1	<b>Stable Expression</b>	of Modified Green	Fluorescent Protein	in Group E	<b>B</b> Streptococcus
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## 2 to Enable Visualization in Experimental Systems

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## 10 SUPPLEMENTAL MATERIAL

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27 Supplementary FIG. S1. Media used for bacteriology exhibited different levels of autofluorescence at the excitation emission wavelengths used for the detection of 28 29 GFPmut3 (A). Autofluorescence detected in THB medium remains stable in the absence 30 of bacteria (squares) but in the presence of GBS (triangles) declines in a manner inversely 31 proportional to culture density (shown in Fig 6). Negative control correction of 32 fluorescence intensity can be used to adjust for the autofluorescence background 33 interference detected in THB medium to provide a fluorescence profile (C) similar to that 34 detected using fluorescence polarization intensity (shown in Fig 6C).



35 Supplementary FIG. S2. Effect of increasing concentrations of sodium hypochlorite 36 on GBS cell morphology and fluorescence. GBS strain 874391 (2, 4, 5) were rendered 37 non-viable following exposure to sodium hypochlorite  $\geq 0.02\%$ , according to colony 38 count assays (Fig 7B). However, GBS continued to emit high levels of fluorescence at 39 this level of sodium hypochlorite (upper panels). Sodium hypochlorite 0.4% abolished 40 the fluorescence and destroyed the bacteria (bottom panel), while 0.1% attenuated 41 fluorescence in intact GBS cells (Fig 7A), which were rendered non-viable, according to 42 colony count assays.



Supplementary FIG. S3. Relative quantitation of cell-associated fluorescence in 43 44 assays comparing the adhesion of WT GBS and CovR mutant to human cells. 45 Inoculation of 5637 uroepithelial cells with WT and CovR mutant GBS (3) (both carrying 46 plasmid pGU2664) shows the attenuated adherence phenotype of the CovR mutant based 47 on the fluorescence detection of GFPmut3; representative cell masks used to quantitate 48 pixels is shown in (A). Quantitation of the fluorescence signals in areas co-located with 49 the human cells (cell-associated fluorescence) was achieved using ImageJ software 1.51 50 (1), and compared to acellular areas (slide-associated fluorescence); six fields-of-view 51 (one representative shown for WT and mutant) were analyzed to generate quantitative 52 data and revealed significantly more fluorescence signal co-located with epithelial cells 53 inoculated with WT GBS versus the CovR mutant (B). Scale bars =  $50 \mu m$ ; % shown is 54 cell-associated fluorescence detected for that image.

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56	Supplementary Movie. S1. Visualization of adhesion of GBS to human cells. 5637				
57	uroepithelial cells were inoculated with GU2666 (GFP+) containing plasmid pGU2664				
58	and video was captured after 2 h of static incubation and subsequent initiation of flow.				
59	The video (30 s total, looped 5x) shows human cells with bound GBS (red arrows) amid a				
60	background of media and bacteria flowing through the chamber. The video highlights				
61	many chains of non-adhered GBS passing in the flow-through (red box). The video				
62	represents ~0.8 chamber-volumes of media using medium flow conditions, as described				
63	in Materials and Methods.				
64					
65	Supplemental References				
66	1.	Schneider, C. A., W. S. Rasband, and K. W. Eliceiri. 2012. NIH Image to			
67		ImageJ: 25 years of image analysis. Nat. Methods 9:671-675.			
68	2.	Sullivan, M. J., B. M. Forde, D. W. Prince, D. S. Ipe, N. L. Ben Zakour, M. R.			
69		Davies, G. Dougan, S. A. Beatson, and G. C. Ulett. 2017. Complete Genome			
70		Sequence of Serotype III Streptococcus agalactiae Sequence Type 17 Strain			
71		874391. Genome Announcements 5.			
72	3.	Sullivan, M. J., S. Y. Leclercq, D. S. Ipe, A. J. Carey, J. P. Smith, N. Voller,			
73		A. W. Cripps, and G. C. Ulett. 2017. Effect of the Streptococcus agalactiae			
74		Virulence Regulator CovR on the Pathogenesis of Urinary Tract Infection. J.			
75		Infect. Dis. <b>215:</b> 475-483.			
76	4.	Takahashi, S., Y. Nagano, N. Nagano, K. Fujita, F. Taguchi, and Y.			
77		Okuwaki. 1993. Opsonisation of group B streptococci and restriction			
78		endonuclease digestion patterns of their chromosomal DNA. J. Med. Microbiol.			
79		<b>38:</b> 191-196.			
80	5.	Takahashi, S., Y. Nagano, N. Nagano, O. Hayashi, F. Taguchi, and Y.			
81		Okuwaki. 1995. Role of C5a-ase in group B streptococcal resistance to			
82		opsonophagocytic killing. Infect. Immun. 63:4764-4769.			
83					