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A Chemical Recipe for Generation of Clinical-Grade Striatal Neurons from hESCs

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SUMMARY

Differentiation of human pluripotent stem cells (hPSCs) into striatal medium spiny neurons (MSNs) promises a cell-based therapy for Huntington's disease. However, clinical-grade MSNs remain unavailable. Here, we developed a chemical recipe named XLSBA to generate clinical-grade MSNs from embryonic stem cells (ESCs). We introduced the γ-secretase inhibitor DAPT into the recipe to accelerate neural differentiation, and replaced protein components with small molecules. Using this optimized protocol we could efficiently direct regular human ESCs (hESCs) as well as clinical-grade hESCs to lateral ganglionic eminence (LGE)-like progenitors and striatal MSNs within less than half of the time than previous protocols (within 14 days and 21 days, respectively). These striatal cells expressed appropriate MSN markers and electrophysiologically acted like authentic MSNs. Upon transplantation into brains of neonatal mice or mouse model of Huntington's disease, they exhibited sufficient safety and reasonable efficacy. Therefore, this quick and highly efficient derivation of MSNs offers unprecedented access to clinical application.

INTRODUCTION

Neurological diseases such as Huntington's disease (HD) are progressive neurodegenerative disorders that threaten humans in the modern age. HD is caused by CAG trinucleotide expansion in exon 1 of the Huntington gene (HTT) (MacDonald et al., 1993), and exemplifies diseases that affect specific brain regions and subtypes of neurons in the brain. The cardinal neuropathological manifestation of HD includes the atrophy of basal ganglia, particularly the caudate nucleus and putamen (Lange et al., 1976; Vonsattel and DiFiglia, 1998), and the progressive loss of the major type of striatal GABAergic projection neurons named medium spiny neurons (MSNs). Although some symptoms of HD can be alleviated by medication, no effective treatment is currently available except the intrastriatal transplantation of fetus-derived ganglionic eminence cells reported in a trial (Rosser and Bachoud-Lévi, 2012).

Human pluripotent stem cells (hPSCs) can differentiate into desired neuronal subtypes including MSNs, providing an unlimited and readily accessible source of MSNs for neural replacement therapy of HD. Protocols including those employing dual-SMAD inhibition have been exploited to produce striatal MSNs (Arber et al., 2015; Aubry et al., 2008; Delli Carri et al., 2013; Ma et al., 2012; Nicoleau et al., 2013). Such endeavors advocate significantly for the understanding of neural differentiation and have shed light on the potential of human embryonic stem cell (hESC)-based cell replacement therapy. These protocols, however, are not designed for generation of clinical-grade cells and most of them are time-consuming, which increases the variability, yielding variable production of MSNs. In either protocol, recombinant proteins such as SHH, DKK1, or ACTIVIN A are used (Arber et al., 2015; Delli Carri et al., 2013; Nicoleau et al., 2013). Protein components in recipe increase the difficulties of quality control, and in addition increase the cost. Chemical compounds are cost-effective and easily quality controlled, exhibiting significant advantages over proteins. They can replace proteins of classic signaling to produce functional cortical neurons and nociceptors from hESCs (Chambers et al., 2012; Qi et al., 2017), suggesting that a chemical recipe is also possible for generating MSNs from hESCs.

Recipes free of proteins are particularly preferred for the manufacture of clinical-grade cells for therapy. We and colleagues have developed xeno-free hESCs with defined quality characteristics fitting for clinical therapy (Gu et al., 2017). To generate MSNs from these clinical-grade hESCs, we invented a small-molecule-based recipe and a streamlined protocol for efficient induction of lateral ganglionic eminence (LGE)-like cells and MSNs. This protocol is further adapted to the clinical-compatible conditions to generate clinical-grade

MSNs, which facilitate the industrial manufacture and clinical use.

RESULTS

DAPT Expedites the Differentiation of Striatal Projection Neurons from hESCs

The ability to produce specific subtypes of neurons such as MSNs from hPSCs has made considerable progress. However, existing hPSC differentiation protocols with extrinsic patterning factors require extended culture periods of as long as 80 days (Arber et al., 2015; Aubry et al., 2008; Delli Carri et al., 2013; Ma et al., 2012). For example, the well-established embryoid body (EB)-based differentiation strategy needs 40 days to generate striatal progenitors for cell transplantation (Figure 1A) (Ma et al., 2012). The protracted *in vitro* differentiation of hPSCs increases variabilities and costs for cell products in clinical application. Therefore, accelerating the differentiation is important for the generation of readily accessible clinical-grade MSNs used for therapy.

N-[N-(3,5-Difluorophenacetyl)-L-alanyl]-S-phenylglycine t-butyl ester (DAPT) was proposed to block NOTCH signaling as a γ-secretase inhibitor (Dovey et al., 2001) and accelerate the neural differentiation (Chambers et al., 2012; Qi et al., 2017). To expedite the differentiation process for generating striatal MSNs from hESCs, we set up an EB and monolayer combination culture system, and introduced DAPT to the medium from day 11, based on the dual-SMAD inhibition approach (termed the NSBS protocol) (Chambers et al., 2009; Kirkeby et al., 2012a, 2012b) (Figure 1A). Using the NSBS protocol, typical neural rosette structures appeared as early as day 5 of differentiation, as compared with day 15 if using the EB protocol (Figure 1B). Neuroepithelia (NE) expressing SOX1 and PAX6 were readily detected at day 5 of differentiation in the NSBS protocol, 10 days earlier than that in the control (Figures 1B and 1C). LGE-like progenitors expressing telencephalic marker FOXG1 and subpallial telencephalic marker DLX2 were detected on day-14 cultures, with a similar efficiency as that in the EB protocol on day 26 (Figures 1B and 1C). A similar amount of MSNs expressing TUJ1, GABA, and DARPP32 were generated at day 21, less than half of that used in the EB protocol (47 days) (Figures 1B and 1C).

DAPT significantly accelerated neural specification and neuronal maturation, as was also shown with qPCR. Administration of DAPT significantly increased levels of the neuroblast gene DCX, pan-neuronal gene TUJ1, mature neurons marker MAP2, and forebrain markers FOXG1 and SIX3, but not neural progenitor markers NESTIN, SOX1, and SOX2 (Figure 1D). It also increased cells expressing post-mitotic neuronal marker DLX5 (Eisenstat et al., 1999; Liu et al., 1997; Panganiban and Rubenstein, 2002) (Figure 1D).

Therefore, using a simple yet efficient NSBS protocol, we successfully expedited the generation of NE, LGE-like cells, and MSNs within 5 days, 14 days, and 21 days as compared with 15 days, 26 days, and 47 days for the EB protocol.

Optimizing SHH Pathway Activity for Robust Generation of Striatal MSNs

Appropriate activity of Sonic hedgehog (SHH) signaling is critical to induce LGE and can ventralize the hPSC derivatives (Campbell, 2003; Ma et al., 2012). Since DAPT significantly accelerates the MSN specification, fine-tuning the SHH activity is crucial in obtaining authentic MSNs in the optimized protocol. To determine an optimal level of SHH, we treated hESCs with 0, 100, 200, and 500 ng/mL SHH from day 0 to day 9 (Figure S1A). On day 14, while SHH concentration did not affect the expression of telencephalic marker FOXG1, 100 ng/mL SHH was sufficient to increase the expression of telencephalic marker OTX2, early developing striatal markers DLX2, and post-mitotic MSNs markers DLX5, FOXP2, and MEIS2 (Figure S2B). Immunostaining at day 17 indicated that cells expressing FOXP1, CALBINDIN, and DARPP32 were significantly increased upon treatment with 100 ng/mL SHH (Figures S2C and S2D). Among the TUJ1$^+$ neurons, 66% were FOXP1$^+$, 91% CALBINDIN$^+$, and 77% DARPP32$^+$ neurons (Figure S2D). However, SHH did not significantly alter the efficiency of neural differentiation as each group yielded ~92% MAP2$^+$ cells among TUJ1$^+$ cells (Figures S2C and S2D).

The NSBS protocol favored the generation of ventral telencephalic cells and accelerated the neural differentiation, as was demonstrated by qPCR at three typical stages of MSN differentiation, namely NE, LGE, and MSN. Except for a decreasing gene expression of PAX6 and DLX2 at the NE and LGE stage, respectively, NESTIN, FOXG1, and SIX3 at the NE stage, GSH2, MASH1, DLX5, and DLX6 at the LGE stage, and FOXP1, MEIS2, CALBINDIN, DRD2, and ARPP21 at the MSN stage were all increased (Figures S2A–S2C).

Overall, these results indicated that 100 ng/mL SHH could enrich neuronal subtype MSN without compromising the efficiency of neural specification and neuronal maturation.

Generation of GABAergic MSNs from hESCs by Chemical Cocktails

A protein-free recipe is critical to obtain subtype-specific neurons from hPSCs for clinical applications. Small molecules such as the ALK2/ALK3 inhibitor LDN-193189 (L) and BMP inhibitor dorsomorphin (D) can replace NOGGIN for neuralization of hPSCs (Chambers et al., 2012; Kim et al., 2010; Morizane et al., 2011; Qi et al., 2017; Yu et al., 2008;

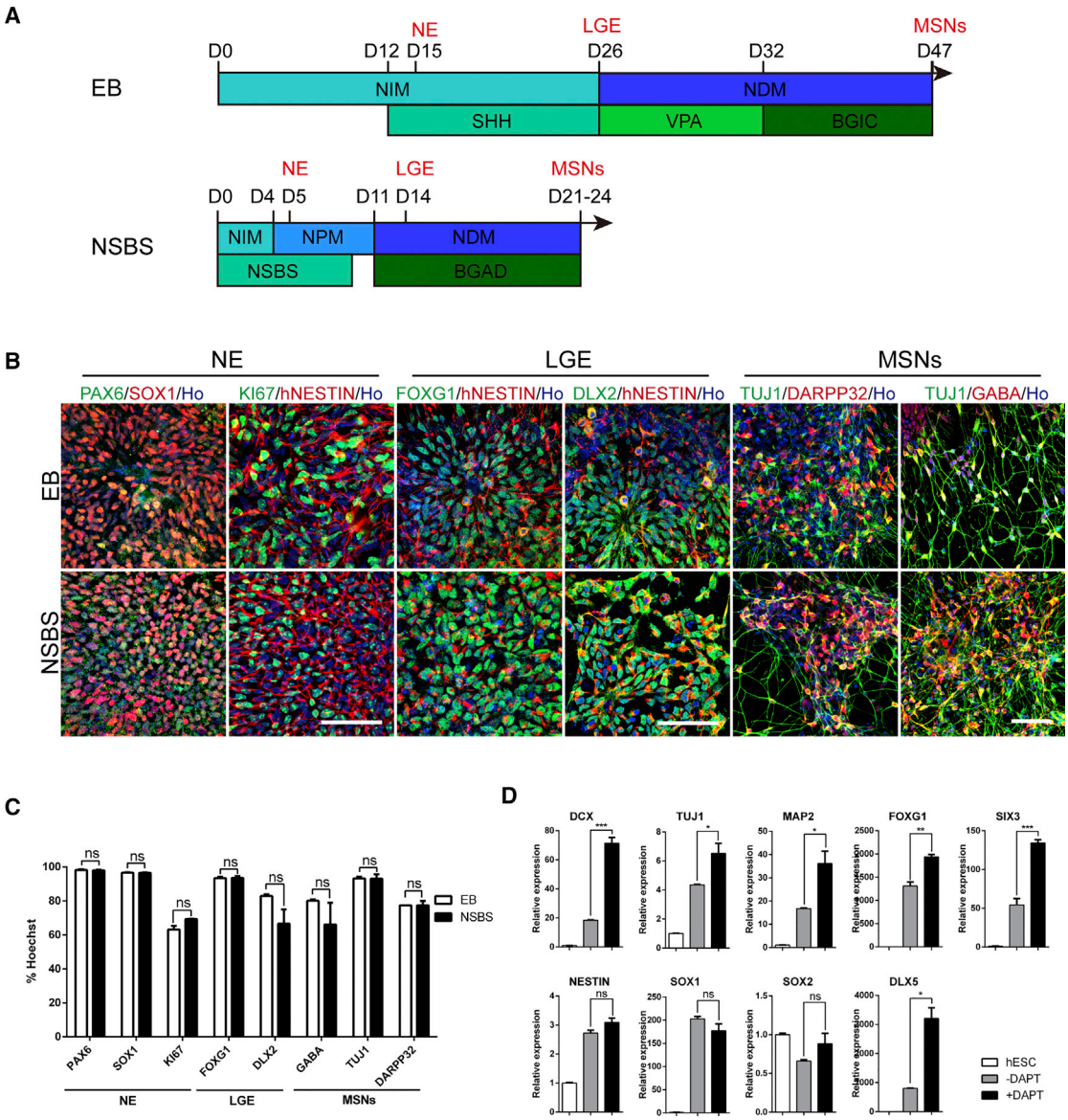


Figure 1. DAPT Accelerates the Differentiation of Striatal Projection Neurons from hESCs

(A) Schematic overview of the differentiation strategies using either the EB or NSBS protocol to obtain striatal MSNs. BGIC represents BDNF, GDNF, IGF-1, and dibutyryl-cAMP. NSBS represents Noggin, SB431542, and SHH. BGAD represents BDNF, GDNF, AA, and DAPT.

