

ISCI, Volume 3

Supplemental Information

HOTf-Catalyzed Alkyl-Heck-type Reaction

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Supplemental Figures for ^1H NMR, ^{13}C NMR, and ^{19}F NMR Spectra

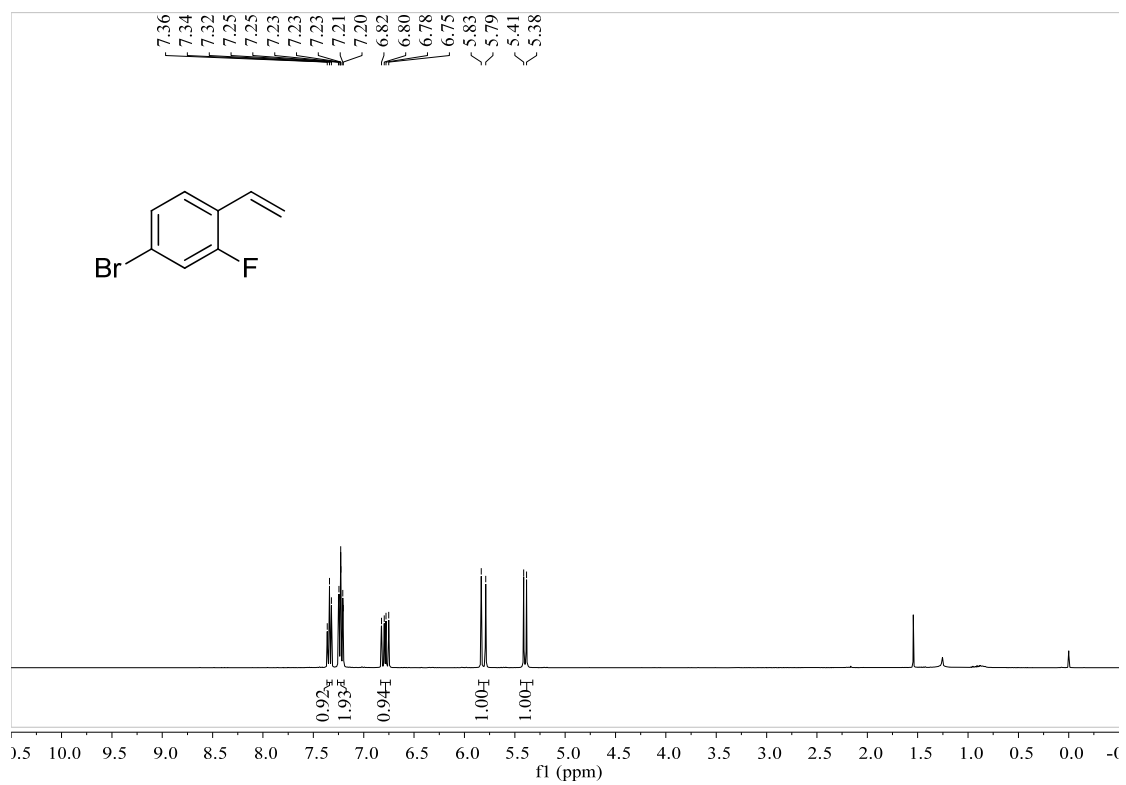


Figure S1. ^1H NMR spectrum of 4-bromo-2-fluoro-1-vinylbenzene, Related to **Figure 1**.

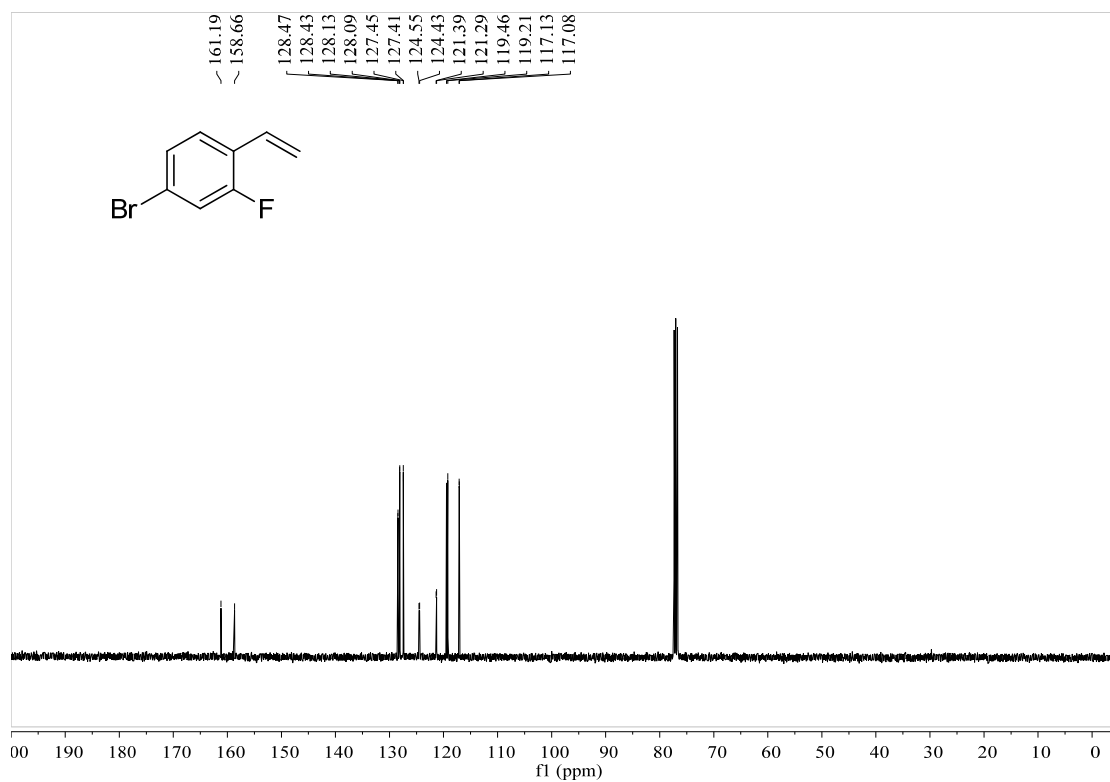


Figure S2. ^{13}C NMR spectrum of 4-bromo-2-fluoro-1-vinylbenzene, Related to **Figure 1**.

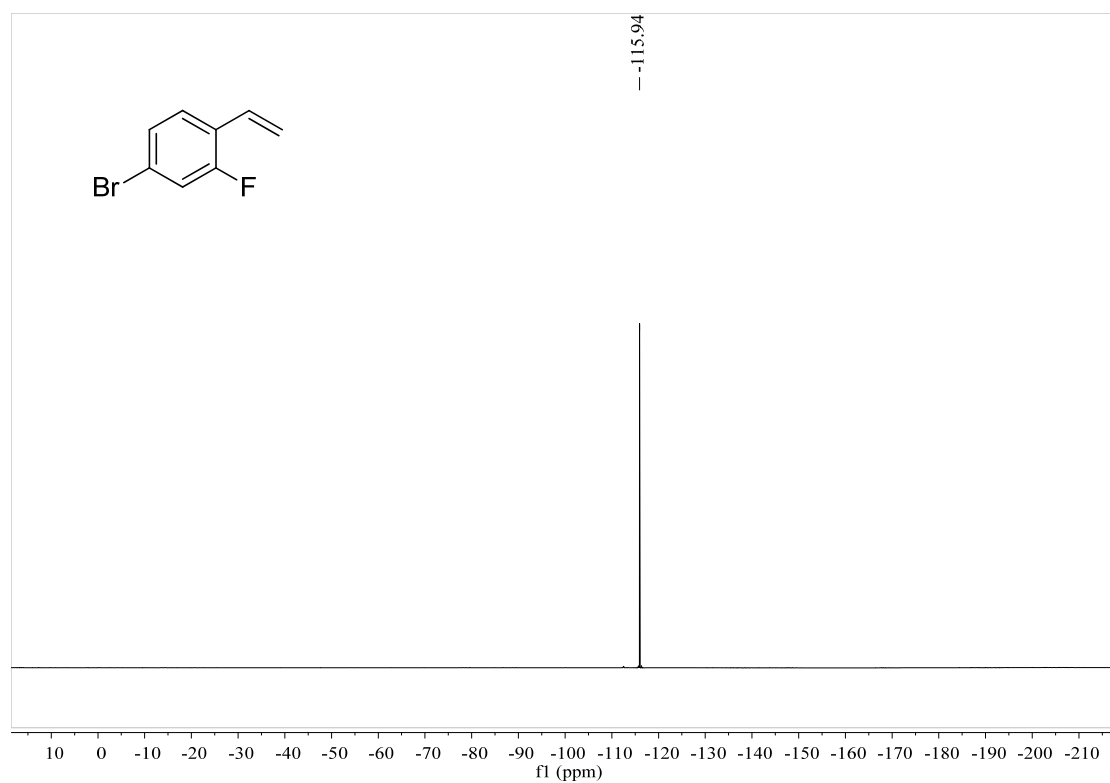


Figure S3. ^{19}F NMR spectrum of 4-bromo-2-fluoro-1-vinylbenzene, Related to **Figure 1**.

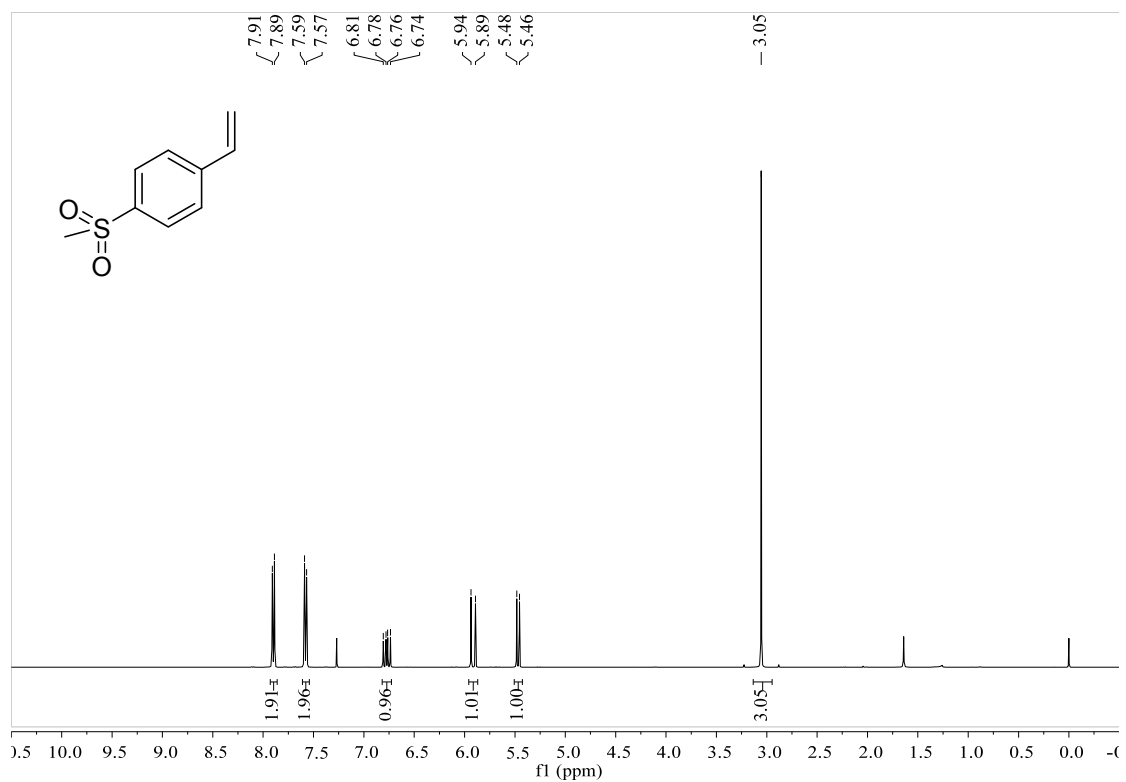


Figure S4. ^1H NMR spectrum of 1-(methylsulfonyl)-4-vinylbenzene, Related to **Figure 1**.

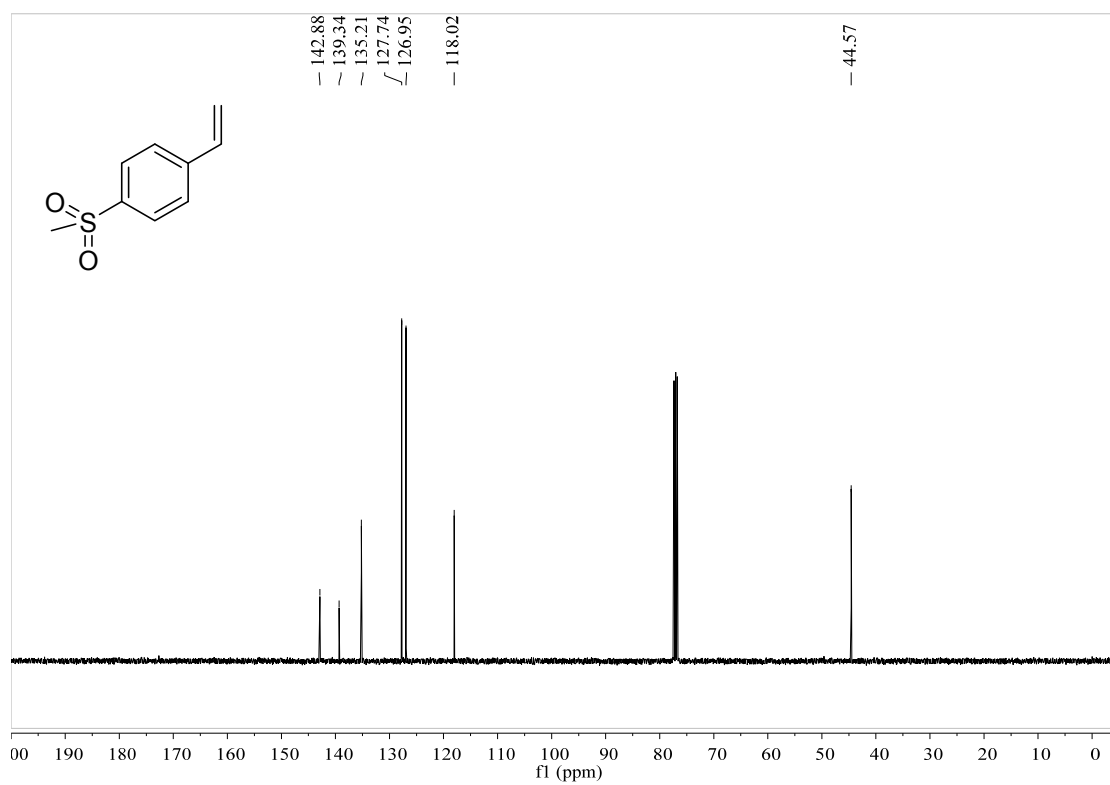


Figure S5. ^{13}C NMR spectrum of 1-(methylsulfonyl)-4-vinylbenzene, Related to **Figure 1**.

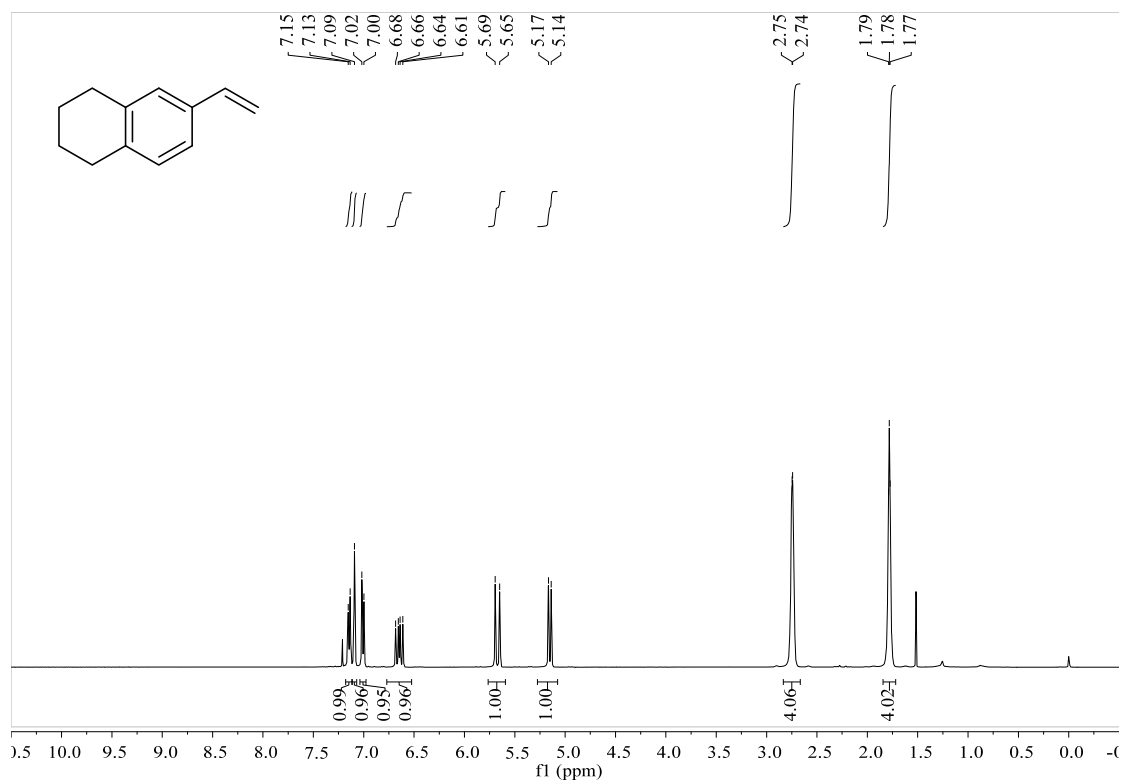


Figure S6. ¹H NMR spectrum of 6-vinyl-1,2,3,4-tetrahydronaphthalene, Related to **Figure 1**.

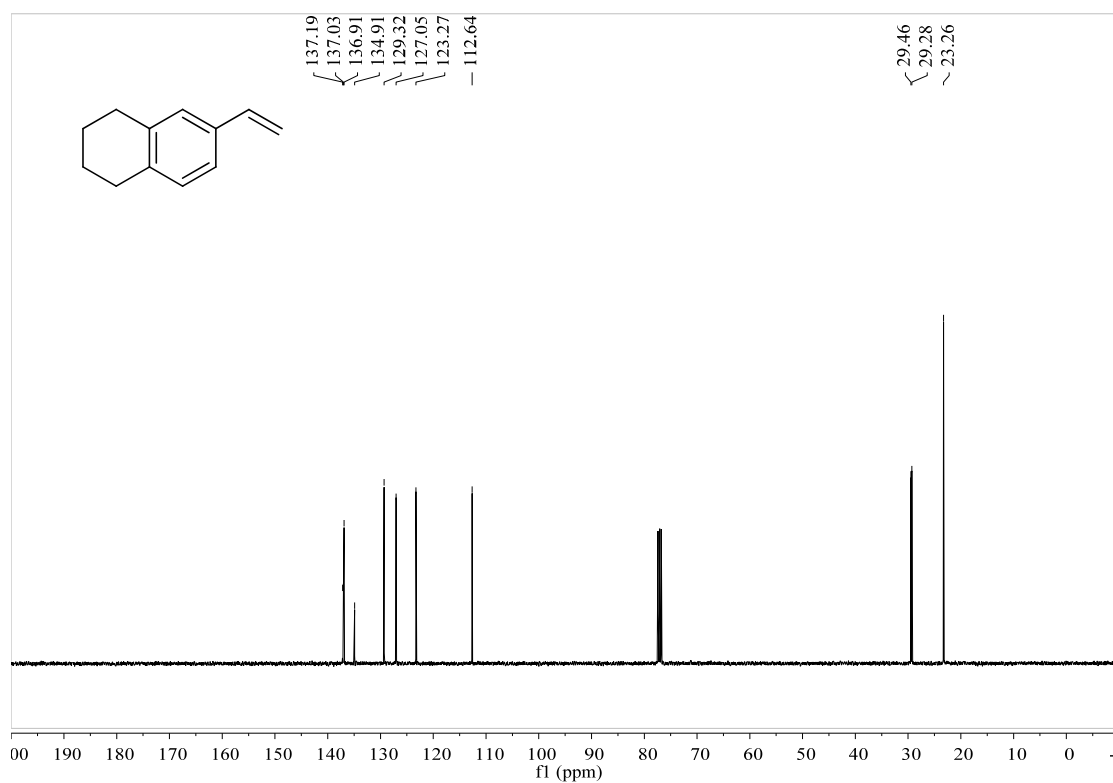


Figure S7. ¹³C NMR spectrum of 6-vinyl-1,2,3,4-tetrahydronaphthalene, Related to **Figure 1**.

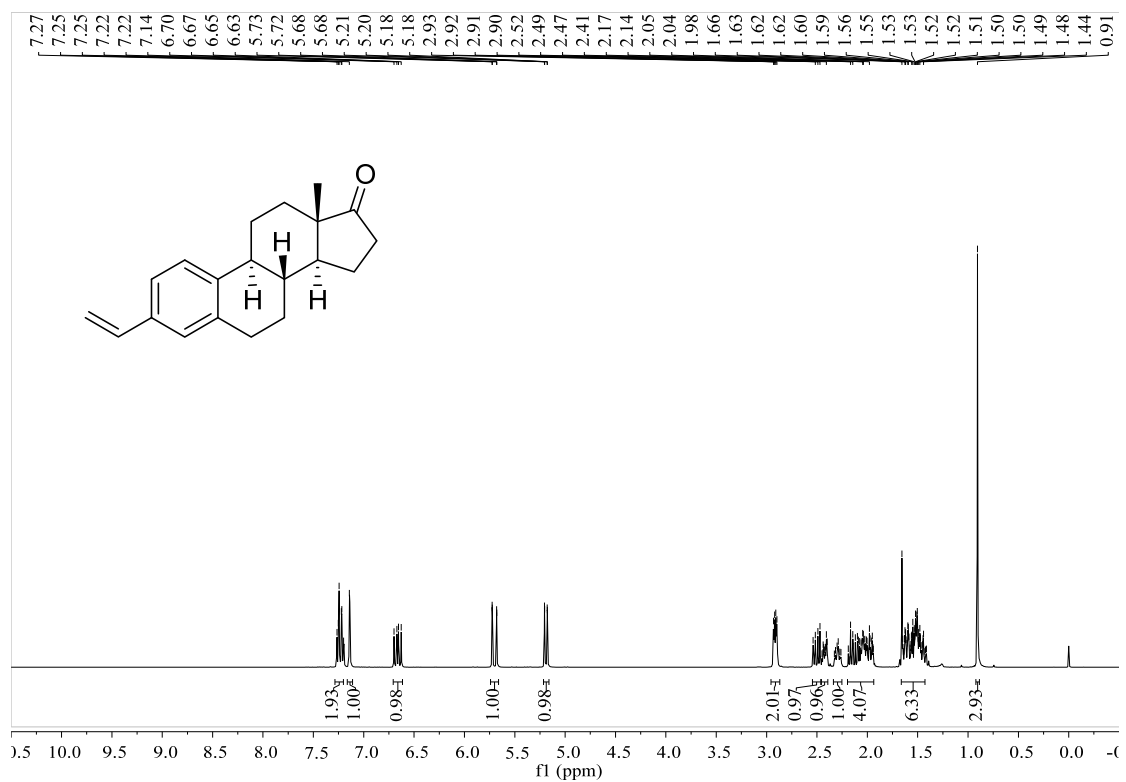


Figure S8. ¹H NMR spectrum of compound 57, related to scheme 2.

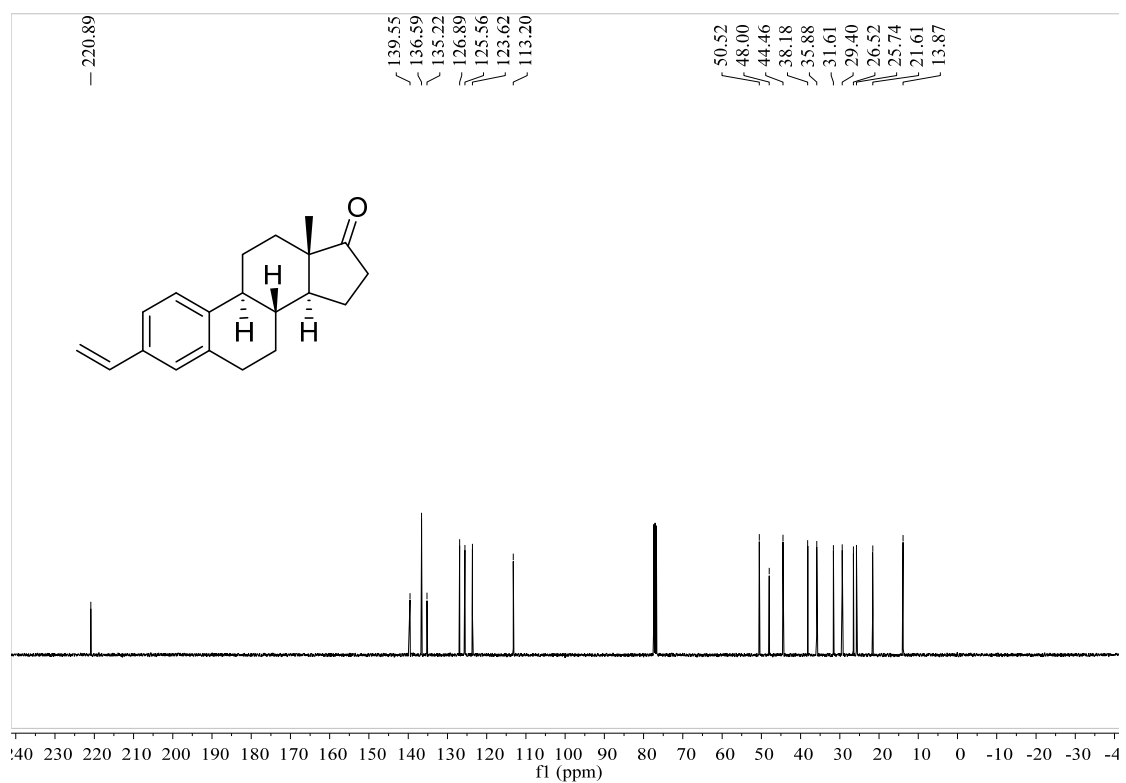


Figure S9. ¹³C NMR spectrum of compound 57, related to Scheme 2.

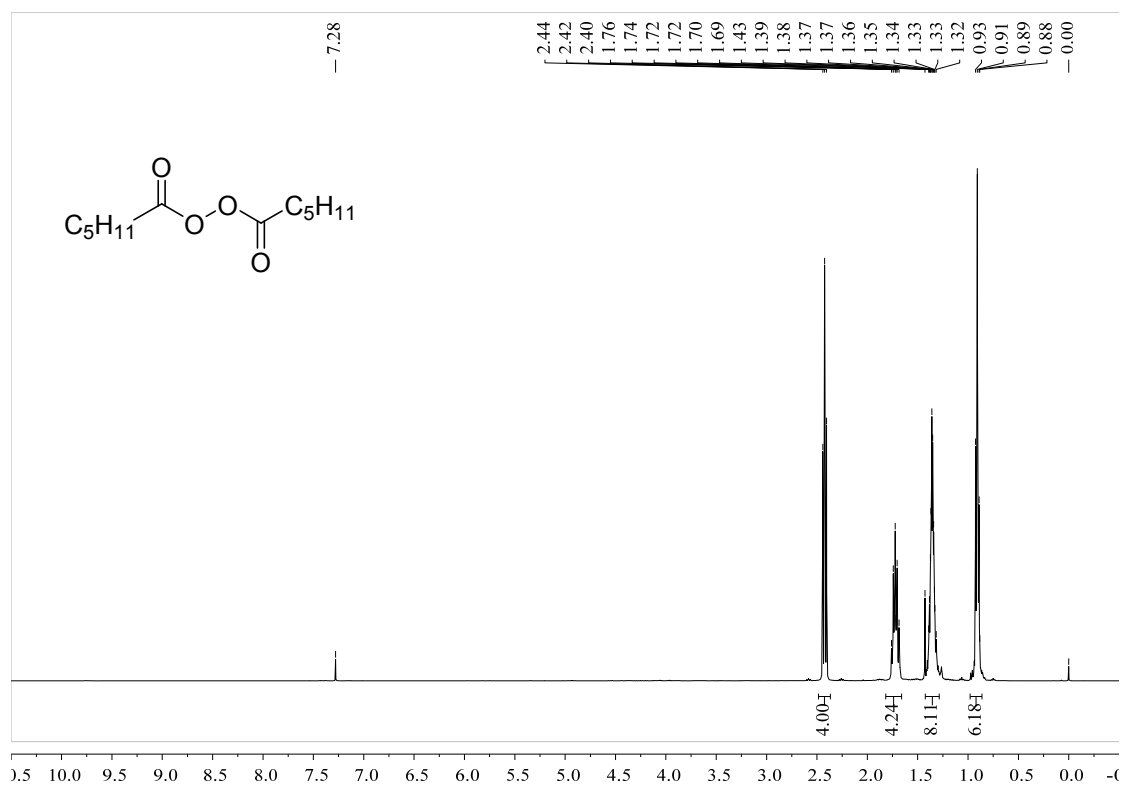


Figure S10. ¹H NMR spectrum of hexanoic peroxyanhydride, related to **Figure 3**.

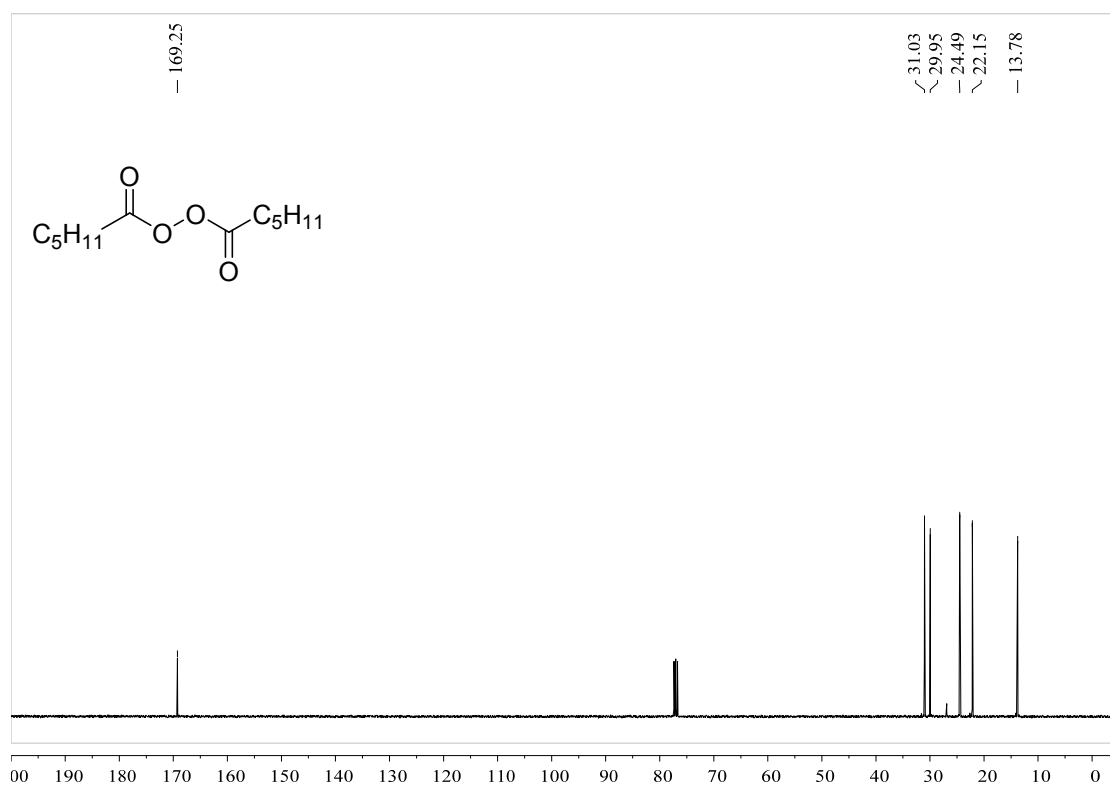


Figure S11. ¹³C NMR spectrum of hexanoic peroxyanhydride, related to **Figure 3**.

