographs. c) Averaged autocorrelation functions of EGFP-UCHD intensity. DMSO, n=14 kymographs; Latrunculin B, n=16 kymographs; NSC23766, n=16 kymographs. d) Comparison of autocorrelation lifetime (tau parameter). Means and standard deviations are plotted. ANOVA and Dunnet's post-hoc test was used.



a) Sequencing chromatogram of wild-type *ve-cadherin* (encoded by *cdh5* gene) sequence (exon 12) and respective truncation mutant $cdh5^{ubs25}$ sequence. b) The wild-type and mutant DNA sequences and their respective translations. The $cdh5^{ubs25}$ mutation leads to a premature stop. c) Schematic illustration of the domains in both full-length wild-type VE-cad and in truncated VE-cad (Cdh5^{ubs25}). d) Example of genotyping PCR and different $cdh5^{ubs25}$ allelic combinations. e) Plot showing colocalization of ZO1 and truncated Cdh5 at endothelial cell junctions. Pearson correlation coefficients of colocalization analysis of anti-VEC and anti-ZO1 in heterozygote ($cdh5^{ubs25+/.}$) and homozygote ($cdh5^{ubs25/ubs25}$) VE-cad mutants. $cdh5^{ubs25/4}$, n=30 cell-cell junctions (2 embryos) ; $cdh5^{ubs25/ubs25}$, n=20 cell-cell junctions (1 embryo).n.s., non-significant (p=0.75, non-parametric Mann-Whitney test). f) Quantitation of the first JBL to emergence of new JBL in the same spot was measured ; wild-type n=15 (2 embryos) and $cdh5^{ubs25/ubs25}$ n=36 (4 embryos). All embryos carried EGFP-UCHD transgene Tg(*fli:GFF*; *UAS:EGFP-UCHD*). Non-parametric Mann-Whitney statistical test was used.



Supplementary Figure 5. *Phenotypic analysis of the ve-cad truncation mutant zebrafish.* a) Table of blood flow phenotype of 56hpf VE-cadherin null mutant ($cdh5^{ubs8/ubs8}$) and the truncation mutant ($cdh5^{ubs8/ubs8}$) embryos. b) Stereomicroscope images of the embryos. Arrows point to pericardiac area, pronounced oedema is observed in mutant embryos. Scale bar, 400µm. c) Fluorescence images of Tg($kdrl:EGFP^{s843}$) embryos at 48hpf. Arrow points of enlarged cardinal vein in $cdh5^{ubs25}$ homozygote embryo. Scale bar, 200 µm. d) Confocal images illustrating the tip cell /stalk cell disconnection (arrow) in the $cdh5^{ubs25}$ homozygote embryo. In normal SeA the connection between tip cell and stalk cell is intact. Scale bar, 20 µm. e) Quantification of tip cell stalk cell disconnection; $cdh5^{ubs25/+}$, n=7 SeA; $cdh5^{ubs25/ubs25}$, n=36 SeA; wild-type n=22 SeA.