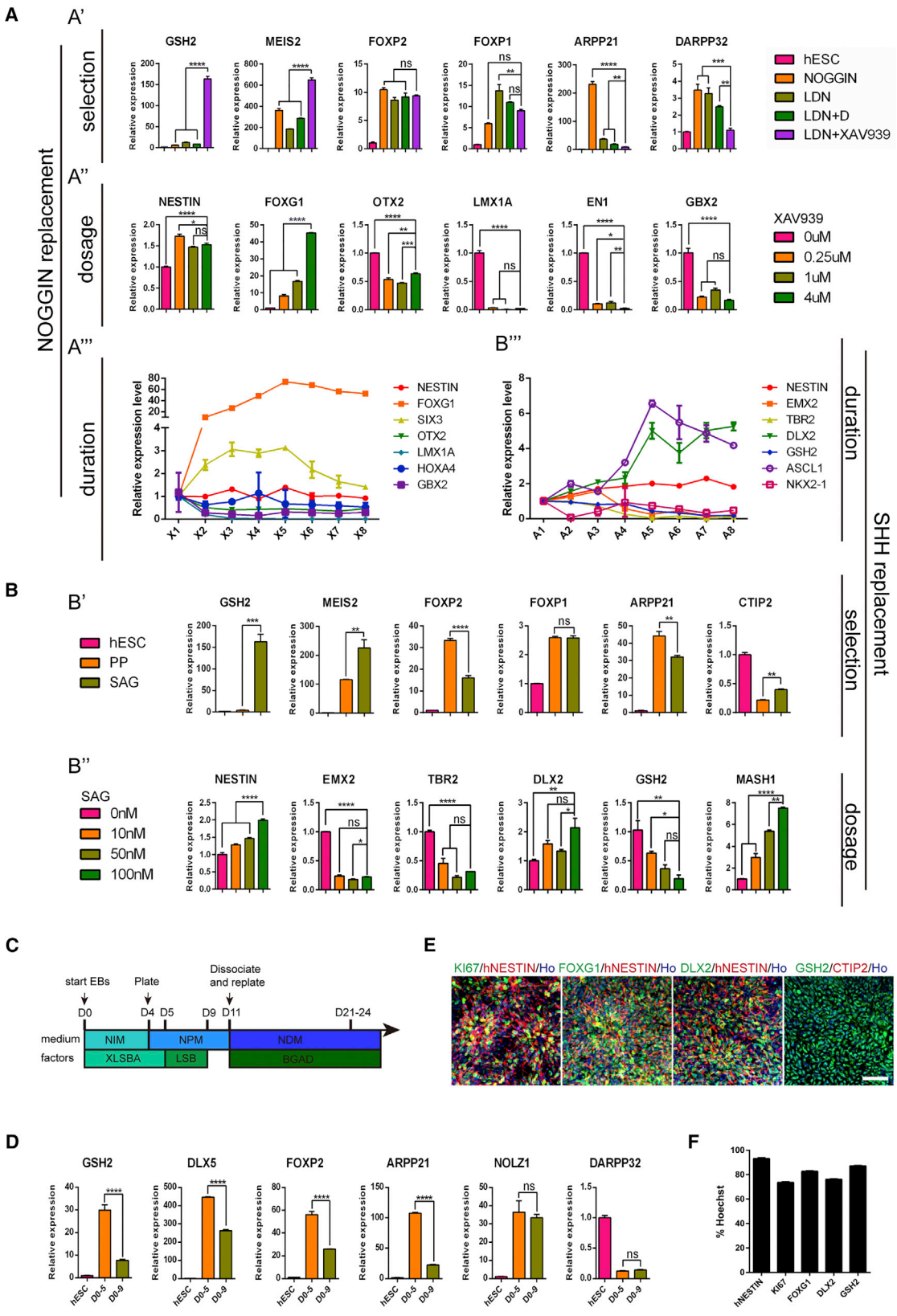
(B) Representative images of immunofluorescence for particular stage markers during EB and NSBS differentiation. PAX6, SOX1, hNESTIN, and KI67 for neuroepithelia; FOXG1 and DLX2 for LGE-like progenitors; DARPP32, GABA, and TUJ1 for MSN GABA neurons. Ho, Hoechst 33258. Scale bars, 50 μm.

(C) Quantification for particular differentiation stage markers from (B). Efficiencies are presented as the percentage of positive cells ±SEM of all fields counted. ns, not significant; Student's t test.

(D) Relative mRNA expression for neuroblasts (DCX, TUJ1), mature neurons (MAP2), forebrain markers (FOXG1, SIX3), neural progenitors (NESTIN, SOX1, SOX2, PAX6), and LGE markers (DLX5) in H9-hESC derivatives generated with or without DAPT (day 18). Data are presented as mean ± SEM (n = 3). *p < 0.05, **p < 0.01, ***p < 0.001, ns, not significant.

See also Figures S1 and S2.

Zhou et al., 2010). We thus chose L and D together with WNT signaling inhibitor XAV939(X), and SHH signaling agonists SAG or purmorphamine (PP), to coordinate the anterior-posterior and dorsal-ventral patterning, respectively, and direct hESCs into LGE-like progenitors. A combination of four small molecules (XAV939, LDN-193189, SB431542,



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and SAG, termed the XLSBA protocol) facilitated the induction of LGE-like precursors that expressed striatal and neuronal markers such as GSH2, MEIS2, FOXP1, FOXP2, NOLZ1, ARPP21, CTIP2, and DARPP32 (Figures 2A′ and 2B′). We further adjusted the duration and concentration of small molecules and optimized the protocol for efficient production of LGE-like cells (Figures 2A″–2A‴ and 2B″–B‴). Under such a condition, the transcript levels of GSH2, DLX5, FOXP2, and APRR21 were significantly increased (Figures 2C and 2D). In addition, abundant NESTIN$^+$ (93.1% ± 1.0%), KI67$^+$ (73.7% ± 0.7%), FOXG1$^+$ (82.7% ± 0.7%), DLX2$^+$ (76.3% ± 0.4%), and GSH2$^+$ (87.2% ± 0.4%) cells were detected (Figures 2E and 2F). Thus, the XLSBA protocol can efficiently and quickly induce hESCs into LGE-like striatal progenitors.

The hESC-induced LGE-like progenitors efficiently converted into MSNs on day 21 of differentiation, as was shown by immunofluorescence of subtype-specific neuronal markers (Figures 3A–3D). Over 87% of the MAP2$^+$ neurons expressed DARPP32 and more than 89% of the TUJ1$^+$ neurons were GABA$^+$ (Figures 3A and 3B), and a proportion of the TUJ1$^+$ cells co-expressed other genes with enriched expression in the striatum such as CALBINDIN, demonstrating their striatal GABAergic MSN identity (Figures 3C and 3D). More mature neurons possessing dendritic spines also expressed PSD95 and SYNAPSIN 1 (Figure 3E). Except for the GABAergic neurons (89.5% ± 5.0%), 8.2% ± 1.2% of cells expressed tyrosine hydroxylase (TH; a marker for dopaminergic neurons). None of the differentiated cells expressed other neuronal subtype markers, such as choline acetyltransferase (CHAT; a marker of cholinergic neurons), 5-hydroxytryptamine (5-HT; a marker for serotonergic neurons), and vesicular glutamate transporter 1 (VGlut1; a marker for excitatory glutamatergic neurons) (Figures 3F and 3G). Thus, the XLSBA protocol primarily induces DARPP32$^+$ GABAergic striatal projection neurons.

XLSBA worked efficiently in additional hESC lines, particularly biosafe clinical-grade hESC line (Gu et al., 2017). Using the xeno-free Clinical Therapy Systems (CTS) reagents, the clinical-grade hESC line was efficiently differentiated into DARPP32-expressing MSNs, and the Q-CTS-hESC-2 line generated even more DARPP32$^+$ cells (~94.9% of MAP2$^+$ neurons) than the regular hESC lines (Figures 3A and 3B). Therefore, the chemical XLSBA protocol is superior in rapidly and robustly inducing clinical-grade LGE/striatal progenitors and a homogeneous population of MSNs from hESCs.

hESC-Derived MSNs Progressively Mature and Act as Authentic GABAergic Neurons

To examine the functional properties of hESC-derived MSNs, we performed whole-cell patch-clamp recordings on hESC-derived MSNs at 20–24 days after differentiation. Current-clamp recording showed that all cells elicited single or multiple action potentials upon the injection of depolarizing currents (Figure 4A). As depolarizing voltage steps in voltage-clamp mode it elicited fast inward currents followed by slow outward currents, indicating opening of voltage-activated sodium and potassium channels, respectively (Figure 4B). Spontaneous action potentials were frequently detected (Figure 4C).

Figure 2. Generation of LGE-like Progenitors from hESCs by Chemical Cocktails

(A) Relative mRNA expression of striatal lineage genes to identify optimal small-molecule combination to replace noggin. (A′) Striatal lineage gene expression in H9-hESCs cultures differentiated in the presence of LDN-193193 (100 nM), dorsomorphin (100 nM), XAV939 (1 μM), or their combination from days 1–9. All expression levels are normalized to levels detected in hESCs. (A″) Anterior-posterior axis gene expression in H9-hESC cultures differentiated after treatment with different concentrations of XAV939 (0, 0.25, 1, and 4 μM). (A‴) Anterior-posterior axis gene expression in H9-hESCs cultures differentiated after treatment with 4 μM XAV939 for different time periods. X1 represents cell cultures treated with XAV939 for 1 day, X2 represents cell cultures treated for 2 days, and so on. Data are presented as mean ± SEM (n = 3). ns, not significant. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001; one-way ANOVA, followed by Tukey's multiple comparisons tests.

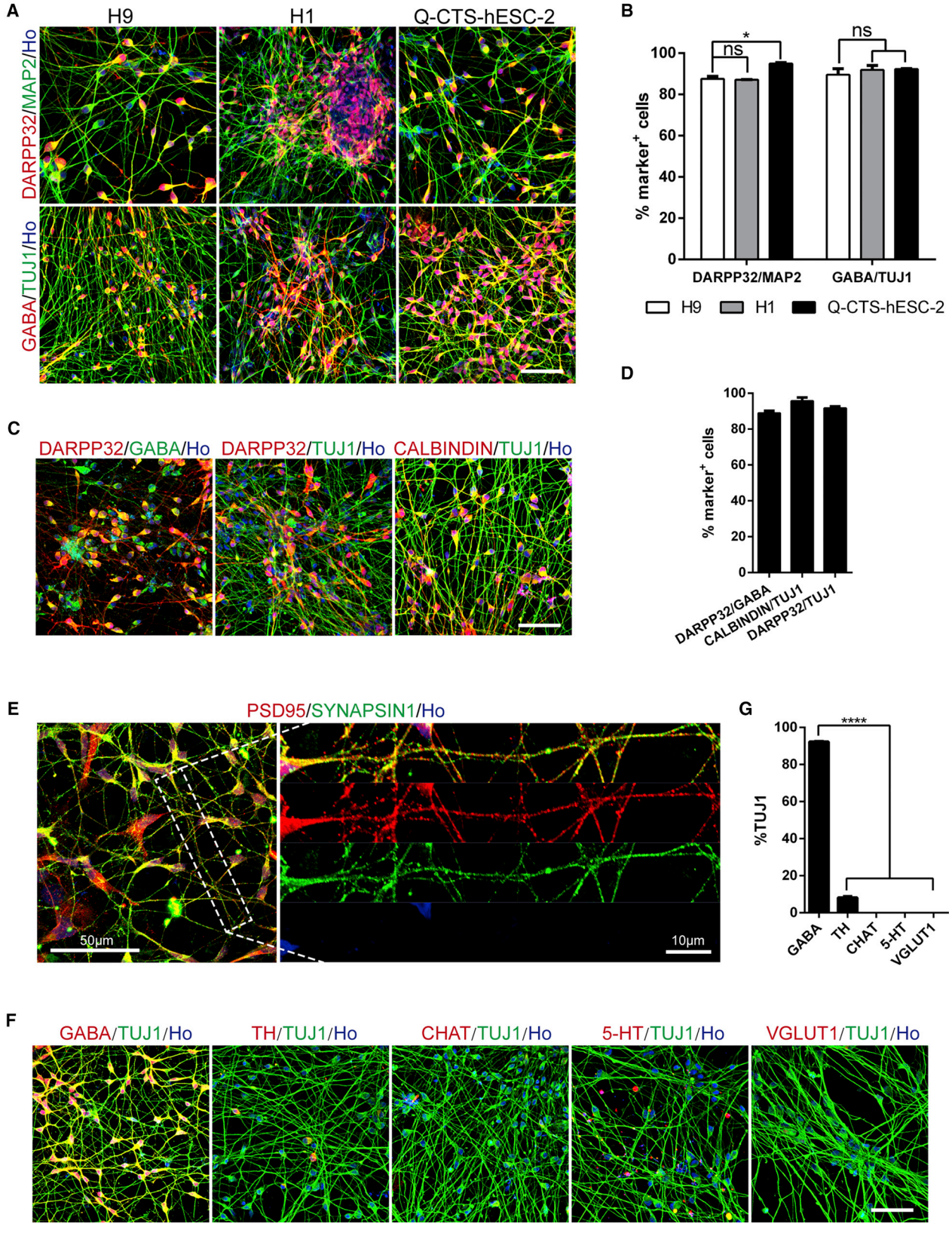
(B) Relative mRNA expression of striatal lineage genes to identify optimal small molecule to replace SHH. (B′) Striatal lineage gene expression in H9-hESC cultures differentiated in the presence of purmorphamine (PP, 1 μM) or SAG (100 nM). All expression levels are normalized to levels detected in hESCs. (B″) Dorsal-ventral axis gene expression in H9-hESCs cultures differentiated after treatment with different concentrations of SAG (0, 10, 50, and 100 nM). (B‴) Dorsal-ventral axis gene expression in H9-hESCs cultures differentiated after treatment with 100 nM SAG for different time periods. A1 represents cell cultures treated with SAG for 1 day. A2 represents cell cultures treated for 2 days, and so on. Data are presented as mean ± SEM (n = 3). ns, not significant. *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001; one-way ANOVA followed by Tukey's multiple comparisons tests.