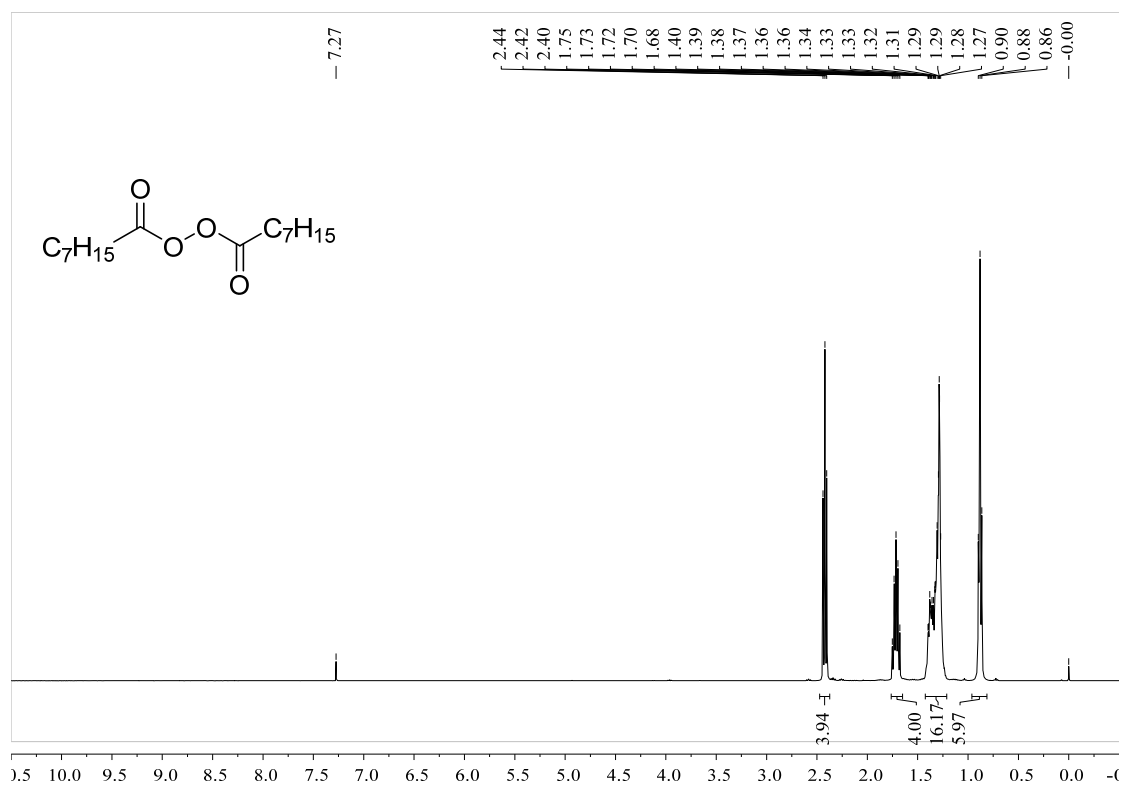


Figure S12. ¹H NMR spectrum of octanoic peroxyanhydride, related to **Figure 3**.

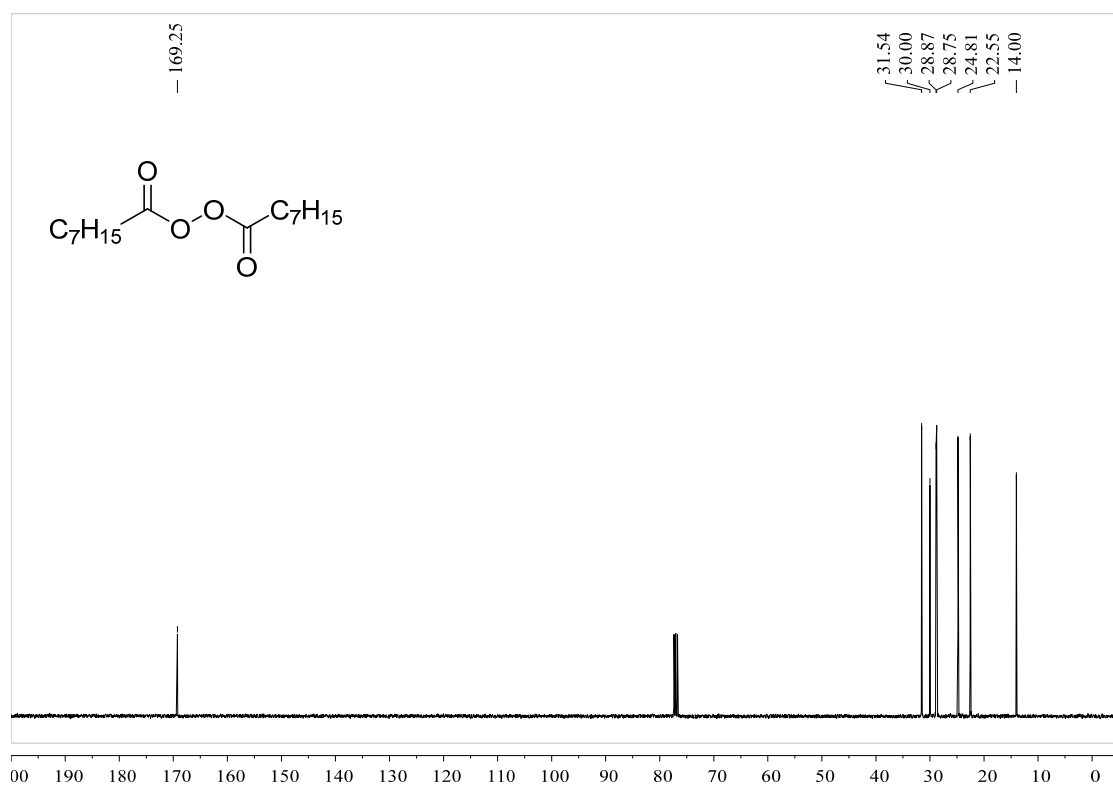


Figure S13. ¹³C NMR spectrum of octanoic peroxyanhydride, related to **Figure 3**.

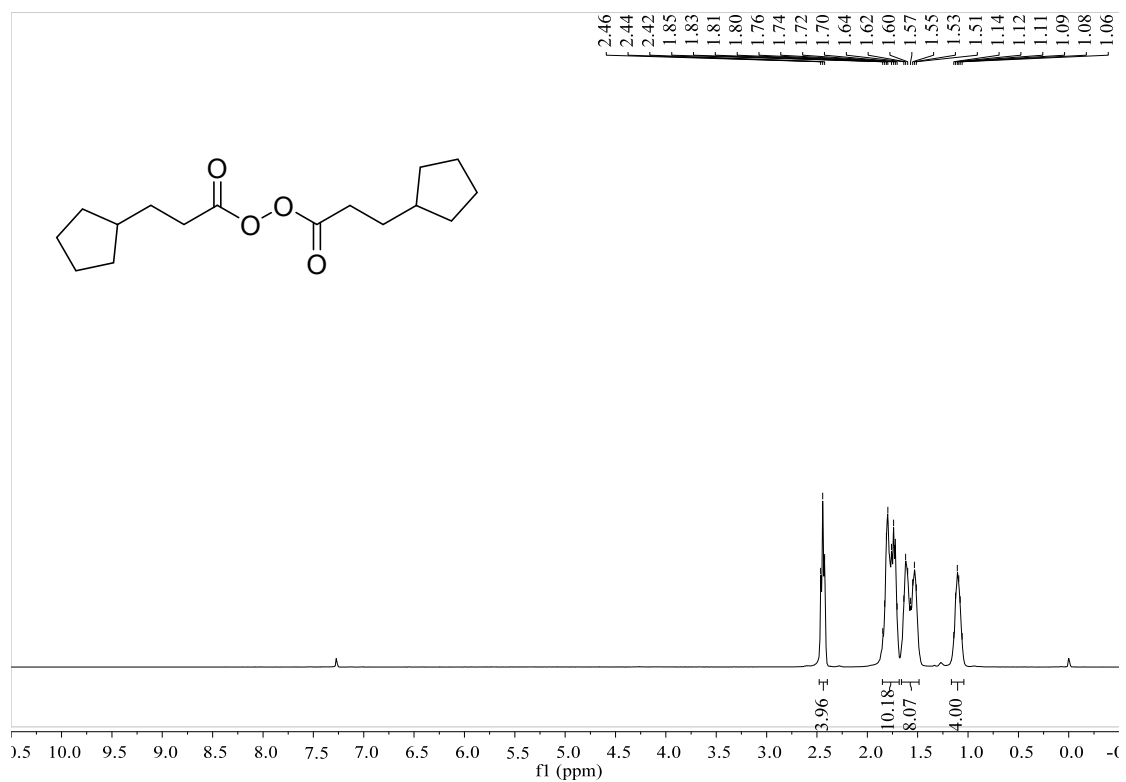


Figure S14. ^1H NMR spectrum of 3-cyclopentylpropanoic peroxyanhydride, related to **Figure 3**.

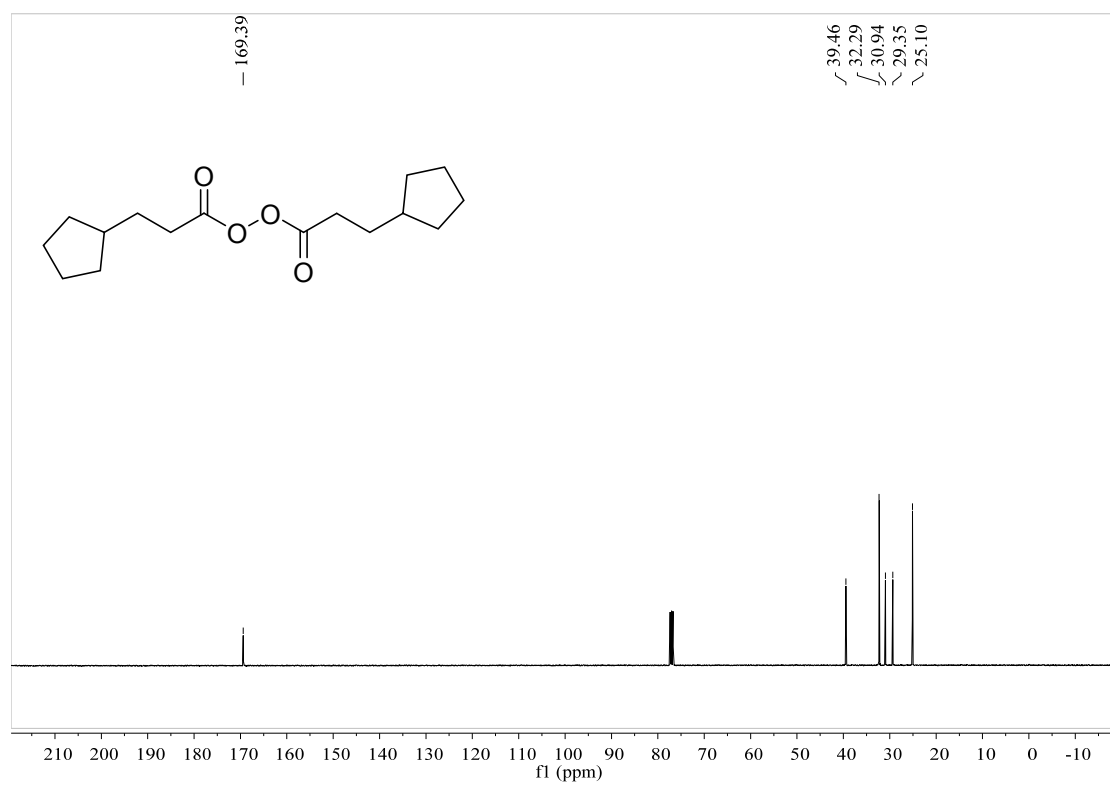


Figure S15. ^{13}C NMR spectrum of 3-cyclopentylpropanoic peroxyanhydride, related to **Figure 3**.

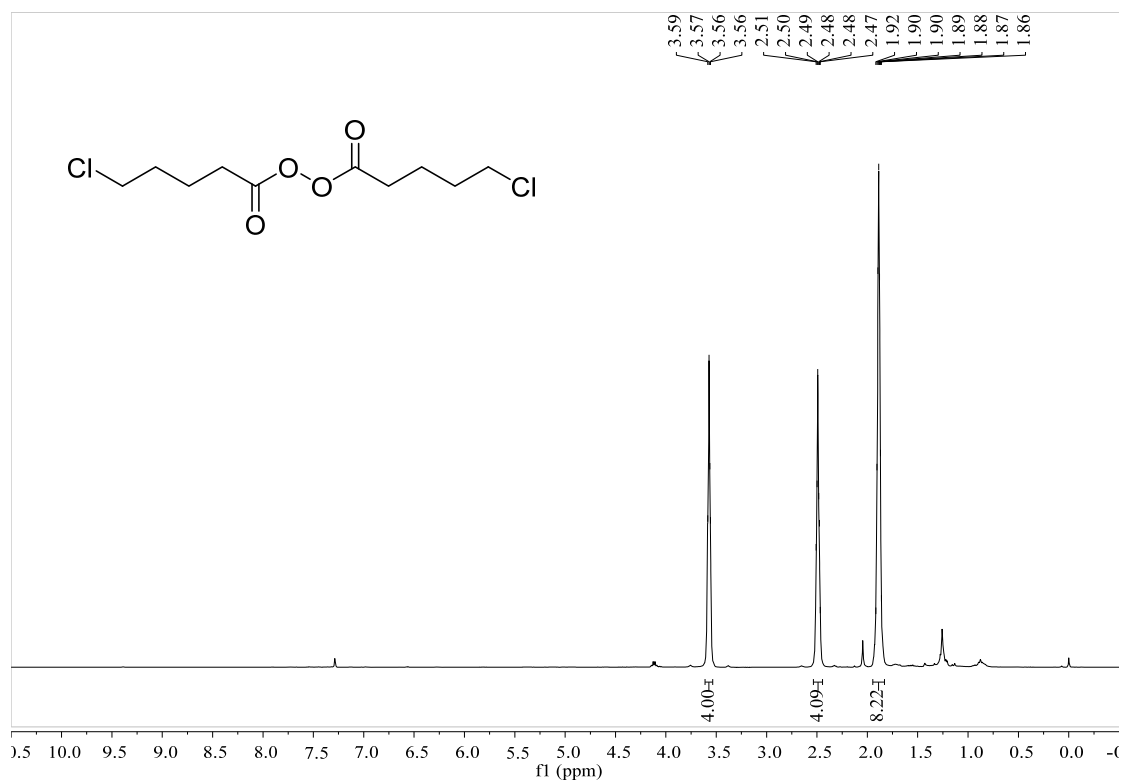


Figure S16. ¹H NMR spectrum of 5-chloropentanoic peroxyanhydride, related to **Figure 3**.

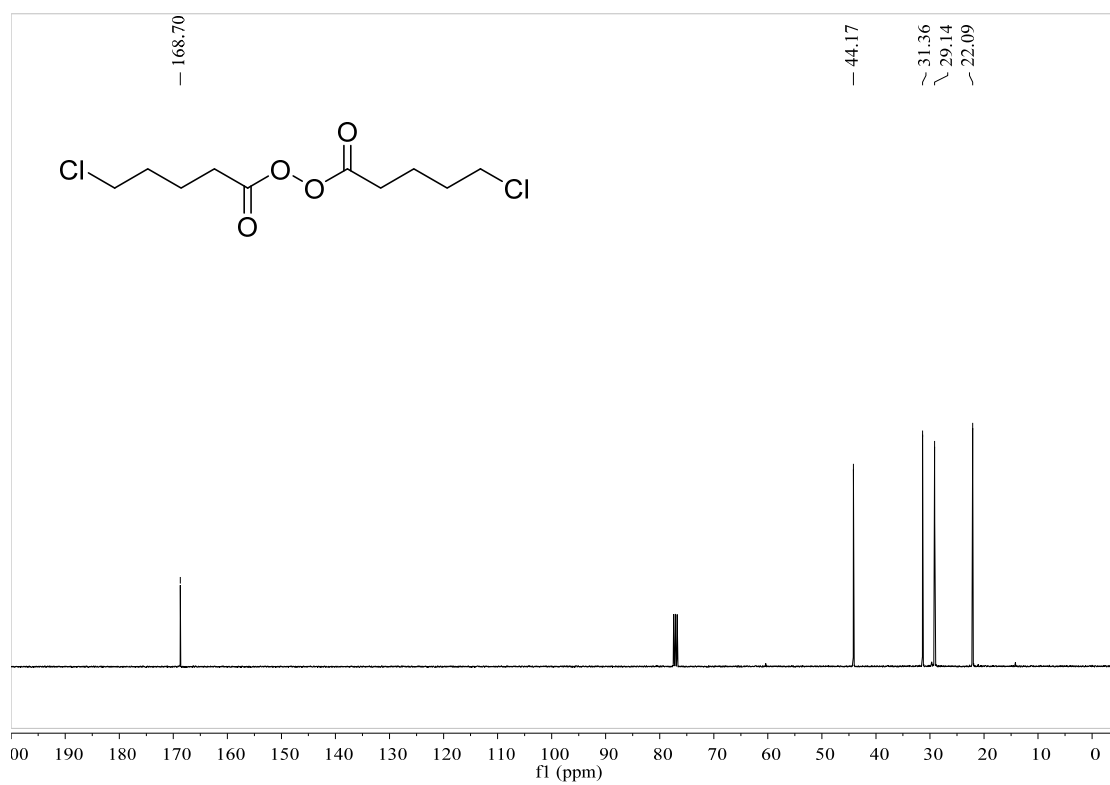


Figure S17. ¹³C NMR spectrum of 5-chloropentanoic peroxyanhydride, related to **Figure 3**.

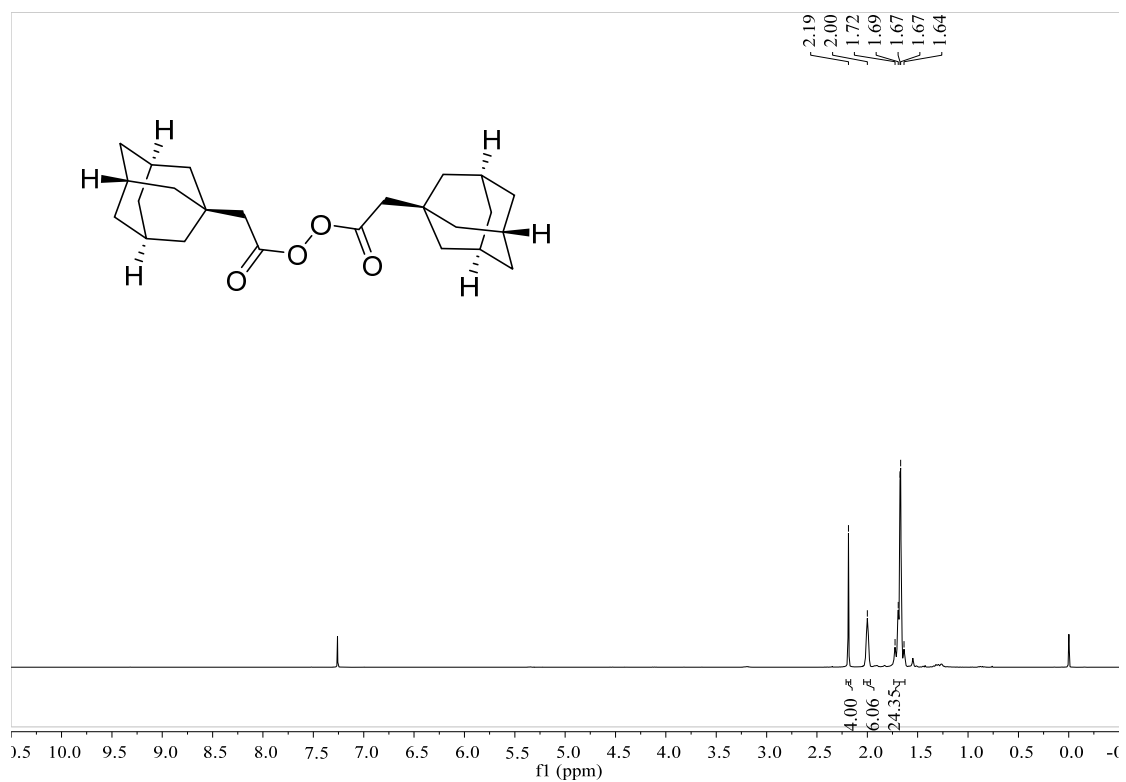


Figure S18. ¹H NMR spectrum of 2-((3*r*,5*r*,7*r*)-adamantan-1-yl)acetic peroxyanhydride, related to **Figure 3**.

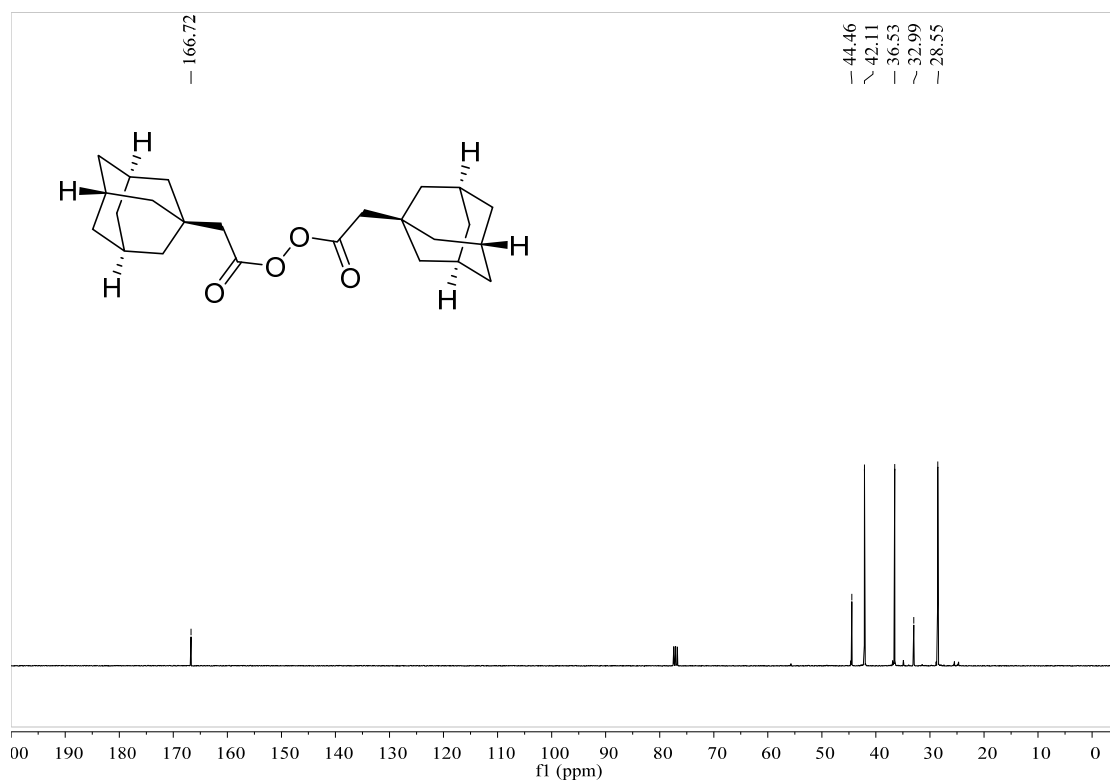


Figure S19. ¹³C NMR spectrum of 2-((3*r*,5*r*,7*r*)-adamantan-1-yl)acetic peroxyanhydride, related to **Figure 3**.

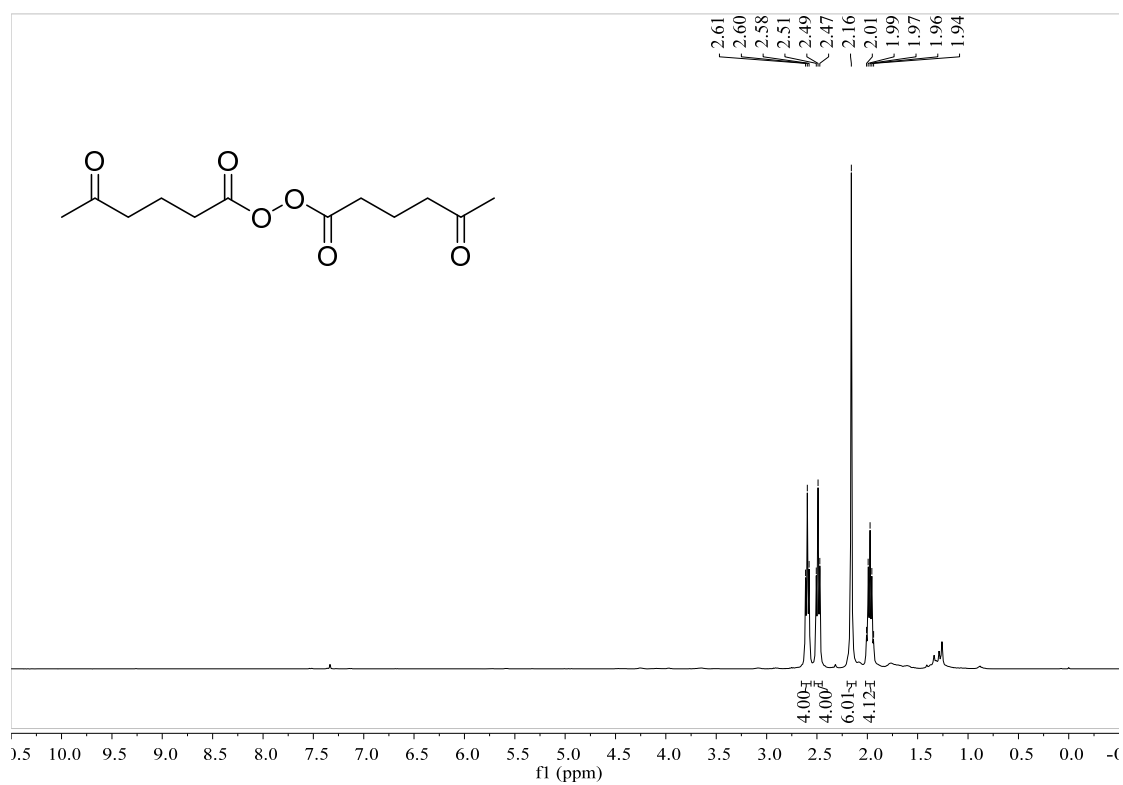


Figure S20. ^1H NMR spectrum of 5-oxohexanoic peroxyanhydride, related to **Figure 3**.

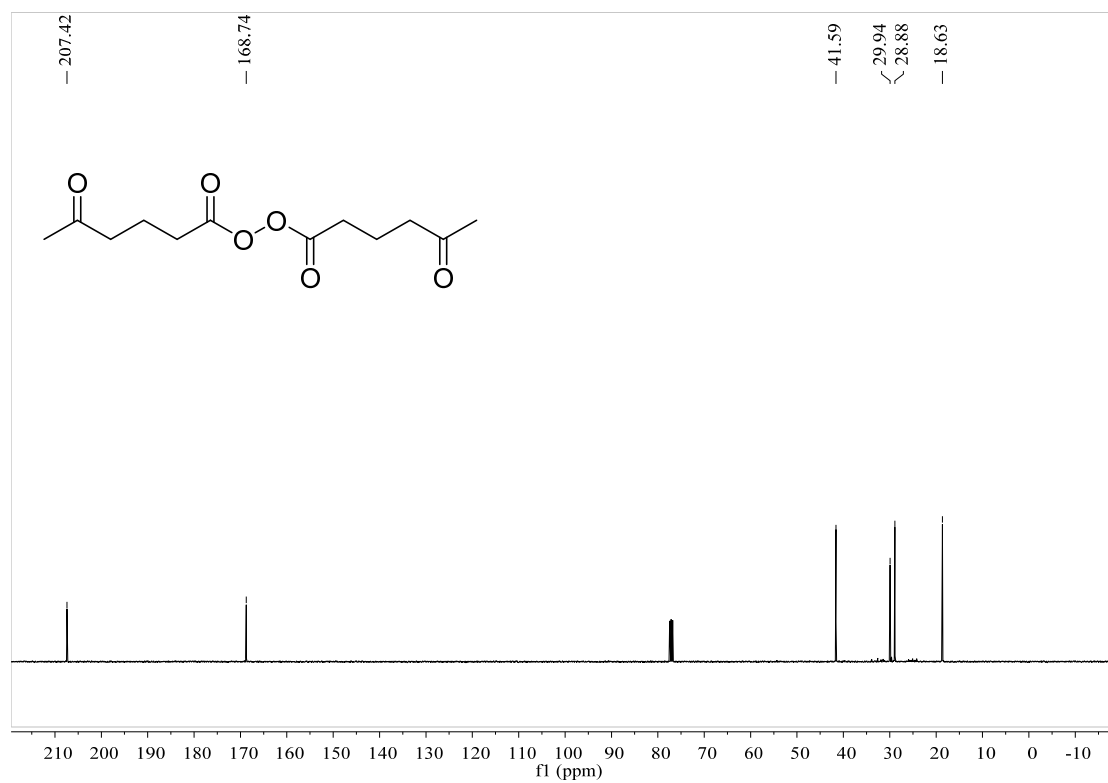


Figure S21. ^{13}C NMR spectrum of 5-oxohexanoic peroxyanhydride, related to **Figure 3**.

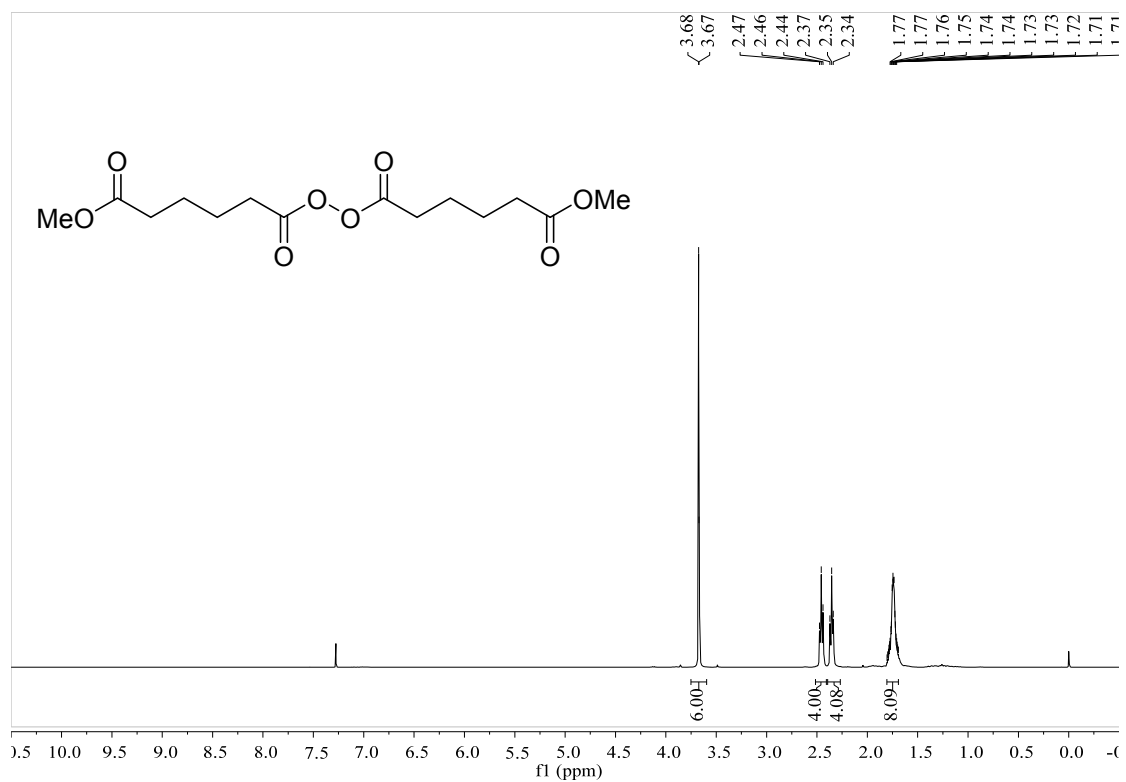


Figure S22. ¹H NMR spectrum of 6-methoxy-6-oxohexanoic peroxyanhydride, related to Figure 3.

