

Supplemental information

A colicin M-type bacteriocin from *Pseudomonas aeruginosa* targeting the HxuC heme receptor requires a novel immunity partner

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Table S1. Strains used in this study.

Species	Strain	Characteristics/Source	Growth inhibition by <i>PaeM4</i> ^a	Presence <i>paeM4</i> locus	References
<i>Escherichia coli</i>	BL21(DE3)	Production of His-tagged bacteriocins			Novagen
	DH5 α	Plasmid propagation			Invitrogen
<i>Pseudomonas aeruginosa</i>	CF_PA1	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA4	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA12	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA17	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA20	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA28	CF patient	+	No	(1)
	CF_PA33	CF patient	+	No	(1)
	CF_PA36	CF patient	+	No	(1)
	CF_PA37	CF patient	+	No	(1)
	CF_PA38	CF patient	+	No	(1)
	CF_PA41	CF patient	+	<i>pmiC</i> (orphan)	(1)
	CF_PA57	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA58	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA59	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA62	CF patient	+	No	(1)
	CF_PA79	CF patient	+	No	(1)
	CF_PA86	CF patient	+	No	(1)
	CF_PA87	CF patient	-	<i>pmiC</i> (orphan)	(1)
	CF_PA94	CF patient	+	No	(1)
	CF_PA103	CF patient	+	No	(1)
	CF_PA116	CF patient	+	No	(1)
	CF_PA117	CF patient	+	No	(1)
	Pr335	Hospital environment	+	No	(2)
	W5 Aug28	River water	+	No	(3)
	LMG 2107	Shallow well water	+	No	(2)
	W15 Apr4	River water	+	No	(4)
	Bo559	Burn wound	+	No	(2)
Jp97	Sea water	T	No	(5)	
Jp100	Sea water	T	No	(4)	
TA08	Sputum	+	No	(5)	
Is580	Burn wound	+	No	(2)	
A19	Wound	+	No	(5)	
W15 Dec4	River water	+	No	(3)	
W15 Oct31	River water	+	No	(4)	
Br776	Throat	-	<i>pmiC</i> (orphan)	(5)	
Br670	Sputum	+	No	(2)	
Br906	Nose	+	No	(2)	

	Li004	CF patient	-	<i>pmiC</i> (orphan)	(2)
	Is579	Burn wound	-	<i>pmiC</i> (orphan)	(2)
	Br908	Throat	+	No	(2)
	CPHL 6750	Urine	+	No	(5)
	CPHL 2000	Wound	+	<i>pmiC</i> (orphan)	(5)
	Us448	Urine	+	No	(2)
	TuD47	Ascite	-	<i>pmiC</i> (orphan)	(2)
	Br262	Tap water room burn wound center	+	No	(5)
	Bu002	Wound	-	<i>pmiC</i> (orphan)	(5)
	Pr317	Burn	-	<i>pmiC</i> (orphan)	(5)
	Pr334	Hospital environment	-	<i>pmiC</i> (orphan)	(5)
	W15 Dec14	River water	-	<i>pmiC</i> (orphan)	(3)
	Is582	Burn wound	-	<i>pmiC</i> (orphan)	(5)
	Is573	Burn wound	-	<i>pmiC</i> (orphan)	(2)
	So092	Burn wound	-	<i>pmiC</i> (orphan)	(2)
	Aa246	Burn wound	+	No	(2)
	Br678	Burn wound	-	<i>pmiC</i> (orphan)	(5)
	So123	Hospital environment	+	No	(5)
	Br817	Wound	-	<i>pmiC</i> (orphan)	(5)
	Bo562	Blood	-	<i>pmiC</i> (orphan)	(5)
	So098	Wound	-	<i>pmiC</i> (orphan)	(2)
	Br993	Sputum	+	No	(2)
	Br229	Hospital	+	No	(2)
	Br680	Burn wound	+	No	(2)
	PA7	Wound	+	No	(6)
	Us451	Burn wound	-	<i>paeM4-pmiC</i>	(5)
	ATCC 27853	Blood	+	No	(2)
	Bu007	Burn wound	-	<i>pmiC</i> (orphan)	(2)
	PAO1	Wound	+	No	(7)
	PA229	Clinical isolate	-	<i>pmiC</i> (orphan)	(8)
	PA62	Clinical isolate	-	<i>pmiC</i> (orphan)	(9)
	PA63	Clinical isolate	-	<i>pmiC</i> (orphan)	(10)
	PABL056	Blood	-	<i>pmiC</i> (orphan)	(11)
	HB15	Sputum	-	<i>pmiC</i> (orphan)	(12)
	SCV20265	CF patient	-	<i>paeM4-pmiC</i>	(13)
	LMG 1242	Source unknown	-	<i>pmiC</i> (orphan)	(14)
	LMG 1272	Cultivated mushroom	+	No	(10)
	PA45	Blood	+	No	(15)
Transposon mutants <i>P. aeruginosa</i> PAO1					
	PA0151	PAO1 mutant hit in PA0151	+	No	(16)
	PA0192	PAO1 mutant hit in PA0192	+	No	(16)
	PA0434	PAO1 mutant hit in PA0434	+	No	(16)
	PA0470	PAO1 mutant hit in PA0470 (<i>fiuA</i>)	+	No	(16)

