

SUPPORTING INFORMATION

Unveiling sequential late-stage methyltransferase reactions in the meleagrins/oxalins biosynthetic pathway

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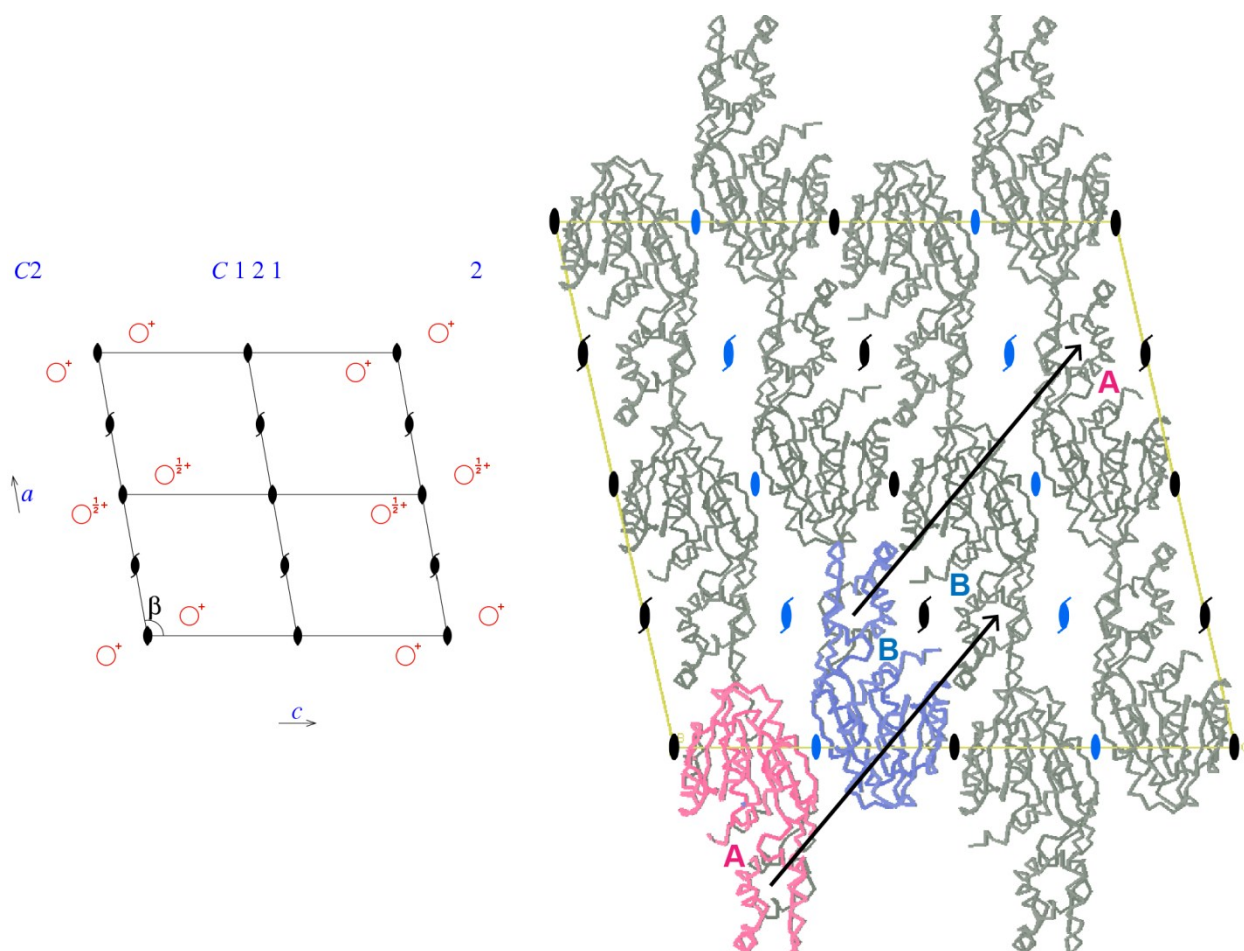
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Figure S1.

OxaG	1	MTRATNFTELYAGKGILETYMIAEKITRYFTRDLIELSGLLESELSPLKLLDLACGTGVV	60
		MTRATNFTELYAGKGIL+TYM+AEKITRY+T+DLI+LSGL ES L+PL +LDLACGTGVV	
RoqN	1	MTRATNFTELYAGKGILD TYMVAEKITRYYTQDLIQLSGLSESSLTPLVILD LACGTGVV	60
OxaG	61	SERLHEMLASKAPASWELICGDISAELTGHVKRKIIIEEGWTNSSARVMDAQNTE LATAEL	120
		S+ LH+ML + +WEL CGDIS ELTGHVK+KI+E GW NS A+V+DAQNTE L T	
RoqN	61	SDALHDMLNFQPKGNWELTCGDIS TELTGHVKQKILERGWENSI AKVVDAQNTE LPTGHY	120
OxaG	121	THVFAALAWTSFPD TYAALKDSL RILRPGGTLTISTWQKTEWLG VLEAAVKTIPTRLPFP	180
		THVFAALA+TSFPD TYAA+K+ +RIL+PGGTLTISTWQ+TEW L V+EAAV IP LPFP	
RoqN	121	THVFAALAF TSFPD TYAAMKEVMRILQP GGTLTISTWQ RTEWLAVVEAAVAIIPADLPFP	180
OxaG	181	TTKEFMSCMNP GWD DEN YVRGRLEEAGFVHVYSTTISKEFQISTADLYKIAAPV IPII VS	240
		TTKEFMSCMNP GWD E+YV R EEAGF V TTISK+F+ S DLYKIA PVIPII VS	
RoqN	181	TTKEFMSCMNP GWDSE DYVHSRFEEAGFHSVQVTTI SKQFETSVEDLYKIAQPVIPII VS	240
OxaG	241	KWWTTEQKEAHEHE I LPALAKHLEATYGETGLVPQKWTAVFAKGEK	286
		KWW EQ++ +E++ILPAL +HL TYGE GLVPQ+WTAVFA G+K	
RoqN	241	KWWNQEQRD KYEND I LPALQRHLNETYGENGLVPQEW TAVFATGQK	286

Figure S2.



A space-group diagram for $C2$ (left). The crystallographic symmetry operators are shown in black. Notice in the OxaG unit cell (right) a parallel twofold (colored blue) relates the two subunits of the asymmetric unit. As a result, subunits A and B are also related by pseudotranslational symmetry (black arrows).

Figure S3.