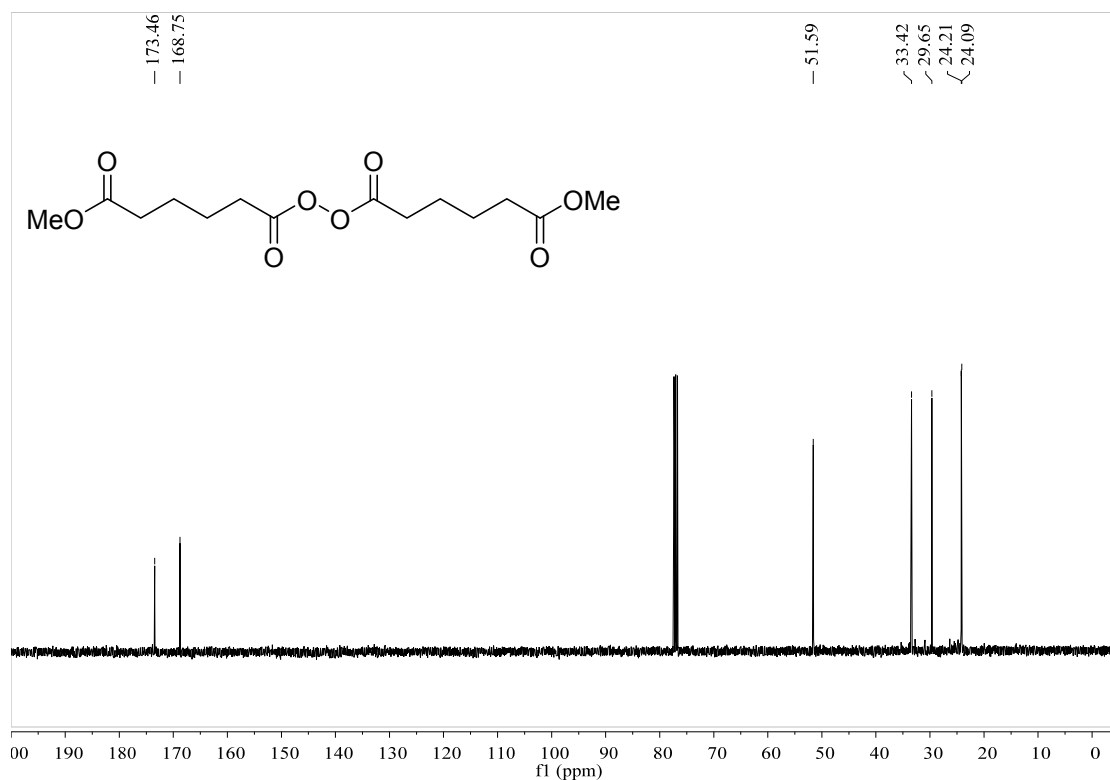


Figure S23. ¹³C NMR spectrum of 6-methoxy-6-oxohexanoic peroxyanhydride, related to Figure 3.

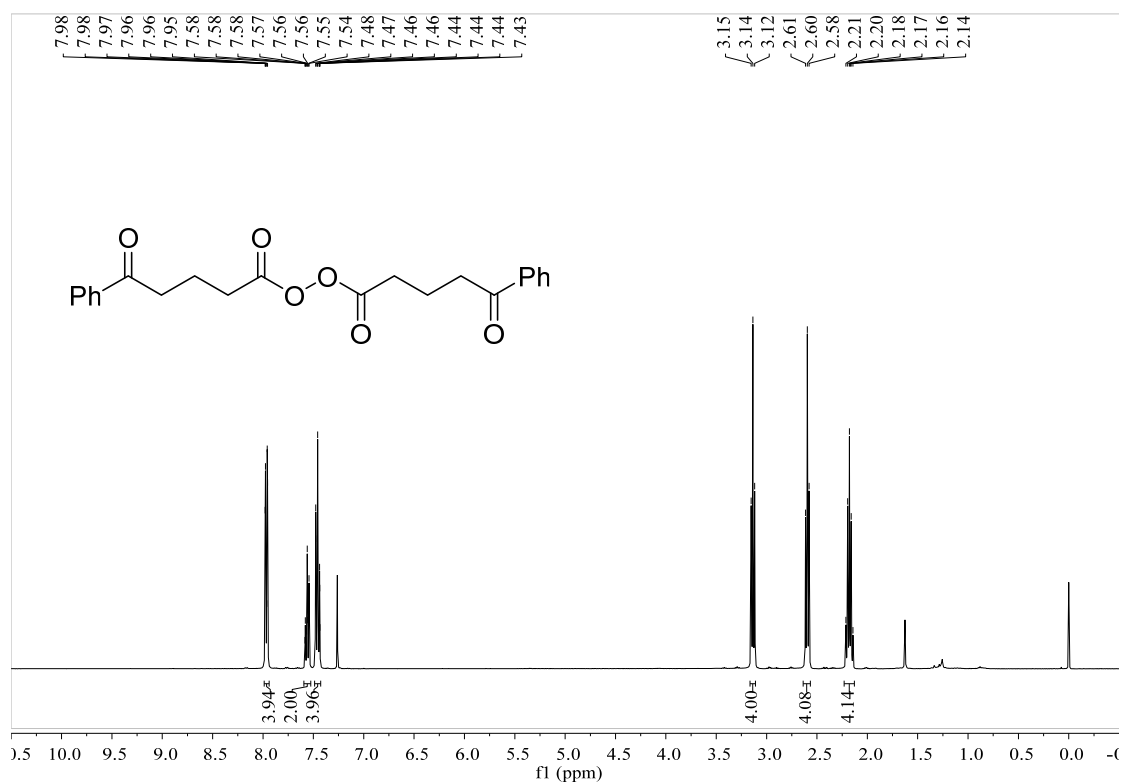


Figure S24. ¹H NMR spectrum of 5-oxo-5-phenylpentanoic peroxyanhydride, related to **Figure 3**.

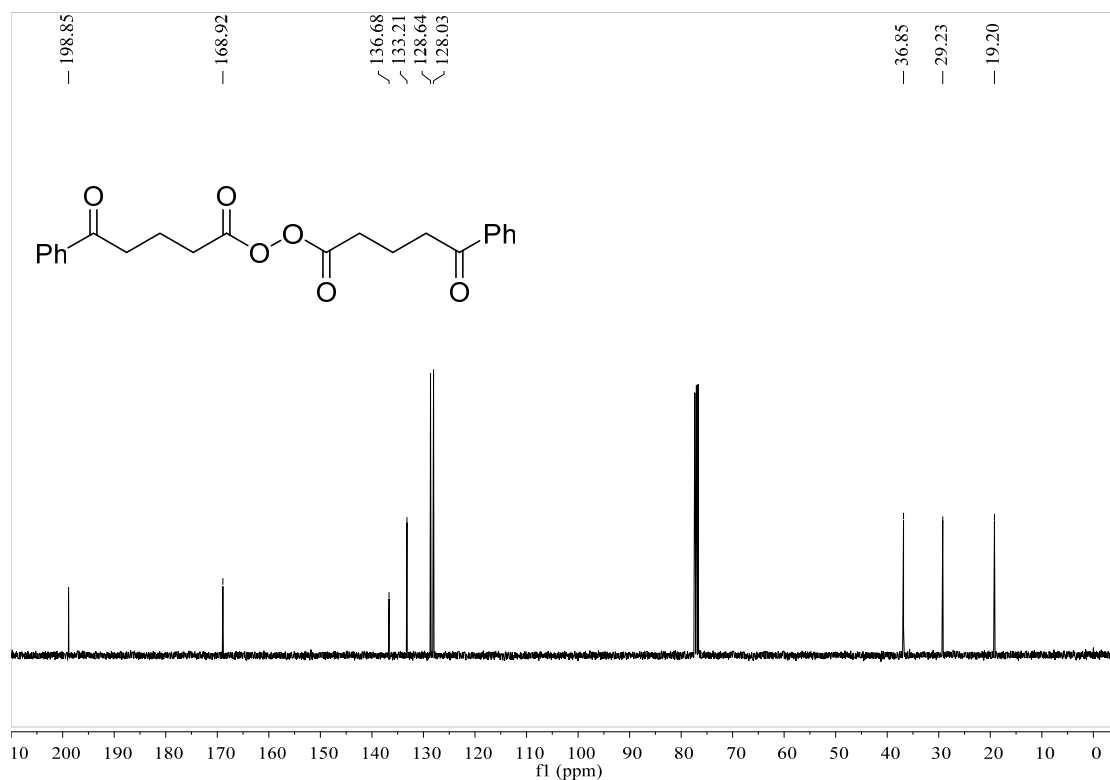


Figure S25. ¹³C NMR spectrum of 5-oxo-5-phenylpentanoic peroxyanhydride, related to **Figure 3**.

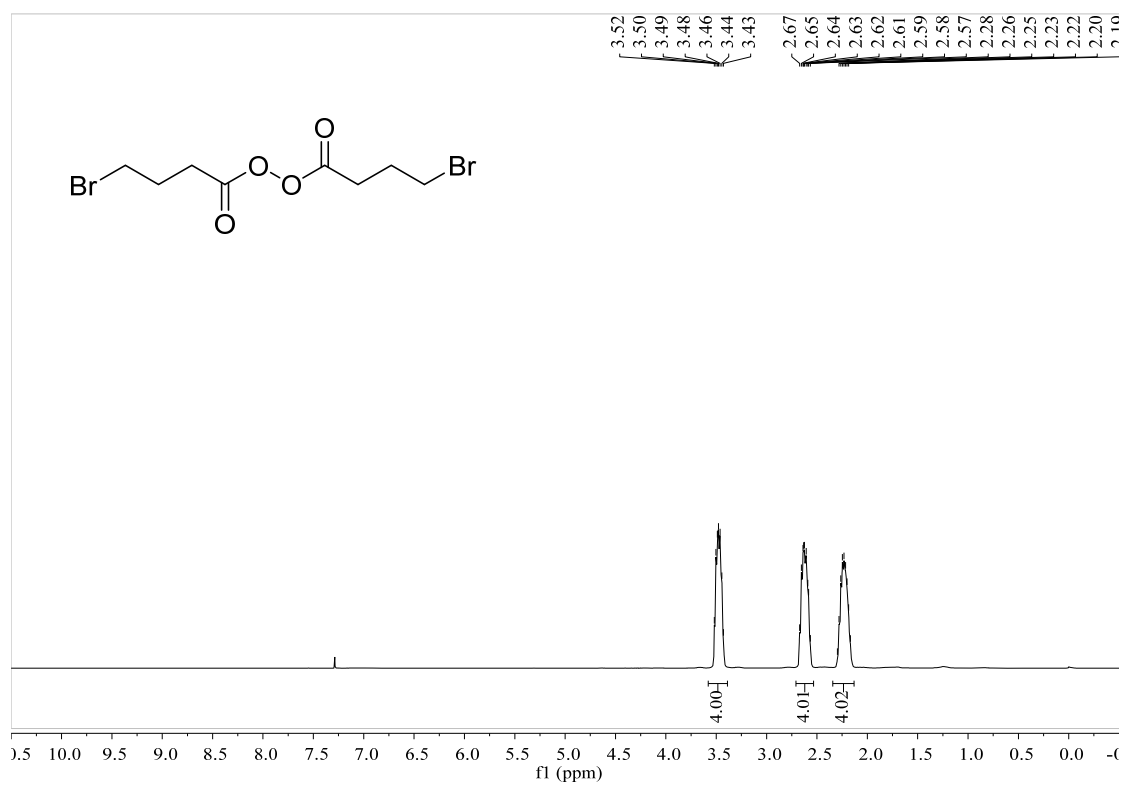


Figure S26. ^1H NMR spectrum of 4-bromobutanoic peroxyanhydride, related to **Figure 3**.

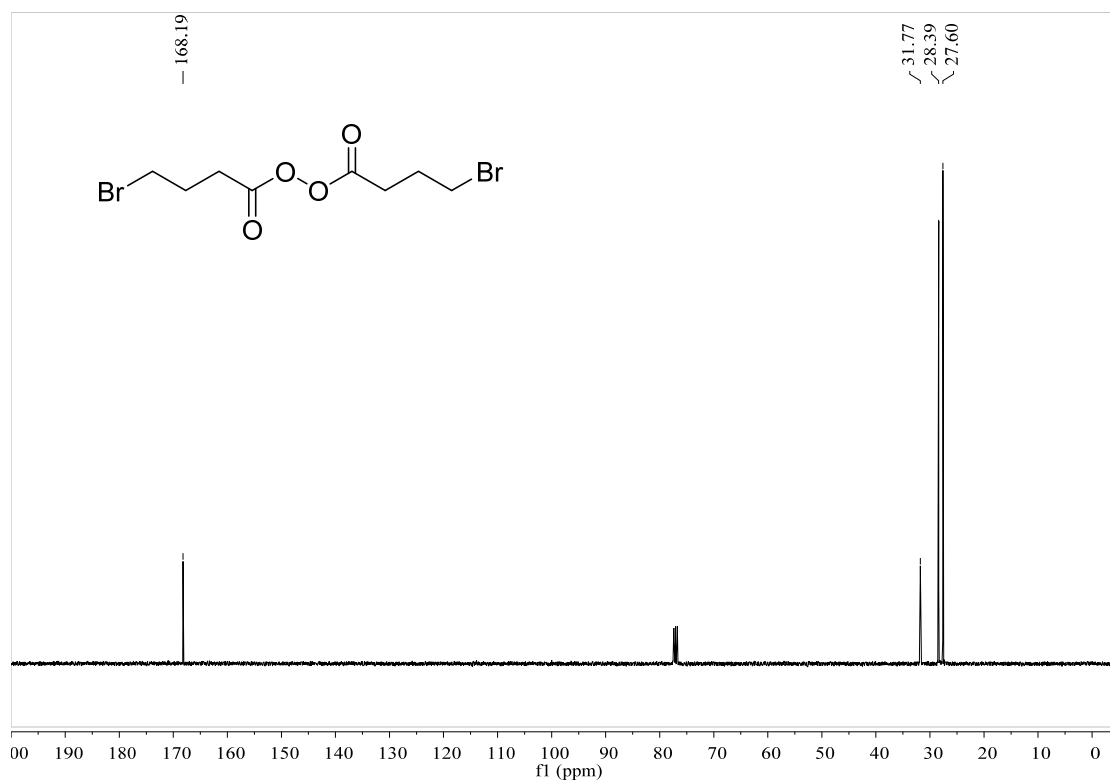


Figure S27. ^{13}C NMR spectrum of 4-bromobutanoic peroxyanhydride, related to **Figure 3**.

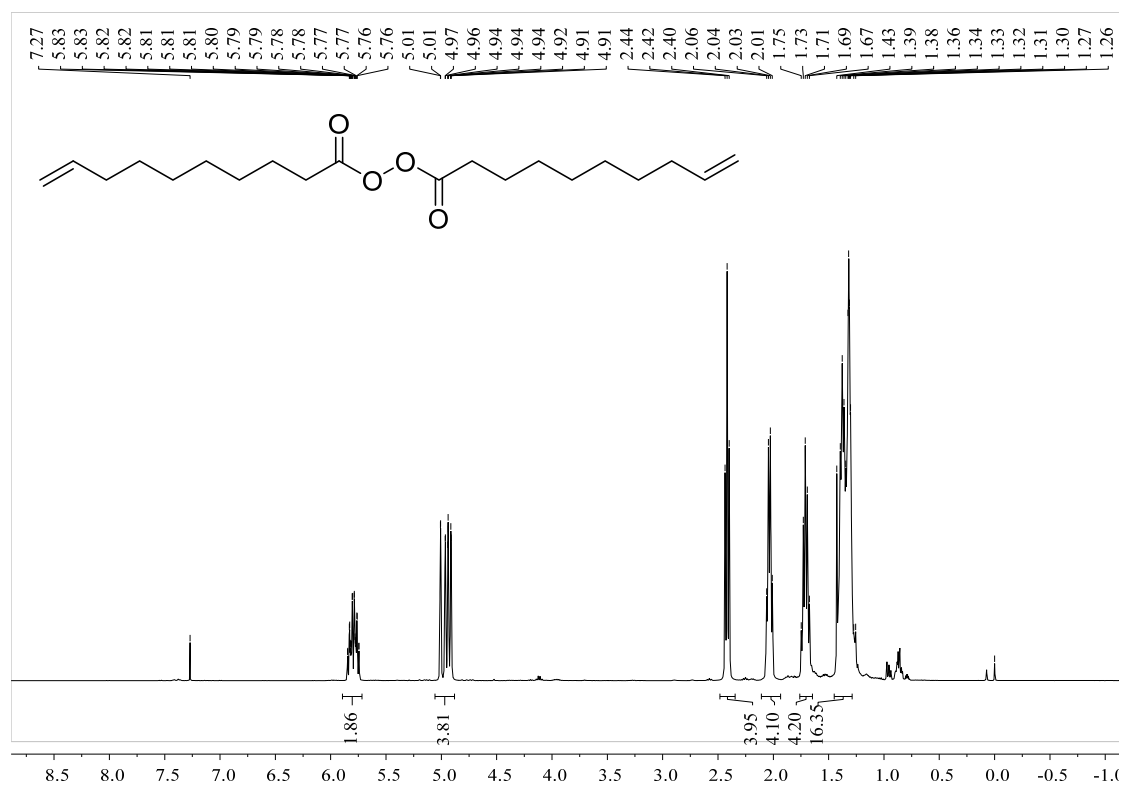


Figure S28. ¹H NMR spectrum of dec-9-enoic peroxyanhydride, related to **Figure 3**.

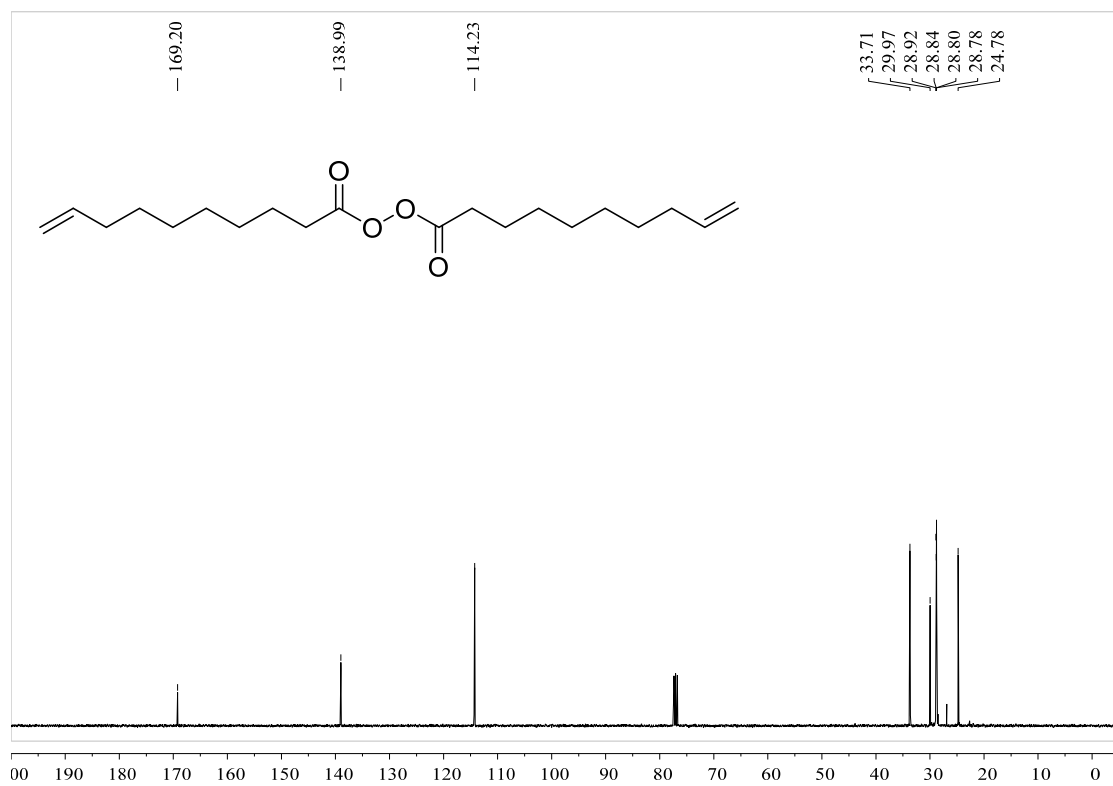


Figure S29. ¹³C NMR spectrum of dec-9-enoic peroxyanhydride, related to **Figure 3**.

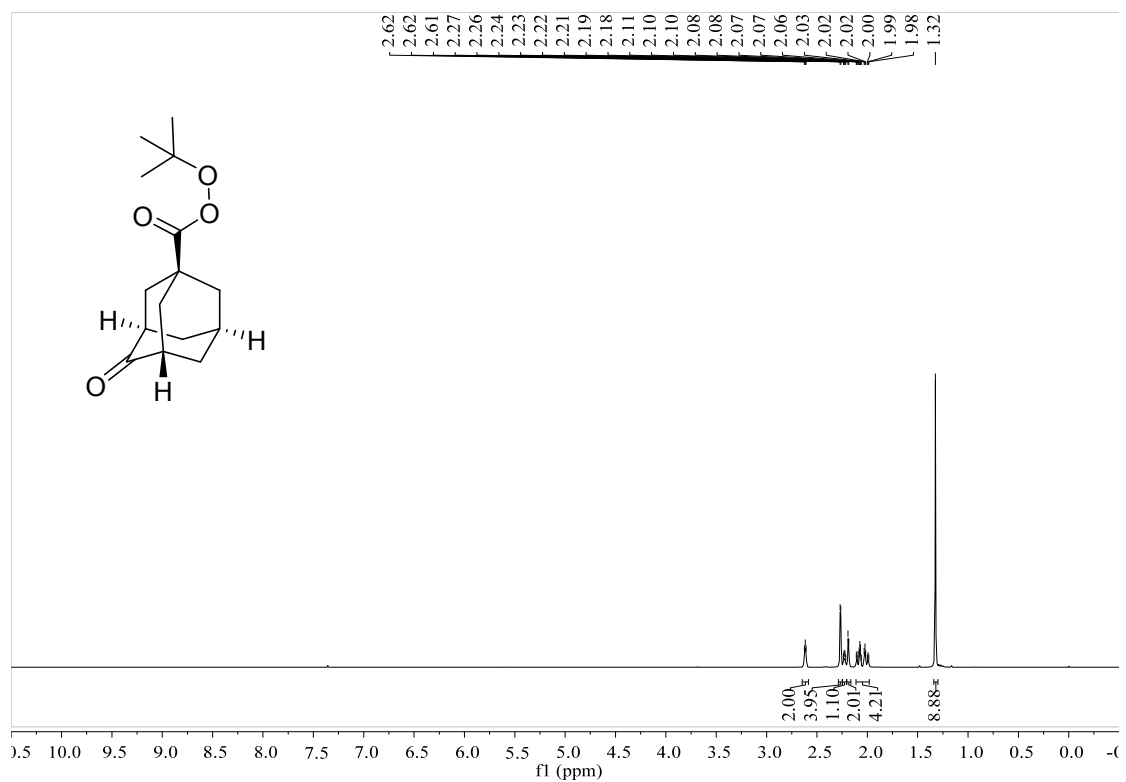


Figure S30. ¹H NMR spectrum of *tert*-butyl(1*s*,3*r*,5*s*,7*s*)-4-oxadamantane-1-carboperoxoate, related to **Figure 2**.

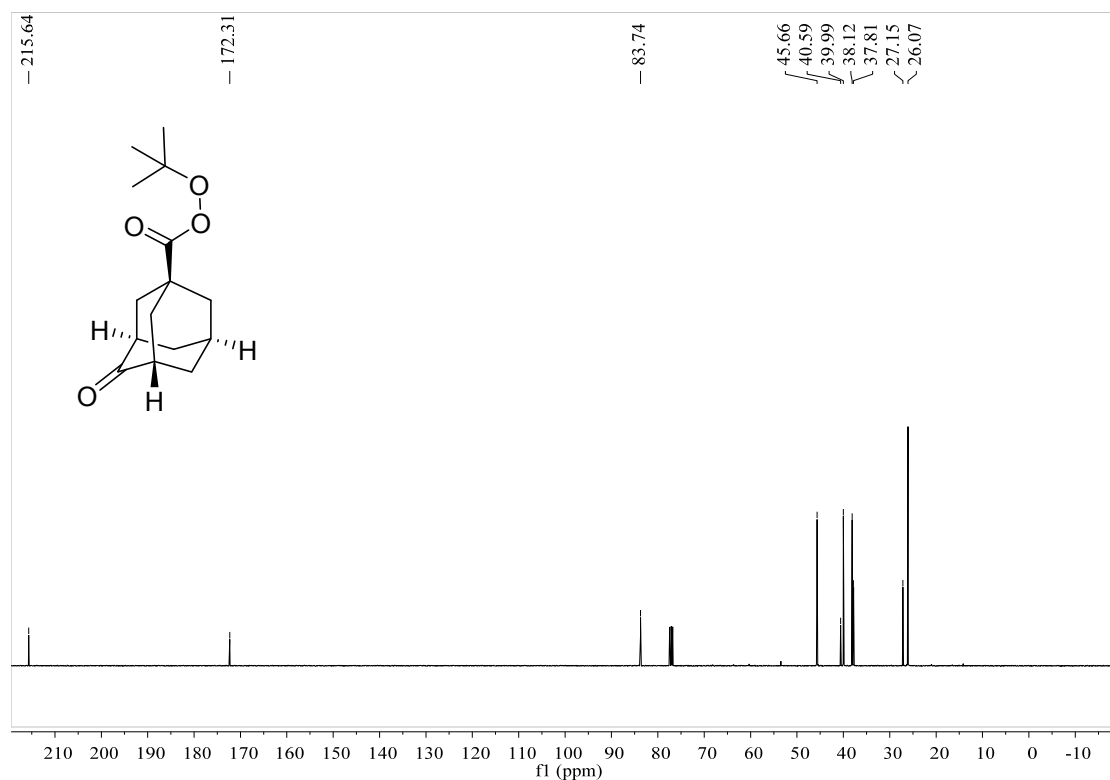


Figure S31. ¹³C NMR spectrum of *tert*-butyl(1*s*,3*r*,5*s*,7*s*)-4-oxadamantane-1-carboperoxoate, related to **Figure 2**.

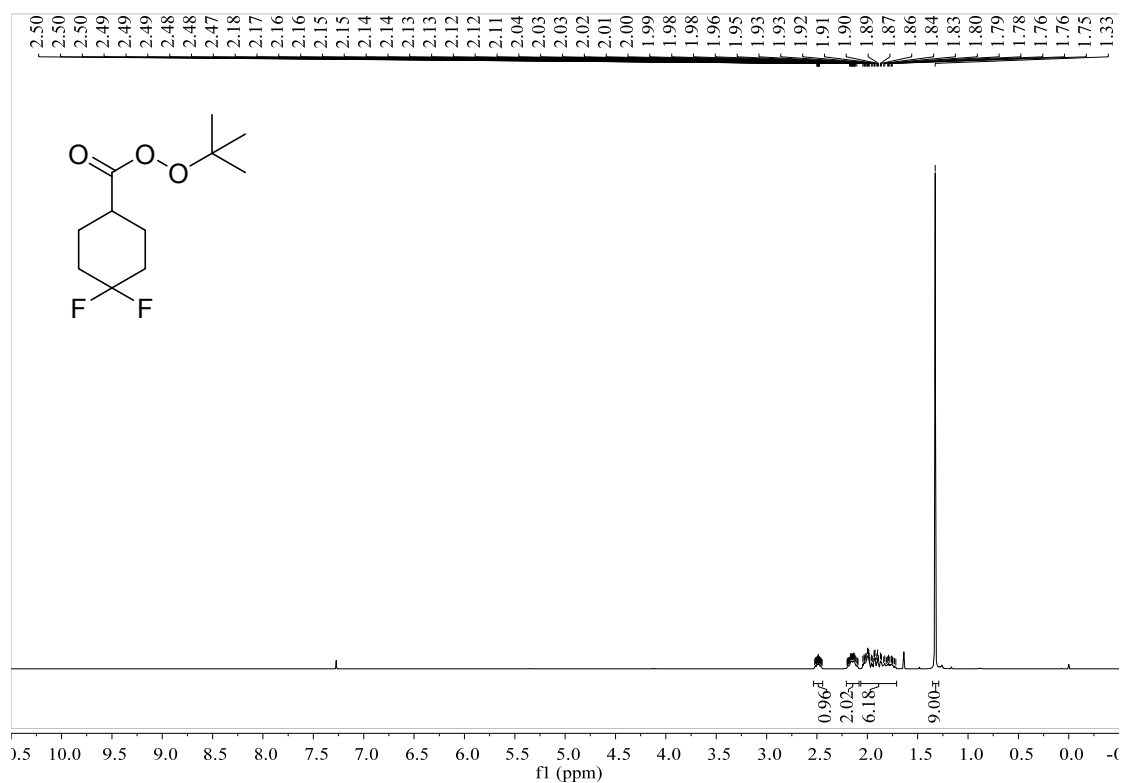


Figure S32. ¹H NMR spectrum of *tert*-butyl 4,4-difluorocyclohexane-1-carboxylate, related to **Figure 2**.