	PA0674	PAO1 mutant hit in PA0674 (<i>pigC</i>)	+	No	(16)
	PA0781	PAO1 mutant hit in PA0781	+	No	(16)
	PA0931	PAO1 mutant hit in PA0931 (<i>pirA</i>)	+	No	(16)
	PA1271	PAO1 mutant hit in PA1271	+	No	(16)
	PA1302	PAO1 mutant hit in PA1302 (<i>hxuC</i>)	-	No	(16)
	PA1322	PAO1 mutant hit in PA1322 (<i>pfuA</i>)	+	No	(16)
	PA1365	PAO1 mutant hit in PA1365	+	No	(16)
	PA1613	PAO1 mutant hit in PA1613	+	No	(16)
	PA1910	PAO1 mutant hit in PA1910 (<i>femA</i>)	+	No	(16)
	PA1922	PAO1 mutant hit in PA1922	+	No	(16)
	PA2057	PAO1 mutant hit in PA2057	+	No	(16)
	PA2070	PAO1 mutant hit in PA2070	+	No	(16)
	PA2089	PAO1 mutant hit in PA2089	+	No	(16)
	PA2289	PAO1 mutant hit in PA2289	+	No	(16)
	PA2335	PAO1 mutant hit in PA2335 (<i>optO</i>)	+	No	(16)
	PA2398	PAO1 mutant hit in PA2398 (<i>fpvA</i>)	+	No	(16)
	PA2466	PAO1 mutant hit in PA2466 (<i>optS</i>)	+	No	(16)
	PA2590	PAO1 mutant hit in PA2590	+	No	(16)
	PA2688	PAO1 mutant hit in PA2688 (<i>pfeA</i>)	+	No	(16)
	PA2911	PAO1 mutant hit in PA2911	+	No	(16)
	PA3268	PAO1 mutant hit in PA3268	+	No	(16)
	PA3408	PAO1 mutant hit in PA3408 (<i>hasR</i>)	+	No	(16)
	PA3790	PAO1 mutant hit in PA3790 (<i>oprC</i>)	+	No	(16)
	PA3901	PAO1 mutant hit in PA3901 (<i>fecA</i>)	+	No	(16)
	PA4156	PAO1 mutant hit in PA4156	+	No	(16)
	PA4168	PAO1 mutant hit in PA4168 (<i>fpvB</i>)	+	No	(16)
	PA4221	PAO1 mutant hit in PA4221 (<i>fpvA</i>)	+	No	(16)
	PA4514	PAO1 mutant hit in PA4514 (<i>piuA</i>)	+	No	(16)
	PA4675	PAO1 mutant hit in PA4675	+	No	(16)
	PA4710	PAO1 mutant hit in PA4710 (<i>phuR</i>)	+	No	(16)
	PA4837	PAO1 mutant hit in PA4837	+	No	(16)
	PA4897	PAO1 mutant hit in PA4897 (<i>optI</i>)	+	No	(16)

^a Growth inhibition by Paem4, +; no growth inhibition by Paem4, -. A non-transparent halo indicative of reduced cell density compared with the cell lawn is marked with T (turbid halo).

Table S2. List of plasmids used in this study.