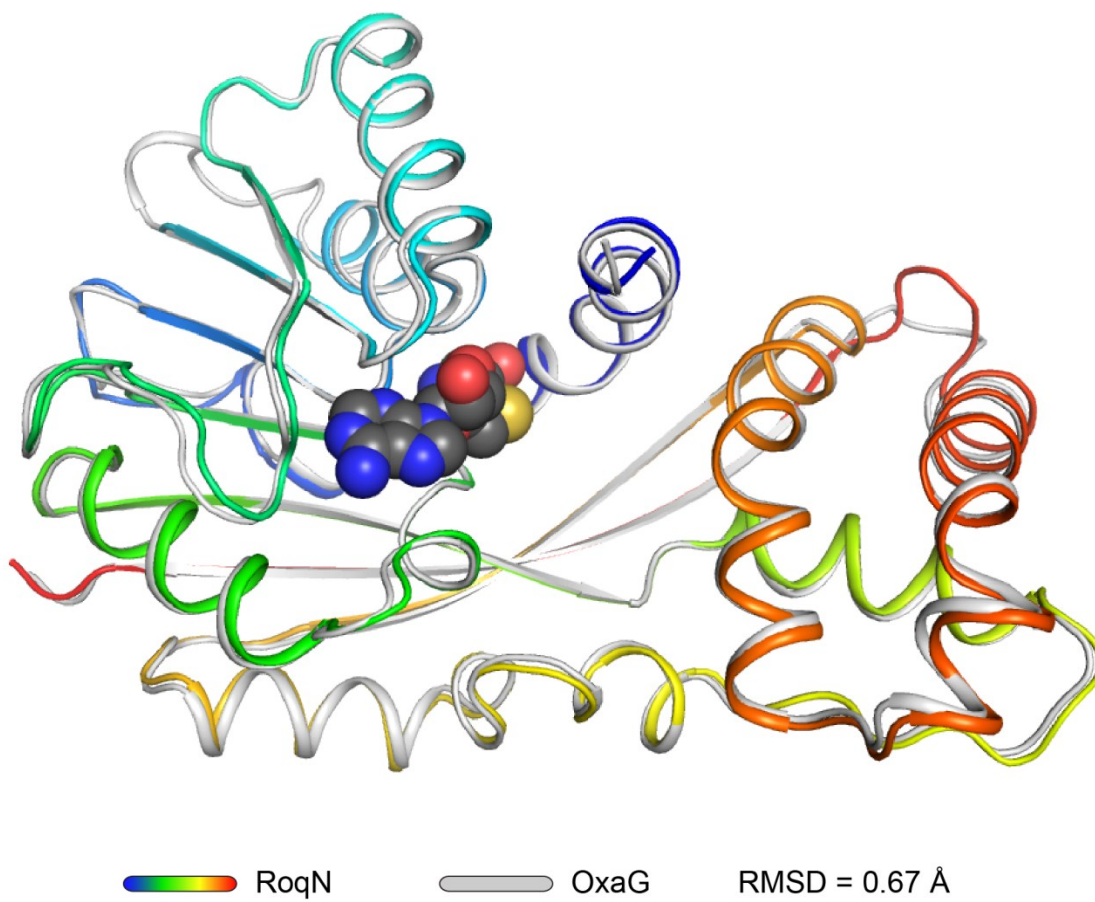
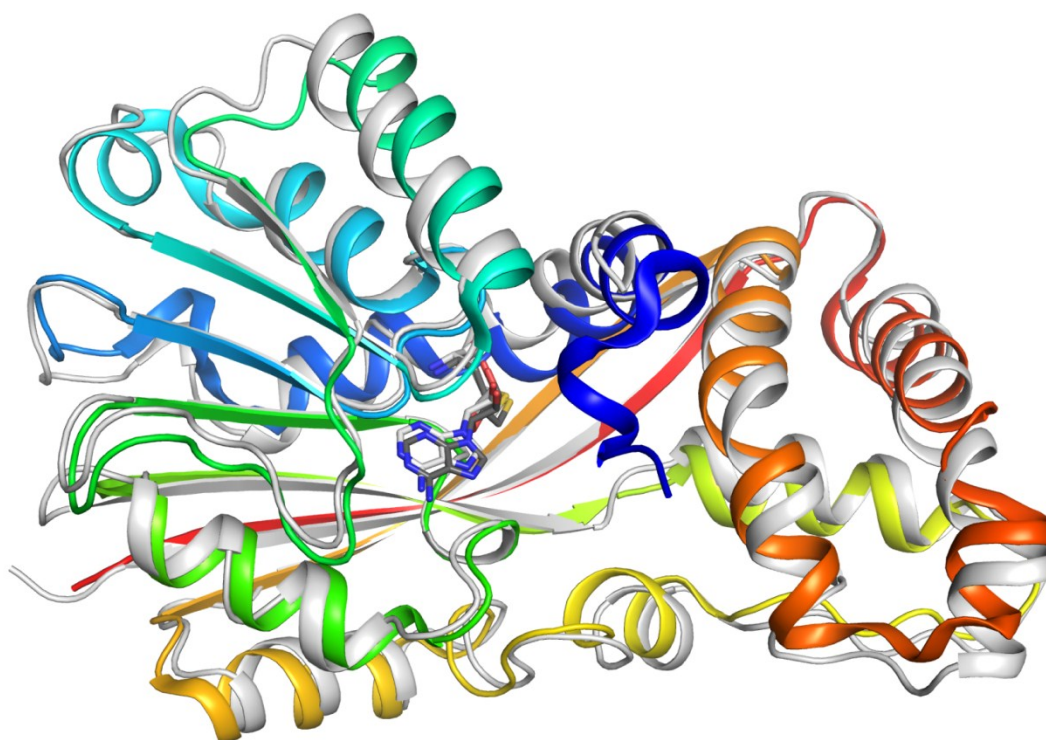


Figure S4.