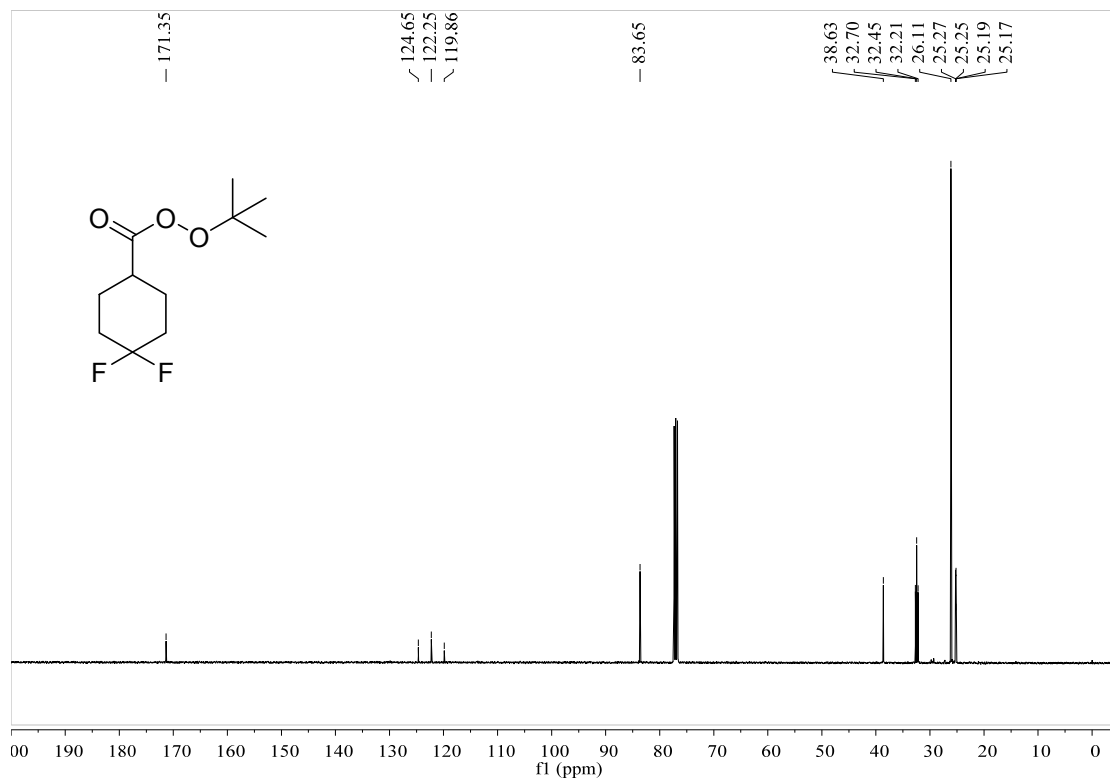


Figure S33. ¹³C NMR spectrum of *tert*-butyl 4,4-difluorocyclohexane-1-carboxylate, related to **Figure 2**.

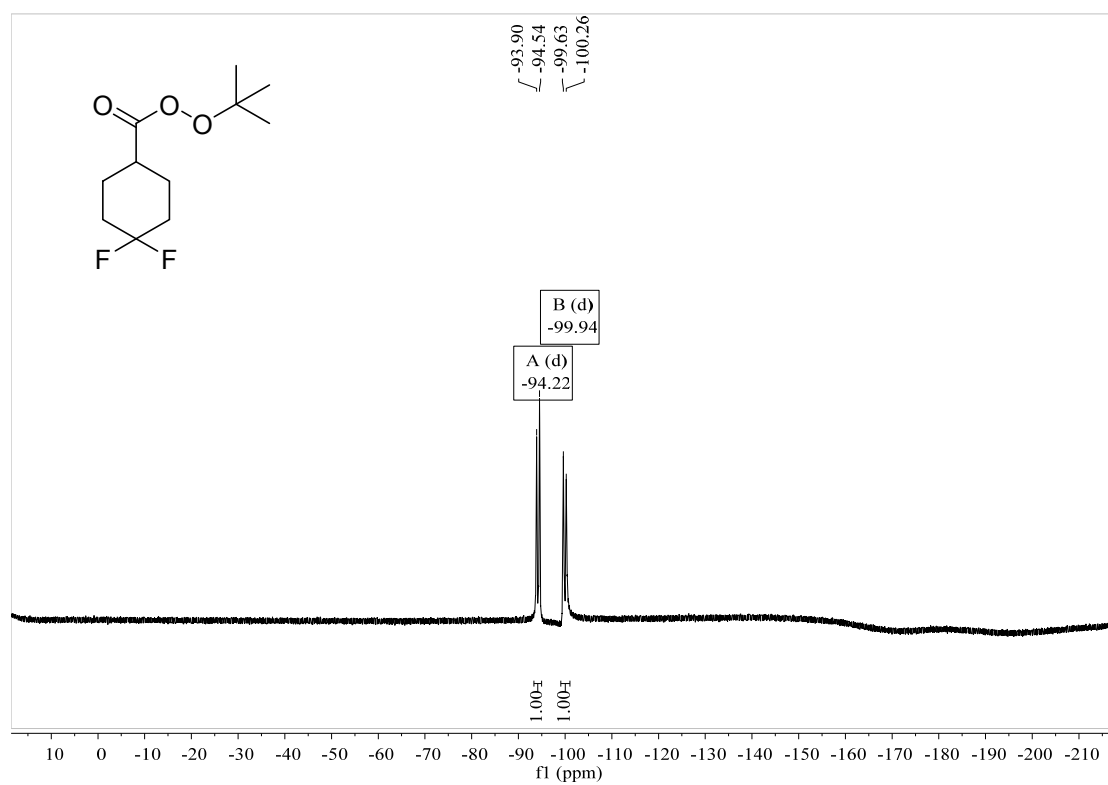


Figure S34. ¹⁹F NMR spectrum of *tert*-butyl 4,4-difluorocyclohexane-1-carboperoxoate, related to **Figure 2**.

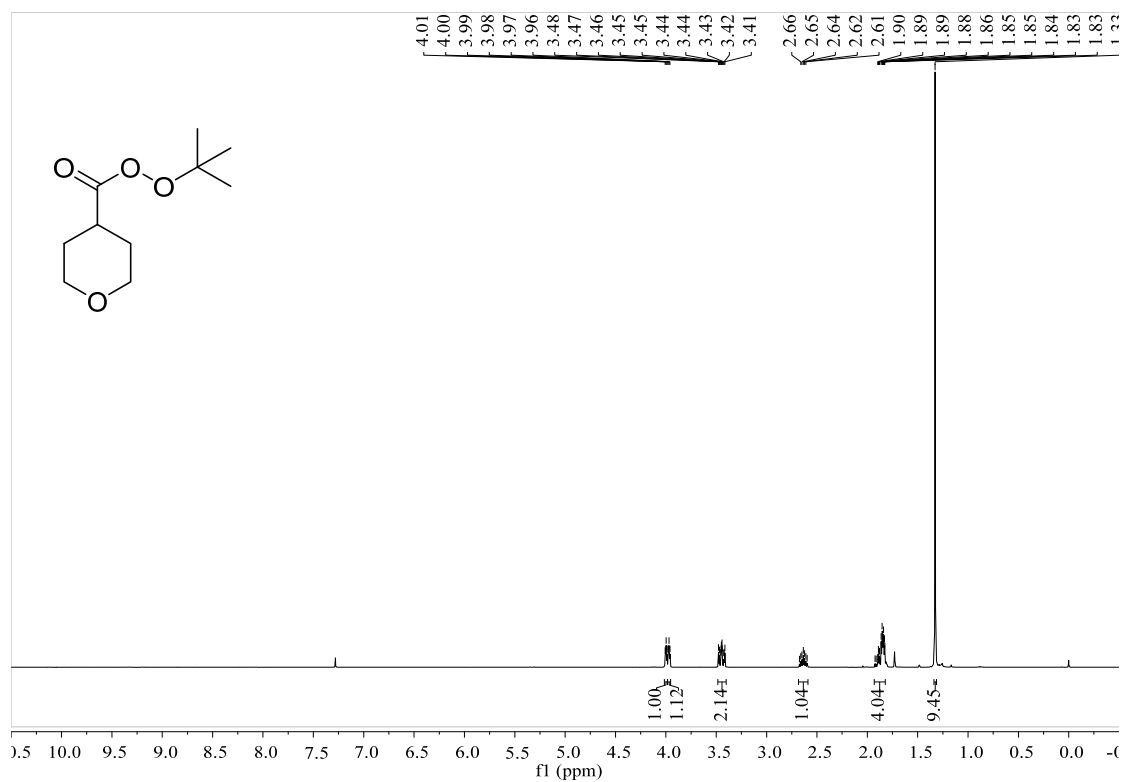


Figure S35. ^1H NMR spectrum of *tert*-butyl tetrahydro-2*H*-pyran-4-carboperoxoate, related to Figure 2.

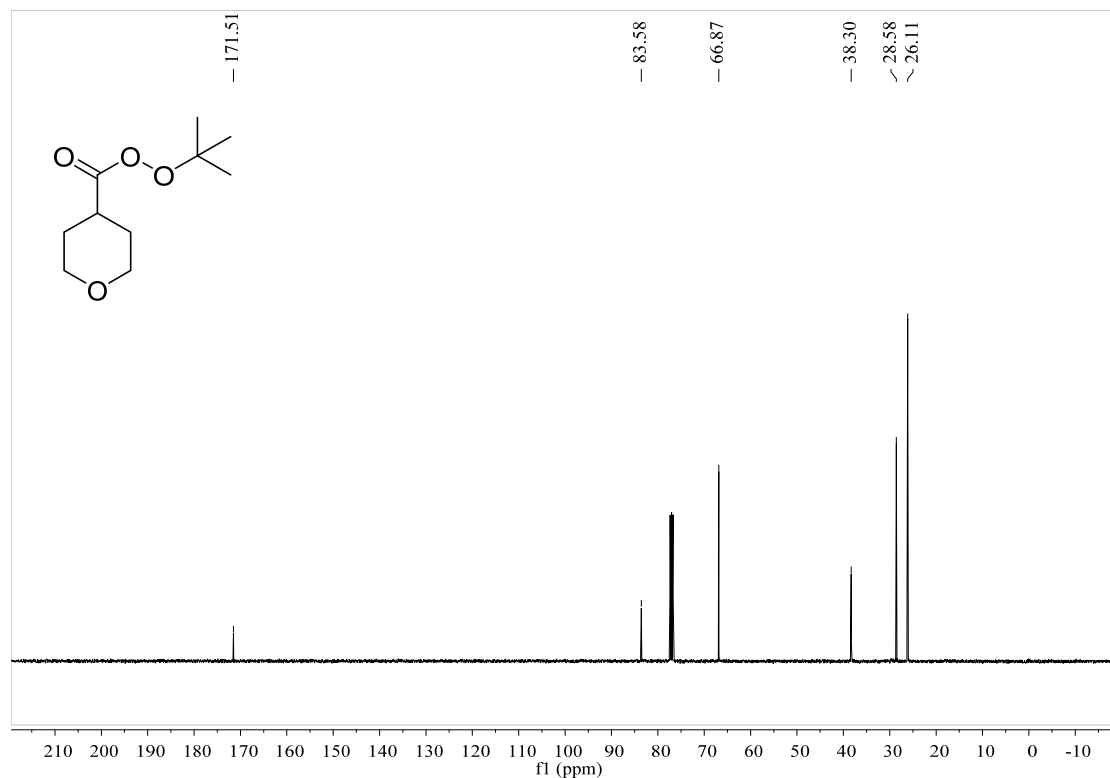


Figure S36. ^{13}C NMR spectrum of *tert*-butyl tetrahydro-2*H*-pyran-4-carboperoxoate, related to Figure 2.

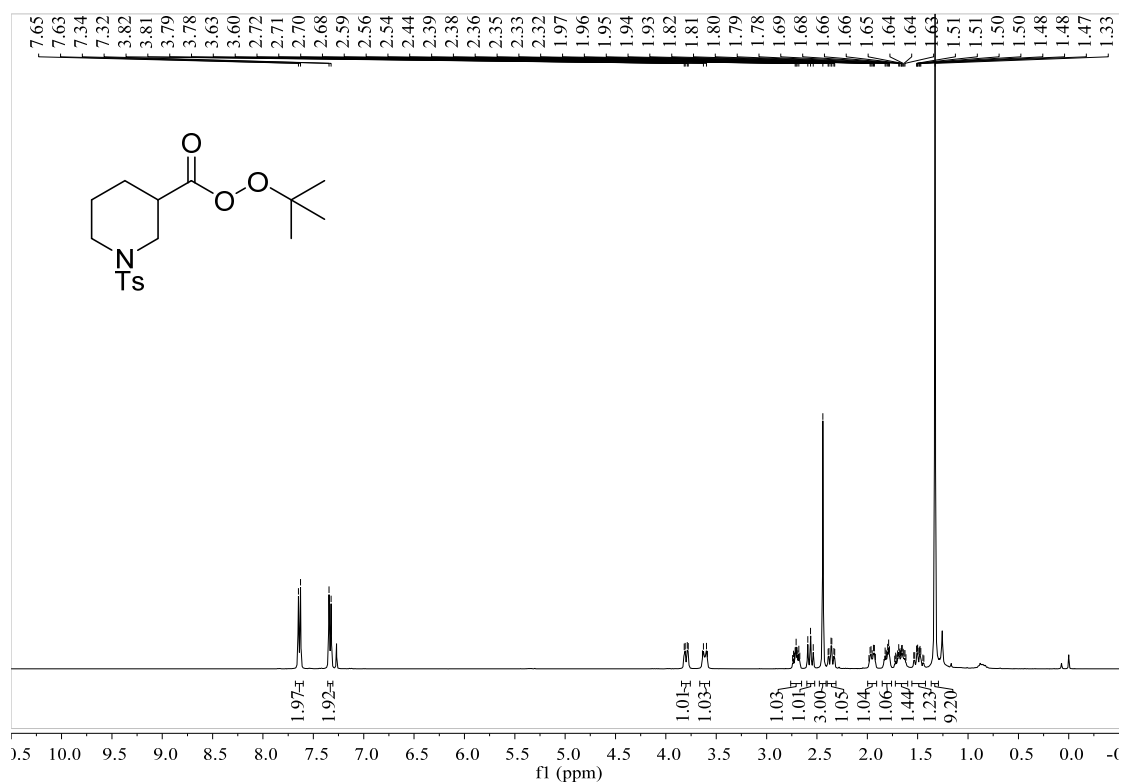


Figure S37. ^1H NMR spectrum of *tert*-butyl 1-tosylpiperidine-3-carboperoxoate, related to Figure 2.

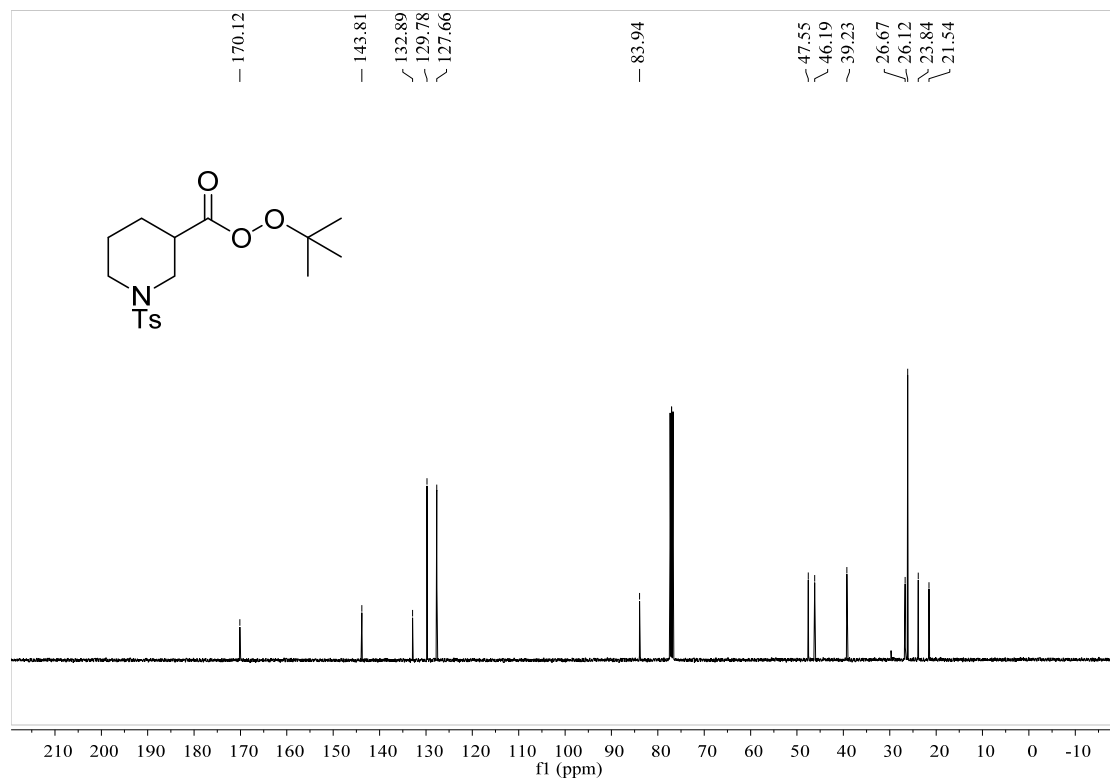


Figure S38. ^{13}C NMR spectrum of *tert*-butyl 1-tosylpiperidine-3-carboperoxoate, related to Figure 2.

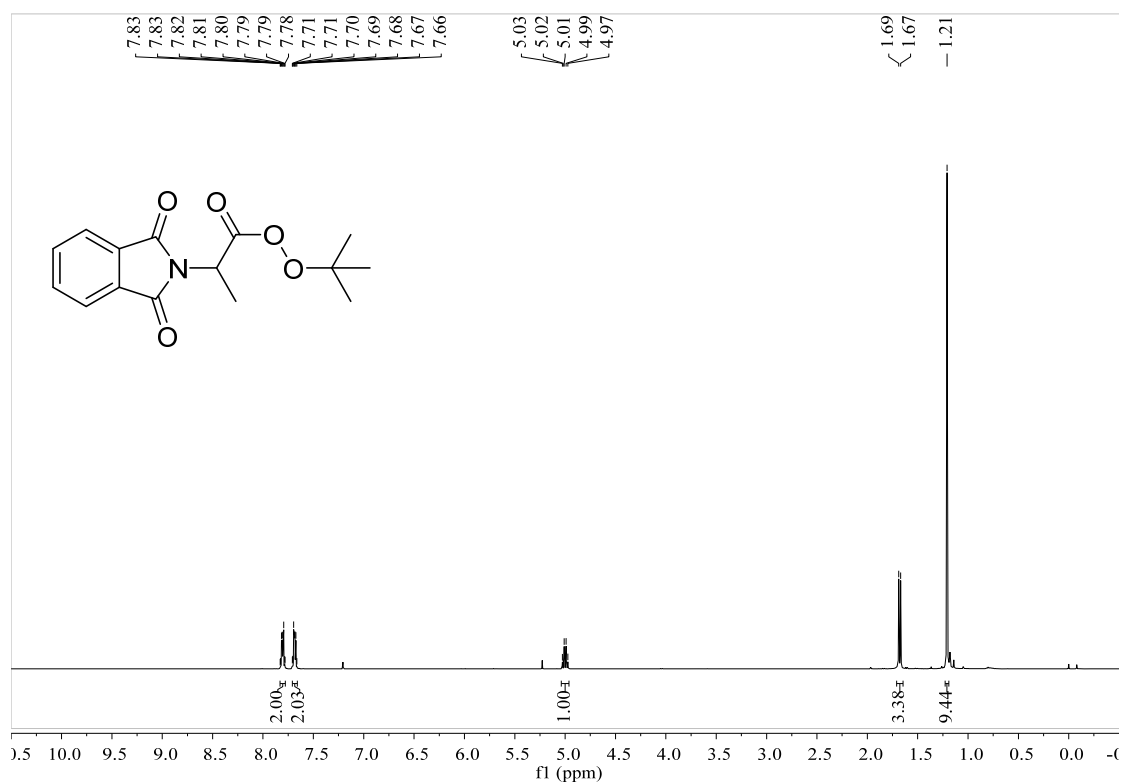


Figure S39. ¹H NMR spectrum of *tert*-butyl 2-(1,3-dioxisoindolin-2-yl)propaneperoxoate, related to **Figure 2**.

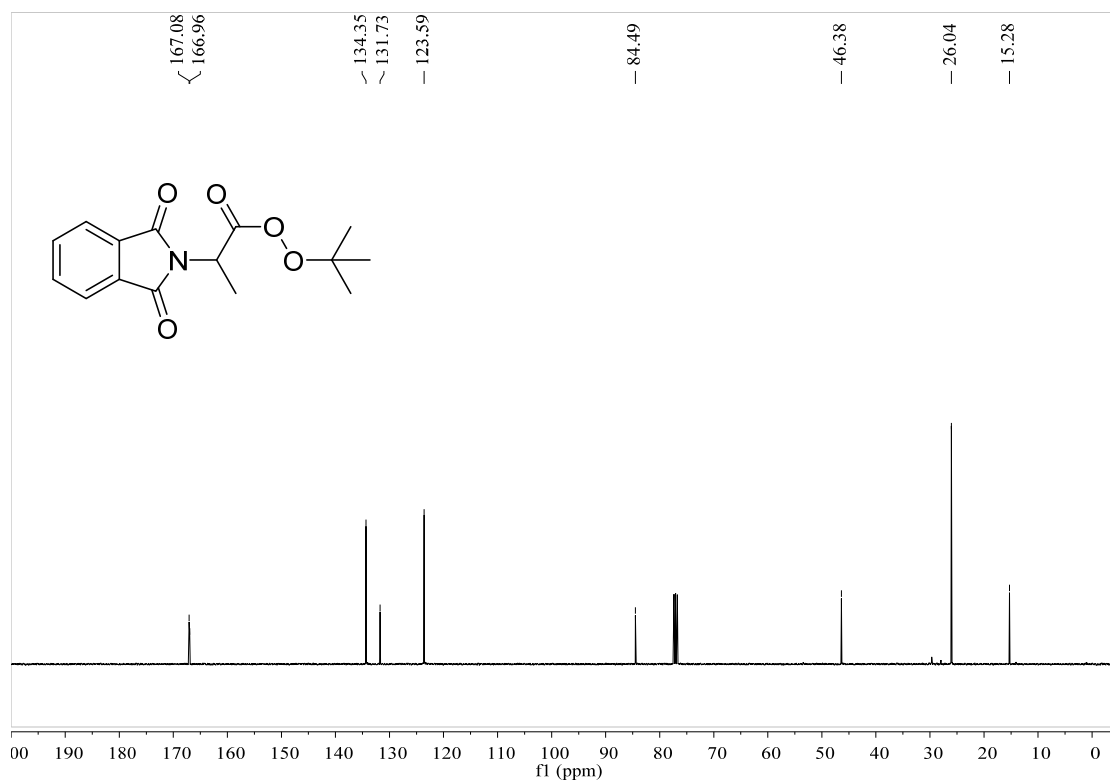


Figure S40. ¹³C NMR spectrum of *tert*-butyl 2-(1,3-dioxisoindolin-2-yl)propaneperoxoate, related to **Figure 2**.

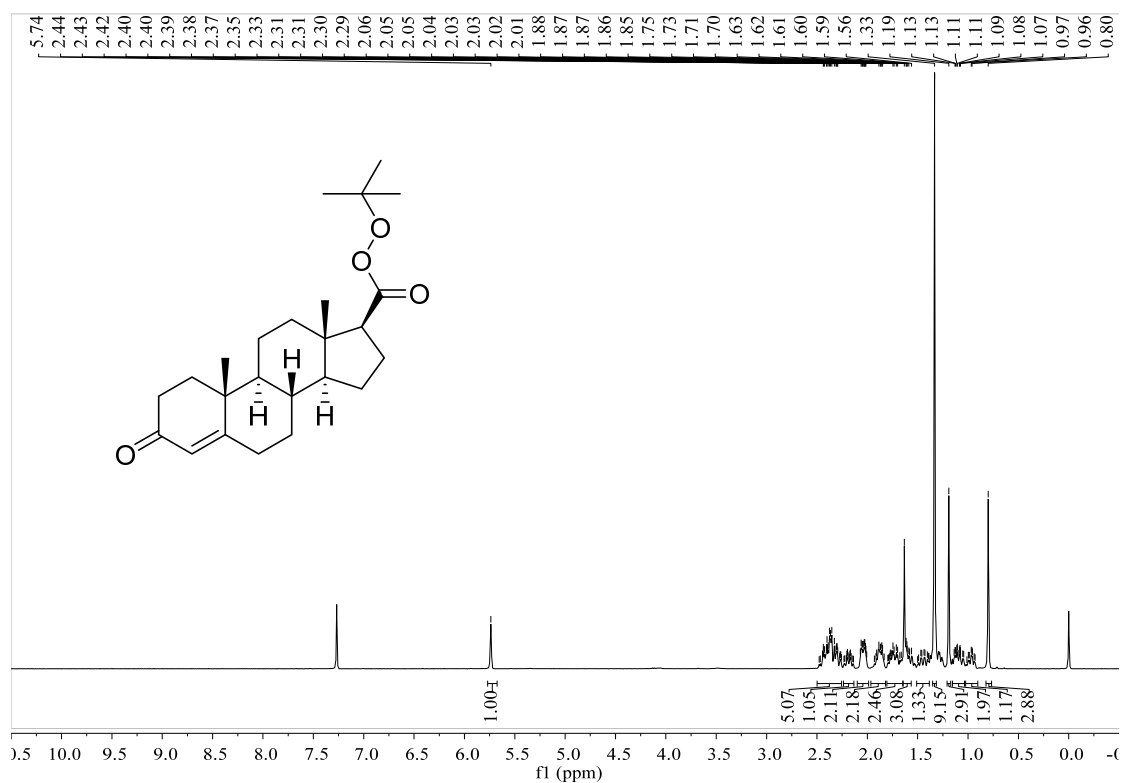


Figure S41. ^1H NMR spectrum of compound 55, related to scheme 2.

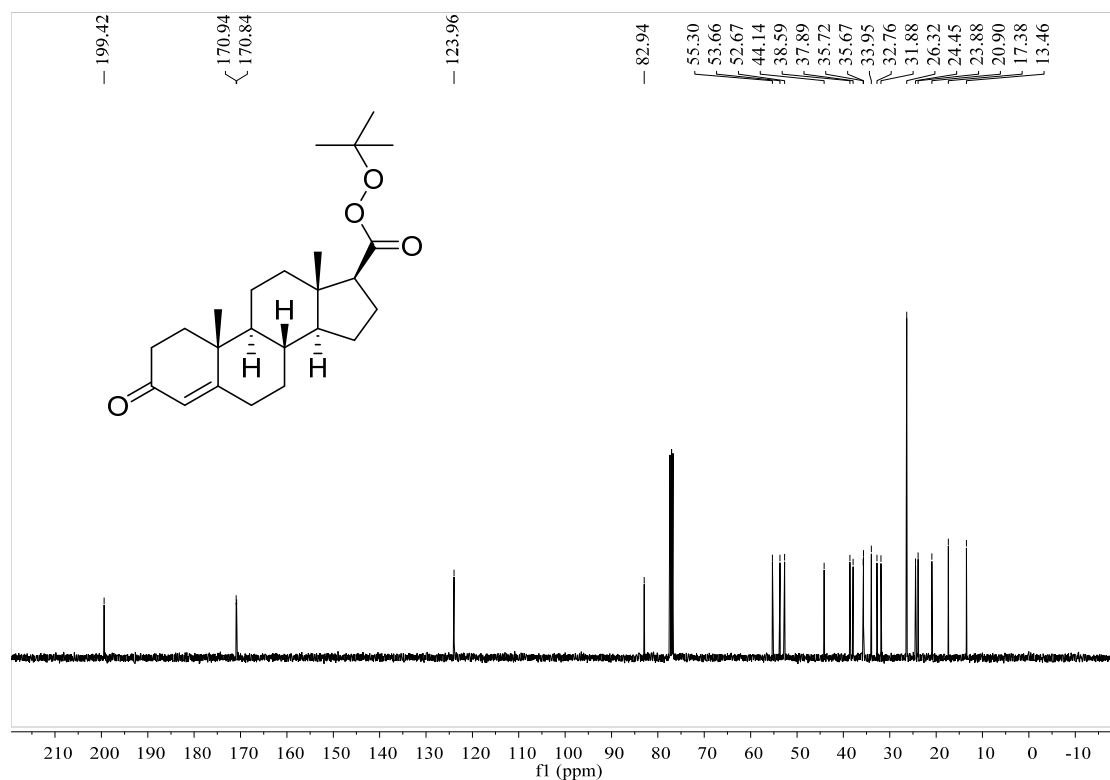


Figure S42. ^{13}C NMR spectrum of compound 55, related to scheme 2.

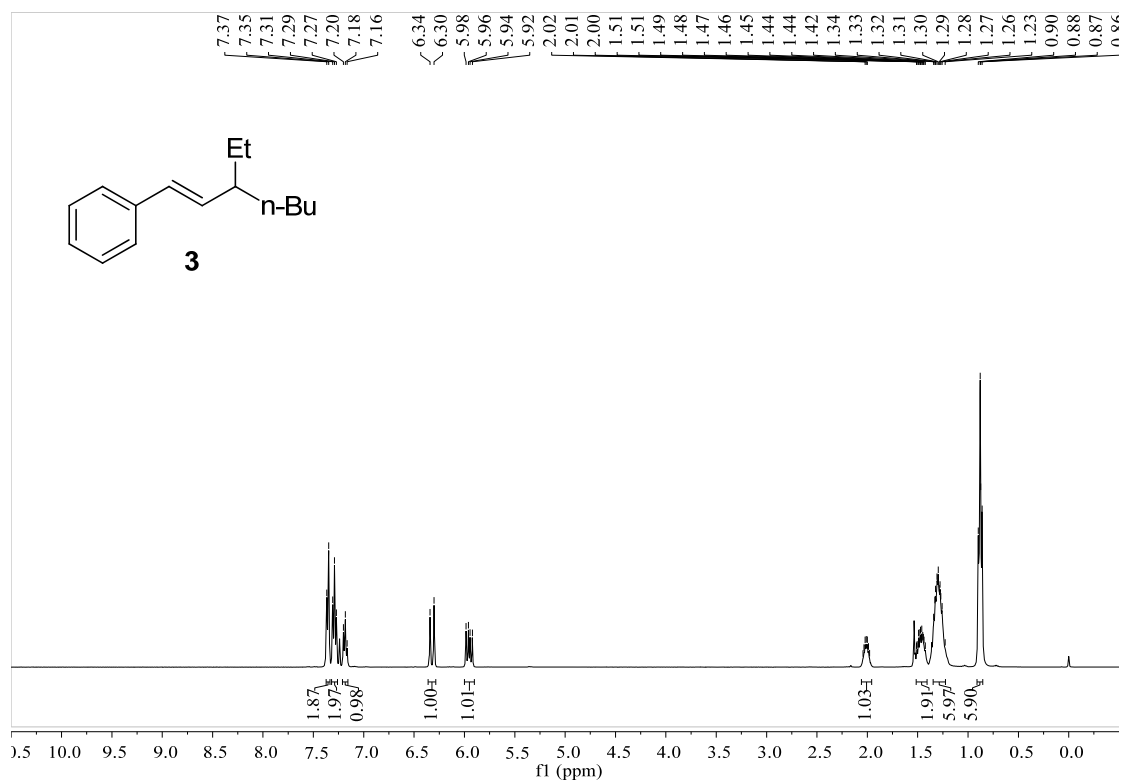


Figure S43. ¹H NMR spectrum of compound 3, related to Table 1.