(C) Schematic overview of the optimized differentiation protocol for hESCs.

(D) Relative mRNA expression of striatal lineage genes treated with small molecules from days 0–5 and days 0–9. All expression levels are normalized to levels detected in hESCs. Data are presented as mean ± SEM. ns, no significant. ****p < 0.0001; Student's t test.

(E) Immunostaining images for neural progenitor marker NESTIN, proliferative marker KI67, forebrain marker FOXG1, LGE progenitor markers GSH2 and DLX2, and post-mitotic MSN marker CTIP2 on day 11. Scale bar, 50 μm.

(F) Quantification for immunostaining of markers in (E). Error bar represents SEM.



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Upon local administration of tetrodotoxin (TTX), traces of membrane action potentials were not detected in representative neurons and the inward currents were ablated (Figures 4D and 4E), indicating the involvement of TTX-sensitive voltage-gated sodium channels. These neurons became functionally mature over time. Whereas they initially displayed modest action potentials at 20 days, approximately 46% and 34% of the recorded cells fired at 22 and 24 days of differentiation, respectively (Figure 4F). The passive membrane properties of differentiating neurons (Table S1) exhibited negative resting membrane potential (RMP; −40 to −50 mV, Figure 4G). The input resistance values (~2–4 GΩ, Figure 4H) decreased while capacitance values (20–25 pF, Figure 4I) were stable during their maturation in culture (days 20–24). Addition of GABA in culture elicited inward currents when cells were held at −70 mV (n = 19, Figure 4J), whereas addition of H_2O could not (n = 12, Figure 4J). The currents were blocked by the GABA type A (GABA-A) receptor antagonist bicuculline (Figure 4J). Therefore, these results confirmed that hESC-derived neurons exhibited GABAergic identity.

Since astrocytes could promote neuron maturation (Johnson et al., 2007), we investigated whether co-culturing differentiated MSNs with human astrocytes would lead to more active electrophysiological characteristics. As anticipated, ~77% of recorded MSNs (17 of 22 total cells) were capable of firing repetitively (Figure 4F) and displayed a more hyperpolarized RMP in the presence of astrocytes compared with those in the absence of astrocytes (Figure 4G), indicating more maturity. However, no significant change in membrane resistance (R_{in}) and membrane capacitance (C_m) was observed (Figures 4H and 4I) and peaks of the voltage-gated Na^+ and K^+ currents also stayed unchanged upon co-culture with human astrocytes (Figures 4K–4N). These results indicated that astrocytes promoted the maturation of hESC-derived striatal MSNs in our culture system to some extent.

hESC-Derived MSNs Become Mature and Functional in Brains of Neonatal and HD Mice

To confirm whether these LGE-like progenitors and MSNs are safe and functional, we transplanted 100,000 LGE-like progenitors into the striatum of P1 neonatal SCID mice (Figure 5A). Immunostaining for human nuclei (hN) indicated that a significant proportion of graft cells expressed DARPP32, FOXP1, FOXG1, and CALBINDIN 12 weeks after transplantation (Figures 5B and 5C). $NESTIN^+$ or $KI67^+$ proliferative cells were rare within the grafts (Figures 5B and 5C). The transplanted LGE-like progenitors differentiated into both the direct and indirect subtypes of MSNs, as evidenced by the presence of substance P^+ (SP^+) and enkephalin$^+$ (ENK^+) cells within the grafts (Figures 5D and 5E). While considerably grafted cells became $TUJ1^+$ neurons (87.4% ± 1.9%), none expressed GFAP (a marker for astrocytes), OLIG2 (a marker for oligodendrocytes), or IBA1 (a marker for microglia) (Figures 5F and 5G). Except for a small population of $CHAT^+$ neurons (9.61% ± 5.21%), GABA neurons (81.7% ± 8.1%) predominated in the population of transplanted cells (Figures 5H and 5I). Taken together, GABAergic MSN neuronal identity was successfully established from grafted progenitors.

To further examine hESC-derived MSNs in an adult and diseased environment, we stereotactically transplanted the same amount of LGE-like progenitors into the striatum of quinolinic acid (QA)-lesioned mice, which mimic some motor deficits of human HD (Sanberg et al., 1989) (Figure 6A). These transplanted cells also gradually matured. Four weeks after transplantation, a vast majority of transplanted hN^+ cells remained $hNESTIN^+$ and $KI67^+$ in the grafts (Figure 6B). $KI67^+$ proliferative cells were reduced at week 8 post transplantation and eventually neither were detectable 16 weeks after transplantation (Figure 6B). Whereas in sham control animals no $NEUN^+$ and $DARPP32^+$ cells were left (Figure 6C), in those that received transplantation hN^+ cells repopulated the lesions (Figure 6D) and 48.7% ± 2.8% of them were $DARPP32^+$

Figure 3. Efficient Generation of $DARPP32^+$ GABAergic Neurons from Clinical-Grade hESCs by Small-Molecule Cocktails

(A) Immunostaining images for TUJ1, MAP2, DARPP32, and GABA of day 21–26 cultures derived from hESC lines H9, H1, and clinical-grade Q-CTS-hESC-2 cell line. Scale bar, 50 μm.

(B) Quantification for the percentage of DARPP32 positive cells among $MAP2^+$ and $GABA^+$ cells among $TUJ1^+$ neurons presented in (A). Data are presented as means ± SEM. ns, not significant, *p < 0.05, Student's t test.

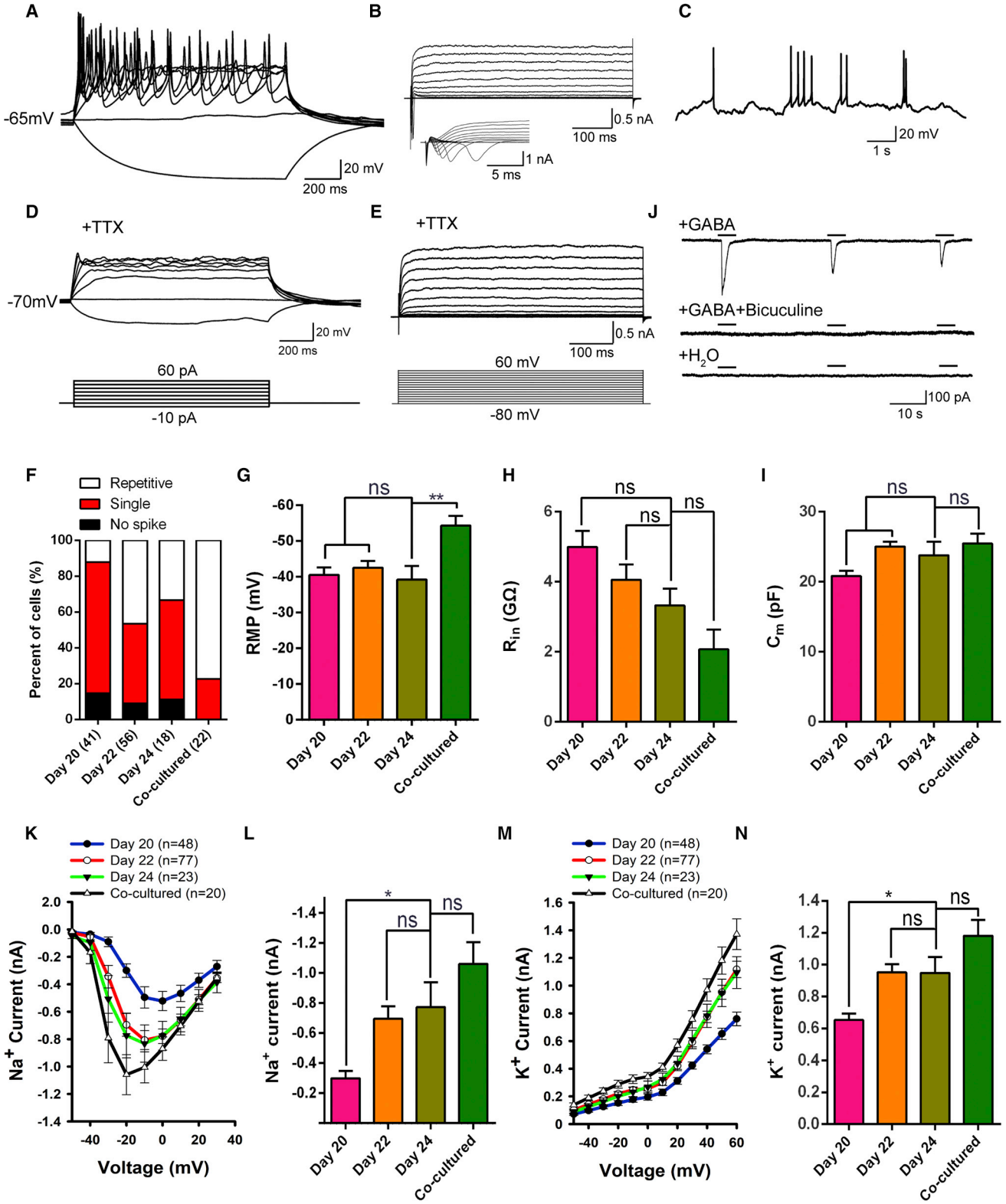
(C) Representative confocal images of GABAergic MSNs markers DARPP32, GABA and CALBINDIN. Scale bar, 50 μm.

(D) Quantification for the percentage of $DARPP32^+$ and $CALBINDIN^+$ cells among $TUJ1^+$ neurons, and $DARPP32^+$ cells among $GABA^+$ neurons presented in (C). Data are presented as means ± SEM.

(E) Immunostaining images of day-21 cultures for mature neuron markers SYNAPSIN1 and post-synaptic protein PSD95 in hESC-derived MSNs. Scale bar, 50 μm. The right panel shows accumulation of PSD95 on spines and dendrites, enlarged from the left image. Scale bar, 10 μm.

(F) Representative images of H9-hESCs cultures immunolabeled with TUJ1 and markers of neuronal subtypes (GABA, TH, CHAT, 5-HT, VGlut1). Scale bar, 50 μm.

(G) Quantification for the percentage of $GABA^+$, TH^+, $CHAT^+$, $5\text{-}HT^+$, and $VGlut1^+$ cells among $TUJ1^+$ cells presented in (F). Efficiencies are presented as the percentage of positive cells ±SEM of all fields counted. ****p < 0.0001, Student's t test.



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(Figure 6E). These graft cells expressed GABA, TUJ1, CTIP2, and FOXP1. A small proportion of them were also SP^+ and ENK^+ cells (Figures 6E–6G). Unsurprisingly, the grafted cells expressed similar proportions of TUJ1, GABA, DARPP32, MEIS2, and KI67 among hN^+ cells between mice transplanted cells by the EB and XLSBA protocols (Figures S3A and S3B). These results demonstrated that hESC-derived MSNs from XLSBA protocol could survive and mature in QA-lesioned mice just as for those from the EB method.

Monthly behavior tests (Figure 6A) showed that compared with the sham transplantation, animals receiving MSN transplantation exhibited significant improvement of behavior rating in open field tests (Figures 6H, 6I, and S4) and rotarod tests (Figure 6J). In addition, a similar degree of motor recovery was observed in mice receiving MSN progenitors generated by the XLSBA protocol and EB method (Figures S5A and S5B). These results indicated that hESC-derived MSNs were functional upon transplantation and could be used for therapy.

DISCUSSION

Cell-based therapy requires readily available cell sources that meet the stringent criteria of human cellular and tissue-based product (HCT/P), namely clinically compliant hESC derivatives in this case. We developed a chemical recipe named XLSBA for rapid and efficient generation of committed striatal LGE-like progenitors and MSNs from hESCs. We also systematically evaluated the safety and efficacy of the XLSBA protocol in neonatal mouse and QA-lesioned mouse model of HD, and found that similar functional improvement after striatal transplantation with cells generated by XLSBA or EB methods. The XLSBA protocol can be used to generate clinical-grade MSNs, and is valuable for standardization and quality control in manufacturing and clinical cell transplantation.

