Supplemental material

Wozniak et al., https://doi.org/10.1085/jgp.201812071

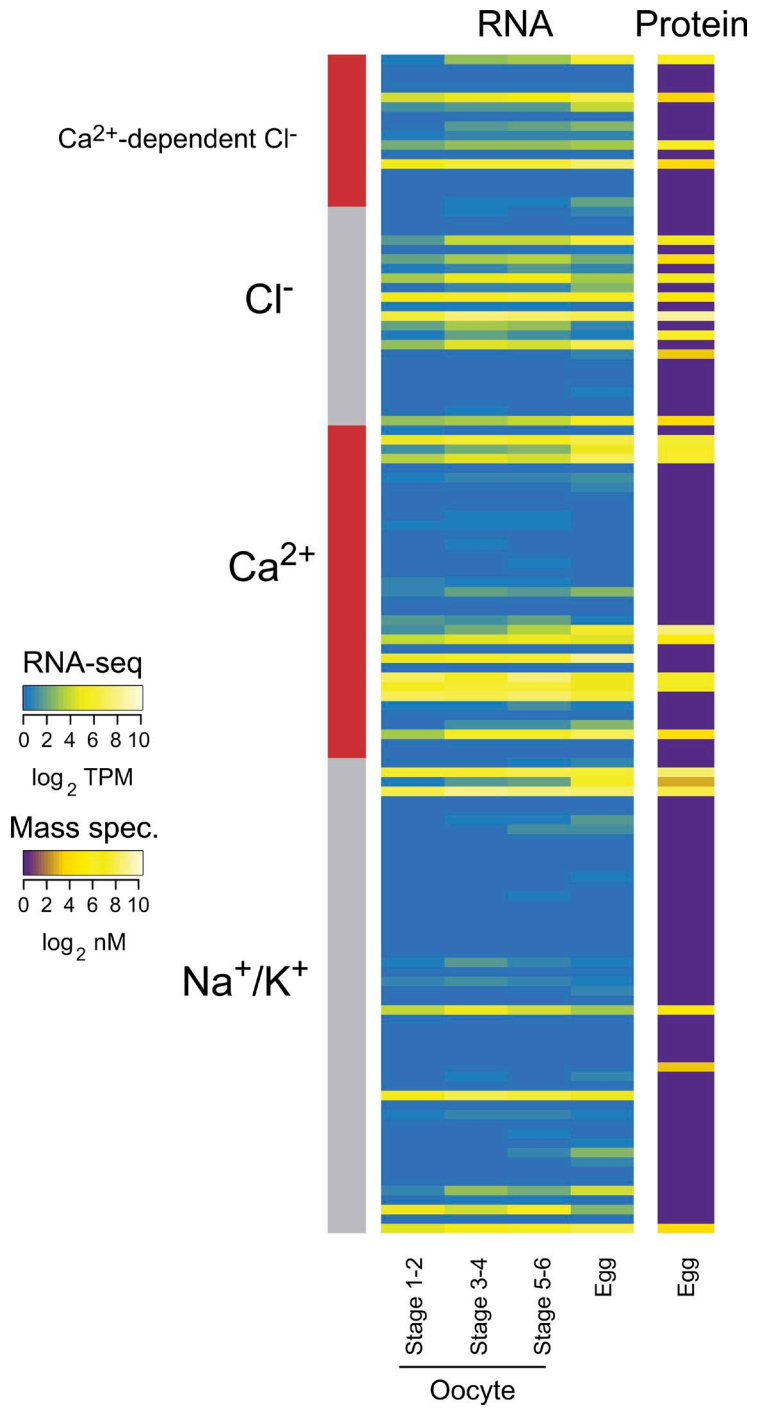

Figure S1. **Ion channel expression in *X. laevis* oocytes and eggs.** Left: Heatmaps showing RNA expression levels (based on RNA-seq from Session et al., 2016) as log2 transcripts per million (TPM). Right: Heatmaps showing protein concentrations (based on mass spectrometry from Wühr et al. 2014) in $\log_2$ nanomolar. Transcripts and proteins are grouped by channel type.