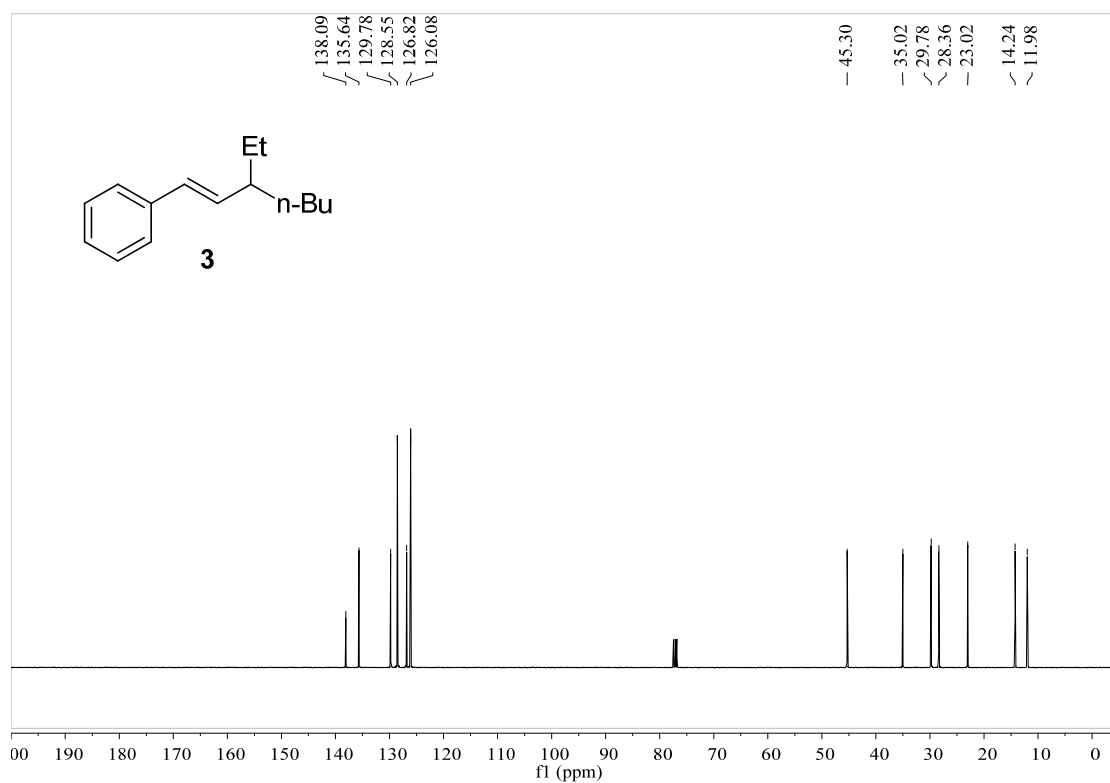


Figure S44. ¹³C NMR spectrum of compound 3, related to Table 1.

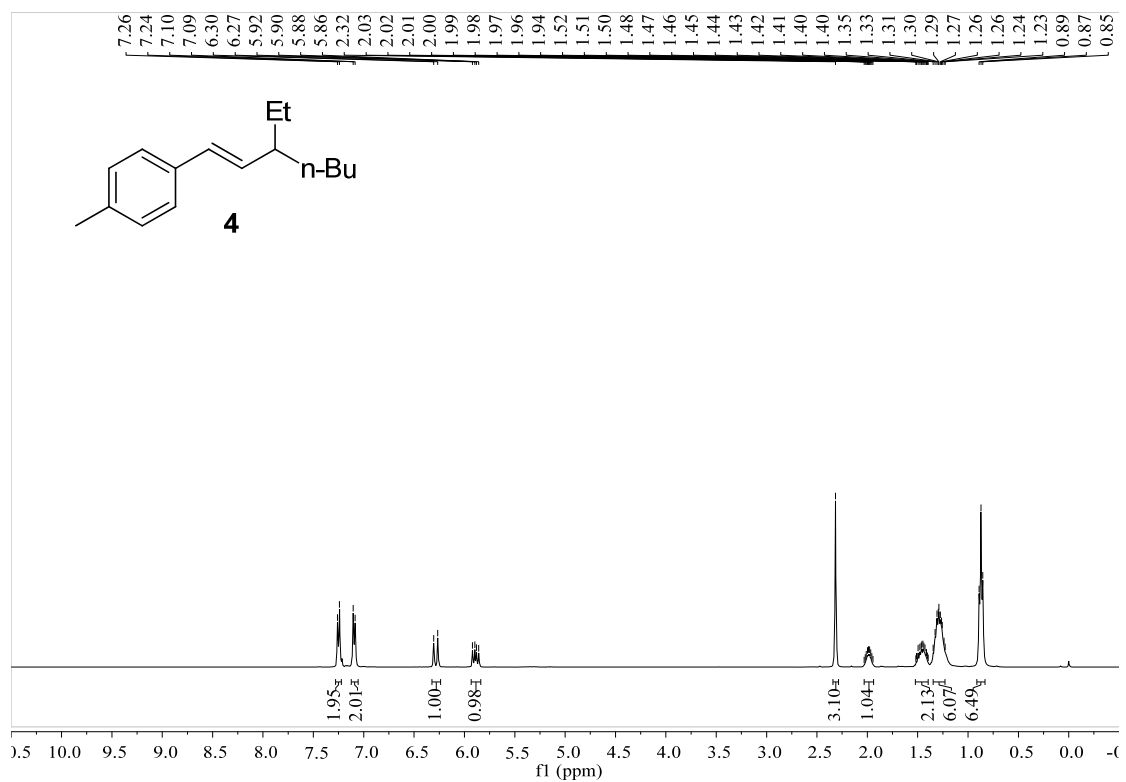


Figure S45. ¹H NMR spectrum of compound 4, related to Figure 1.

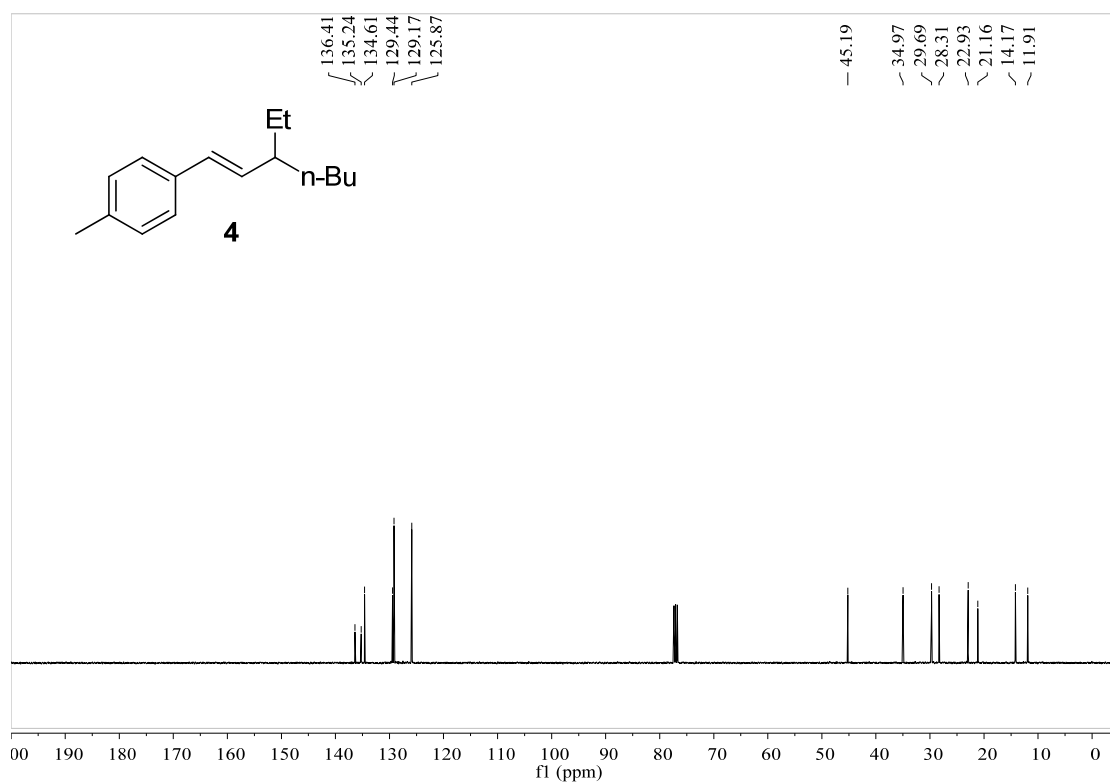


Figure S46. ¹³C NMR spectrum of compound 4, related to Figure 1.

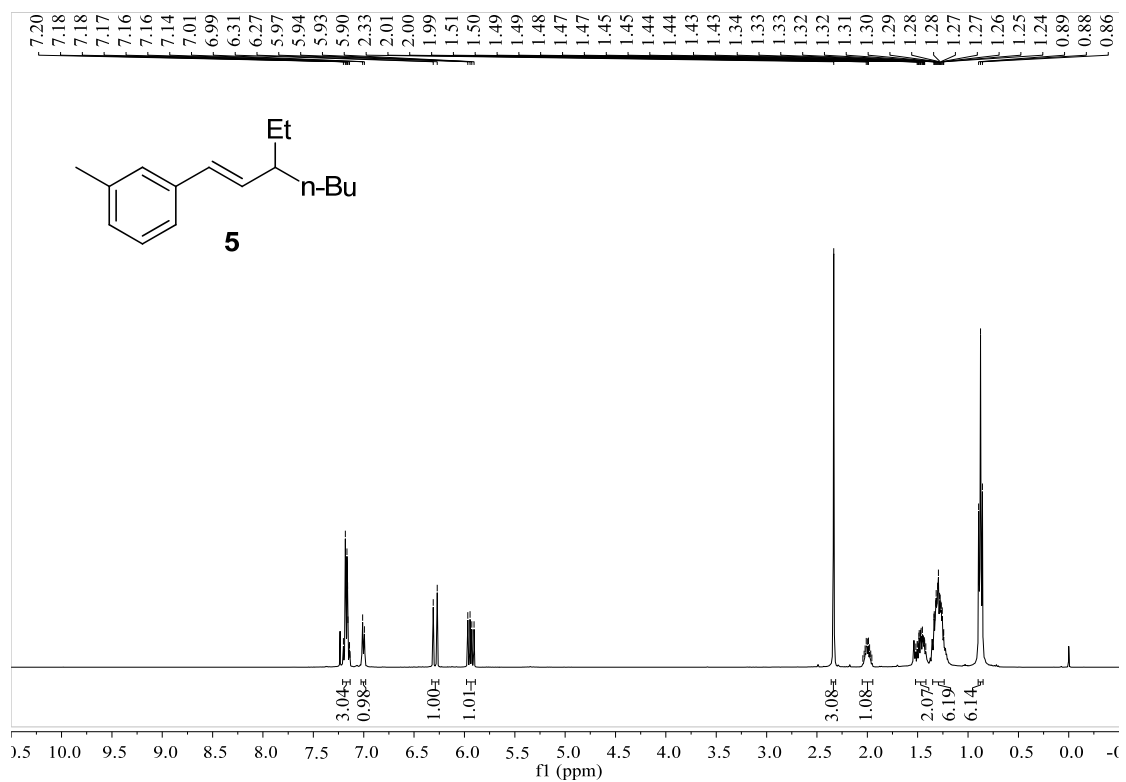


Figure S47. ¹H NMR spectrum of compound **5**, related to Figure 1.

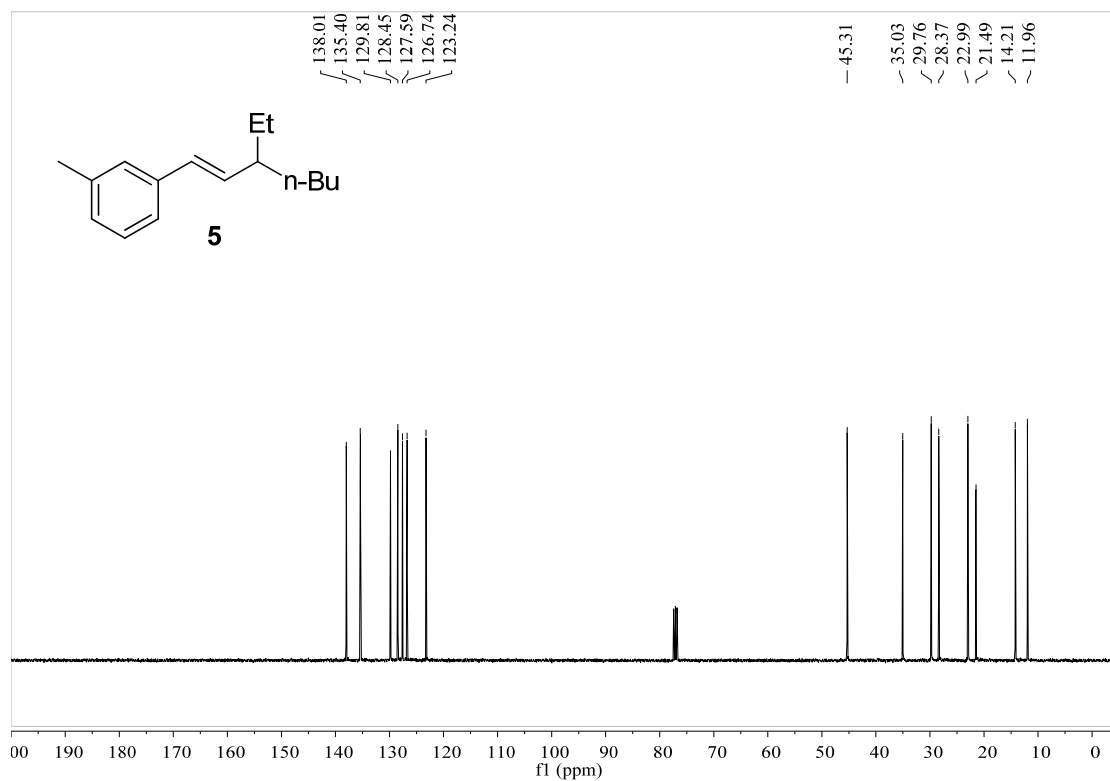


Figure S48. ¹³C NMR spectrum of compound **5**, related to Figure 1.

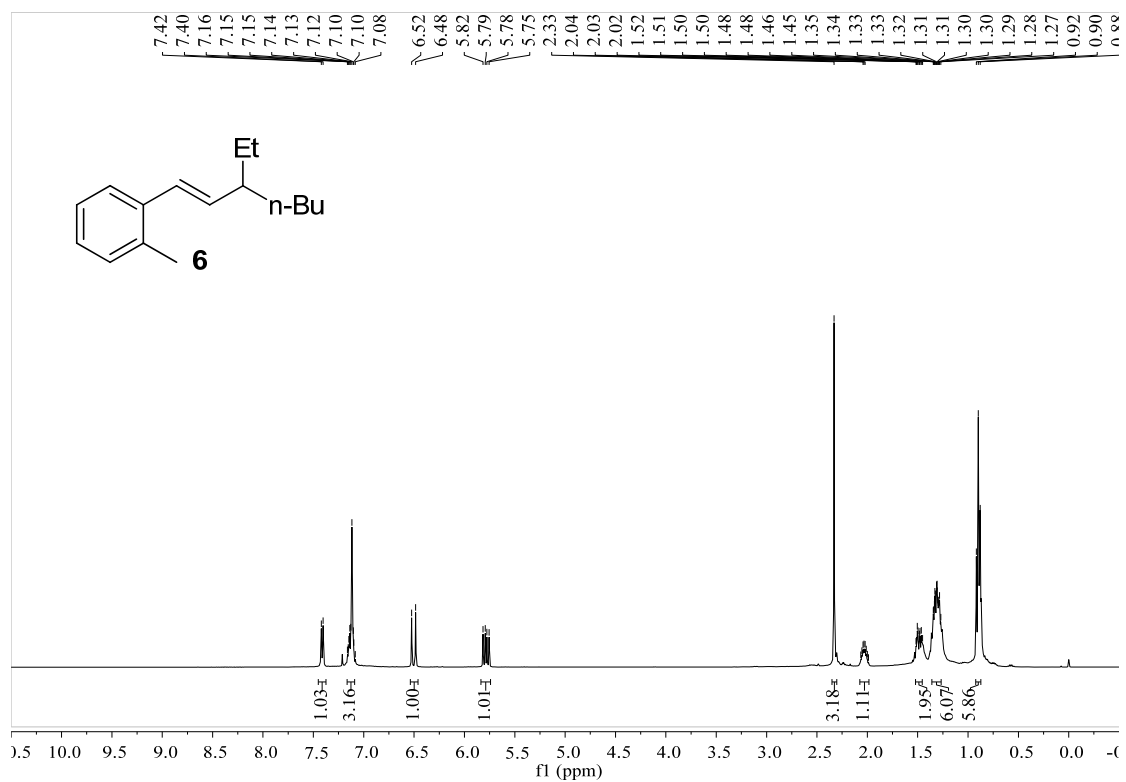


Figure S49. ¹H NMR spectrum of compound **6**, related to Figure 1.

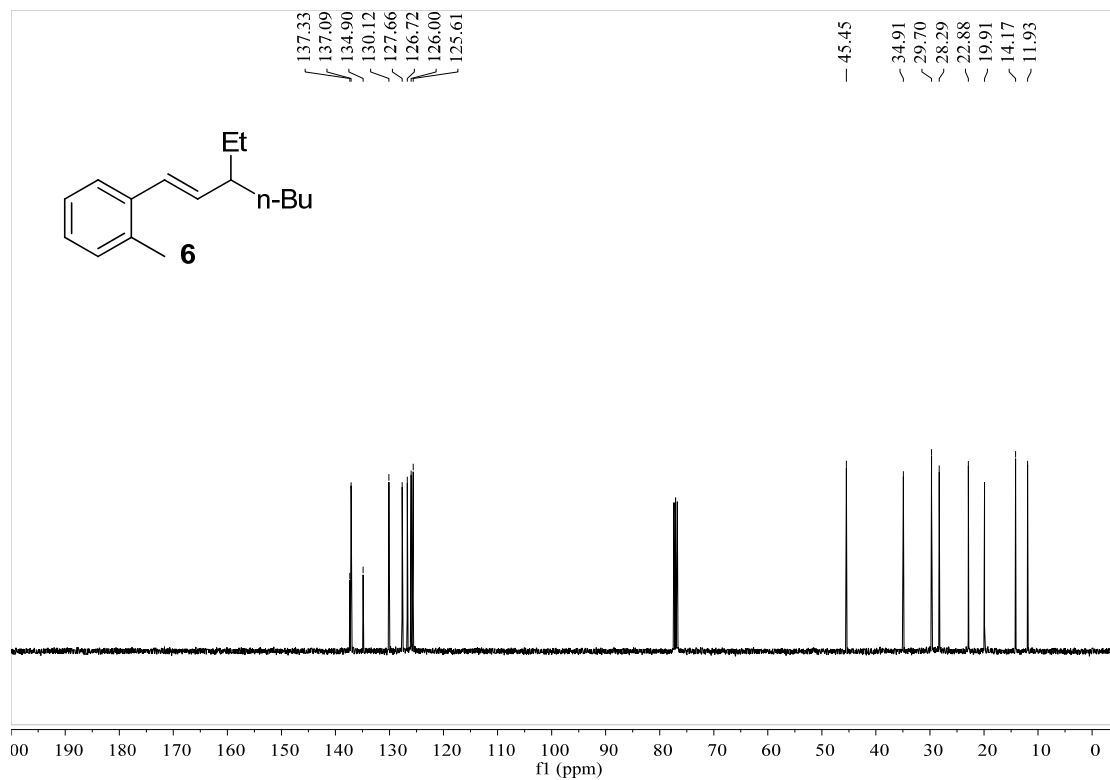


Figure S50. ¹³C NMR spectrum of compound **6**, related to Figure 1.

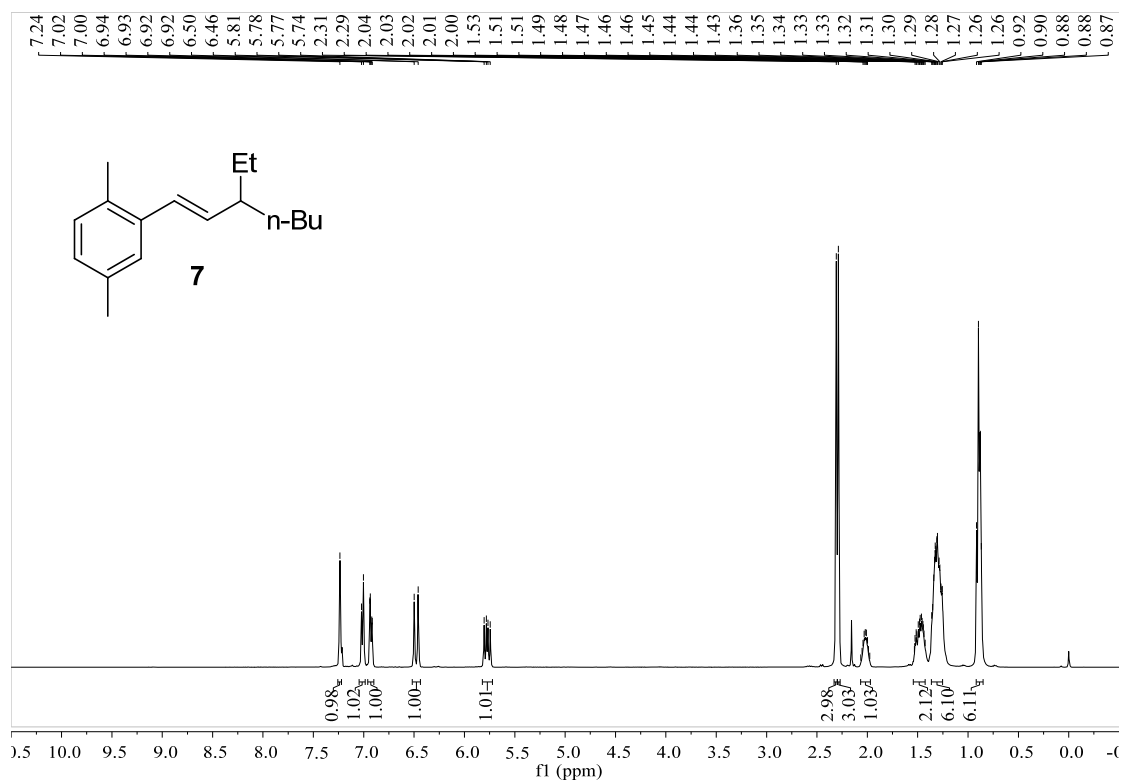


Figure S51. ^1H NMR spectrum of compound **7**, related to **Figure 1**.

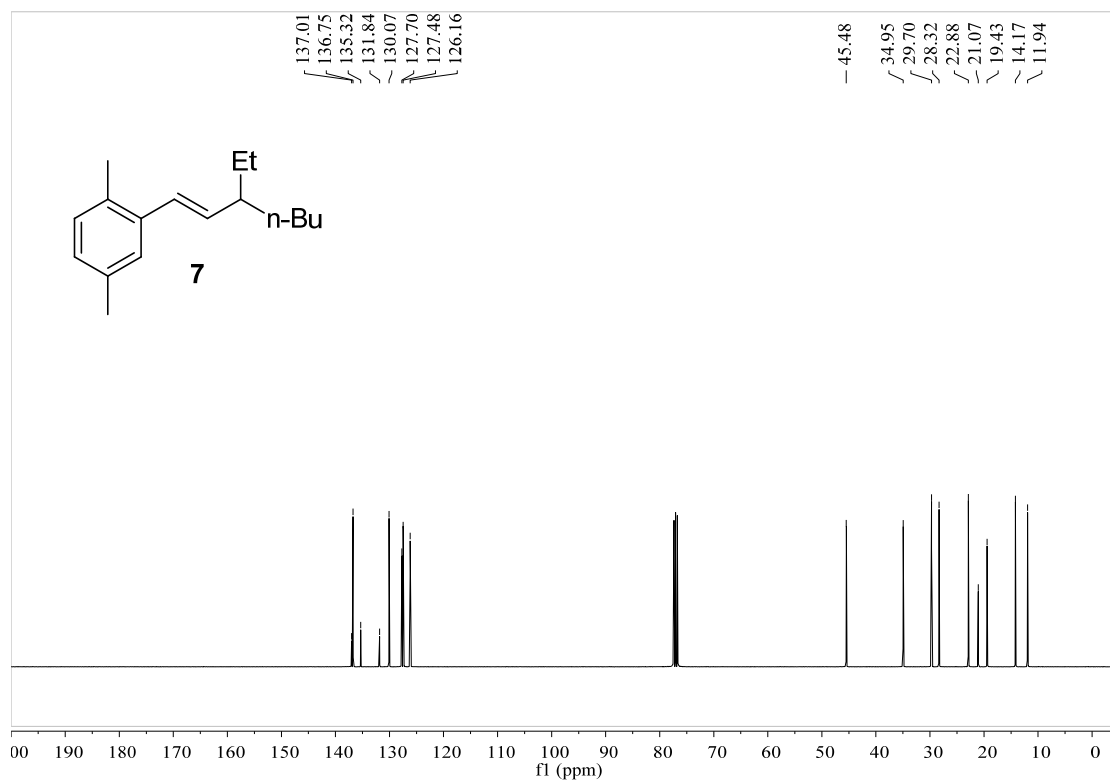


Figure S52. ^{13}C NMR spectrum of compound **7**, related to **Figure 1**.

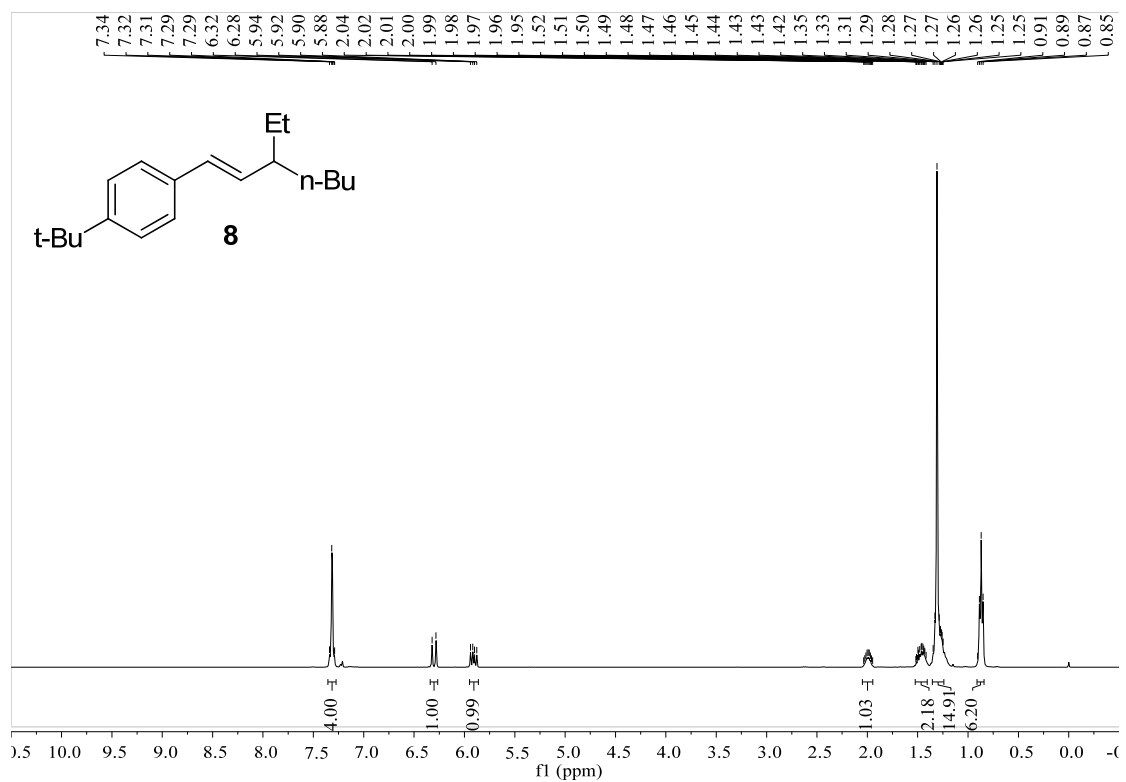


Figure S53. ¹H NMR spectrum of compound **8**, related to Figure 1.

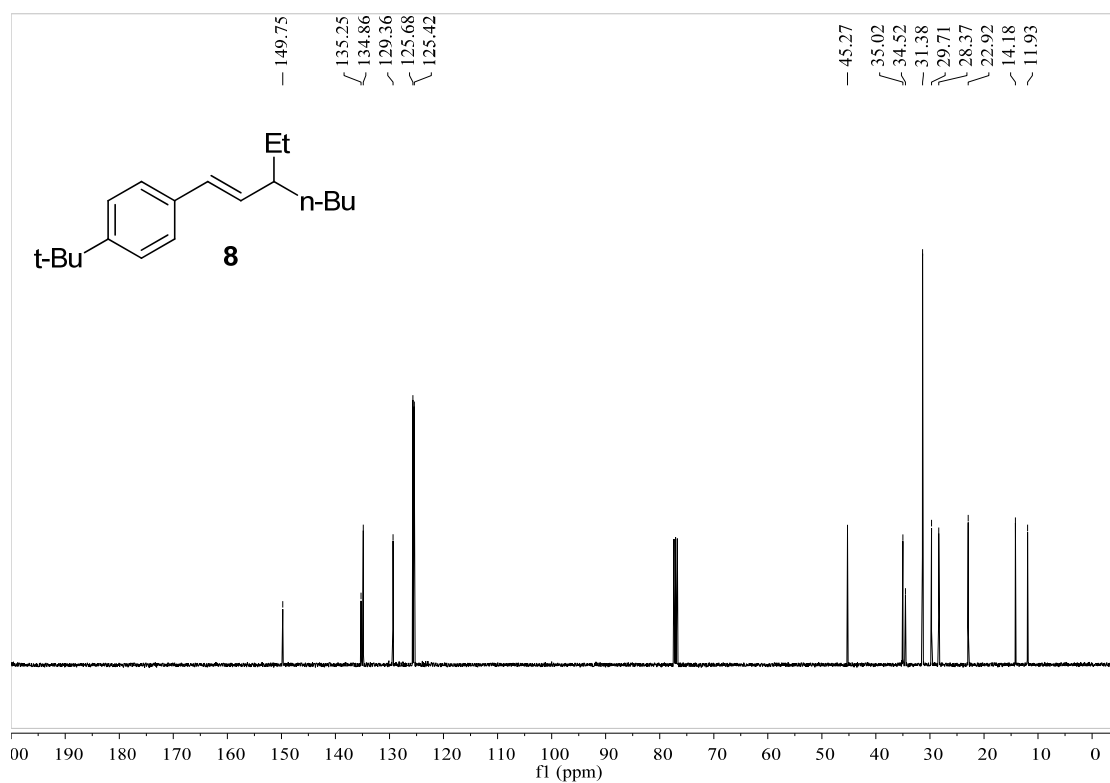


Figure S54. ¹³C NMR spectrum of compound **8**, related to Figure 1.

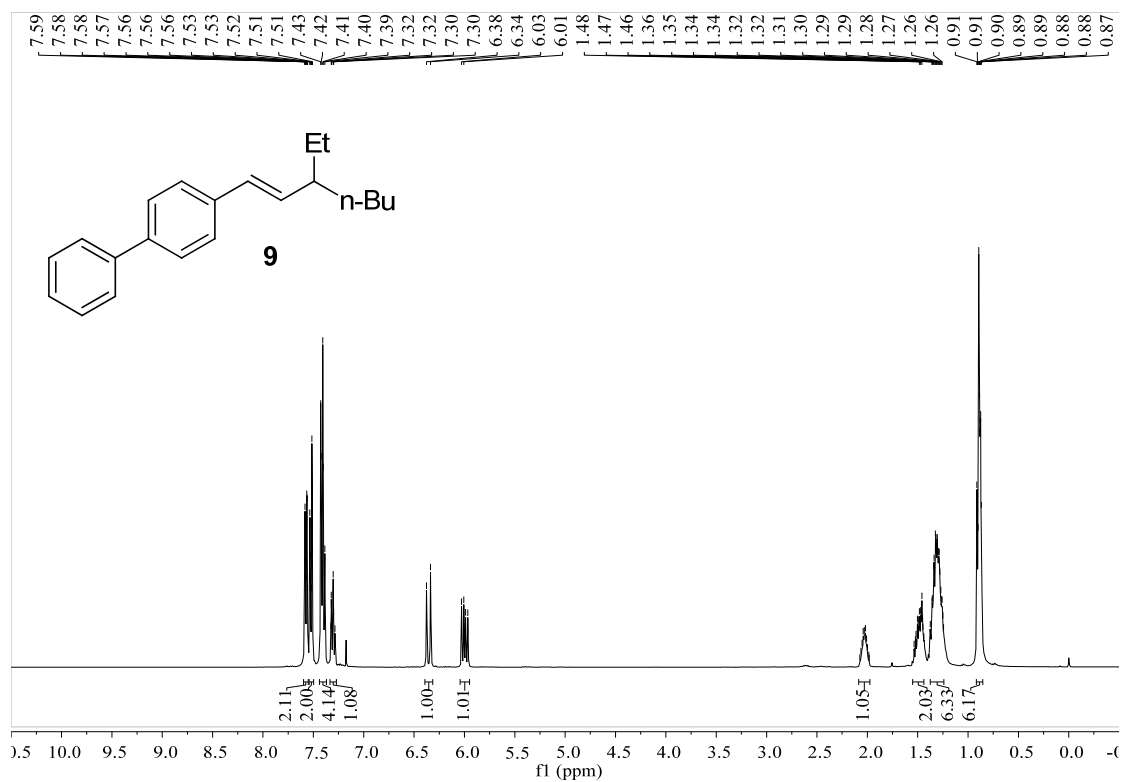


Figure S55. ^1H NMR spectrum of compound **9**, related to **Figure 1**.