A



B

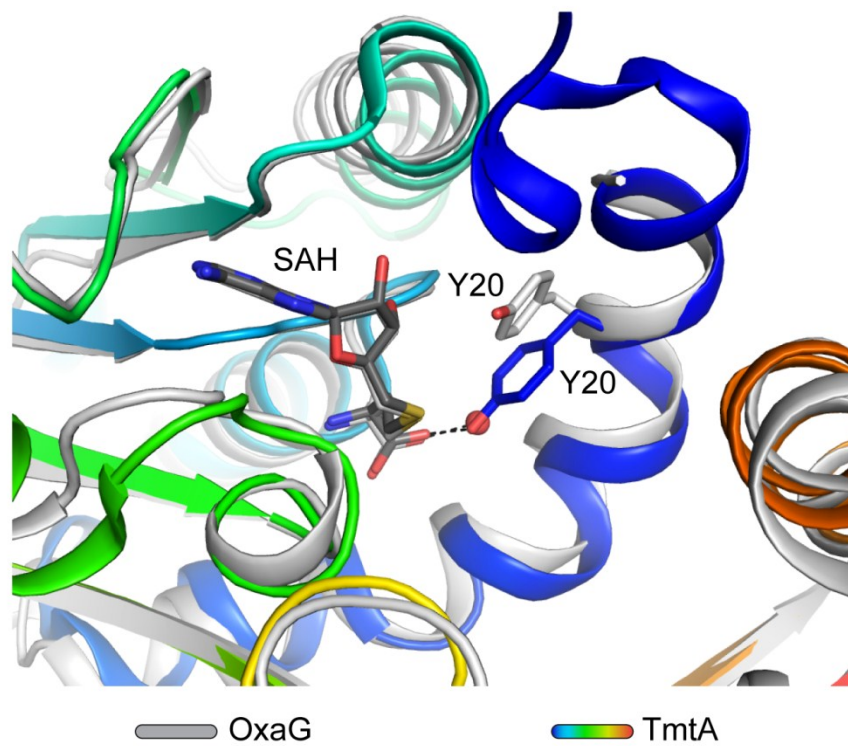


Figure S5.

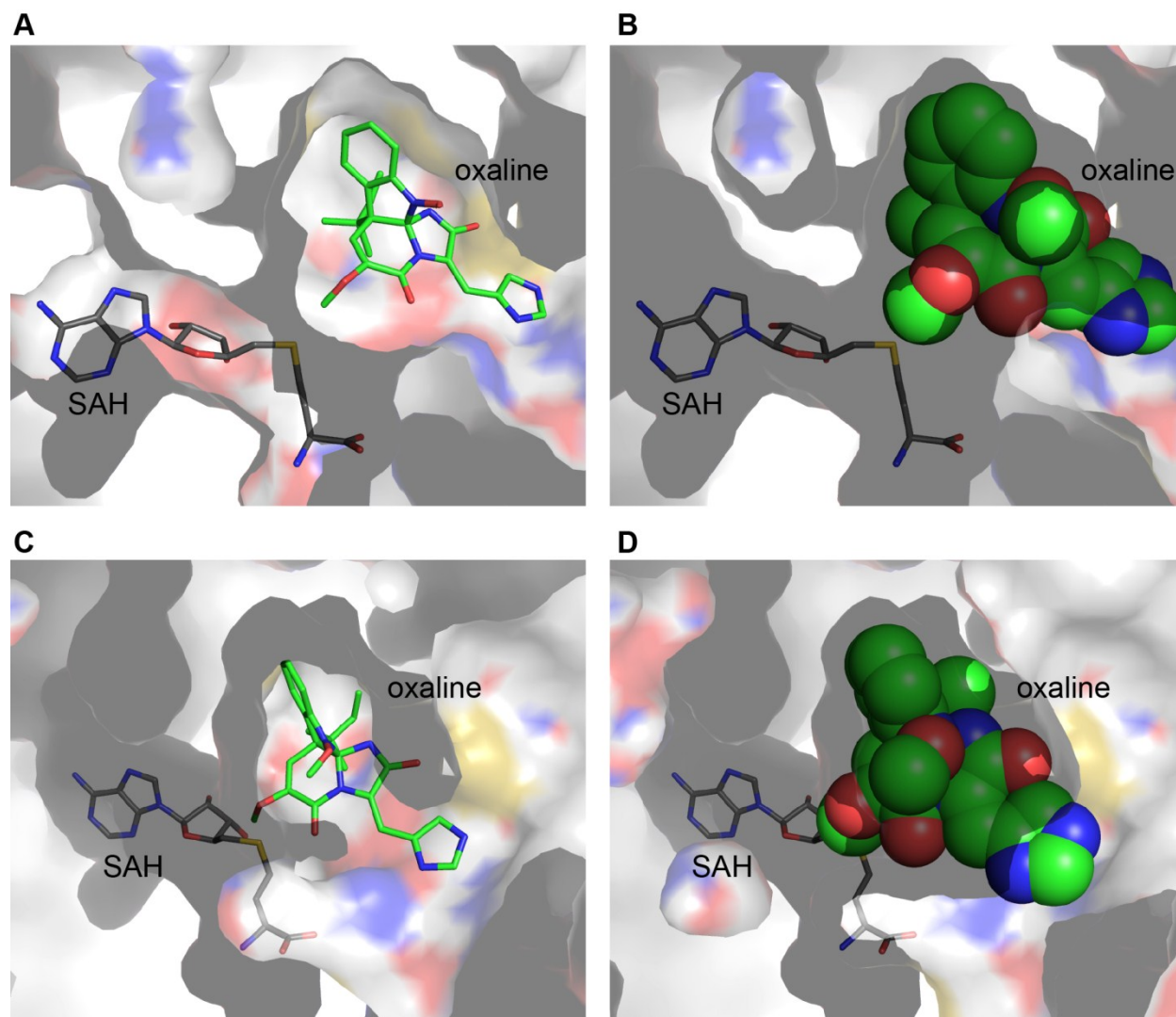
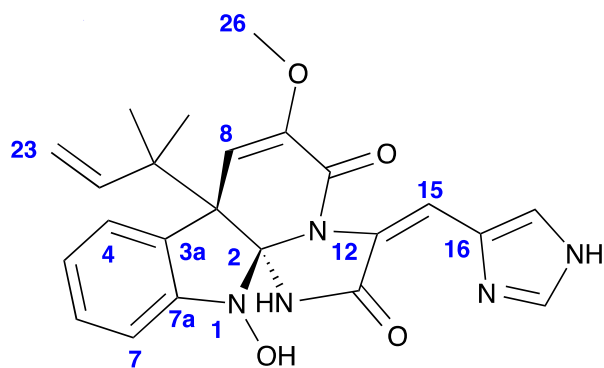


Table S1.