Plasmid	Characteristics	Reference
pET28a(+)	pBR322 origin, His-tag/thrombin/T7 tag; Km ^R	Novagen
pJB3Tc20	Broad-host-range cloning vector, IncP; Ap ^R , Tc ^R	(17)
pMMB67EH	Broad-host-range cloning vector, IncQ; Gm ^R	(18)
pCMPG6250	pET28a(+) with 866-bp PCR-amplified fragment containing <i>paeM</i> (locus tag ERS445055_00256) from <i>P. aeruginosa</i> NCTC 10332 cloned in NcoI/XhoI	(19)
pCMPG6252	pJB3Tc20 with 467-bp PCR-amplified containing <i>pmiA</i> (no annotation of immunity gene downstream of <i>paeM</i>) from <i>P. aeruginosa</i> NCTC 10332 cloned in PstI/EcoRI	(19)
pCMPG6283	pET28a(+) with 1025-bp PCR-amplified fragment containing <i>paeM4</i> (locus tag Q057_04089) from <i>P. aeruginosa</i> BL03 cloned in NcoI/XhoI	This study
pCMPG6287	pJB3Tc20 with 739-bp PCR-amplified containing <i>pmiC</i> (locus tag Q057_04090) from <i>P. aeruginosa</i> BL03 cloned in PstI/EcoRI	This study
pCMPG6293	pMMB67EH with 2727-bp PCR-amplified fragment containing <i>hxuC</i> (locus tag PA1302) from <i>P. aeruginosa</i> PAO1 cloned in EcoRI/HindIII	This study

References

1. Dingemans J, Ye L, Hildebrand F, Tontodonati F, Craggs M, Bilocq F, De Vos D, Crabbé A, Van Houdt R, Malfroot A, Cornelis P. 2014. The deletion of TonB-dependent receptor genes is part of the genome reduction process that occurs during adaptation of *Pseudomonas aeruginosa* to the cystic fibrosis lung. *Pathog Dis* 71:26-38.
2. van Belkum A, Soriaga LB, LaFave MC, Akella S, Veyrieras JB, Barbu EM, Shortridge D, Blanc B, Hannum G, Zambardi G, Miller K, Enright MC, Mugnier N, Brami D, Schicklin S, Felderman M, Schwartz AS, Richardson TH, Peterson TC, Hubby B, Cady KC. 2015. Phylogenetic Distribution of CRISPR-Cas Systems in Antibiotic-Resistant *Pseudomonas aeruginosa*. *mBio* 6:e01796-15.
3. Pirnay JP, Matthijs S, Colak H, Chablain P, Bilocq F, Van Eldere J, De Vos D, Zizi M, Triest L, Cornelis P. 2005. Global *Pseudomonas aeruginosa* biodiversity as reflected in a Belgian river. *Environ Microbiol* 7:969-980.
4. Pirnay JP, Blasdel BG, Bretaudeau L, Buckling A, Chanishvili N, Clark JR, Corte-Real S, Debarbieux L, Dublanchet A, De Vos D, Gabard J, Garcia M, Goderdzishvili M, Górski A, Hardcastle J, Huys I, Kutter E, Lavigne R, Merabishvili M, Olchawa E, Parikka KJ, Patey O, Pouilot F, Resch G, Rohde C, Scheres J, Skurnik M, Vaneechoutte M, Van Parys L, Verbeken G, Zizi M, Van den Eede G. 2015. Quality and safety requirements for sustainable phage therapy products. *Pharm Res* 32:2173-2179.
5. Pirnay JP, Bilocq F, Pot B, Cornelis P, Zizi M, Van Eldere J, Deschaght P, Vaneechoutte M, Jennes S, Pitt T, De Vos D. 2009. *Pseudomonas aeruginosa* population structure revisited. *PLoS One* 4:e7740.
6. Roy PH, Tetu SG, Larouche A, Elbourne L, Tremblay S, Ren Q, Dodson R, Harkins D, Shay R, Watkins K, Mahamoud Y, Paulsen IT. 2010. Complete genome sequence of the multiresistant taxonomic outlier *Pseudomonas aeruginosa* PA7. *PLoS One* 5:e8842.
7. Stover CK, Pham XQ, Erwin AL, Mizoguchi SD, Warrenner P, Hickey MJ, Brinkman FS, Hufnagle WO, Kowalik DJ, Lagrou M, Garber RL, Goltry L, Tolentino E, Westbrook-Wadman S, Yuan Y, Brody LL, Coulter SN, Folger KR, Kas A, Larbig K, Lim R, Smith K, Spencer D, Wong GK, Wu Z, Paulsen IT, Reizer J, Saier MH, Hancock RE, Lory S, Olson MV. 2000. Complete genome sequence of *Pseudomonas aeruginosa* PAO1, an opportunistic pathogen. *Nature* 406:959-964.
8. Buyck JM, Luyckx C, Muccioli GG, Krause KM, Nichols WW, Tulkens PM, Van Bambeke F. 2017. Pharmacodynamics of ceftazidime/avibactam against extracellular and intracellular forms of *Pseudomonas aeruginosa*. *J Antimicrob Chemother* 72:1400-1409.
9. Defraigne V, Verstraete L, Van Bambeke F, Anantharajah A, Townsend EM, Ramage G, Corbau R, Marchand A, Chaltin P, Fauvart M, Michiels J. 2017. Antibacterial Activity of 1-[(2,4-Dichlorophenethyl)amino]-3-Phenoxypropan-2-ol against Antibiotic-Resistant Strains of Diverse Bacterial Pathogens, Biofilms and in Pre-clinical Infection Models. *Front Microbiol* 8:2585.
10. Ghequire MG, Dingemans J, Pirnay JP, De Vos D, Cornelis P, De Mot R. 2014. O serotype-independent susceptibility of *Pseudomonas aeruginosa* to lectin-like pyocins. *Microbiologyopen* 3:875-884.
11. Ozer EA, Allen JP, Hauser AR. 2012. Draft genome sequence of the *Pseudomonas aeruginosa* bloodstream isolate PABL056. *J Bacteriol* 194:5999.
12. Soares-Castro P, Marques D, Demyanchuk S, Faustino A, Santos PM. 2011. Draft genome sequences of two *Pseudomonas aeruginosa* clinical isolates with different antibiotic susceptibilities. *J Bacteriol* 193:5573.
13. Eckweiler D, Bunk B, Spröer C, Overmann J, Häussler S. 2014. Complete Genome Sequence of Highly Adherent *Pseudomonas aeruginosa* Small-Colony Variant SCV20265. *Genome Announc* 2:e01232-13.
14. Nakano K, Terabayashi Y, Shiroma A, Shimoji M, Tamotsu H, Ashimine N, Ohki S, Shinzato M, Teruya K, Satou K, Hirano T. 2015. First Complete Genome Sequence of *Pseudomonas aeruginosa*