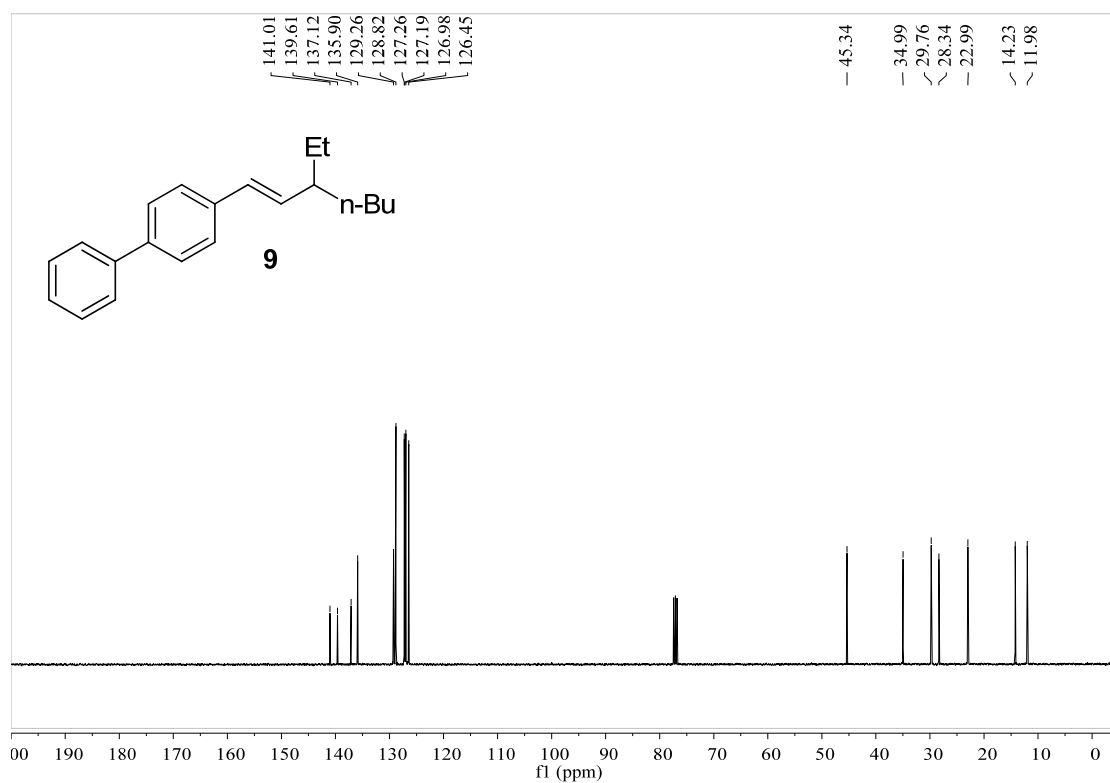


Figure S56. ^{13}C NMR spectrum of compound **9**, related to **Figure 1**.

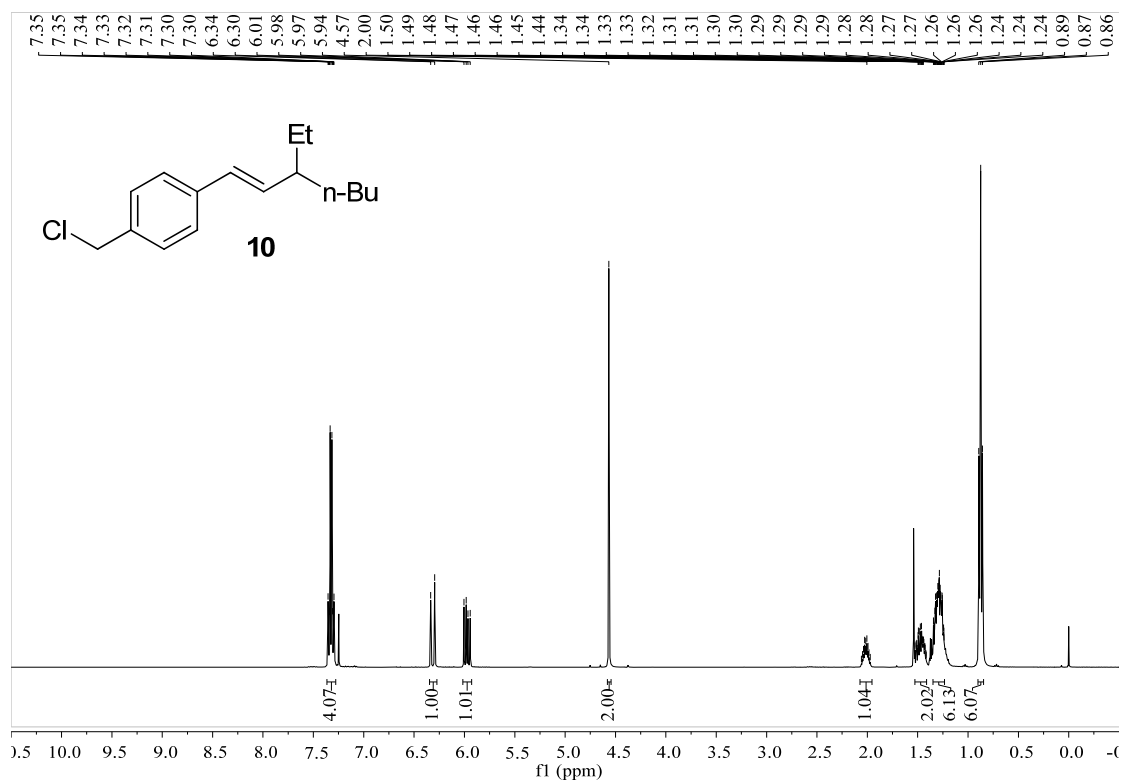


Figure S57. ¹H NMR spectrum of compound **10**, related to Figure 1.

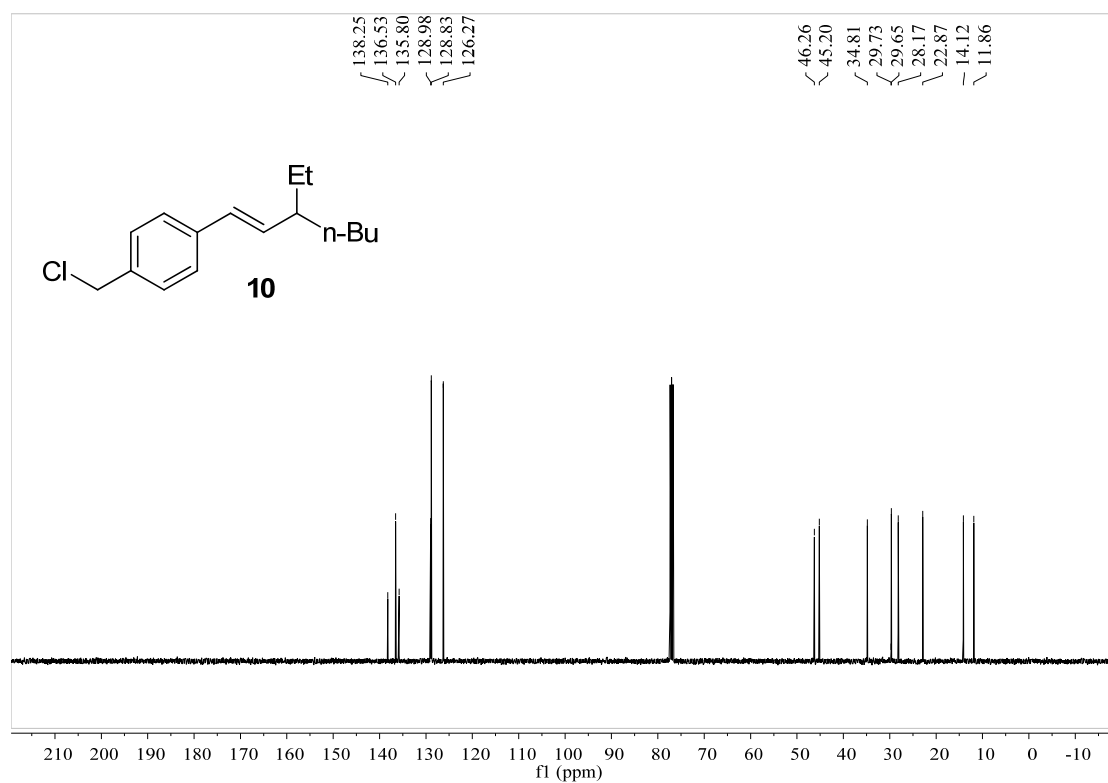


Figure S58. ¹³C NMR spectrum of compound **10**, related to Figure 1.

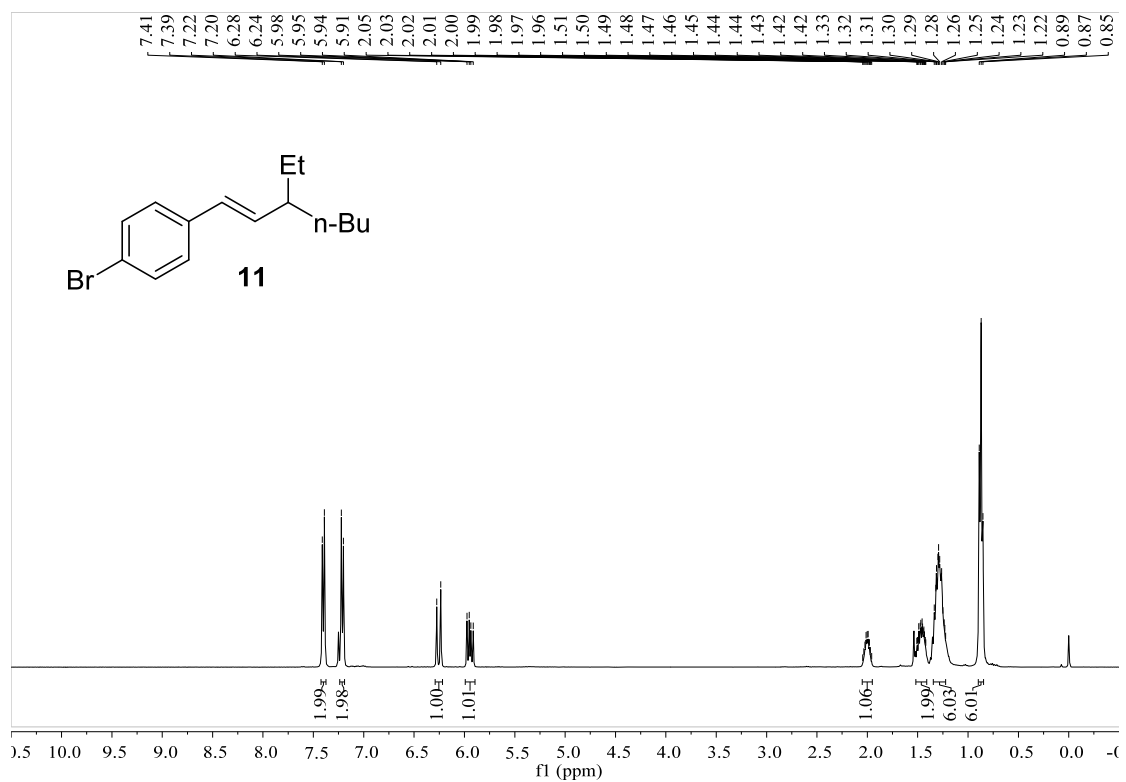


Figure S59. ¹H NMR spectrum of compound **11**, related to **Figure 1**.

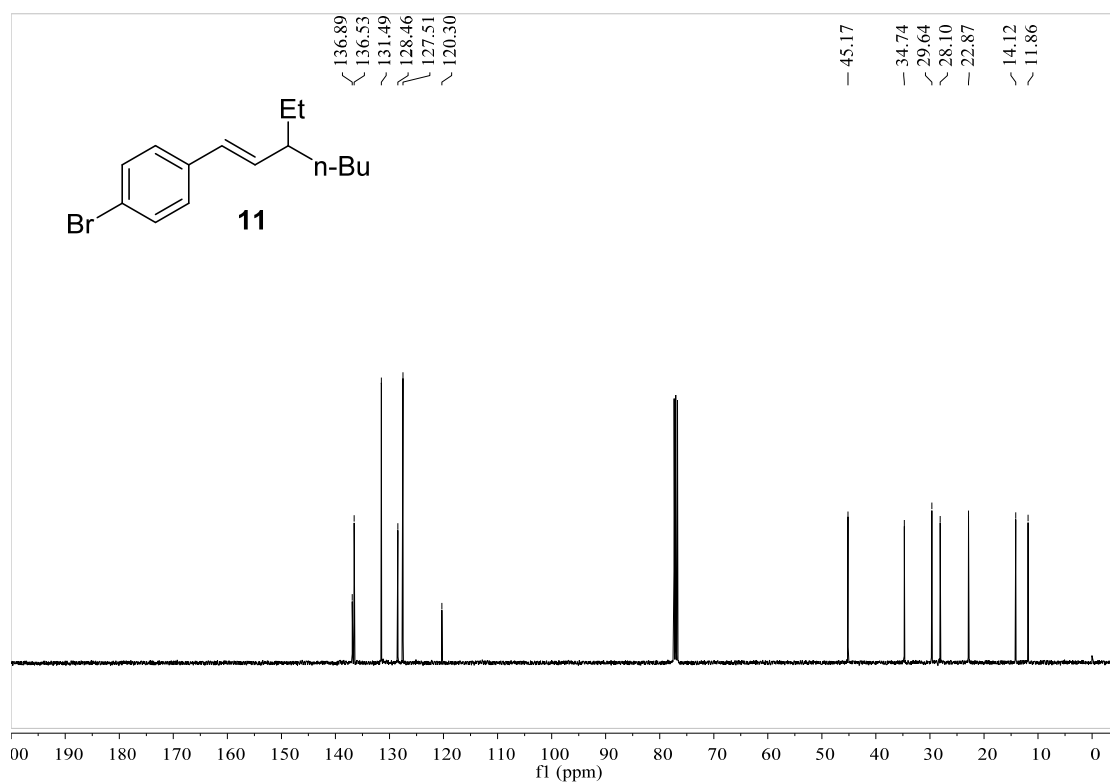


Figure S60. ¹³C NMR spectrum of compound **11**, related to **Figure 1**.

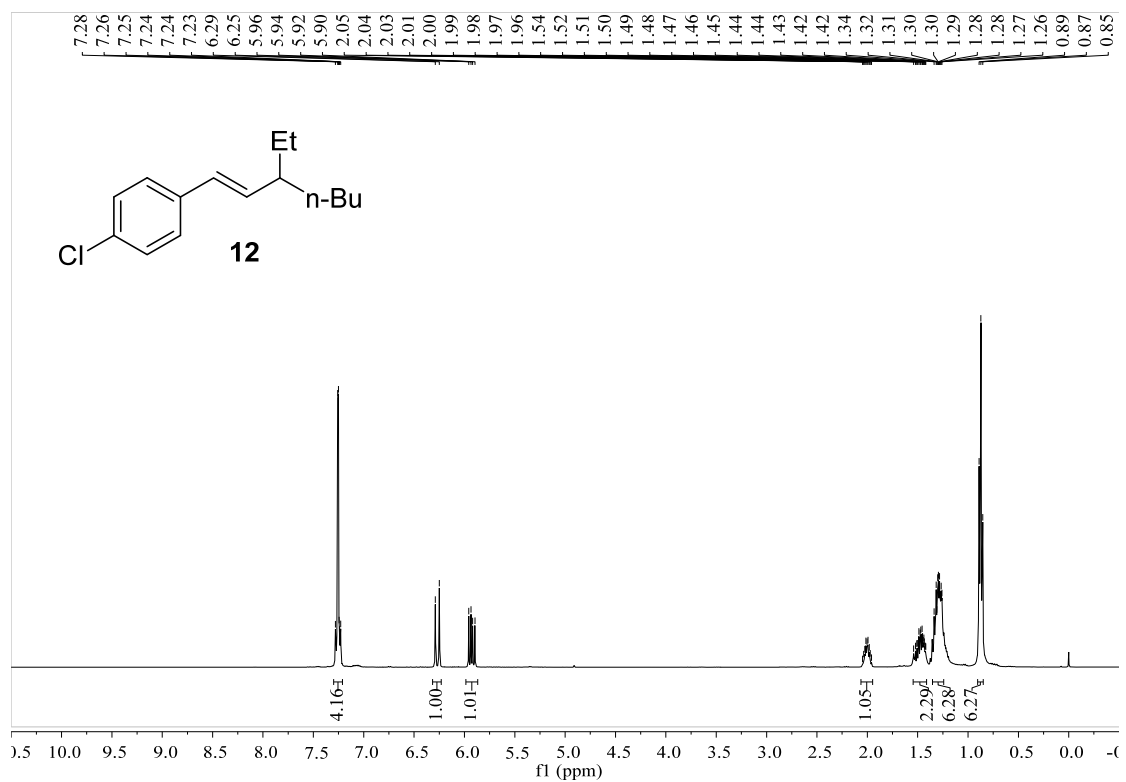


Figure S61. ¹H NMR spectrum of compound **12**, related to **Figure 1**.

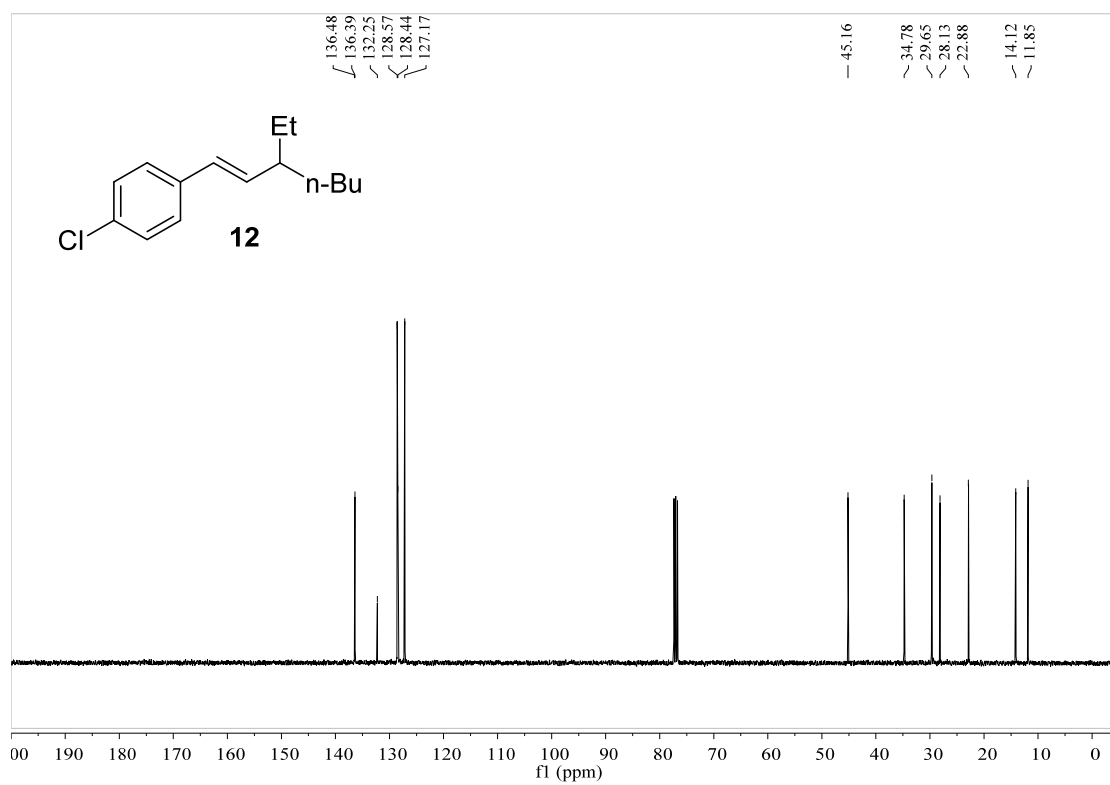


Figure S62. ¹³C NMR spectrum of compound **12**, related to **Figure 1**.

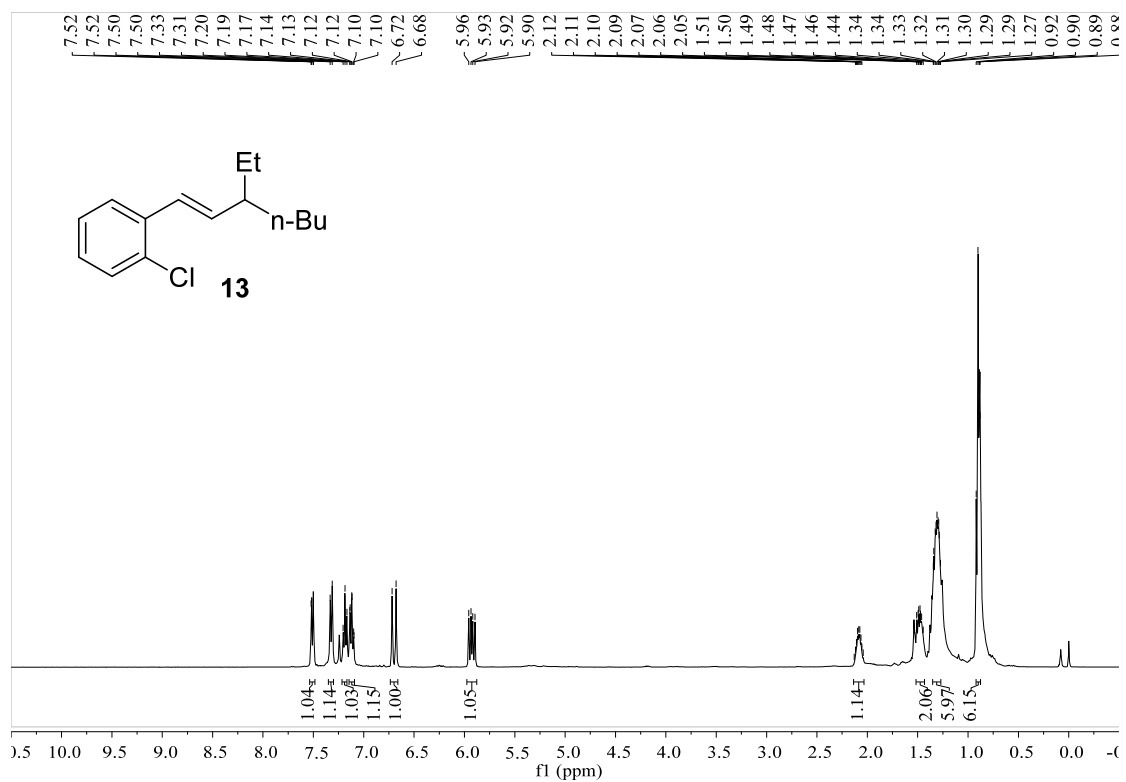


Figure S63. ¹H NMR spectrum of compound **13**, related to **Figure 1**.

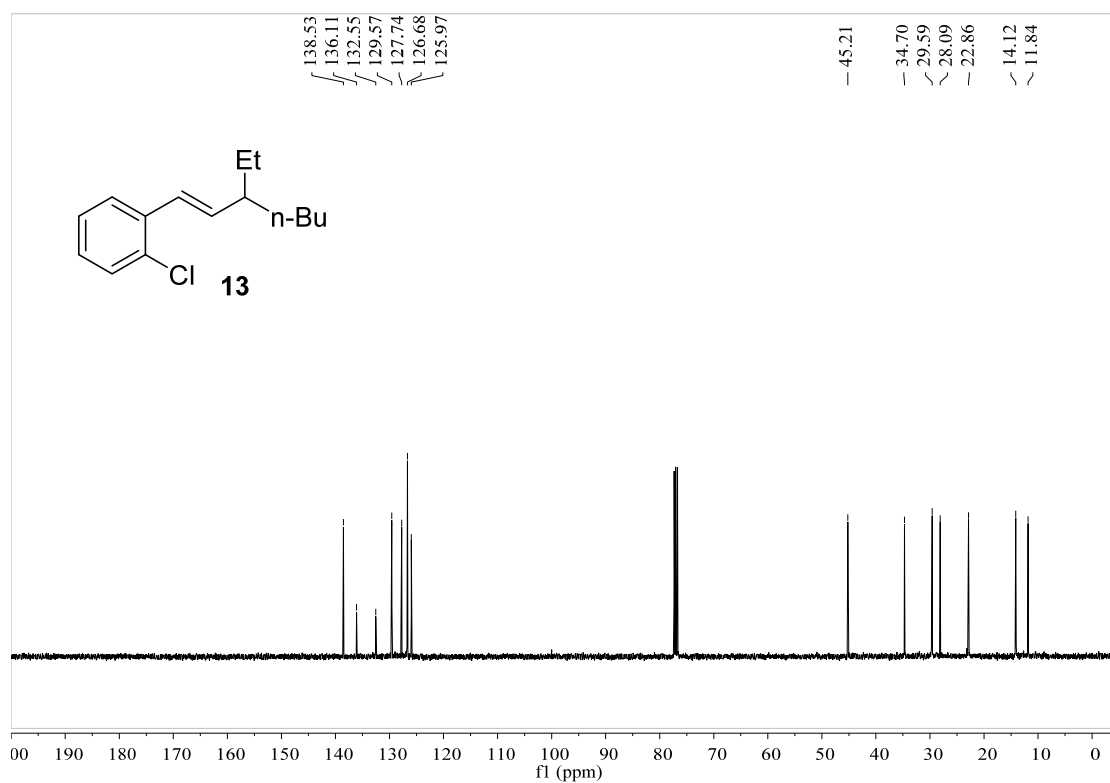


Figure S64. ¹³C NMR spectrum of compound **13**, related to **Figure 1**.

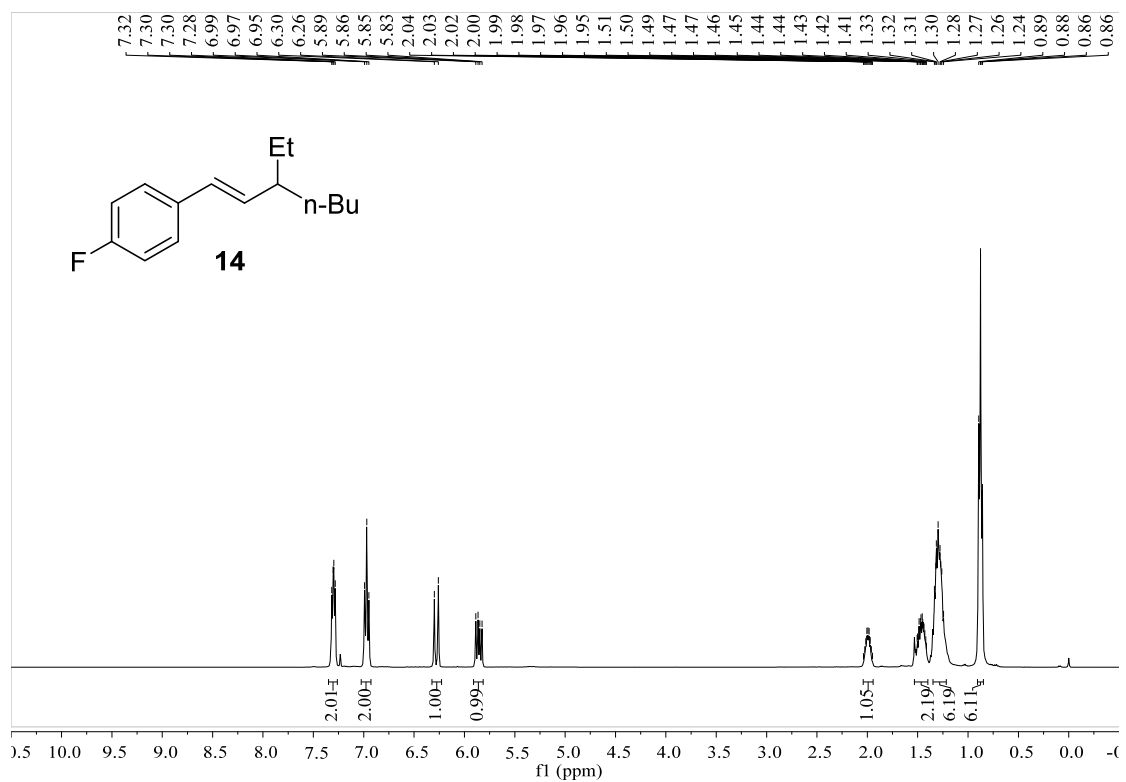


Figure S65. ¹H NMR spectrum of compound **14**, related to Figure 1.

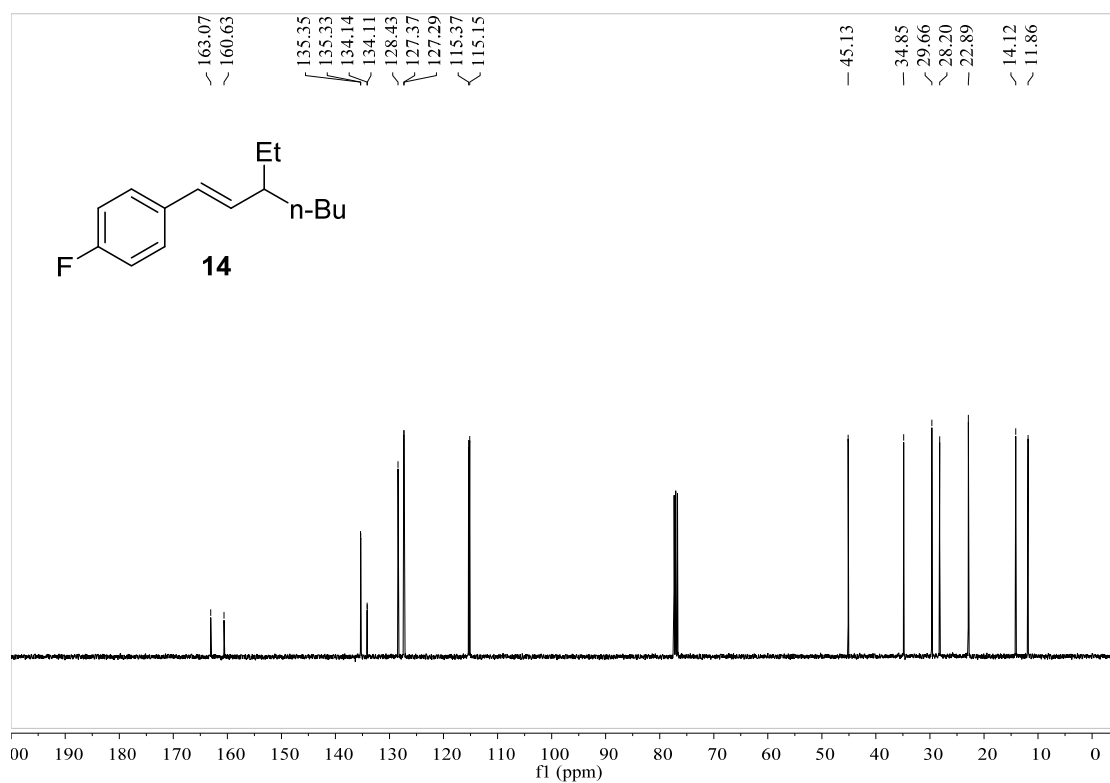


Figure S66. ¹³C NMR spectrum of compound **14**, related to Figure 1.