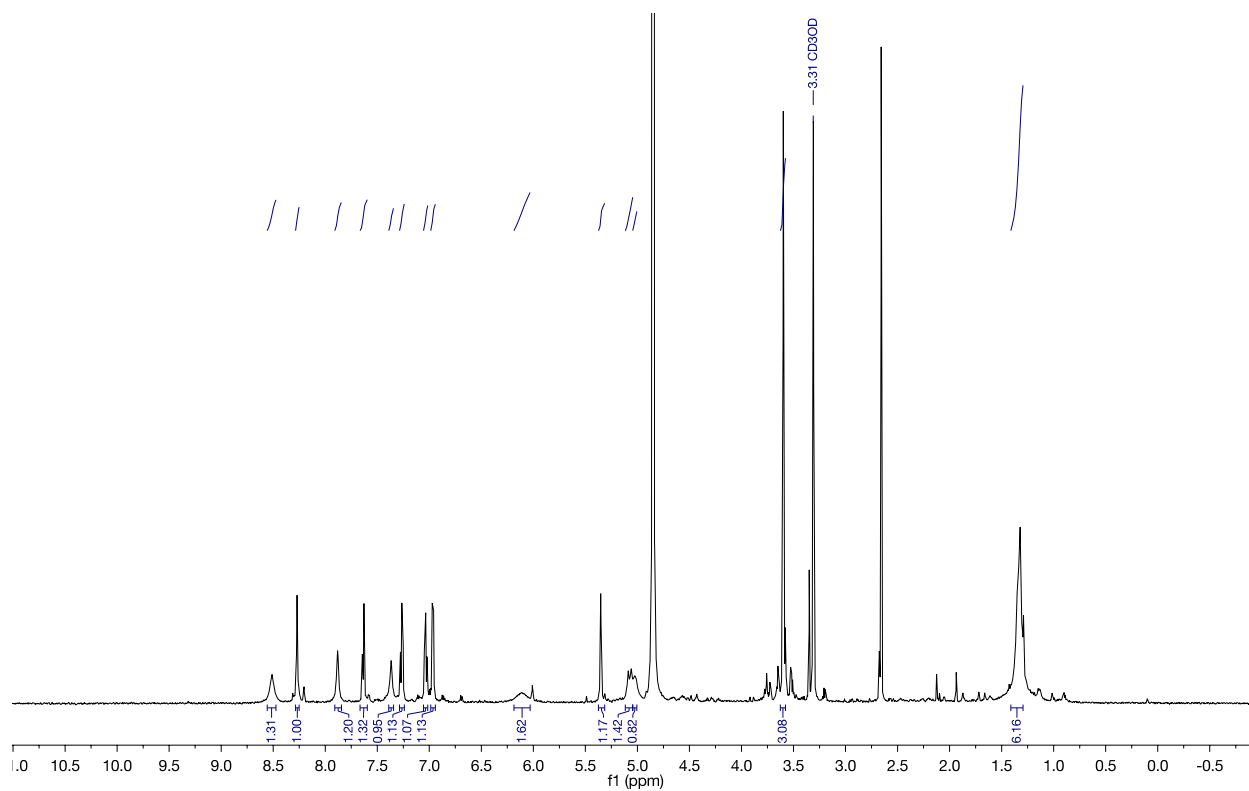
<i>oxaC_for</i>	5'- gagaatctctactccaaggcgctagcATGACATTTTCAAACGCTGATGCGC -3'
<i>oxaC_rev</i>	5'- ggctttgtagcagccggatctcagTTATTTCAACCTGGCCTCGATGATGCC -3'
<i>oxaG_for</i>	5'- gagaatctctactccaaggcgctagcATGACCCGCGCCACCAACTTTAC -3'
<i>oxaG_rev</i>	5'- ggctttgtagcagccggatctcagTCACTTCTCCTTCTCTCCCTTGGCAAATAC -3'
<i>roqN_for</i>	5'- ctgagaatctctactccaaggcGCTAGCATGACTCGCGCCACCAACTTTACCG -3'
<i>roqN_rev</i>	5'- cctttcgggctttgtagcagccggatcCTAAGATCCCTTCTGACCAGTTGC -3'
<i>oxaG_Y20F</i>	5' - GCAAGGGCATCCTGGAGACGTTTATGATCGCCGAGAA GATTACGCGATATTTC -3'
<i>oxaG_Y20A</i>	5'- GCAAGGGCATCCTGGAGACGGCGATGATCGCCGAGAAGA TTACGCGATATTTC -3'
<i>oxaC_D314A</i>	5'- GATTTACTACTTCCGCCGCGTCTTTCACGCGTGGCCT GATTTGCCTGAAGGTAAAAAG -3'