The distinct advantages of small molecules over recombinant proteins include better cell permeability, no immunogenicity, lower cost, and rapid, reversible biological effects. As for the chemical recipe in this study, in addition to the dual-SMAD inhibition by small molecules (Arber et al., 2015; Chambers et al., 2012; Delli Carri et al., 2013; Nicoleau et al., 2013; Qi et al., 2017), all patterning factors were replaced by small molecules such as SAG for SHH signaling (Chen et al., 2002; Danjo et al., 2011), and tankyrase inhibitor XAV939 for inhibition of canonical WNT signaling (Huang et al., 2009). Through carefully titrating the dosage of each small molecule, we obtained an optimal combination of XAV939, SAG, and dual-SMAD inhibition, together with well-designed episodes of administration to efficiently produce committed striatal LGE-like progenitors. Compared with the less efficient and variable proportion of $TUJ1^+$ (22%–93% of total cells) and $DARPP32^+$ (20%–53% of $TUJ1^+$ cells) cells using other protocols (Arber et al., 2015; Aubry et al., 2008; Delli Carri et al., 2013; Golas and Sander, 2016; Ma et al., 2012; Nicoleau et al., 2013), the XLSBA protocol generated ~90% of $DARPP32^+$ GABAergic MSNs and nearly pure $TUJ1^+$ neurons. Such a homogeneous population of defined neurons inevitably advocates the therapeutic potential of cell transplantation.

Accelerating the production of MSNs from hPSCs has considerable potential in modeling development *in vitro*. First, refining the generation process of MSNs from hESCs will provide insight into the signaling programs and decipher key processes of MSN development. Various signalings coordinate temporally and spatially to fine-tune the cellular identity during development. Moderate SHH

Figure 4. Electrophysiological Properties of GABAergic MSNs Differentiated from hESCs

(A) Representative traces of membrane potential responding to step depolarization by current injection steps from −10 pA to +60 pA in 10-pA increments. Membrane potential was current-clamped at around −65 mV.

(B) Representative traces of whole-cell currents in voltage-clamp mode; cells were held at −70 mV; step depolarization from −80 mV to +60 mV at 10-mV intervals was delivered. The inset shows Na^+ currents.

(C) Spontaneous action potentials (APs) recorded from neurons of 24 days differentiation. No current injection was applied (n = 21).

(D) TTX blocked the membrane APs (n = 47).

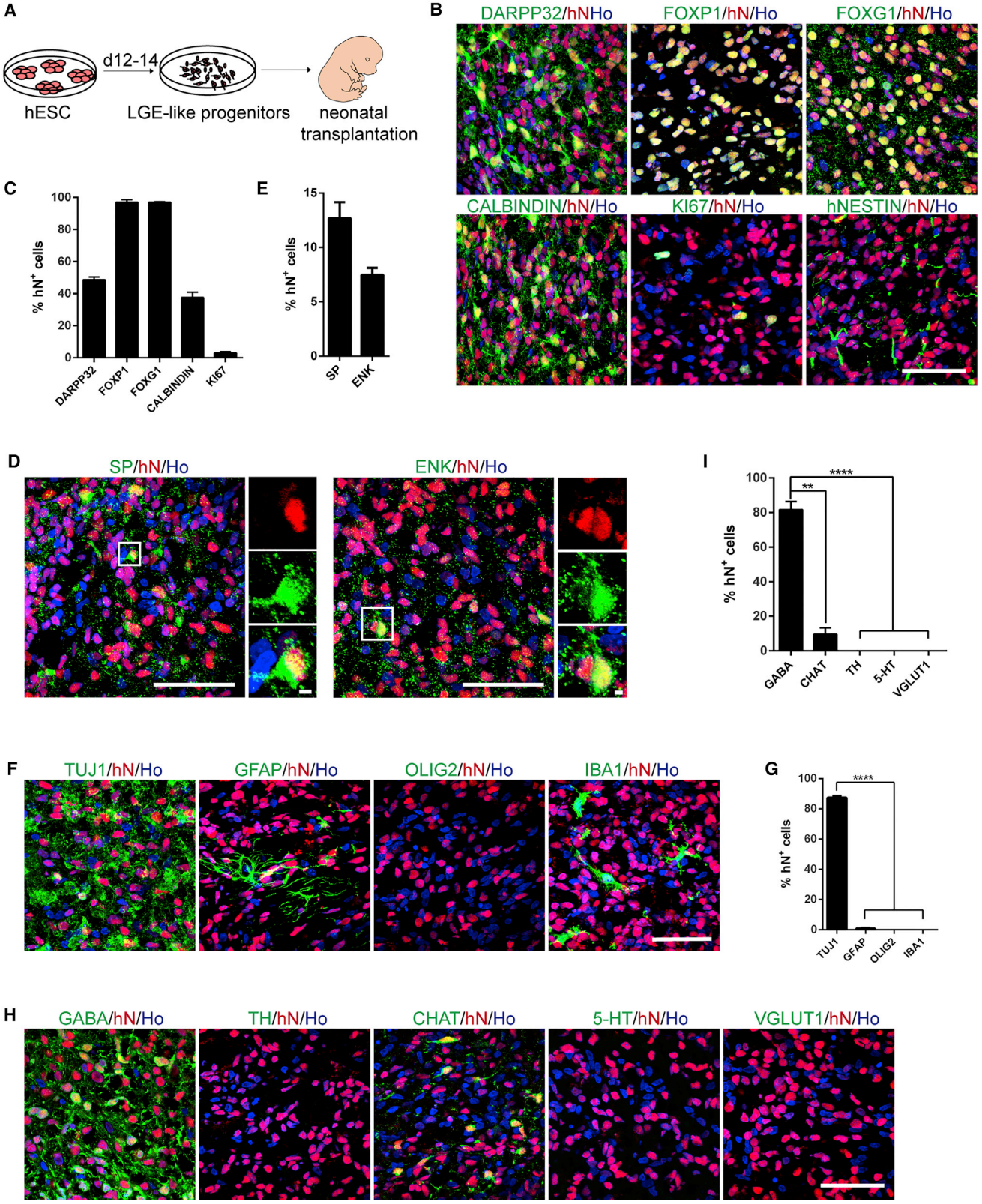
(E) The Na^+ currents of neurons were blocked by TTX. When the cells were treated with 1 μM TTX, the channel of Na^+ currents was blocked completely (n = 47).

(F) Percentages of different AP spikes at 20, 22, and 24 days after differentiation. Increased complexity of AP spikes of differentiated cells over the maturation process. The majority of cells showed single spikes at 24 days differentiation, whereas a larger percentage of cells generated repetitive APs upon co-culture with human glia.

(G–I) Quantification of resting membrane potential (RMP, G), membrane resistance (R_{in}, H) and membrane capacitance (C_m, I) in neurons at 20, 22, and 24 days after differentiation. Error bars indicate ±SEM. ns, not significant. **p < 0.01; one-way ANOVA followed by Tukey's multiple comparisons tests.

(J) Focal application of GABA elicited inward membrane currents (n = 19), which was almost completely eliminated by bicuculline. Focal application of ddH_2O could not induce inward membrane currents (n = 11).

(K–N) Averaged (means ±SEM) current-voltage relationship (I-V curves) for Na^+ and K^+ currents, recorded from hESC-derived neurons. ns, not significant. *p < 0.05; one-way ANOVA followed by Tukey's multiple comparisons tests.



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signaling, for example, is necessary to induce LGE fate (Aubry et al., 2008; Ma et al., 2012). A high dose, however, suppresses the telencephalic fate (Fasano et al., 2010). It is interesting that although a certain level of SHH signaling favors an optimal production of MSNs, extra SHH appears to be dispensable for GABAergic gene expression, as we revealed that LGE-like progenitors are indeed induced even in the absence of SHH. In addition, we found that 100 ng/mL SHH is sufficient for the maximal induction of LGE-like cells compared with that in previous studies, whereas 200 ng/mL SHH is needed to induce LGE identity (Aubry et al., 2008; Ma et al., 2012). This is also supported by a recent study, which reported that hPSCs produce both dorsal cortical glutamatergic and ventral GABAergic neurons by default (Floruta et al., 2017). Differences in the culture methods, such as monolayer versus aggregates, might induce variable intrinsic signaling, and DAPT, which drastically alters the temporal induction, might also reshuffle the orchestration of the cell signaling. Second, the recapitulation of several key developmental stages of MSN with significantly shortened time span would allow us to obtain samples for research more easily and quickly. A combination of hESC differentiation with single-cell transcriptome sequencing technology will encourage us further to identify new markers and probe their function in specific stages of developing MSNs. As human striatum development is a complex process, while neuronal monolayers may lack complexities *in vivo*, it will also be possible in the near future to establish three-dimensional organoids to mirror the process of striatal development more accurately based on the spatial relationships and interplay of multiple cell types during development.

Since human induced PSCs (hiPSCs) possess the same developmental potential as hESCs and carry related pathogenic gene components, they make it possible to model human diseases *in vitro*. Currently many disease-specific hiPSCs such as Parkinson's disease, HD, and Alzheimer's disease have been established (Park et al., 2008). The use of these patient-specific hiPSC lines can help determine the mechanisms of disease initiation, identify the pathogenesis of the disease, and promote the development of early intervention therapies. However, modeling HD with hiPSC-derived neurons often recapitulated partial relevant phenotypes (An et al., 2012; HD iPSC Consortium, 2012; Jeon et al., 2012), and additional cellular stressors are required to trigger essential HD-associated phenotypes such as the formation of mHTT aggregates (Jeon et al., 2012; Nekrasov et al., 2016). Recently, a paper pointed out that the manifestation of HD phenotypes is dependent on cellular age, because age signature retention through direct neuronal conversion could help to exhibit the mHTT aggregates, which are absent in iPSC-derived neurons with erased age marks (Victor et al., 2018). Therefore, it has been challenging to model disease progression using hiPSC-derived MSNs *in vitro* for neurodegenerative diseases with long latency such as HD. However, it becomes possible to capture the phenotypes of late-onset disease when we accelerate the appearance of pathological phenotypes with our short-term culture method *in vitro*. Meanwhile, by establishing a superior method for differentiated cells with minimal variability, HD hiPSCs can also be used for high-throughput drug screening. These may have profound implications for the development of personalized therapeutic interventions for neurodegenerative diseases such as HD.

Cell-based therapy has raised the hope of curing hitherto intractable human neurodegenerative diseases in clinical trials, since targeted differentiation of hPSCs into neuronal subtypes have begun to demonstrate their therapeutic potential in epilepsy, Parkinson's disease, and HD (Cunningham et al., 2014; Kikuchi et al., 2017; Ma et al., 2012).

Figure 5. Survival, Differentiation, and Maturation of hESC-Derived LGE-like Progenitors in the Neonatal Mouse Brain

(A) Schematic overview of the transplantation: LGE-like progenitors from hESCs were differentiated for 12–14 days *in vitro*, and the cell cultures were injected into newborn mouse brain.

(B) Immunostaining of 12-week-old grafts in neonatal mice for human nuclei (hN) and DARPP32, FOXP1, FOXG1, CALBINDIN, KI67, or hNESTIN. Scale bar, 50 μm.

(C) Quantification of immunostaining of markers in (B), showing the percentage of indicated markers among hN^+ cells. Data are presented as mean ± SEM.

(D) Immunostaining of 12-week-old grafts for hN, SP, and ENK. Scale bars, 50 μm. The right panels show enlarged images from the inset. Scale bars, 2 μm.