- (Schroeter 1872) Migula 1900 (DSM 50071^T), Determined Using PacBio Single-Molecule Real-Time Technology. *Genome Announc* 3:e00932-15.
15. Segata N, Ballarini A, Jousson O. 2013. Genome Sequence of *Pseudomonas aeruginosa* PA45, a Highly Virulent Strain Isolated from a Patient with Bloodstream Infection. *Genome Announc* 1:e00289-13.
 16. Jacobs MA, Alwood A, Thaipisuttikul I, Spencer D, Haugen E, Ernst S, Will O, Kaul R, Raymond C, Levy R, Chun-Rong L, Guenther D, Bovee D, Olson MV, Manoil C. 2003. Comprehensive transposon mutant library of *Pseudomonas aeruginosa*. *Proc Natl Acad Sci U S A* 100:14339-14344.
 17. Blatny JM, Brautaset T, Winther-Larsen HC, Haugan K, Valla S. 1997. Construction and use of a versatile set of broad-host-range cloning and expression vectors based on the RK2 replicon. *Appl Environ Microbiol* 63:370-379.
 18. Christen M, Kulasekara HD, Christen B, Kulasekara BR, Hoffman LR, Miller SI. 2010. Asymmetrical distribution of the second messenger c-di-GMP upon bacterial cell division. *Science* 328:1295-1297.
 19. Ghequire MG, Kemland L, De Mot R. 2017. Novel immunity proteins associated with colicin M-like bacteriocins exhibit promiscuous protection in *Pseudomonas*. *Front Microbiol* 8:93.