Table 2. NMR SPECTRA

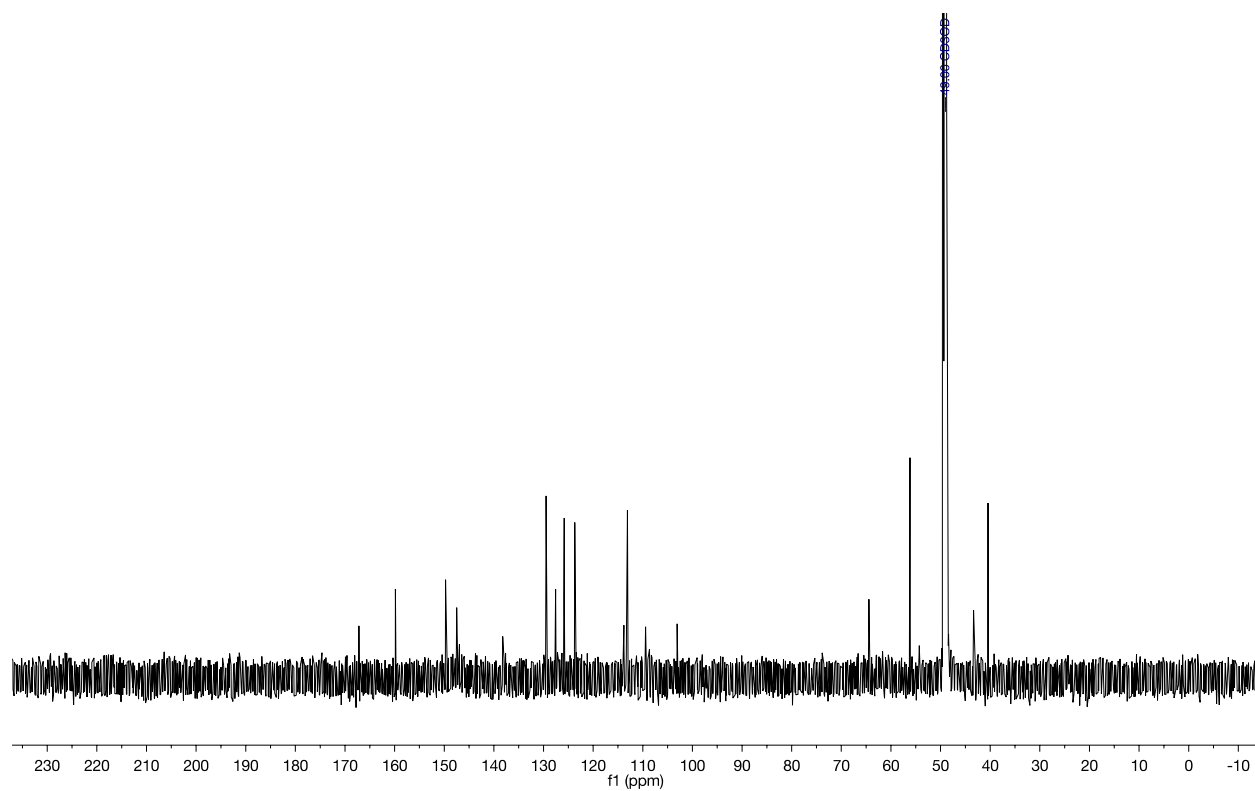


Position	Carbon	Hydrogen <i>J</i> Hz	HMBC correlations
2	103.10		
3	54.37		
3a	127.53		
4	125.84	7.63 (d, <i>J</i> = 7.7 Hz, 1H)	C3, C6, C7a
5	123.77	7.04 (t, <i>J</i> = 7.6 Hz, 1H)	C3a, C7
6	129.49	7.26 (t, <i>J</i> = 7.7 Hz, 1H)	C4, C7a
7	113.09	6.96 (d, <i>J</i> = 7.8 Hz, 1H)	C3a, C5
7a	149.76		
8	109.46	5.35 (s, 1H)	C2, C3a, C9, C10
9	147.51		
10	159.83		
12	127.17		
13	165.74		
15	108.80	8.27 (s, 1H)	C13, C12,(C16) C20
16	127.10		
18	138.31	7.88 (s, 1H)	
20	132.27	7.37 (s, 1H)	
21	41.69		
22	n.d	6.11 (s, 1H)	
23a (Trans)	113.85	5.08 (d, <i>J</i> = 17.0 hz, 1H)	
23b (Cis)	113.85	5.02 (bs, 1H)	
24	n.d	1.29 (s, 3H)	
25	n.d	1.32 (s, 3H)	
26	56.19	3.60 (s, 3H)	C8, C9, C10

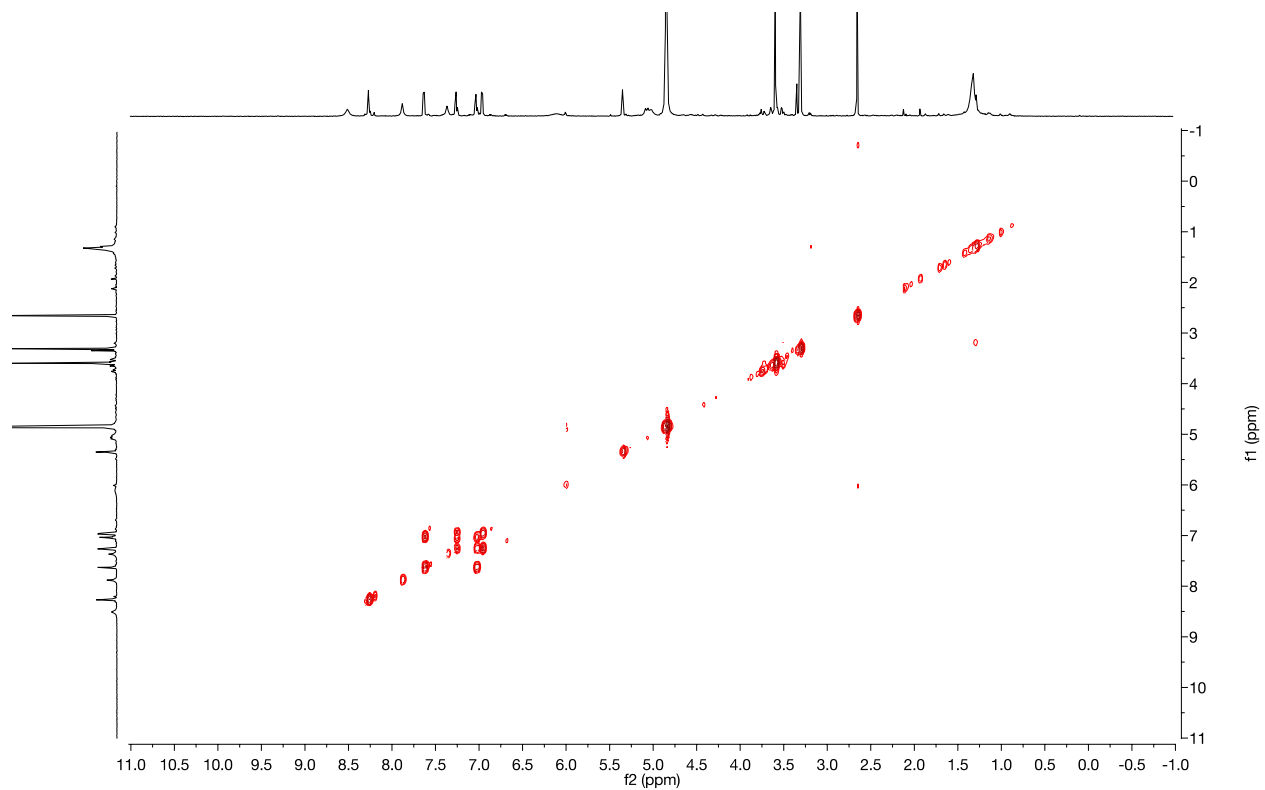
¹H NMR spectrum of glandicoline C in MeOH-d₄ at 600 MHz