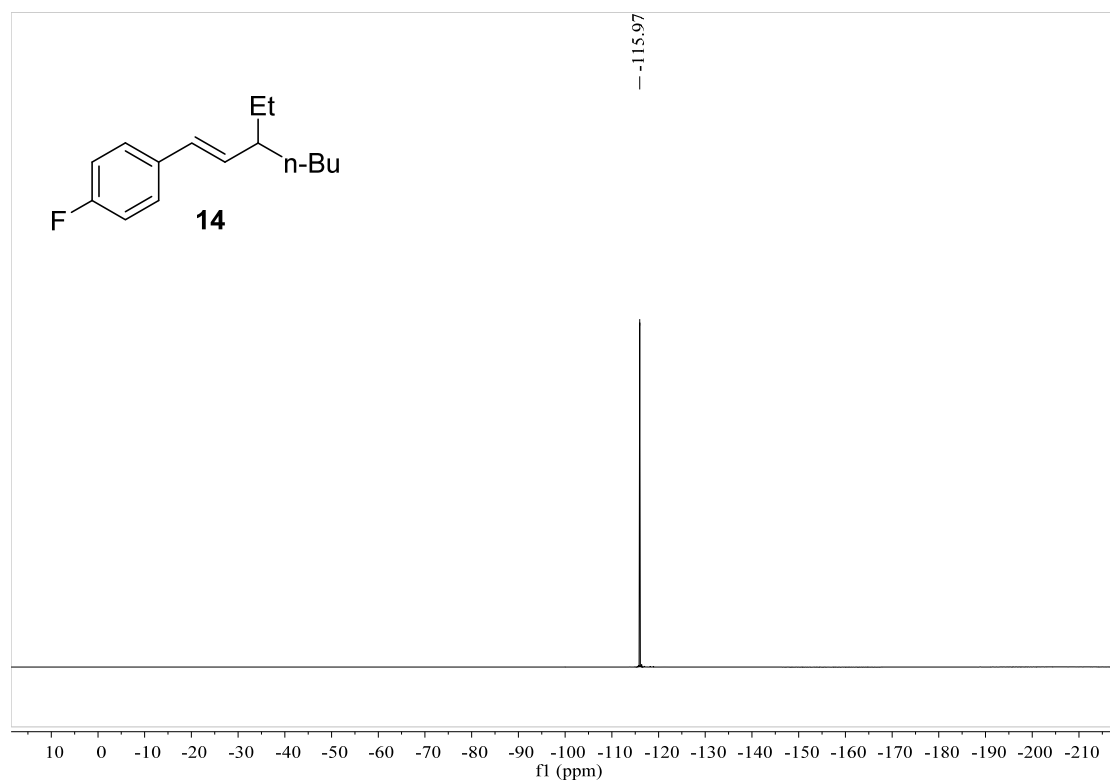


Figure S67. ^{19}F NMR spectrum of compound **14**, related to **Figure 1**.

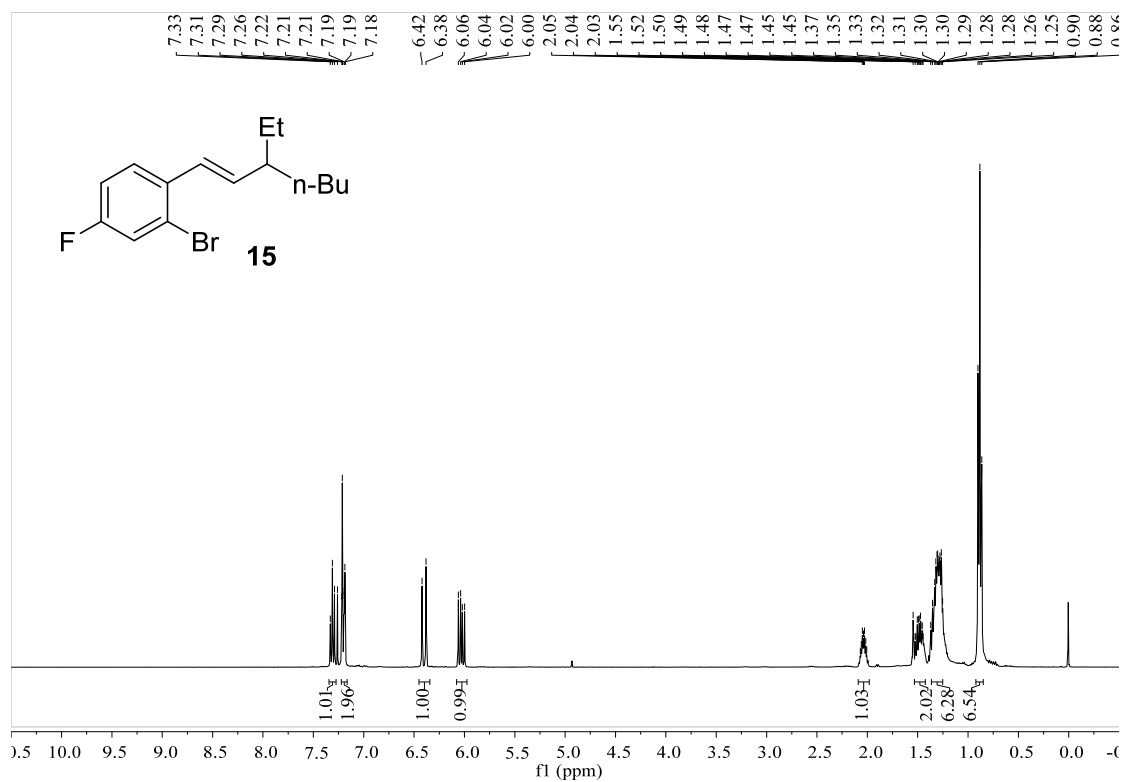


Figure S68. ¹H NMR spectrum of compound **15**, related to **Figure 1**.

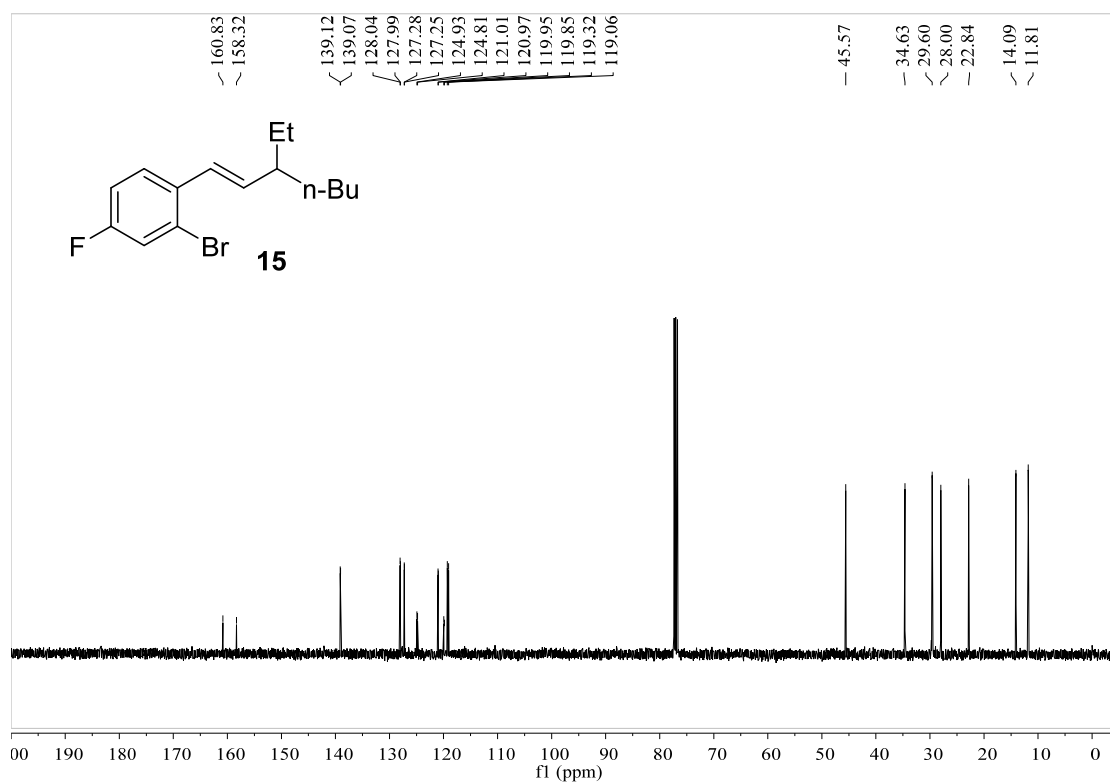


Figure S69. ¹³C NMR spectrum of compound **15**, related to **Figure 1**.

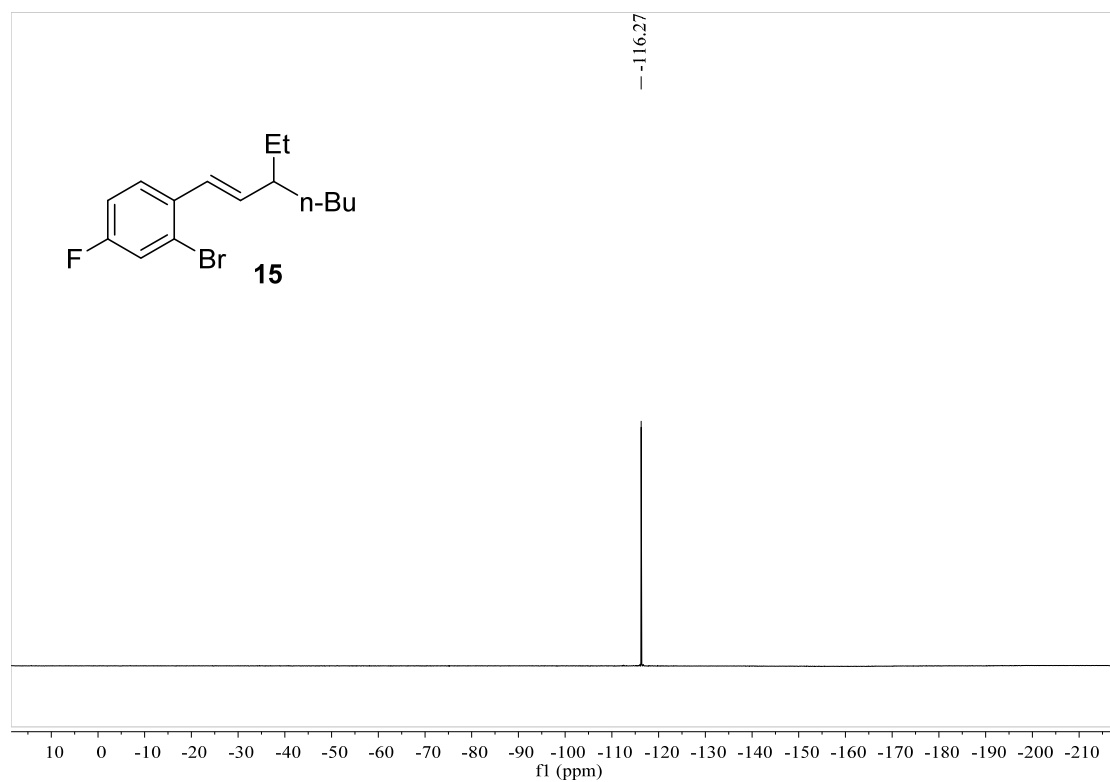
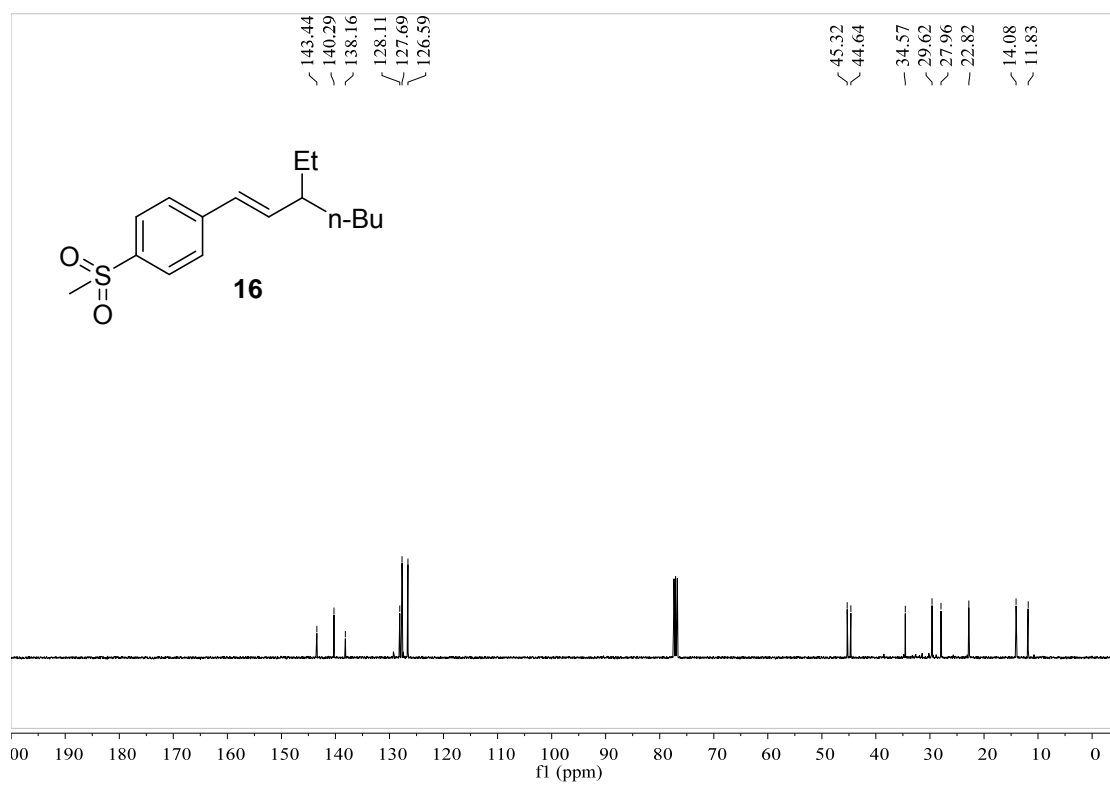
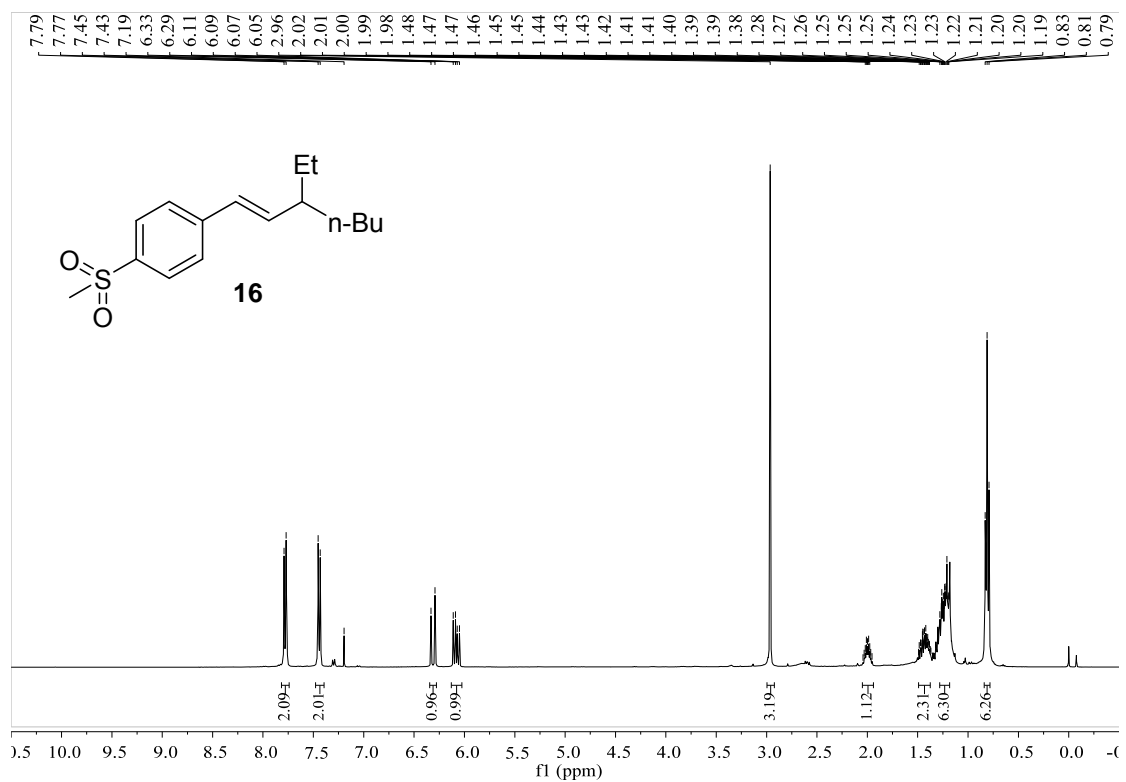


Figure S70. ^{19}F NMR spectrum of compound **15**, related to **Figure 1**.



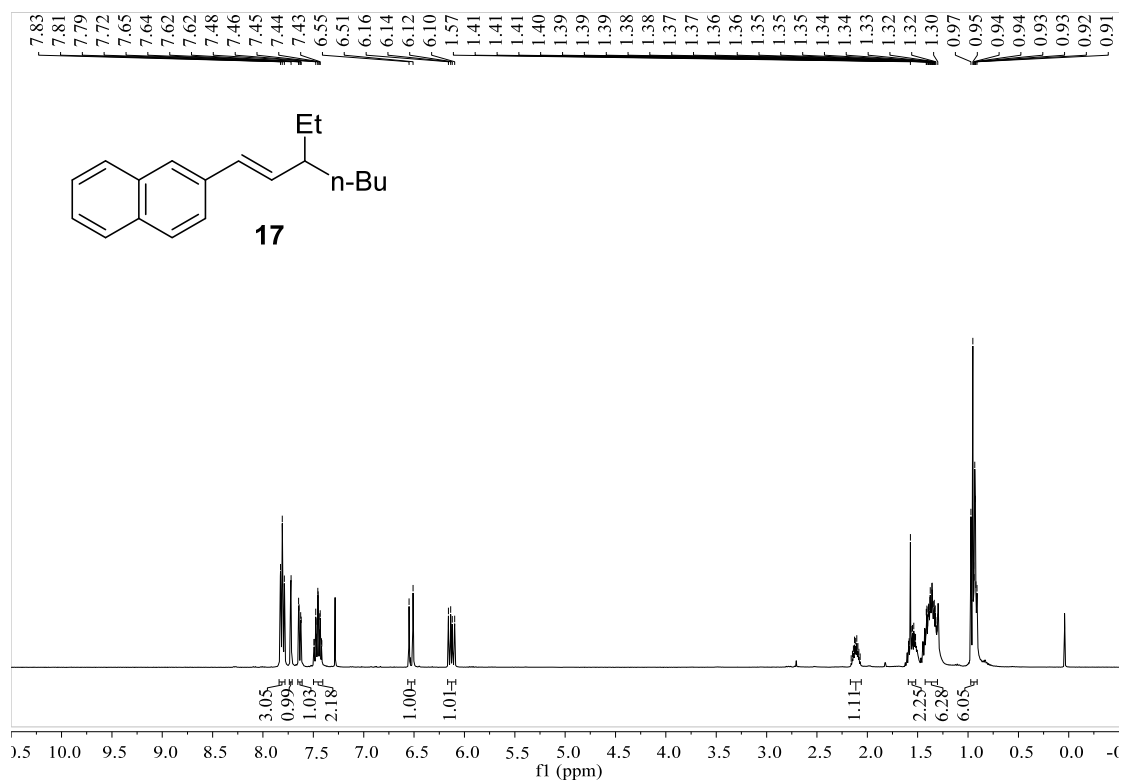


Figure S73. ¹H NMR spectrum of compound 17, related to Figure 1.

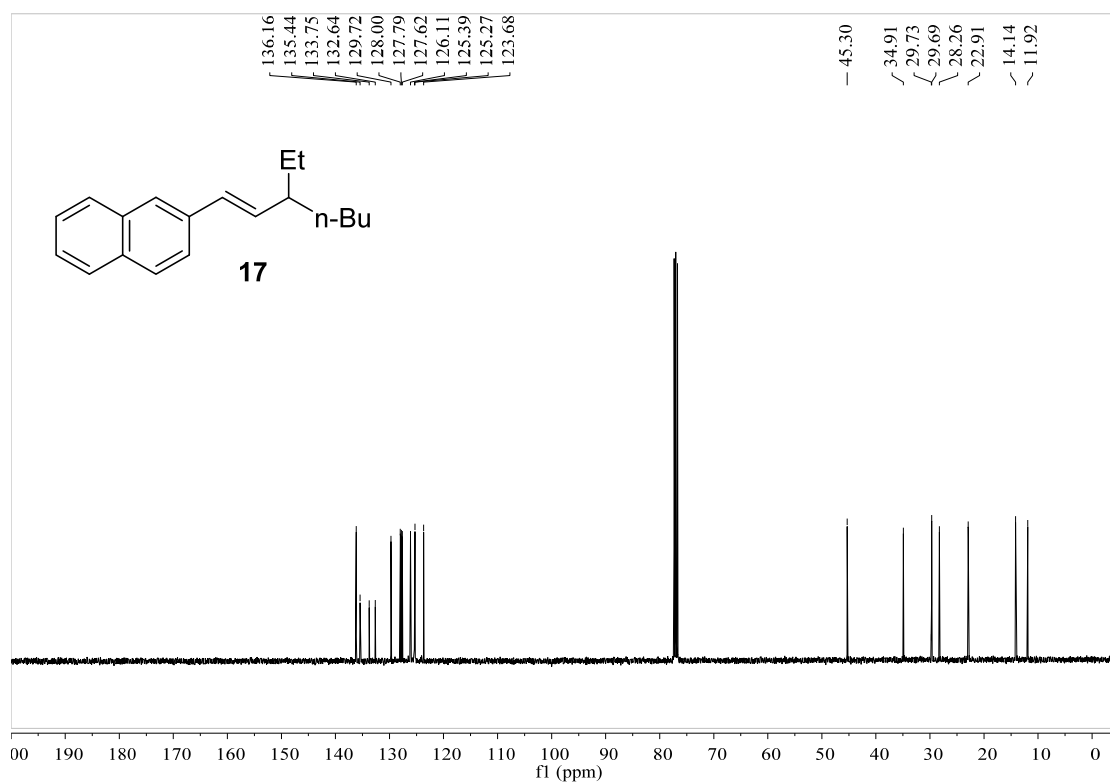


Figure S74. ¹³C NMR spectrum of compound 17, related to Figure 1.

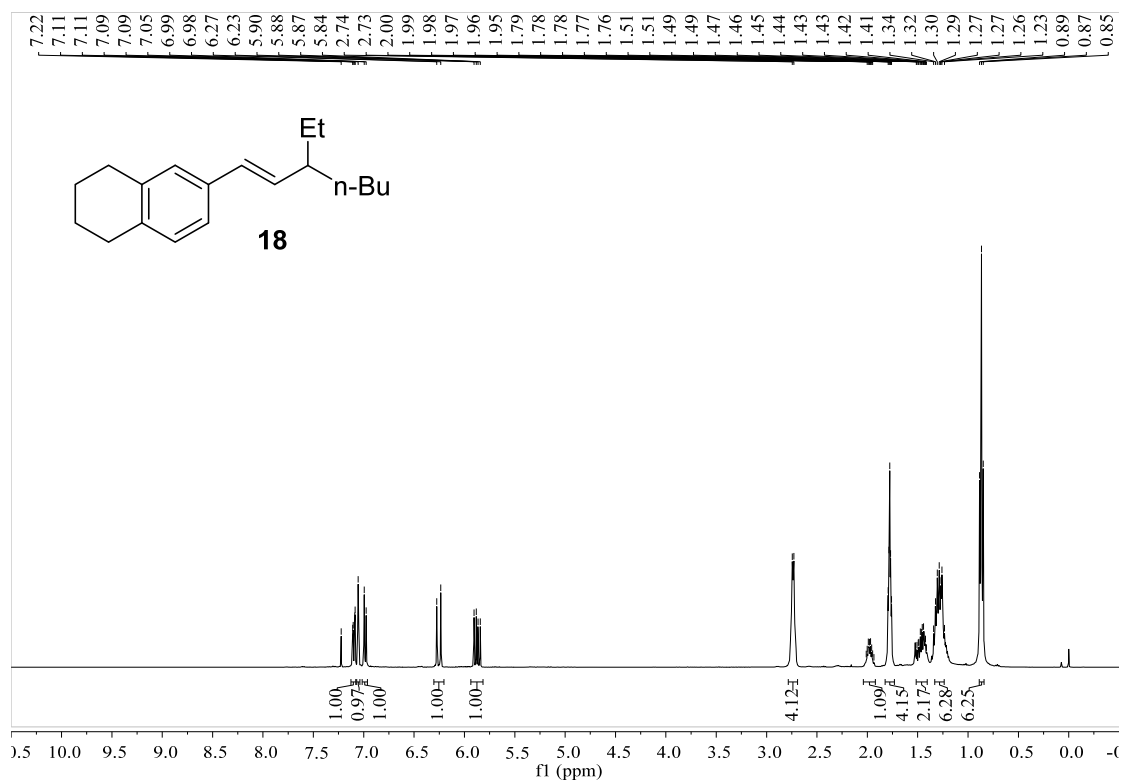


Figure S75. ¹H NMR spectrum of compound **18**, related to Figure 1.

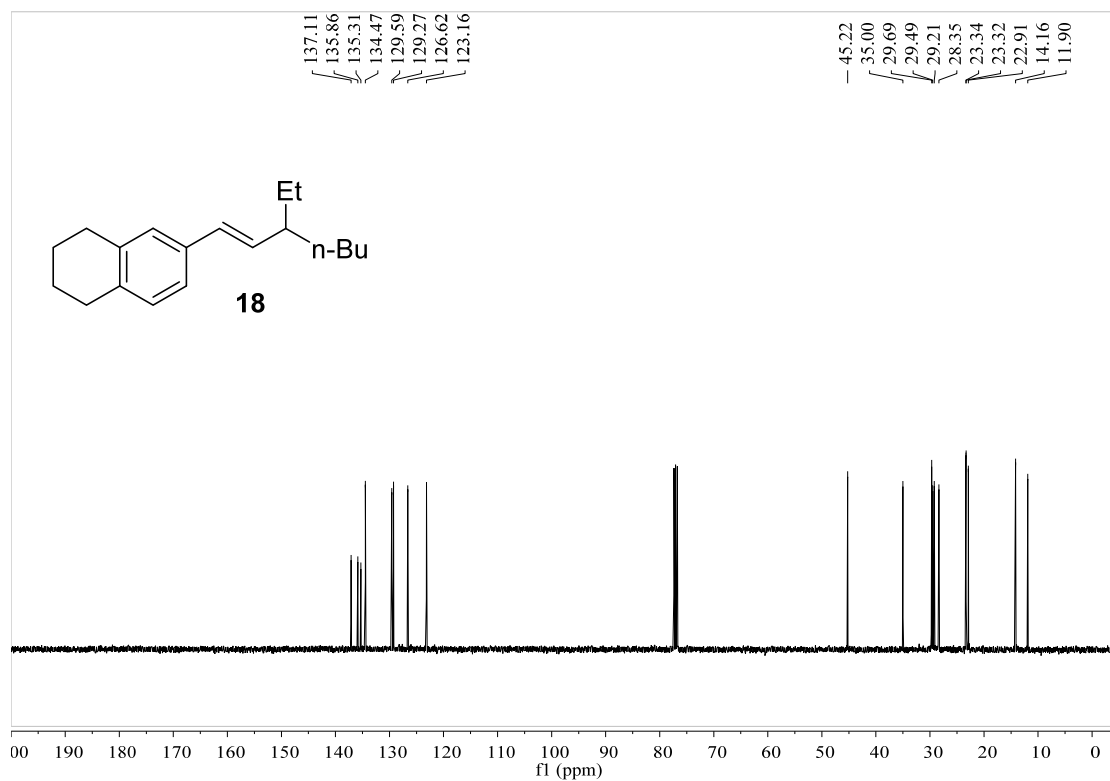


Figure S76. ¹³C NMR spectrum of compound **18**, related to Figure 1.

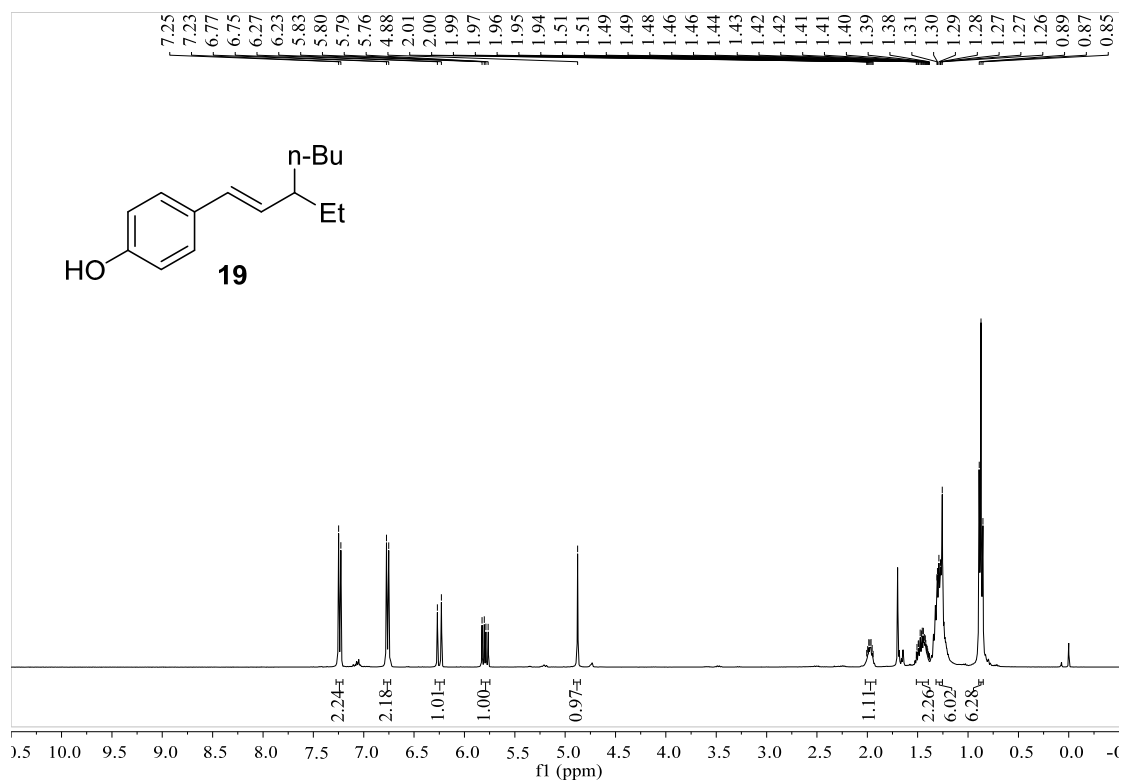


Figure S77. ¹H NMR spectrum of compound **19**, related to **Figure 1**.