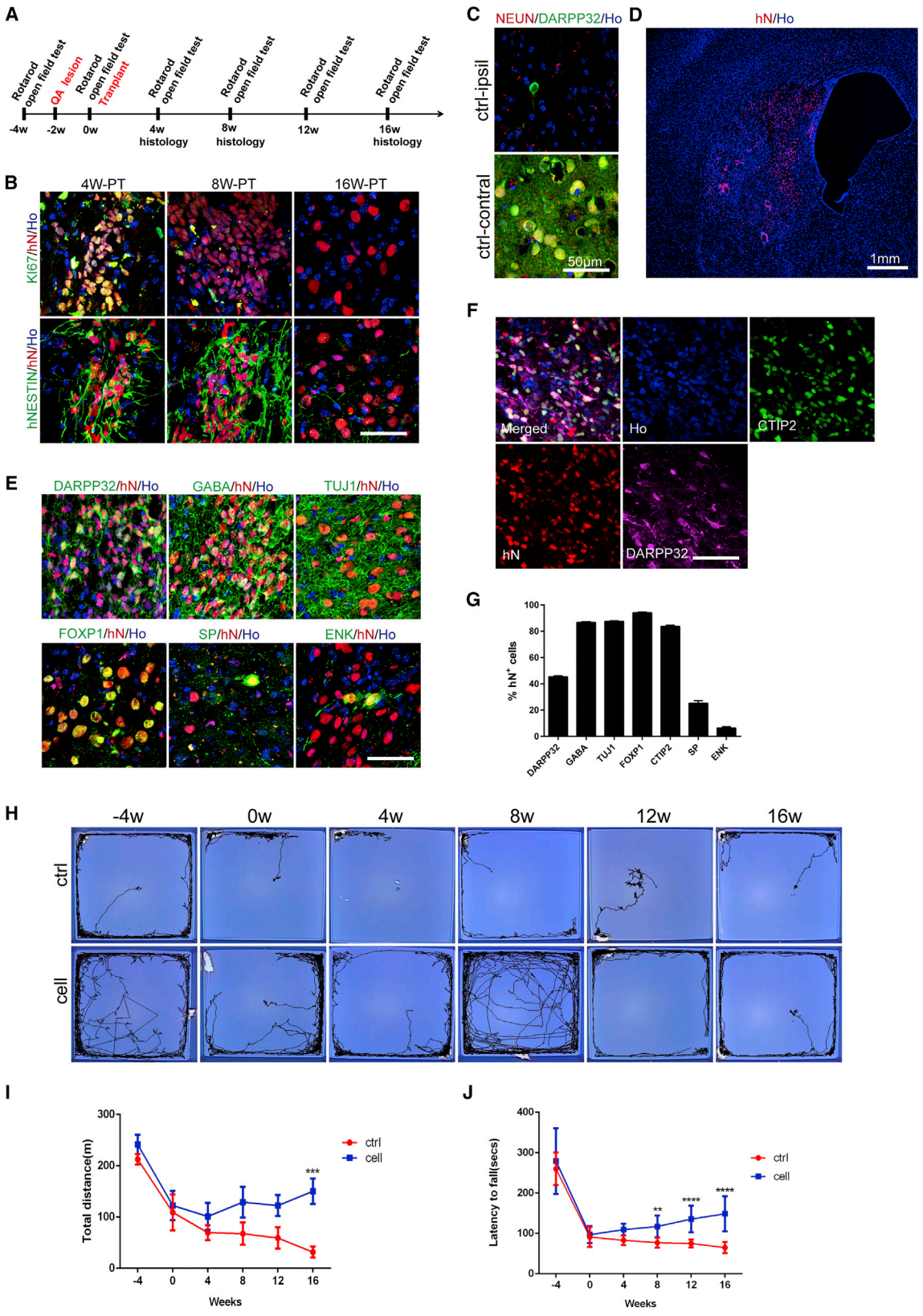
(E) Quantification of immunostaining of markers in (D), shown as a percentage of indicated markers among hN^+ cells. Data are presented as mean ± SEM.

(F) Immunostaining images of 12-week-old grafts for hN and TUJ1, GFAP, OLIG2, and IBA1. Scale bar, 50 μm.

(G) Quantification of immunostaining of markers in (F). The majority of the grafted cell were $TUJ1^+$ neurons with few being GFAP-expressing astrocytes and no $OLIG2^+$ oligodendrocytes and $IBA1^+$ microglia. Data are presented as mean ± SEM. ****$p < 0.0001$.

(H) Representative images of 12-week-old grafts immunolabeled with hN and markers of neuronal subtypes GABA, TH, CHAT, 5-HT, and VGlut1. Scale bar, 50 μm.

(I) Quantification for the proportion of neural subtypes, determined by counting GABA-, TH-, CHAT-, 5-HT-, and VGlut1-positive cells among hN^+ cells presented in (H). Data are presented as mean ± SEM. **$p < 0.01$, ****$p < 0.0001$.



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However, most hESC lines worldwide are research grade, as they were produced in the presence of animal components and without biosafety evaluation. Therefore they are not ideal for use in clinical trials. Currently xeno-free hESCs are available, such as the biosafe clinical-grade hESC lines accredited at the Chinese National Institutes for Food and Drug Control (Gu et al., 2017). In the context of clinical applications, in addition to hESCs their differentiated progeny cells must also be developed in adherence to Good Manufacture Practices, which will guarantee clinical-grade level for all procedures and reagents used for the generation of the cells to be grafted. To date, clinical-grade MSNs remain unavailable for HD therapy. With this goal in mind, we used commercially available CTS reagents that are free of xenogeneic components and obtained MSN precursor cells from clinical-grade hESCs. These would provide therapeutically safe cell sources for HD. For fetal grafts, the limited time for tests of quality control raised a major challenge (Nicoleau et al., 2011). Thus, differentiating hPSCs into neural progenies in an accelerating manner would not only provide timely, reliable, and scalable supply of cell sources but also save enough time to perform rigorous tests to ensure the biosafety of the preparation. The faster time frame and validated quality would enhance the reproducible production and facilitate the routine application of hPSC-derived striatal cells in clinical implementation. Therefore, this would take stem cell-based neural replacement therapy for HD patients one step closer to the clinic.

EXPERIMENTAL PROCEDURES

hESC Culture and Neural Differentiation

hESCs (H9 and H1) were maintained on Matrigel; clinical-grade Q-CTS-hESC-2 line was cultured on vitronectin, and were dissociated with EDTA (Gibco) for passaging or dispase (Gibco) for differentiation (Beers et al., 2012). Differentiation of striatal MSNs by EB protocol is based on the procedures as previously reported (Ma et al., 2012). In brief, hESCs were detached to form EBs with dispase. After adhering to a plastic surface on day 7, definitive neuroepithelial cells with neural-tube-like rosettes were mechanically picked and transferred to dishes to form neurospheres on days 15–17. For immunostaining, neural progenitor clusters were dissociated with Accutase and plated on the Matrigel-coated coverslips. SHH, VPA, brain-derived neurotrophic factor (BDNF), glial cell line-derived neurotrophic factor (GDNF), insulin-like growth factor 1 (IGF-1), and cyclic AMP (cAMP) were added at appropriate times. For differentiation of MSNs by small molecules, differentiation media were described as previously (Kirkeby et al., 2012a, 2012b). On day 0, hESCs were detached with dispase to form EBs in NIM. The EBs were then plated in NPM onto plastic plates coated with Matrigel on day 4. Within days 11–16 of differentiation, the cells were dissociated to small clusters with Accutase and replated onto Matrigel-coated plates in NDM. BDNF, GDNF, ascorbic acid (AA), and DAPT were added for terminal differentiation. For clinical-grade hESC differentiation, all CTS reagents were purchased from Life Technologies. Cells were reseeded on polyornithine/fibronectin/laminin-coated coverslips. Small-molecule compounds were as follows: XAV939 (0–4 μM, Selleck), dorsomorphin (100 nM, Selleck), LDN-193189 (100 nM, Selleck), SB431542 (10 μM, Merck), SAG (0–100 nM, Stemgent), purmorphamine (0–1 μM, Stemgent), AA (0.2 mM, Sigma), and DAPT (1 μM, Tocris). Recombinant growth factors were as follows: SHH (C25II; 0–500 ng/mL), NOGGIN (200 ng/mL), BDNF (20 ng/mL), GDNF (10 ng/mL), all from Peprotech. For detailed differentiation procedures, see Supplemental Experimental Procedures.

hESC-Derived Neural Progenitors and Human Astrocyte Co-cultures

Human astrocytes were purchased from ScienCell. Astrocyte culture medium consisted of DMEM (Gibco) supplemented with 10% fetal bovine serum (Gibco) and non-essential amino acids (Gibco). For electrophysiology, human astrocytes were passaged onto coverslips coated with poly-D-lysine (Sigma) in 24-well plates

Figure 6. hESC-Derived Progenitors Survive and Produce MSNs in QA-Lesioned Mice

(A) Experimental scheme for the transplantation of QA-lesioned mice and behavior tests.

(B) Microtome sections of QA-lesioned adult mouse brains at 4, 8, and 16 weeks post transplantation were analyzed by immunohistochemistry for hN and NESTIN or KI67. Scale bar, 50 μm.

(C) Adult SCID mice injected with QA and without cell transplantation showed a loss of neurons marked by NEUN staining and MSNs marked by DARPP32 staining on the lesion side (ctrl-ipsil), while normal on contralateral side of the striatum (ctrl-contral). Scale bar, 50 μm.

(D) Low magnification of a coronal brain section showing transplanted human cells labeled with hN residing in the striatum. Scale bar, 1,000 μm.

(E) Immunostaining images of 16-week-old grafts for GABA, DARPP32, TUJ1, FOXP1, substance P (SP), and enkephalin (Enk). Scale bar, 50 μm.

(F) Triple staining for DARPP32, CTIP2, and hN on a section of a 16-week-old grafts. Scale bar, 50 μm.

(G) Quantification of immunostaining of striatal markers, shown as a percentage of hN^+ cells. Data are presented as means ± SEM.

(H) Representative tracing images of animals in open field tests.

(I) Open field tests indicated increased total distances in animals receiving cell grafts compared with sham control. Open field behaviors were analyzed by two-way ANOVA (***$p < 0.001$; control group, n = 13; cell group, n = 13).

(J) Rotarod tests showed increased latency in animals transplanted with hESC-derived progenitors compared with sham control. The tests were analyzed by two-way ANOVA (**$p < 0.01$, ****$p < 0.001$; control group, n = 13; cell group, n = 13).

See also Figures S3–S5.

(5×10^4 per well) and incubated overnight at 37°C in the CO_2 incubator. On the second day, ~2×10^4 GABA neural progenitors differentiated from hESCs between 11 and 16 days were dissociated with Accutase and replated on top of human astrocytes in NDM supplemented with BDNF, GDNF, AA, and DAPT. Half of the medium was changed every other day. The cells were maintained in this medium for 8 days or more to detect their electrophysiological characteristics.

Immunocytochemical Analyses

Cultures were fixed in 4% paraformaldehyde, washed with PBS, and incubated in a blocking buffer (10% donkey serum and 0.3% Triton X-100 in PBS) for 60 min at room temperature before being incubated in primary antibodies (see Table S2) overnight at 4°C. Secondary antibodies were species-specific Alexa-dye conjugates (Invitrogen) and nuclei were stained with Hoechst 33258 (Ho in figures). Images were collected with a Zeiss LSM710 or Zeiss LSM780 (Carl Zeiss, Germany).

RNA Isolation and qRT-PCR

Total RNA was collected from cultured cells with TRIzol (Life Technologies). cDNA was generated from 500 ng of total RNA with the PrimeScript RT Reagent Kit with gDNA Eraser (Takara), and used as a template for the qPCR. qPCR was performed with the QuantStudio 6 Flex instrument (Applied Biosystems) by SYBR Green detection. The use of primer sequences and details are summarized in Table S3.

Neural Transplantation and Quinolinic Acid Striatal Lesions

All animal experiments were carried out in accordance with the instructions for the Care and Use of Animals in Research published by the Institute of Zoology of the Chinese Academy of Sciences. Transplantation into the neonatal NOD-SCID mice was conducted as described previously (Danjo et al., 2011). The establishment of QA-lesioned mouse model of HD and cell transplantation were performed as described previously (Ma et al., 2012) and as detailed in Supplemental Experimental Procedures.

Electrophysiology

Whole-cell current and clamp recordings of hESC-derived neurons were performed as described in detail in Supplemental Experimental Procedures.

Behavioral Tests

Behavioral tests were conducted before and after transplantation monthly until the animals were sacrificed.

Open Field Test

Mice were placed in the center of activity chambers (Clever Sys; $48 \times 48 \times 42$ cm^3). Activities were recorded for 10 min under normal conditions of lighting. Quantitative analysis was done on total distance.

Rotarod Test

An accelerating rotarod (YLS-4C, China) was used to test motor coordination. The mouse was placed on a rotating rod that accelerated from 4 to 40 rotations per minute in a period of 300 s. The period of time the mouse stayed on the rod was monitored and the three of a total of five runs in which the mouse performed best were recorded.

Statistical Analysis

For all experiments, analysis was derived from at least three independent experiments. GraphPad Prism 6 software was used for statistical analysis. In all studies, comparison of mean values was conducted with unpaired t test, one-way ANOVA, or two-way ANOVA. In all analyses, statistical significance was determined at the 0.01%, 0.1%, 1%, or 5% level ($p < 0.0001$, $p < 0.001$, $p < 0.01$, $p < 0.05$, respectively).

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, five figures, and three tables and can be found with this article online at https://doi.org/10.1016/j.stemcr.2018.08.005.

AUTHOR CONTRIBUTIONS

M.W. contributed to data collection, analysis, and interpretation, and manuscript writing. D.Z. and C.B. established the HD model, and performed cell transplantation and behavior tests. T.M. performed whole-cell current and clamp recordings. W.Z. and L.X. performed qPCR analysis. Z.T. revised the manuscript. B.H. and Y.W. designed and supervised the research and revised the manuscript.