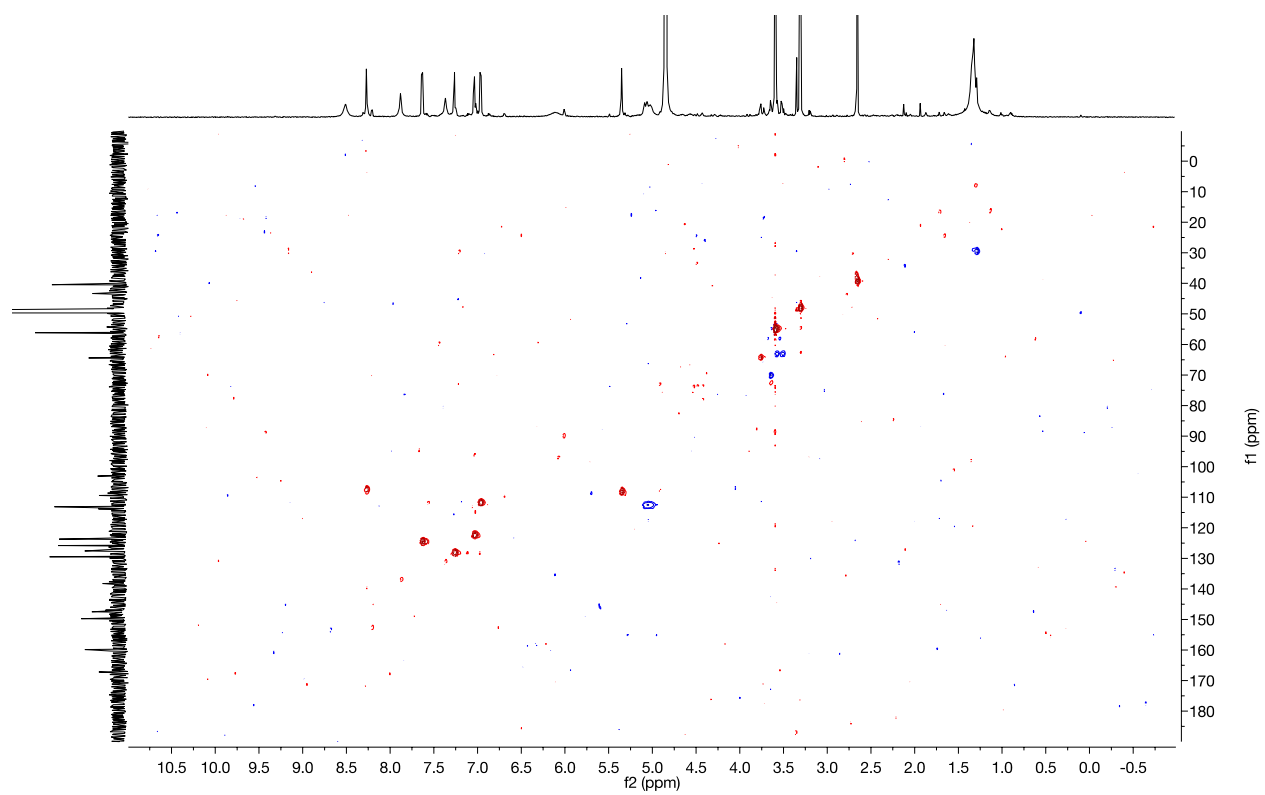
^{13}C NMR spectrum of glandicoline C in $\text{MeOH-}d_4$ at 125 MHz



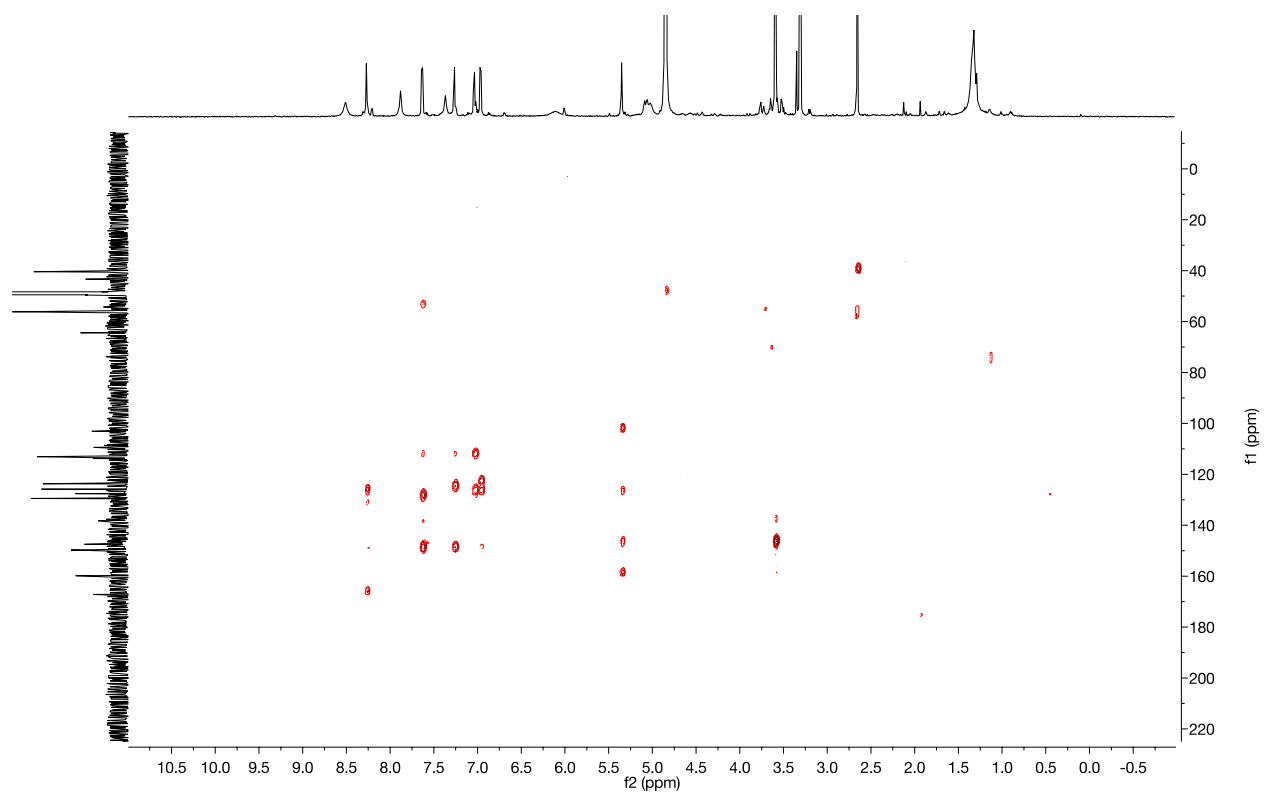
COSY NMR spectrum of glandicoline C in MeOH-*d*₄



