## **Supplementary Information**



FIGURE S1. pCt regulates current amplitude of the TRAAK-TREK1 heterodimer. A-E, Representative whole-cell recordings from HEK cells expressing covalent tandems TREK1-TREK1 (A), TRAAK-TRAAK (B), TRAAK-TREK1 (C), TRAAK-TREK1pCt<sub>TRAAK</sub> (D) and TRAAKpCt<sub>TREK1</sub>-TREK1 (E). F, Current densities at 0 mV for the indicated channels. Data are presented as mean  $\pm$  SEM, \*p<0.05, the number of cells is indicated (one-way Anova with Tukey's test).



FIGURE S2. Fusion of pHluorin to TREK1 and TRAAK does not affect channel activity. A, D, Representative whole-cell recordings of HEK cells expressing TREK1 or TREK1-PH (A) and TRAAK or TRAAK-PH (D). B, E, Current density/voltage curves obtained from currents shown in A and D. C, F, Current densities at 0 mV, Data are presented as mean ± SEM. The number of cells is indicated (Student's t-test).





FIGURE S3. **Deletion or swapping of pCt does not affect channel cellular trafficking. A**, Schematic representation of pHluorin-tagged channels. An additional membrane-spanning segment was added between the channel and pHluorin, to expose pHluorin to the extracellular medium. **B**, Immunolabeling of wild-type channels expressed in HEK cells. Channels are in green. **C**, Representative images of pHluorin-tagged channels in live HEK cells. Acidification of the extracellular medium induces a quenching of pHluorin exposed at the plasma membrane without affecting fluorescence in internal cell compartments. Application of ionophores leads to the dissipation of the pH gradients, leading to a maximal signal at pH 7.4 and to a total fluorescence quenching at pH 6.

1



FIGURE S4. **TREK1 and TRAAK currents are not affected by strong depolarizations in the absence of VSP co-expression.** A, D, Representative perforated patch-clamp recordings from HEK cells expressing TREK1 (A) and TRAAK (D). Same protocol as in Fig. 6A and Fig. 7A. B, E, TREK1 (B) and TRAAK (E) currents before (a) and after (b) the +120 mV/40 s depolarizing pulse. C, E, Current inhibition (%) at 0 mV in cells expressing VSP *versus* cells not expressing VSP. Data are presented as mean  $\pm$  SEM, \*\*\*p<0.001, the number of cells is indicated (Student's t-test).



FIGURE S5. A, B, D, E, Representative patch-clamp recordings from HEK cells expressing wild-type (A, D) or mutant channels (B, E) in the presence of 10  $\mu$ M fluoxetine on basal currents. C, F, Fraction of fluoxetine-sensitive current. Data are presented as mean  $\pm$  SEM. \*\*\*p<0.001, the number of cells is indicated (Student's t-test).

2