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Stem Cell Reports, Volume *11*

Supplemental Information

A Chemical Recipe for Generation of Clinical-Grade Striatal Neurons from hESCs

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SUPPLEMENTAL INFORMATION

SUPPLEMENTAL FIGURES AND TABLES

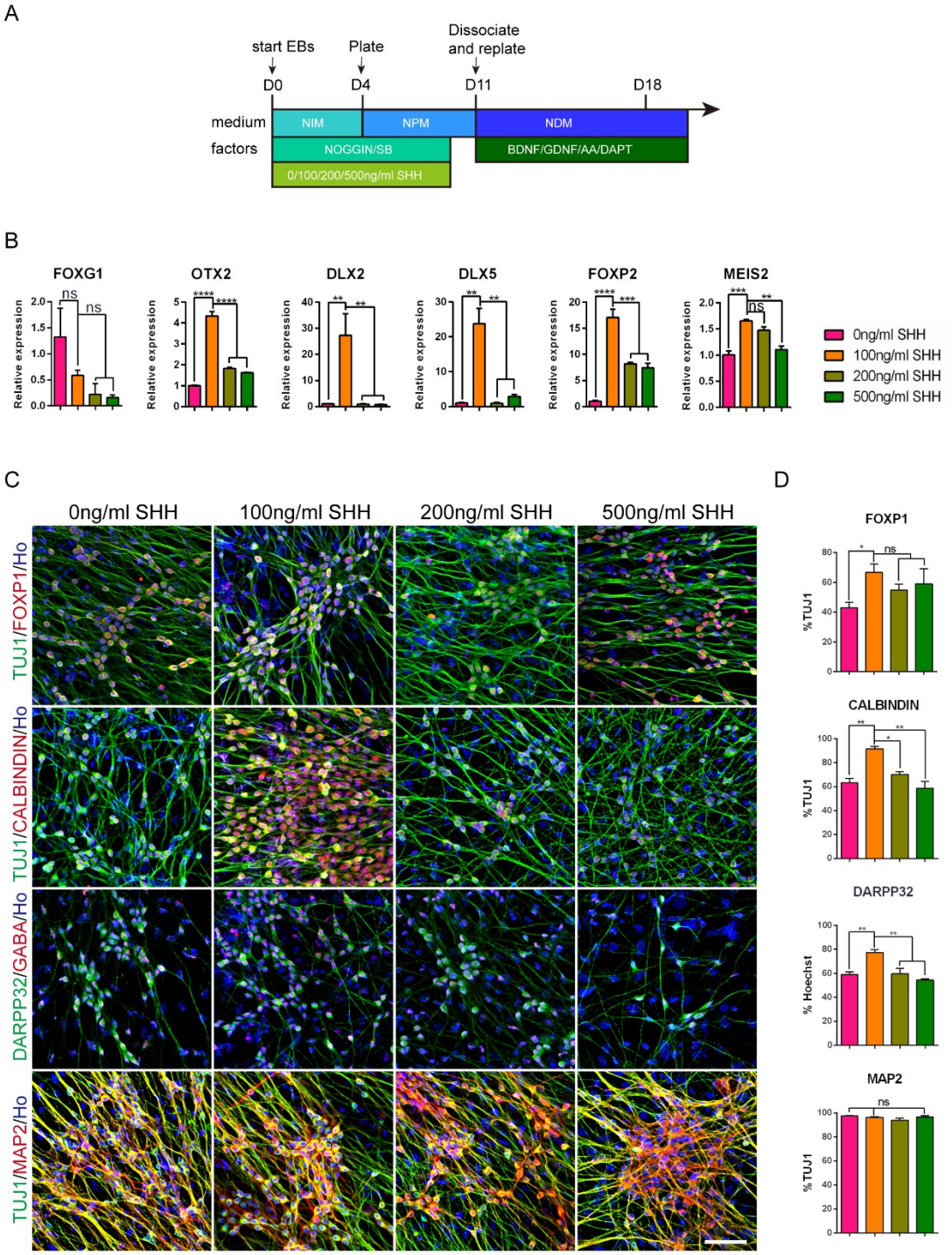


Figure S1. Robust generation of striatal MSNs by optimizing SHH pathway activity.

(A) Overview of differentiation protocol for hESCs. SB represents SB431542; AA represents ascorbic acid.

(B) Gene expression changes of cultures treated with different concentrations of SHH as measured by qRT-PCR on differentiation 14 day. Data are presented as the mean of three independent experiments;

Error bars represent SEM. ns, no significant, **P < 0.01, ***P < 0.001, ****P < 0.0001. One-way ANOVA, followed by Tukey's multiple comparisons tests.

(C) Expression of MSN markers after treatment with different concentrations of SHH, imaged by confocal microscopy. Ho represents Hochest33258. Scale bars, 50 μm.

(D) Efficiencies of MSN differentiation, determined by counting FOXP1, CALBINDIN, DARPP32 and MAP2-positive cells. Efficiencies are presented as the percentage of positive cells plus or minus the SEM of all fields counted. ns, no significant, *P < 0.05, **P < 0.01. One-way ANOVA, followed by Tukey's multiple comparisons tests.

See also Figure 1.

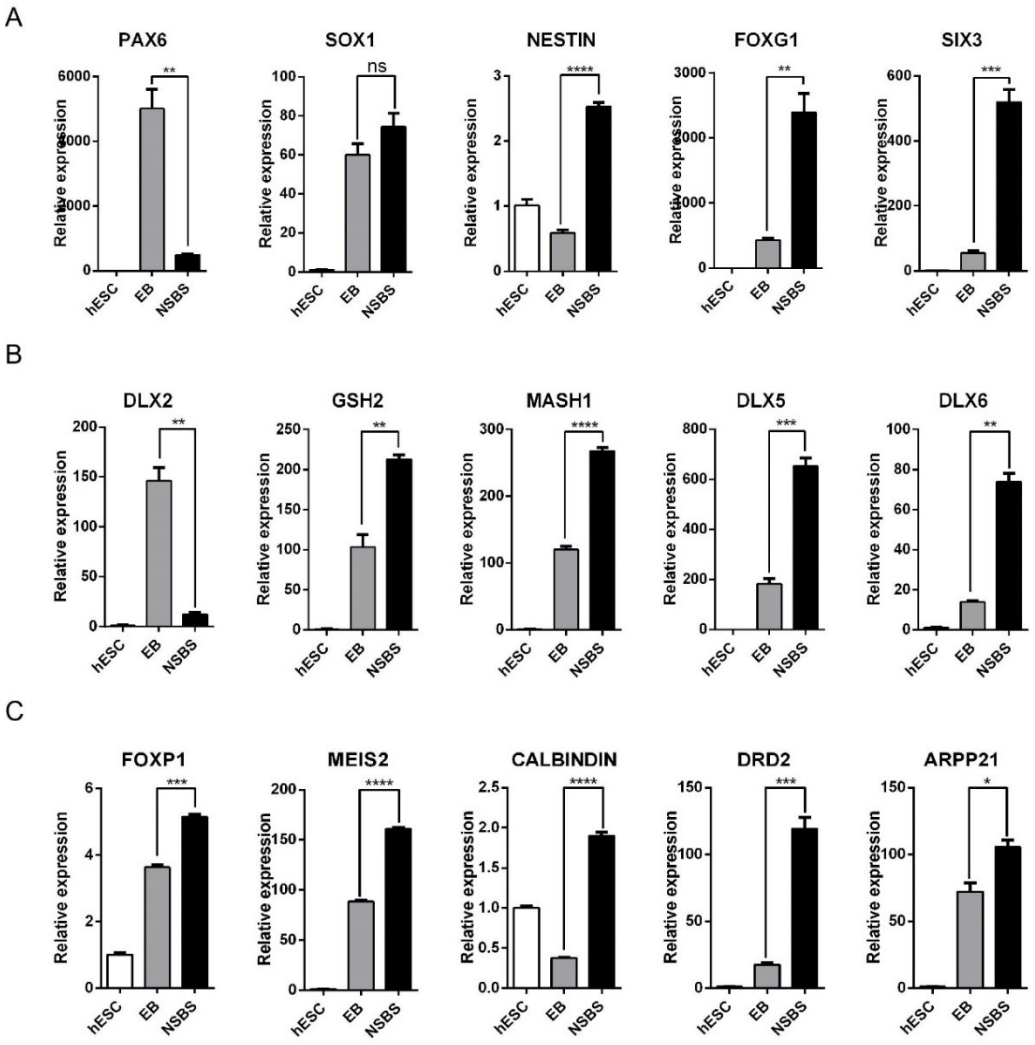


Figure S2. NSBS differentiation protocol is more efficient compared to EB protocol.

(A-C) QPCR analysis of gene expression at different differentiation stages of EB and NSBS protocol. Data are presented as the mean of three independent experiments; error bars represent SEM. ns, no significant, *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. Student's *t*-test.

See also Figure 1.

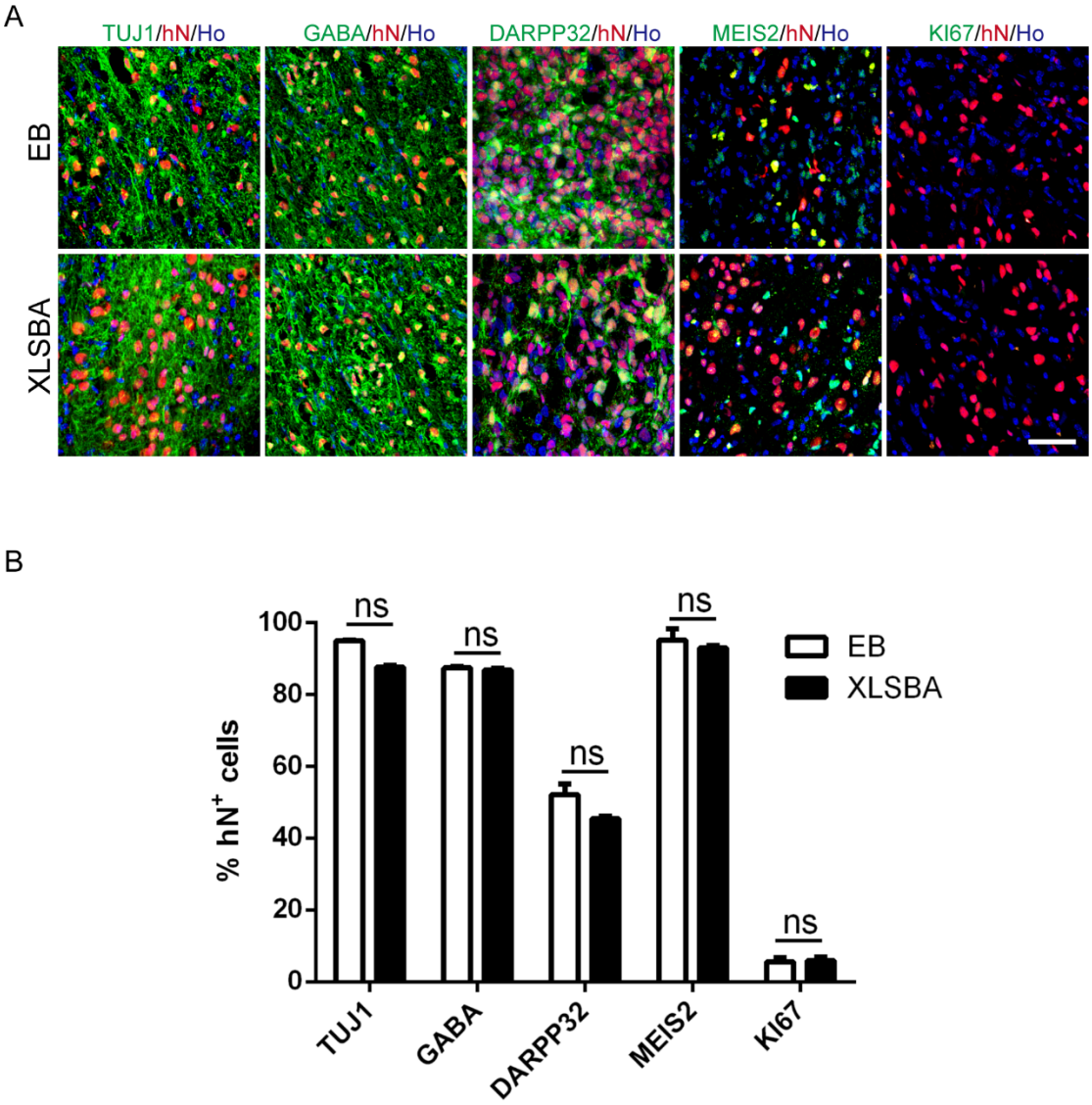
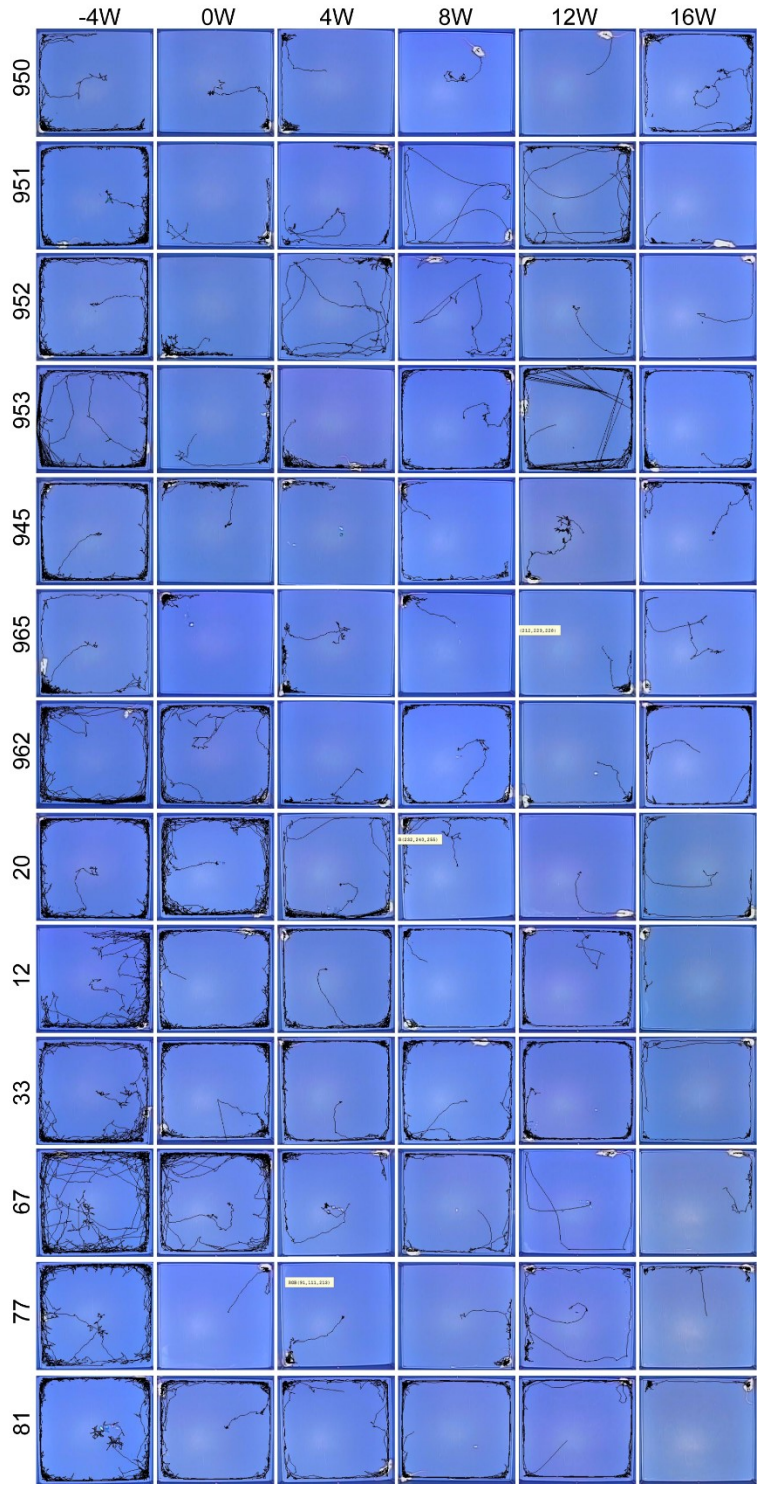