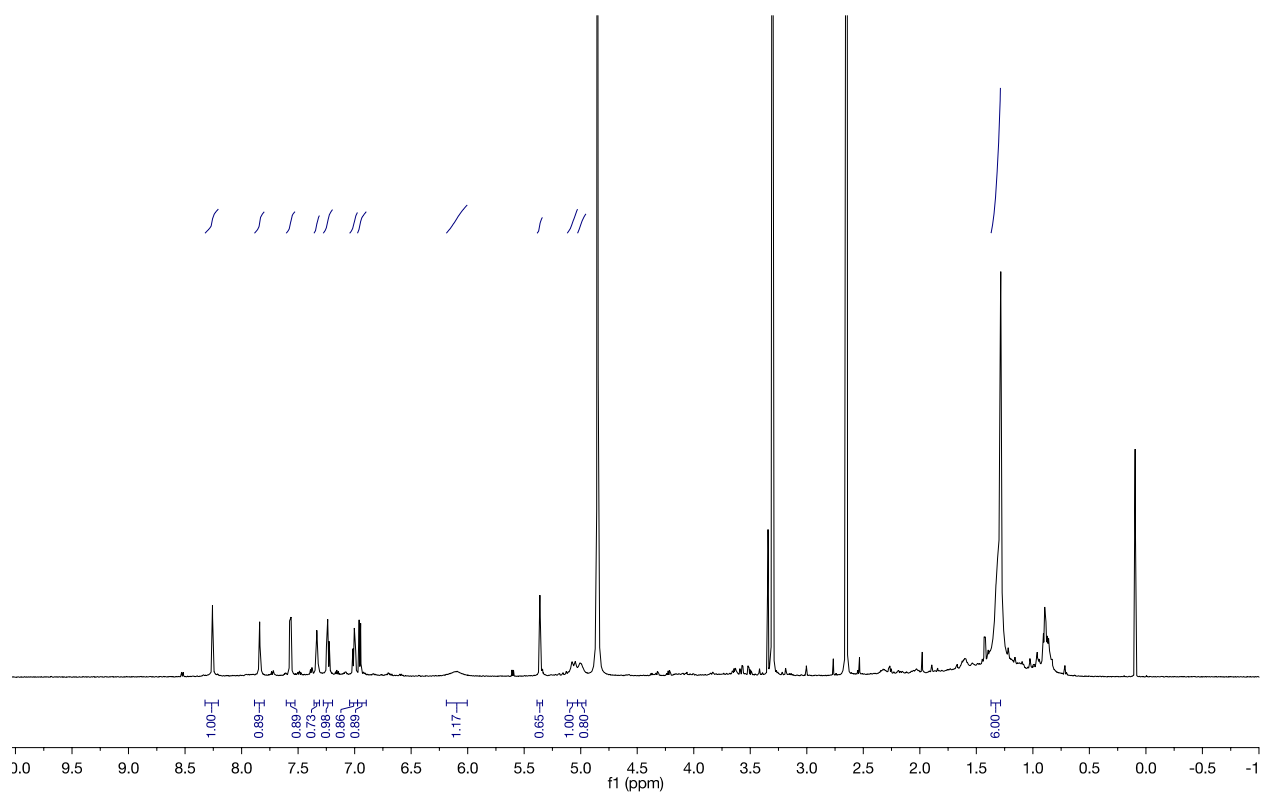
HSQC NMR spectrum of glandicoline C in MeOH- d_4



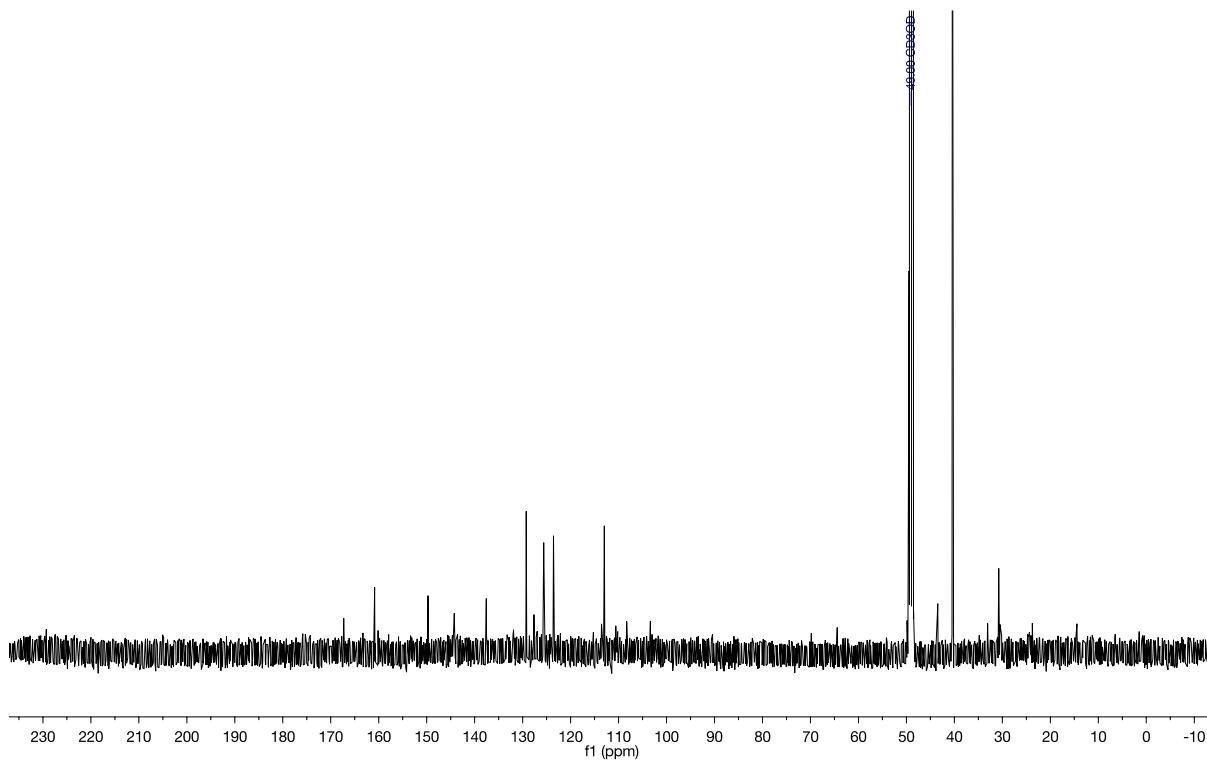
HMBC spectrum of glandicoline C in MeOH-*d*₄