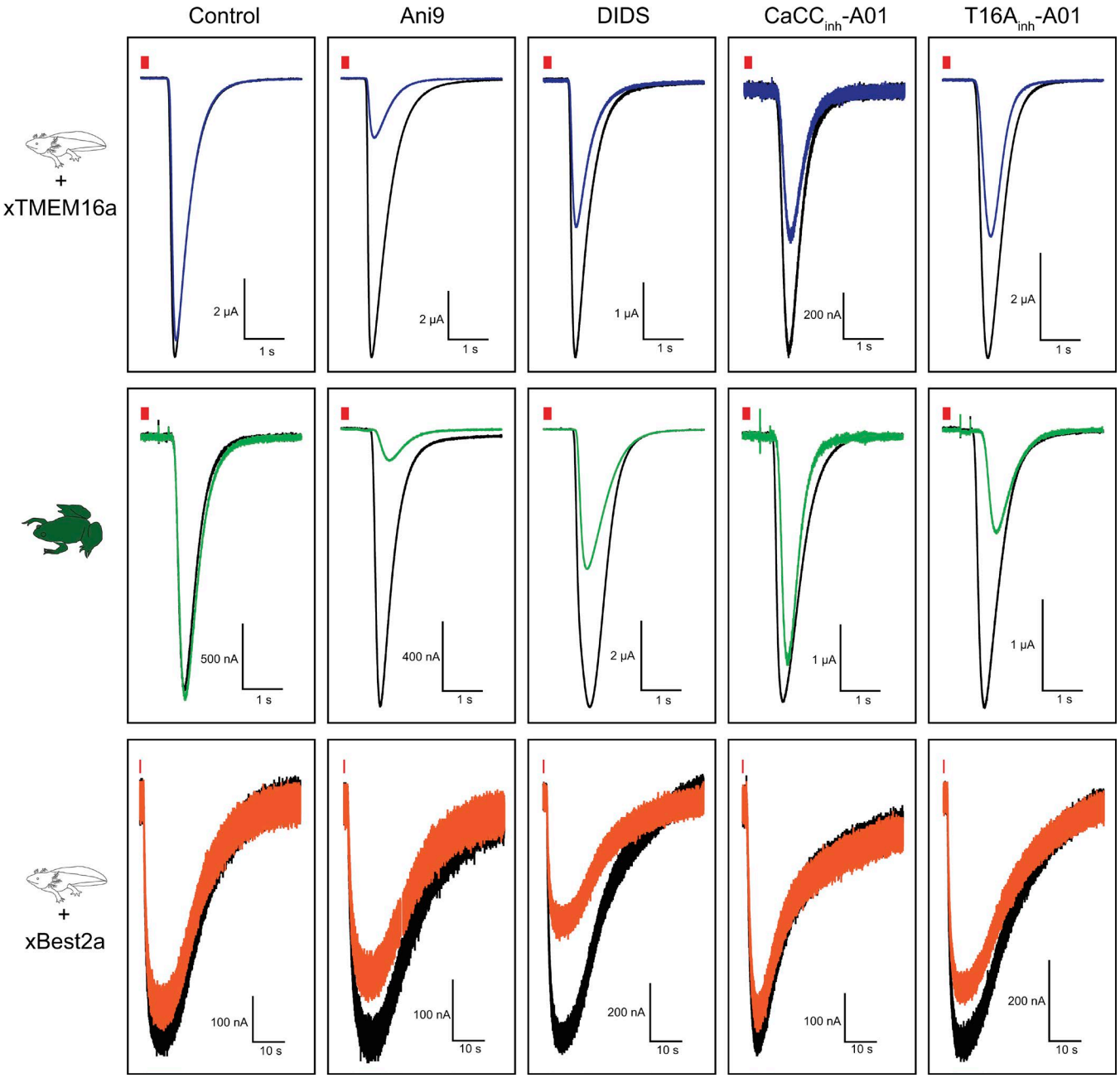

Figure S2. **IP_3-evoked currents in various inhibitors.** Representative current traces evoked by IP_3 uncaging in axolotl oocytes expressing xTMEM16A (top) or xBEST2A (bottom) and in wild-type *X. laevis* oocytes (middle). Shown are typical traces before (black) and after (colored) application of a control solution, Ani9, DIDS, CaCC$_{inh}$-A01, or T16A$_{inh}$-A01. Red bars denote the 250-ms UV exposure.

Table S1. **Inhibition of Ca^{2+}-activated current using Cl^- channel inhibitors**

	Control	10 µM MONNA	1 µM Ani9	30 µM T16a$_{inh}$-A01	10 µM CaCC$_{inh}$-A01	7.5 µM DIDS
xTMEM16A in axolotl oocytes	5 ± 3	72 ± 3	76 ± 5	46 ± 11	35 ± 7	46 ± 2
X. laevis oocytes	2 ± 4	87 ± 2	80 ± 4	45 ± 13	25 ± 11	47 ± 7
xBEST2A in axolotl oocytes	7 ± 4	10 ± 8	22 ± 7	26 ± 4	9 ± 3	29 ± 7

Mean ± SEM percentage of current inhibition seen for uncaging experiments is reported. The number of independent observations for each treatment is n = 8–16 for MONNA, n = 5–8 for Ani9, n = 6–7 for T16a$_{inh}$-A01, n = 6–10 for CaCC$_{inh}$-A01, and n = 6–7 DIDS.