Figure S3. Comparisons of the survival and differentiation of grafted cells between EB and XLSBA group.

(A) Immunostaining images of 16-week-old grafts for TUJ1, GABA, DARPP32, MEIS2 and KI67. Grafted human cells were revealed by human nuclei (hN) staining. Scale bars, 50 μm. (B) Quantification for the proportion of indicated markers in (A). ns, no significant, Student's t-test. Data are presented as mean ±SEM.

See also Figure 6.

A Control group:



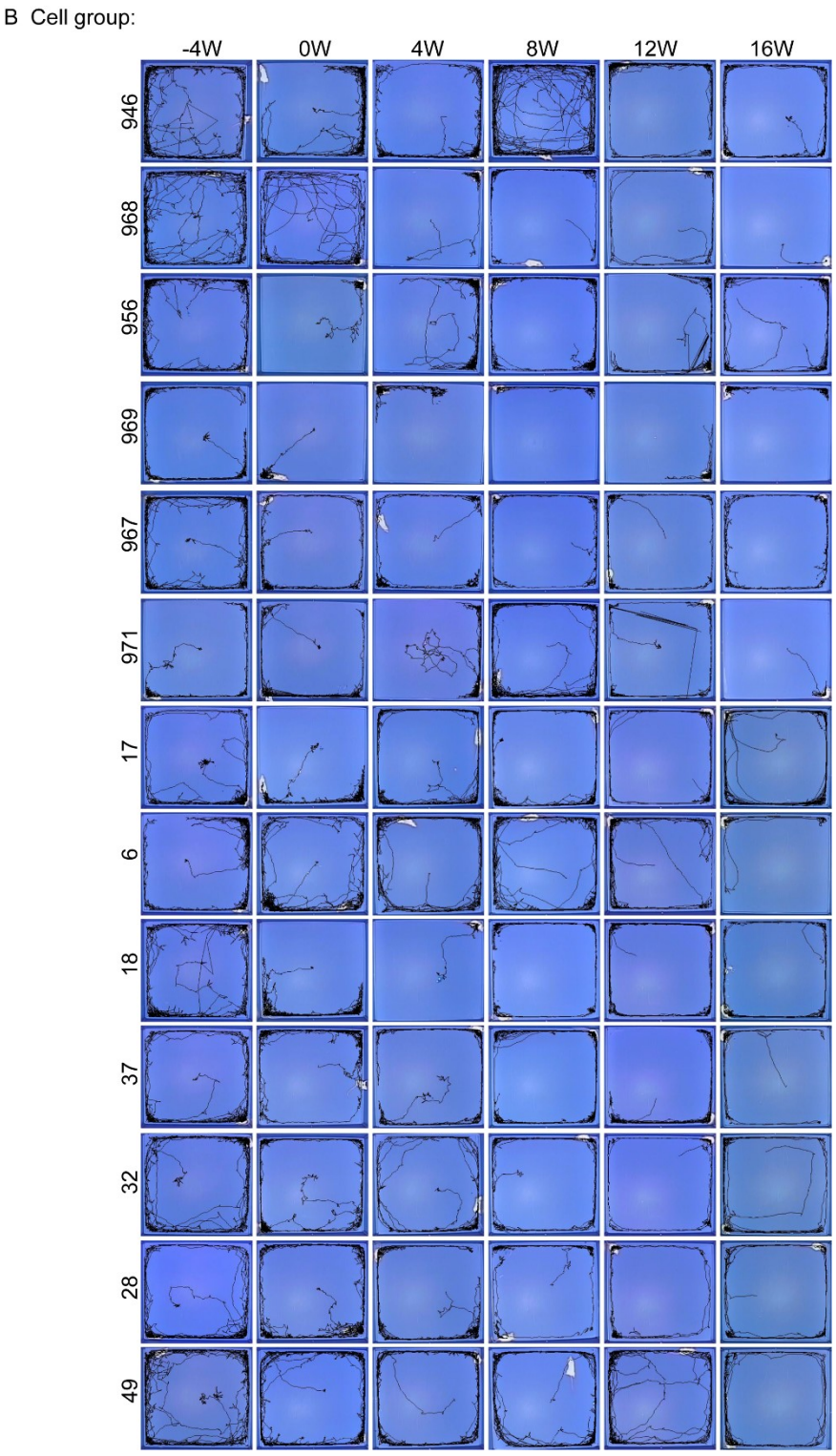


Figure S4. Functional recovery is evaluated by behavior tests after transplantation of hESC-derived LGE progenitors.

(A) Traces of open field test in animals receiving sham operation. (B) Traces of open field test in animals receiving cell grafts. The numbers represent the identifier of each mouse.

See also Figure 6.

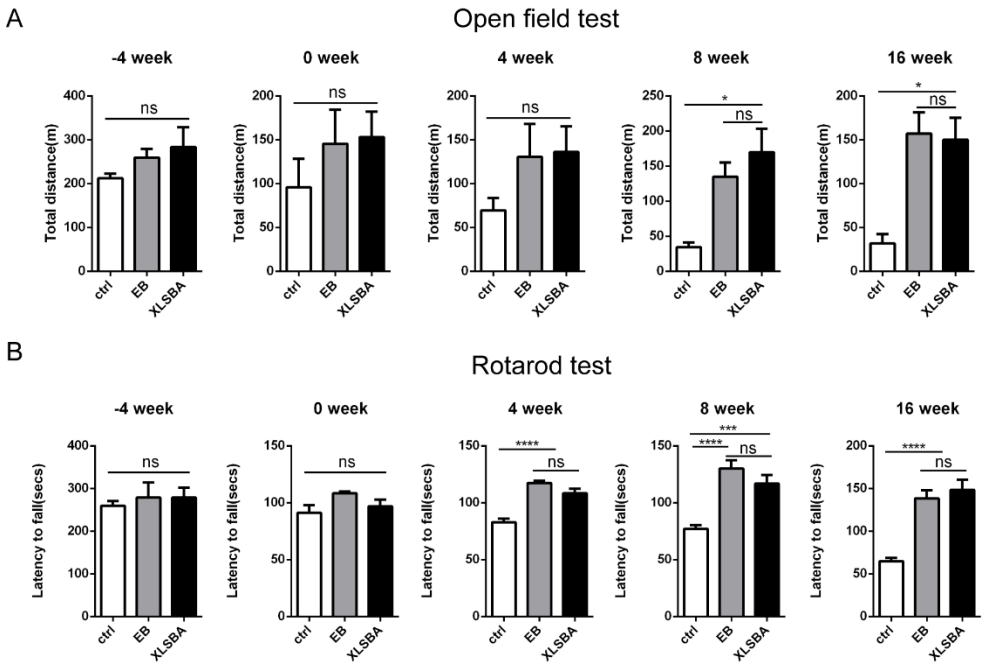


Figure S5. Comparisons of behavior performances between EB and XLSBA group.

(A) Open field tests indicated increased total distance in mice grafted with cells generated by EB and XLSBA protocol but not control group after 8 week and 16 week post-transplantation. The total distance showed no change in mice between EB and XLSBA group. (B) The rotarod test showed increased latency in mice transplanted with cells generated by EB and XLSBA protocol but not control group starting from 4 week post-transplantation. The performance showed no difference between EB and XLSBA group. The tests were analyzed by one-way ANOVA (ns, no significant, *p< 0.05, ***p< 0.001, ****p< 0.0001, control group: n =13, EB group: n=13, XLSBA group: n=13). Data are presented as mean ±SEM.

See also Figure 6.

Table S1. Electrophysiological properties of neurons during differentiation.

Group	D20 (n=39)	D22 (n=54)	D24 (n=16)	Co-culture (n=20)
RMP (mV)	-40.5179±2.1788	-42.5796±1.8247	-39.2188±3.8054	-54.36±2.6579
R_{in} (GΩ)	4.9938±0.4654	4.0574±0.4345	3.3242±0.4852	2.0702±0.568
C_m (pF)	20.8231±0.7453	25.0344±0.7008	23.7881±1.9527	25.467±1.3996

RMP: Resting Membrane Potential; R_{in}: Input Resistance; C_m: Capacitance.

Tables S2. Primary antibodies used for immunofluorescence staining.

Antibody	Species	Catalog Number	Company	Dilution
CALBINDIN	rabbit	AB1778	Millipore	1/1000
ChAT	goat	AB144P	Millipore	1/500
CTIP2	rat	ab18465	Abcam	1/500
DARPP32	rabbit	ab40801	Abcam	1/500
DLX2	rabbit	ab5726	Millipore	1/500
Met-Enkephalin	rabbit	ab5026	Millipore	1/1000
FOXG1	rabbit	ab18259	Abcam	1/500
FOXP1	rabbit	ab16645	Abcam	1/500
GABA	rabbit	A2052	Sigma	1/500
GABA	mouse	A0310	Sigma	1/200
GFAP	rabbit	180063	Invitrogen	1/200
GSH2	rabbit	ABN162	Millipore	1/500
hNuclei	mouse	MAB1281	Millipore	1/200
5-HT	rabbit	20080	Immunostar	1/500
IBA1	rabbit	019-19741	Wako	1/200
KI67	rabbit	AB9260	Millipore	1/500
MAP2	mouse	ab11267	Abcam	1/1000
NESTIN	mouse	MAB5326	Millipore	1/500
NESTIN	rabbit	ABD69	Millipore	1/1000
NEUN	mouse	MAB377	Millipore	1/500
OLIG2	goat	SC-19969	Santa Cruz	1/200
PAX6	rabbit	ab5790	Abcam	1/500
PAX6	mouse	MAB5552	Millipore	1/500
PSD95	goat	ab12093	Abcam	1/1000
PV	mouse	MAB1572	Millipore	1/2000
SOX1	goat	AF3369	R&D	1/2000
SOMATOSTATIN	rabbit	ab5494	Millipore	1/1000
Substance P	rabbit	AB1566	Millipore	1/1000
Synapsin1	mouse	106011	Synaptic systems	1/500
Synaptotagmin1	mouse	105011	Synaptic systems	1/500
TH	rabbit	AB152	AB152	1/200
TUJ1	mouse	T8660	Sigma	1/1000
TUJ1	rabbit	T2200	Sigma	1/1000
VGLUT1	rabbit	135302	Synaptic systems	1/2000

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Table S3. Sequence of qRT-PCR primers used in this study.