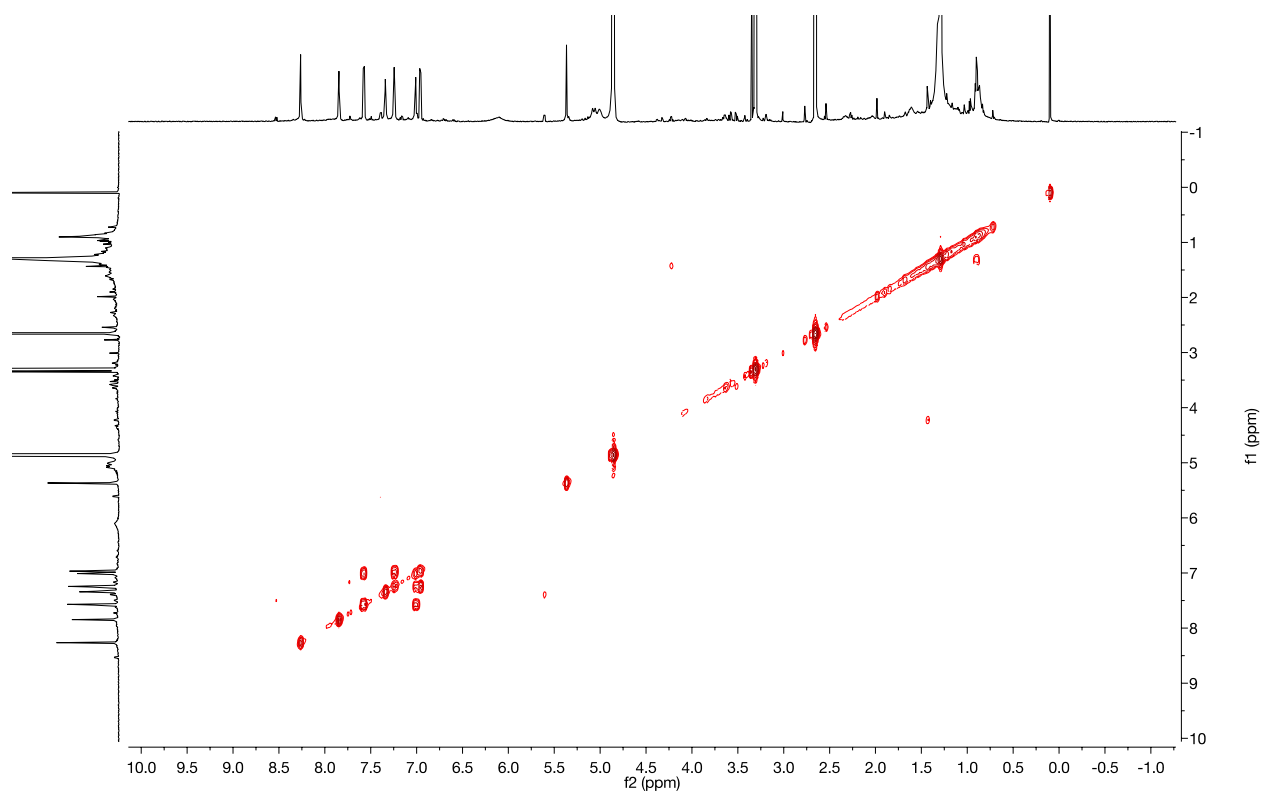
¹H NMR spectrum of glandicoline B in MeOH-d₄ at 600 MHz



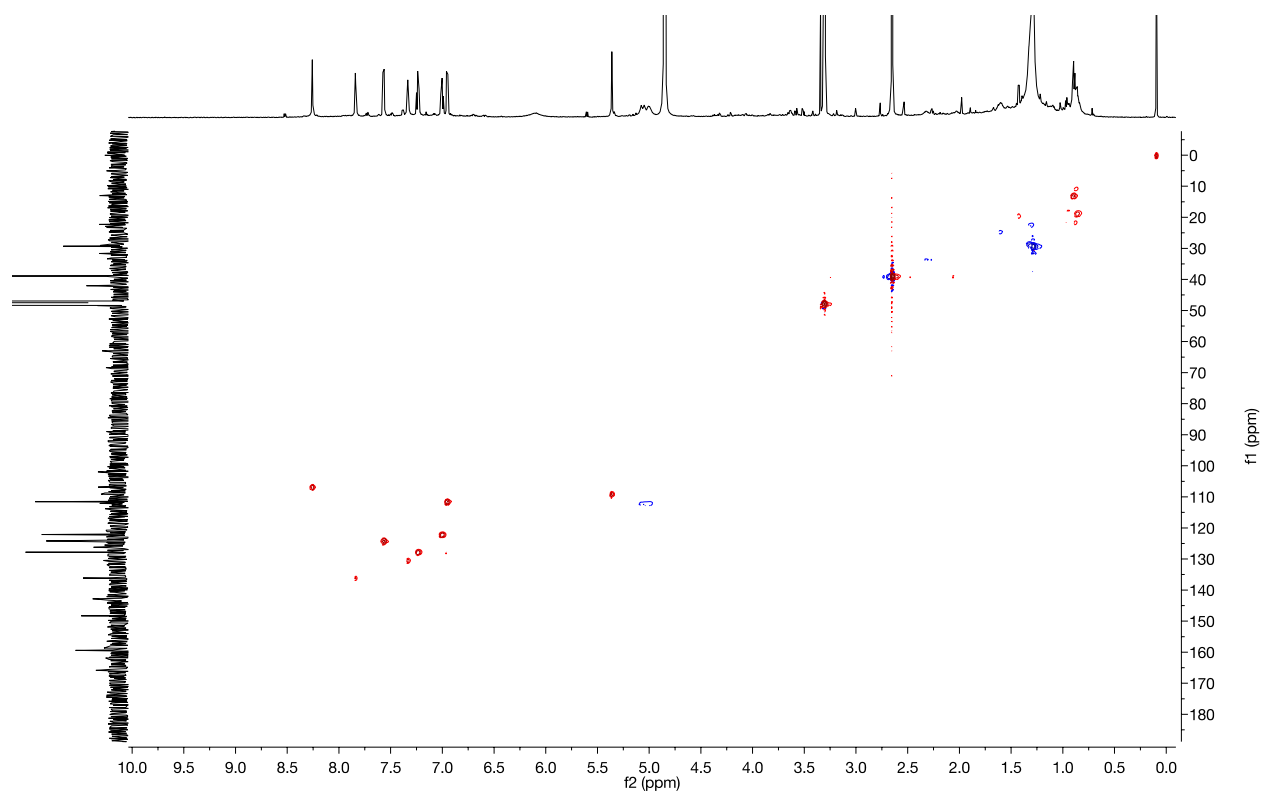
^{13}C NMR spectrum of glandicoline B in $\text{MeOH-}d_4$ at 125 MHz



COSY NMR spectrum of glandicoline B in MeOH- d_4



HSQC NMR spectrum of glandicoline B in MeOH-*d*₄



HMBC spectrum of glandicoline B in MeOH-*d*₄