Target gene	Primer forward	Primer reverse
ASCL1	GTCCTGTCGCCCACCATCTC	CCCTCCCAACGCCACTGAC
ARPP21	GTGCAAAGCGTGATGGTTTCC	CCTTGACCTGCCTGGTTAGG
CALBINDIN1	ATCAGGACGGCAATGGATAC	TAAGAGCAAGATCCGTTCGG
CTIP2	ATCCTCAGCCCCTTTTGTTT	GCCGTTGTTCCTGAATTGTT
DARPP32	CCTGAAGGTCATCAGGCAGT	GGTCTTCCACTTGGTCCTCA
DCX	CAAGTCTAAGCAGTCTCCCATC	ATAGCCCTGTTGGACACTTG
DLX2	ACCAGACCTCGGGATCCGCC	CTGCGGGGTCTGAGTGGGGT
DLX5	TTCAGAAGACTCAGTACCTCGC	GAGTTACACGCCATTGGGTC
DLX6	TACCTCCAGTCCTACCACAAC	AATAAATGGTCCGAGGCTTCCG
DRD2	CTGAGGGCTCCACTAAAGGAG	CATTCTTCTCTGGTTTGGCG
EAR	GAGGCTGAGGCAGGAGAATCG	GTCGCCCAGGCTGGAGTG
EMX2	GGGATCCGTCCACCTTCTAC	CTCAAAGGCGTGTTCCAGCC
EN1	CGTGGCTTACTCCCCATTTA	TCTCGCTGTCTCTCCCTCTC
FOXG1	TGTTGACTCAGAACTCGCTGG	CTGCTCTGCGAAGTCATTGAC
FOXP1	CTACCGCTTCCATGGGAAATC	CTGTTGTCACTAAGGACAGGG
FOXP2	AATGTGGGAGCCATACGAAG	GCCTGCCTTATGAGAGTTGC
GAPDH	GAAGGTGAAGGTCGGAGTC	GAAGATGGTGATGGGATTTC
GBX2	CTCGCTGCTCGCCTTCTC	GCCAGTCAGTCAGATTGTCATCCG
GSH2	TATGTCGACTCGCTCATCATC	CAAGCGGGATGAAGAAATCC
HOXA4	ACGCTCTGTTTGTCTGAGCGCC	AGAGGCCGAGGCCGAATTGGA
LMX1A	CGCATCGTTTCTTCTCCTCT	CAGACAGACTTGGGGCTCAC
MAP2	AAAGCTGATGAGGGCAAGAA	GGCCCCTGAATAAATTCCAT
MEIS2	GATGAAAGAGACGGCAGCTCC	GGGTTGAGGTTGCATCATCG
NESTIN	GGAAGAGAACCTGGGAAAGG	CTTGGTCCTTCTCCACCGTA
NKX2.1	ACCGGGTTCAGACTCAGTTC	ATCGACATGATTCGGCGTCGG
NOLZ1	ACATTTTGCACCCCGAGTAC	GGAGTACGGCTTGAAACTCG
OTX2	TCAACTTGCCCGAGTCGAGG	CAATGGTCGGGACTGAGGTG
PAX6	TCCATCAGTTCCAACGGAGAA	GTGGAATTGGTTGGTAGACAC
SIX3	CCGGAAGAGTTGTCCATGTT	CGACTCGTGTTTGTTGATGG
SOX1	ATGCACCGCTACGACATGG	CTCATGTAGCCCTGCGAGTTG
SOX2	CAAGATGCACAACTCGGAGA	CGGGGCCGGTATTTATAATC
TBR2	CACCGCCACCAAACTGAGAT	CGAACACATTGTAGTGGGCAG
TUJ1	AACGAGGCCTCTTCTCACAA	GGCCTGAAGAGATGTCCAAA

SUPPLEMENTAL EXPERIMENTAL PROCEDURES

Generation of striatal MSNs from hESCs with chemical cocktails

Differentiation media were described as previously (Kirkeby et al., 2012a; Kirkeby et al., 2012b): neural induction medium (NIM) consisted of DMEM/F12: Neurobasal (1:1), N2 supplement (1:100), B27 supplement (1:50) and GlutaMAX (1:100); neural proliferation medium (NPM) consisted of DMEM/F12: Neurobasal (1:1), N2 supplement (1:200), B27 supplement (1:100) and GlutaMAX (1:100); and neural differentiation medium (NDM) consisted of Neurobasal, B27 supplement (1:50) and GlutaMAX (1:100). For differentiation, hESCs were detached with dispase to form EBs in NIM. Then the EBs were plated in NPM onto plastic plate coated with Matrigel on day 4. From D0 to D9, SB431542 and LDN-193189, Dorsomorphin or NOGGIN was present in the medium. Patterning factors SHH-C24II, SAG or Purmorphamine were also in present in the medium from d0-d9 or d0-d5. On D11 of differentiation, the cells were dissociated to small clusters with Accutase and replated onto the matrigel-coated plates in NDM. Brain-derived neurotrophic factor (BDNF), glial cell line-derived neurotrophic factor (GDNF), ascorbic acid (AA) and DAPT were added for terminal differentiation. For clinical-grade hESC differentiation, all Clinical Therapy Systems (CTS) reagents were purchased from Life Technologies, and cells were replated on the PO/FN/lam-coated plates (Fibronectin, Gibco; Laminin, Sigma; Polyornithine, Sigma). Small-molecule compounds were as follows: XAV939 (0-4μM, Selleck), Dorsomorphin (100 nM, Selleck), LDN-193189 (100 nM, Selleck), SB431542 (10 μM, Merck), SAG (0-100nM, Stemgent), Purmorphamine (0-1μM, Stemgent), ascorbic acid (200 mM, sigma) and DAPT (1μM, Tocris). Recombinant growth factors were as follows: SHH (C25II, 0–500 ng/ml), NOGGIN (200 ng/ml), BDNF (20 ng/ml) and GDNF (10 ng/ml), all from Peprotech.

1. **Day 0**: Check that the colonies appear pluripotent by visual criteria, and if needed, remove differentiated colonies from the culture. Aspirate E8 medium and add appropriate volume of dispase (2U/ml) to the cells (i.e., 1 ml for a 6-well) and incubate at 37°C for 8-12 min. When the edges of hPSC colonies begin to curl, aspirate off the dispase.
2. Add 2 ml of fresh E8 medium to each well. Gently pipette colonies off the dish. Collect hESC colonies into a 15-ml tube, and centrifuge at 600rpm for 2 min. Alternatively, let the hESC colonies settle down by letting the tube rest for 3–5 min. Aspirate off the supernatant without disturbing the cells.
3. To start differentiation, resuspend the cells in differentiation medium: NIM + LDN193189 (100nM) + SB431542 (10 μM) +XAV939 (4 μM) +SAG (100 nM). Plate cell suspension in non-treated cell culture dishes for EB formation.
4. **Day 1**: Transfer EB suspensions to a tube and centrifuge at 600rpm for 2 min. Alternatively, let the EBs settle down by letting the tube rest for 3–5 min. Aspirate the medium and re-suspend the EBs in new NIM medium + LDN193189 (100nM) + SB431542 (10 μM) +XAV939(4 μM)+SAG (100 nM). Transfer EBs into the same dishes.
5. **Day 2-3**: Change the medium as step 4 every day.
6. **Day 4**: Transfer EB suspensions to a tube and spin down gently at 600rpm for 2 min. Resuspend EBs in NPM medium+ LDN193189 (100 nM) + SB431542 (10 μM) +XAV939(4 μM)+SAG (100 nM). Replate the EBs into wells coated with PO/lam/FN or Matrigel.
7. **Day 5**: Change the medium with NPM medium+ LDN193189 (100 nM) + SB431542 (10 μM).
8. **Day 7**: Change the medium with NPM medium+ LDN193189 (100 nM) + SB431542 (10 μM).

9. **Day 9**: Change the medium with NPM medium without any small molecules.
10. **Day 11-16**: Change the medium with NDM medium+BDNF (20 ng/ml) +GDNF (10 ng/ml) +AA (0.2 mM) +DAPT (1μM). Or passage cells: wash cells with PBS, and add Accutase to the wells. Incubate the cells at 37℃ for 3-5 min. Dissociate cells with a pipette and spin down at 600rpm for 3 min (before spinning, take an aliquot out for cell counting). Resuspend cells in NDM medium + BDNF (20 ng/ml) +GDNF (10 ng/ml) +AA (0.2 mM) +DAPT (1μM). Then replate the cells into wells coated with PO/lam/FN or Matrigel in a density of 2.5–5x10^6 cells for one well of a 6-well plate.
11. **Day 17-24**: Change the medium with NDM medium+BDNF (20 ng/ml) +GDNF (10 ng/ml) +AA (0.2 mM) +DAPT (1μM) every other day.

Reagent setup:

Neural induction medium (NIM):

H9/H1 hESC	Q-CTS-hESC-2	Volume
DMEM/F12	CTS™ KnockOut™ DMEM/F-12	240 ml
Neurobasal	CTS™ Neurobasal® Medium	240 ml
N2 (100X)	CTS™ N-2 Supplement	5 ml
B27 (50X)	CTS™ B-27® Supplement	10 ml
GlutaMAX (100X)	CTS™ GlutaMAX™-I	5 ml
Total		500 ml

Neural proliferation medium (NPM):

H9/H1 hESC	Q-CTS-hESC-2	Volume
DMEM/F12	CTS™ KnockOut™ DMEM/F-12	243.75 ml
Neurobasal	CTS™ Neurobasal® Medium	243.75 ml
N2 (100X)	CTS™ N-2 Supplement	2.5 ml
B27 (50X)	CTS™ B-27® Supplement	5 ml
GlutaMAX (100X)	CTS™ GlutaMAX™-I	5 ml
Total		500 ml

Neural differentiation medium (NDM):

H9/H1 hESC	Q-CTS-hESC-2	Volume
Neurobasal	CTS™ Neurobasal® Medium	485 ml
B27 (50X)	CTS™ B-27® Supplement	10 ml
GlutaMAX (100X)	CTS™ GlutaMAX™-I	5 ml
Total		500 ml

Electrophysiological assessment. Whole-cell current and clamp recordings were performed at 22 °C in artificial cerebral spinal fluid, bubbled with 95% O2 and 5% CO2. The extracellular fluid consisted of (in mM) 124 NaCl, 3.3 KCl, 2.4 MgSO4, 1.2 KH2PO4, 26 NaHCO3, 2.5 CaCl2 and 10 Glucose (at pH 7.4). TTX (100 nM), bicuculline (30 μM), or GABA (10 μM) were used in the bath solution. Borosilicate glass electrodes (resistance 6–10 MΩ) were filled with an intracellular solution containing 135 mM potassium gluconate, 7 mM NaCl, 10 mM HEPES, 2 mM MgATP, 0.3 mM Na2GTP and 2 mM MgCl2,

adjusted to pH 7.4 with KOH. Cell visualization and patch pipette micromanipulation were performed by video-microscopy, employing a 40× water-immersion objective mounted on an upright microscope equipped with infrared differential interference contrast (Nikon, Eclipse fn1, Japan). Intracellular membrane electrical potentials were recorded in current-clamp mode, using a Multi-clamp 700B amplifier (Molecular Devices, Palo Alto, CA, USA). For voltage clamp recordings, cells were held at -70 mV. (-)-Bicuculline methochloride and tetrodotoxin (TTX) were purchased from Shanghai yuanye Bio-Technology Co., Ltd; GABA were purchased from Solarbio (Beijing, China). All other compounds were obtained from Sigma (St. Louis, MO). Data were digitized at 10 kHz with a 2 kHz low-pass filter. Data processing and analysis were performed using Clampfit 10.6 (Axon Instruments).

Neural transplantation and quinolinic acid striatal lesions. All animal experiments were carried out in accordance with the instructions for the Care and Use of Animals in Research published by the Institute of Zoology of the Chinese Academy of Sciences. Transplantation into the neonatal NOD-SCID mice was conducted as described previously (Danjo et al., 2011). In brief, transplantation was performed by anesthetizing day 1 postnatal NOD-SCID mice on ice for 3 min, and injecting the cell suspension (~$5x10^4$ cells per μl) into the striatum (coordinated 1 mm anterior, 1 mm lateral, and 2.5 mm deep from the bregma). The establishment of quinolinic acid (QA)-lesioned mouse model of HD and cell transplantation were performed as previously described (Ma et al., 2012). To establish unilateral lesion of mouse model of Hunting's disease, adult male SCID mice (8 weeks of age) were anesthetized and received a stereotaxic injection of 1ul of 60nM quinolinic acid (QA, P63204; Sigma, in saline with 0.2 mg/ml ascorbic acid) into the right striatum on a coordinator (anterior-posterior [AP] = +0.7 mm, lateral [L] = +1.7 mm, vertical [V] =-3.5 mm). Differentiated LGE-like progenitors (12-14 days of differentiation from hESCs) were dissociated with Accutase and prepared at approximately $5x10^4$ cells/ul in PBS containing B27, 0.2 mM AA, 20 ng/ml BDNF, and 10 ng/ml GDNF. Cell suspension (2 μl) was injected into the lesioned striatum (AP = +0.8 mm, L = +1.8 mm, V = -3.5 mm) of anesthetized animals 2 weeks after QA lesion with a glass pipette (0.3–0.5 mm in diameter) over a period of 5 min. The QA-lesioned animals receiving the same surgery and injection of 2 μl of PBS solution (without cells) served as controls. For analysis, mice were perfused transcardially with 4% paraformaldehyde, and 20μm-thick brain sections were taken for immunostaining.