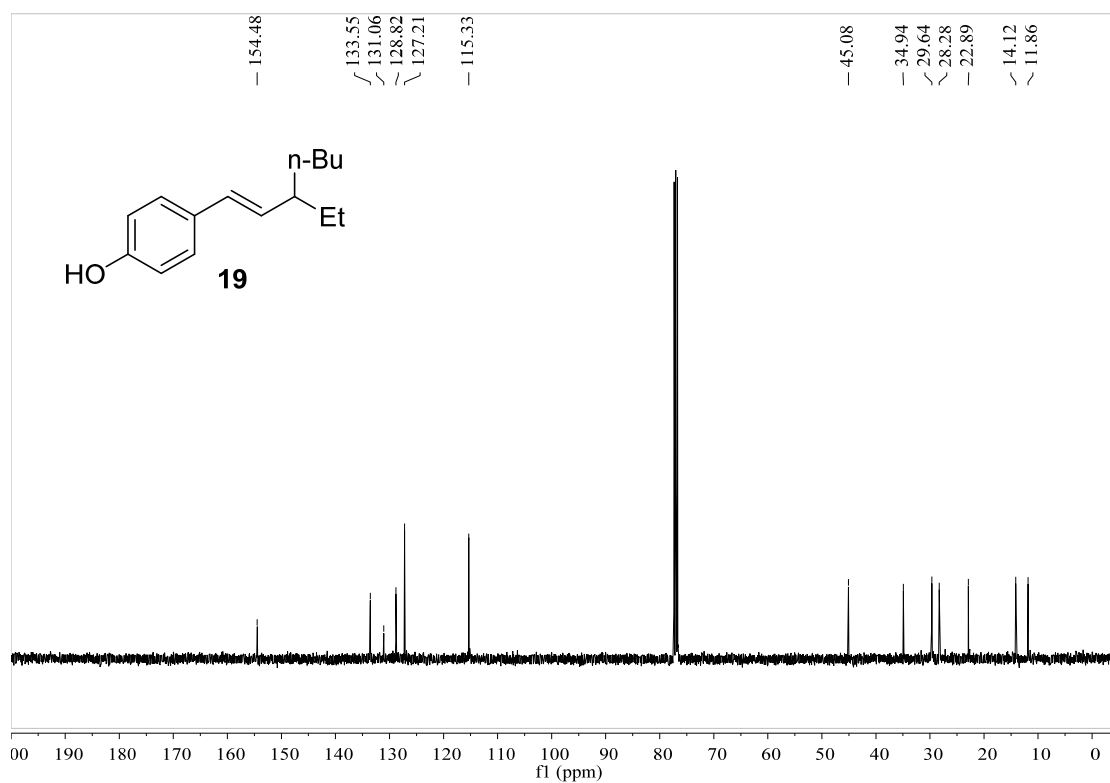


Figure S78. ¹³C NMR spectrum of compound **19**, related to **Figure 1**.

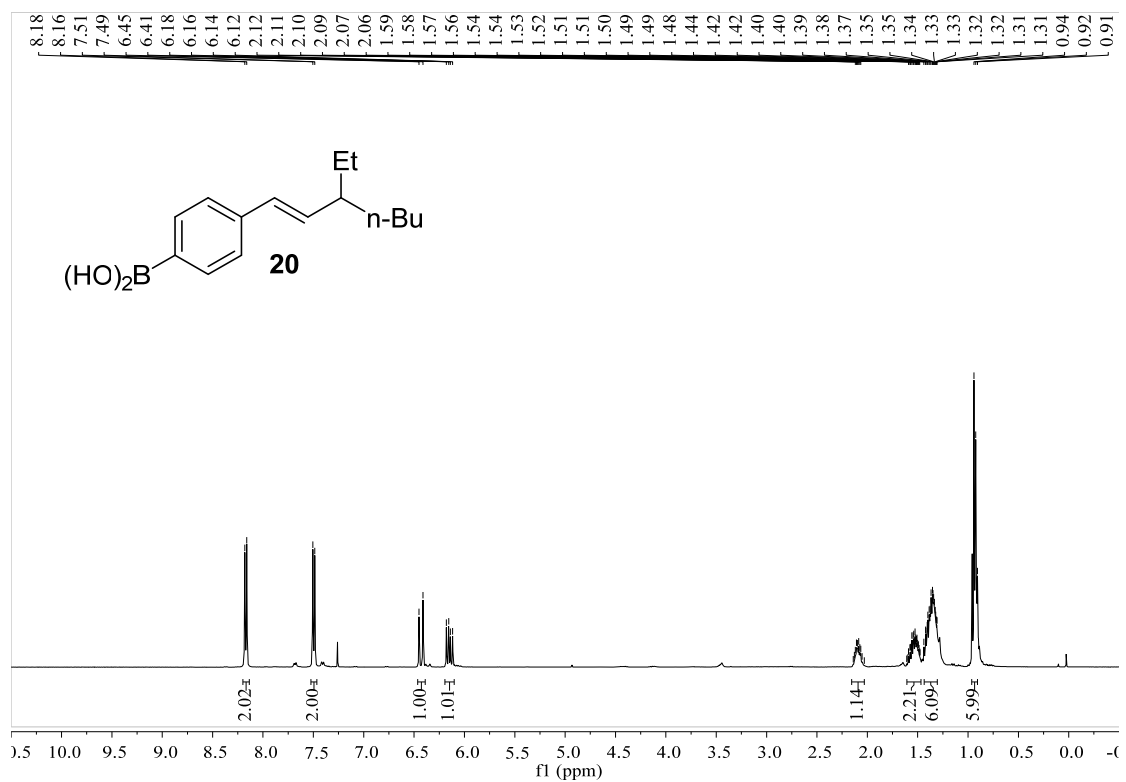


Figure S79. ¹H NMR spectrum of compound **20**, related to **Figure 1**.

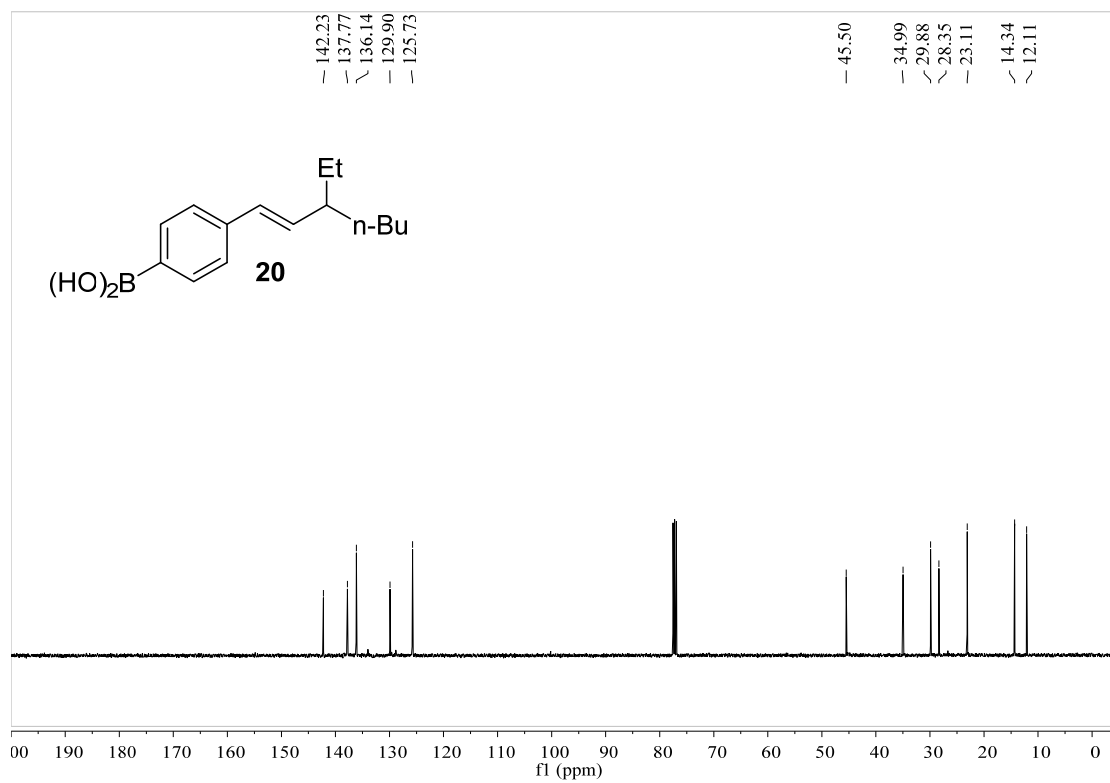


Figure S80. ¹³C NMR spectrum of compound **20**, related to **Figure 1**.

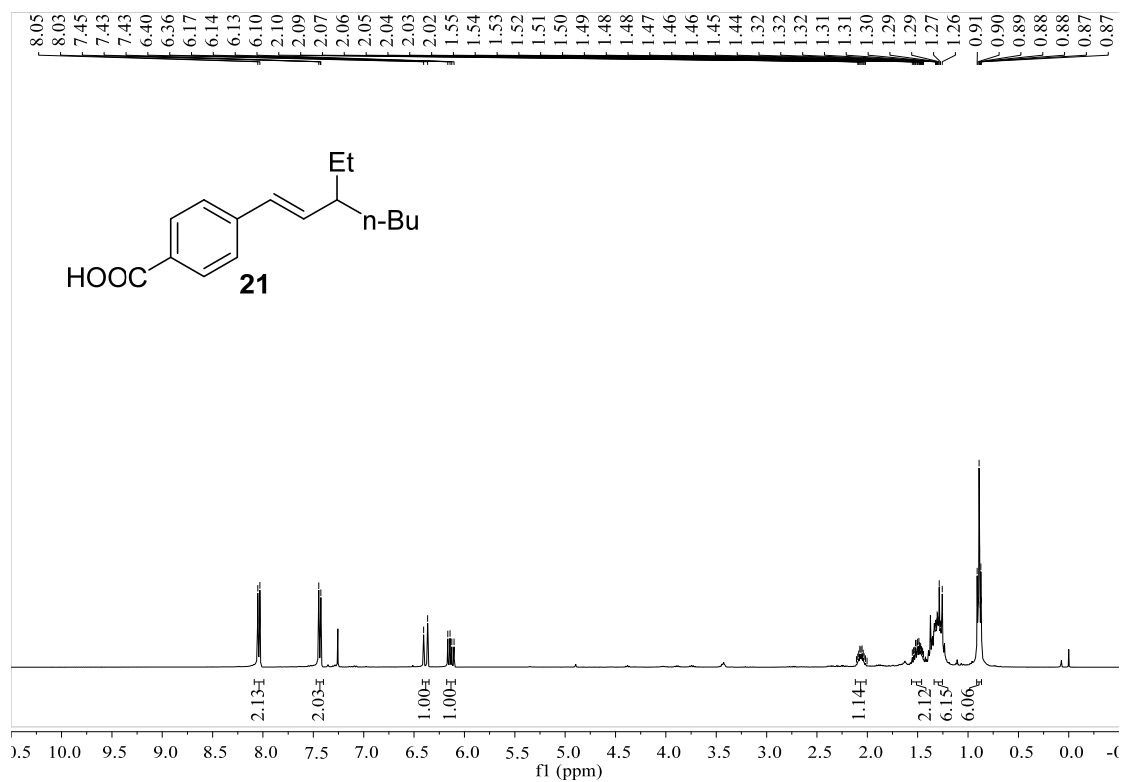


Figure S81. ¹H NMR spectrum of compound **21**, related to **Figure 1**.

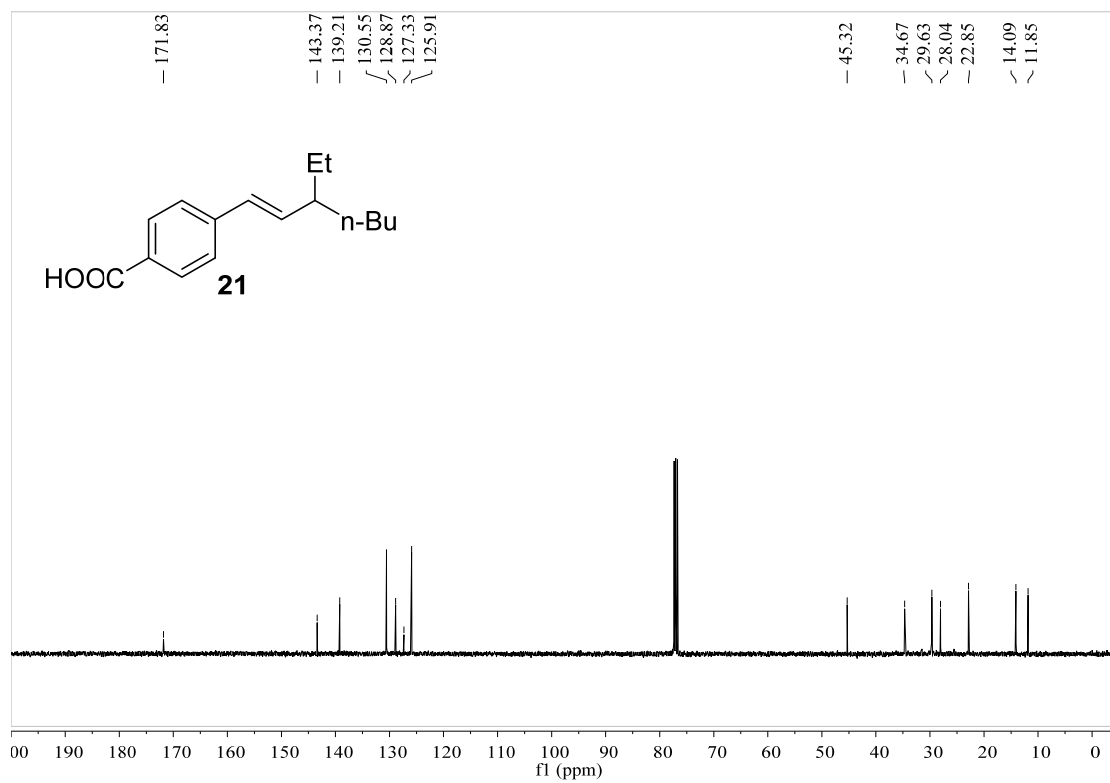


Figure S82. ¹³C NMR spectrum of compound **21**, related to **Figure 1**.

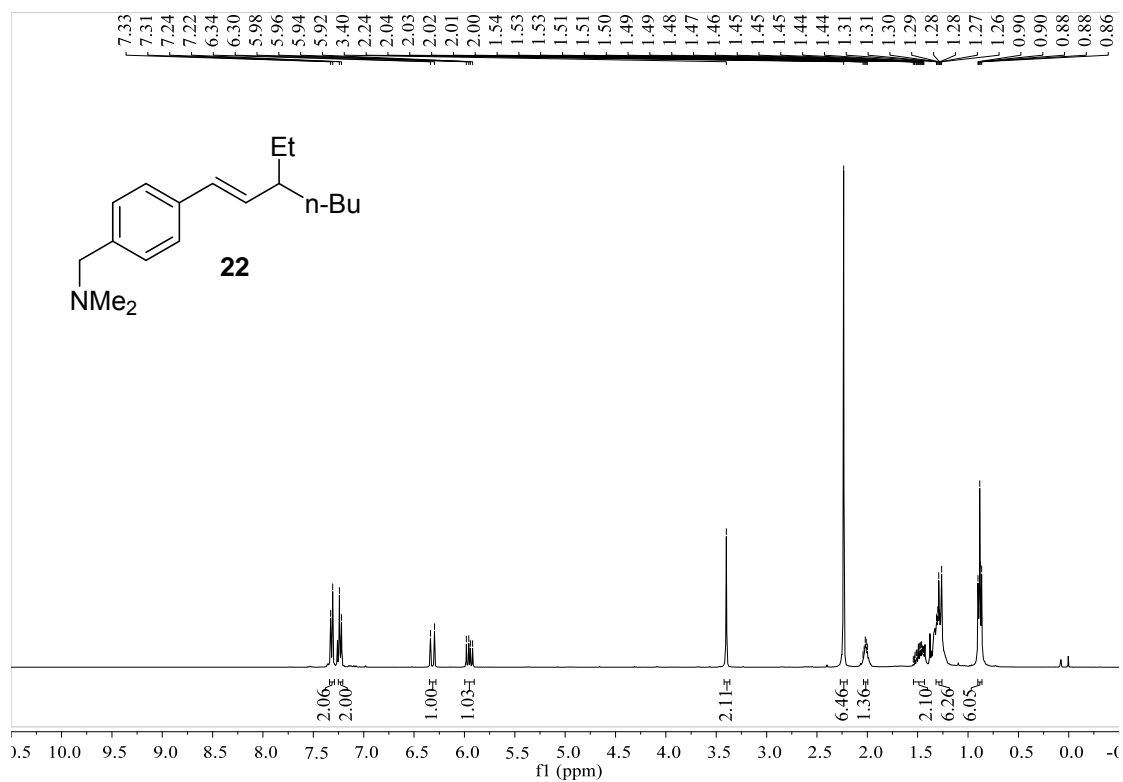


Figure S83. ¹H NMR spectrum of compound **22**, related to **Figure 1**.

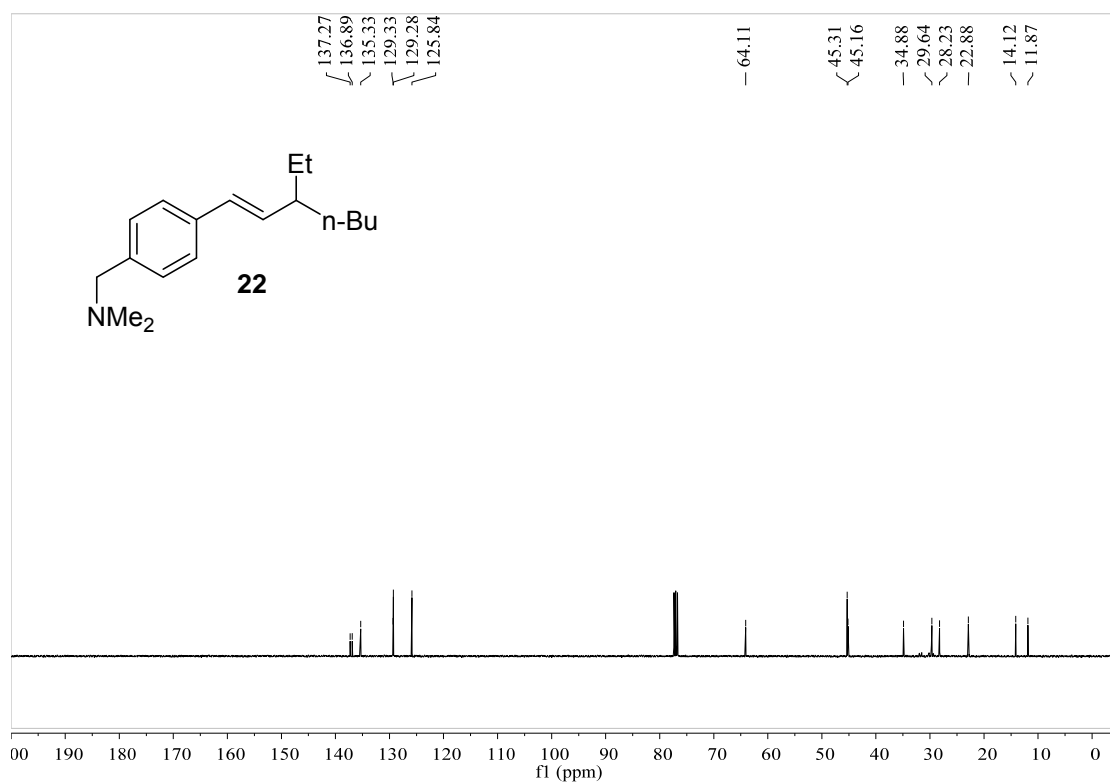


Figure S84. ¹³C NMR spectrum of compound **22**, related to **Figure 1**.

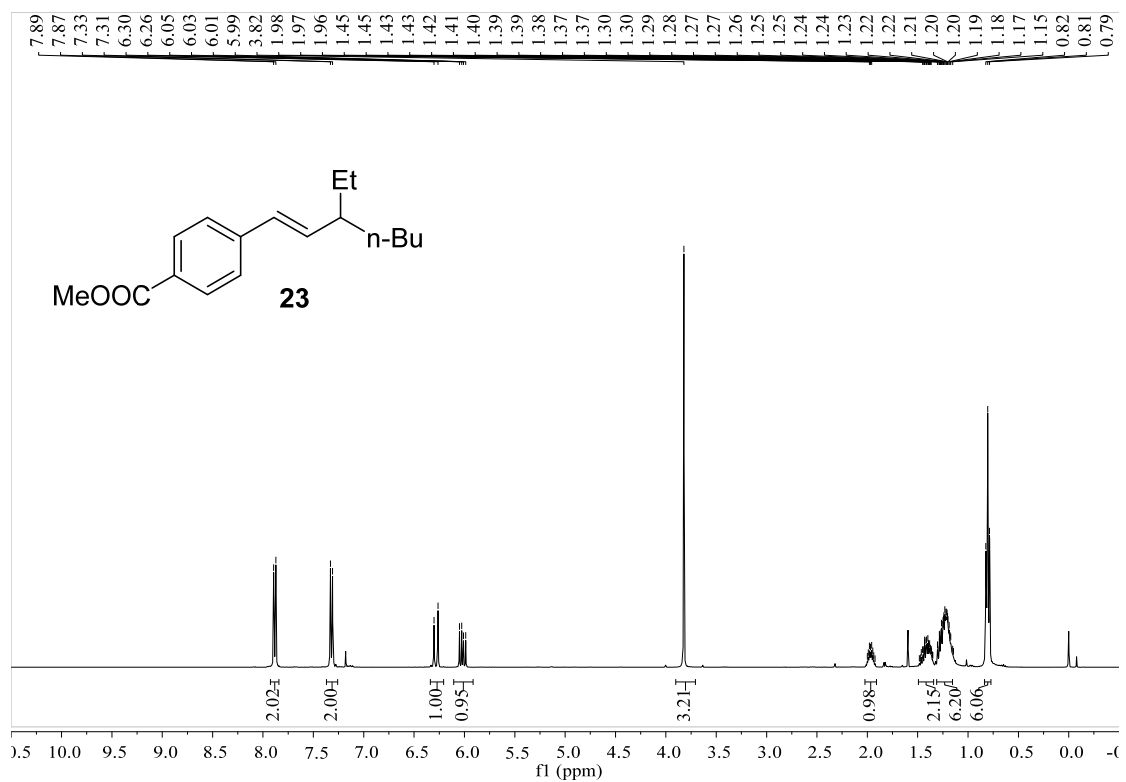


Figure S85. ¹H NMR spectrum of compound **23**, related to Figure 1.

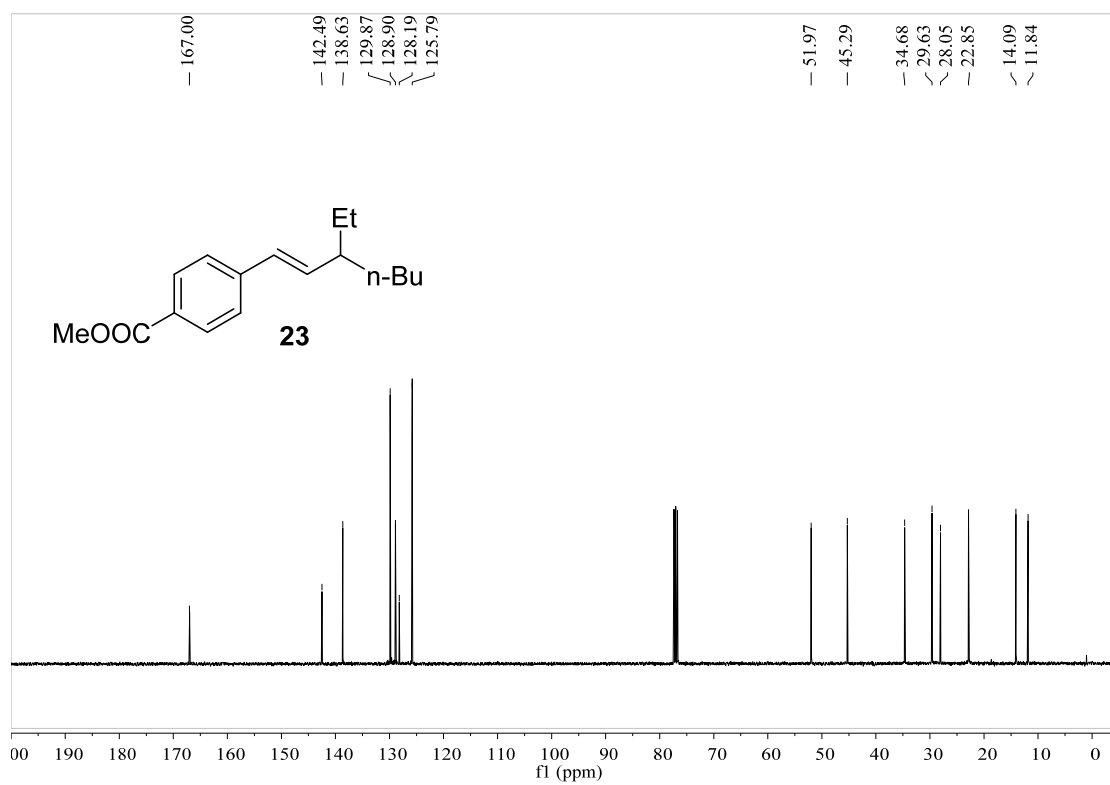


Figure S86. ¹³C NMR spectrum of compound **23**, related to Figure 1.

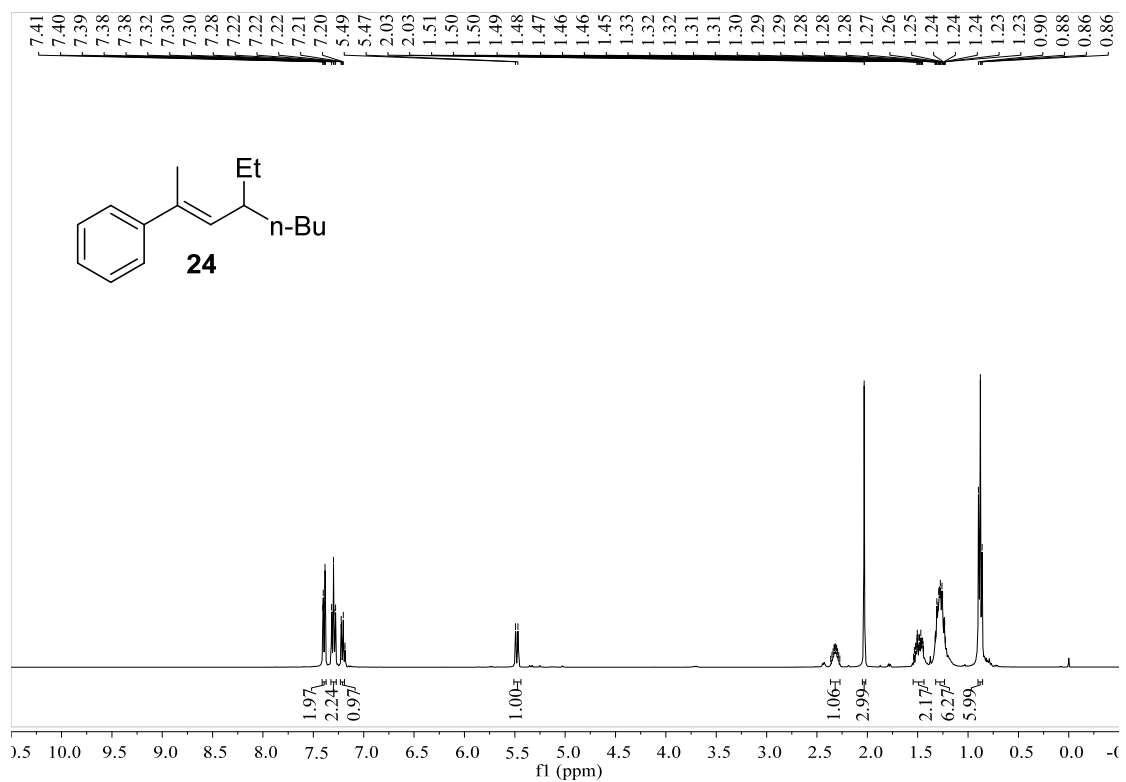


Figure S87. ¹H NMR spectrum of compound **24**, related to **Figure 1**.

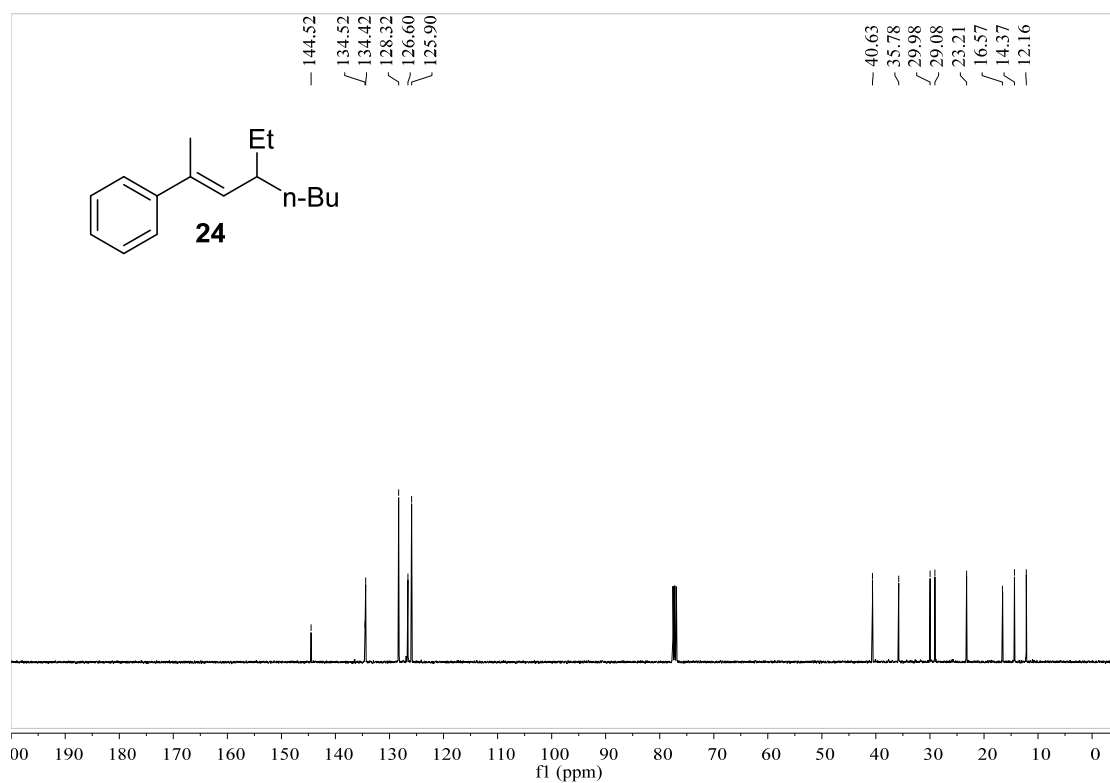


Figure S88. ¹³C NMR spectrum of compound **24**, related to **Figure 1**.

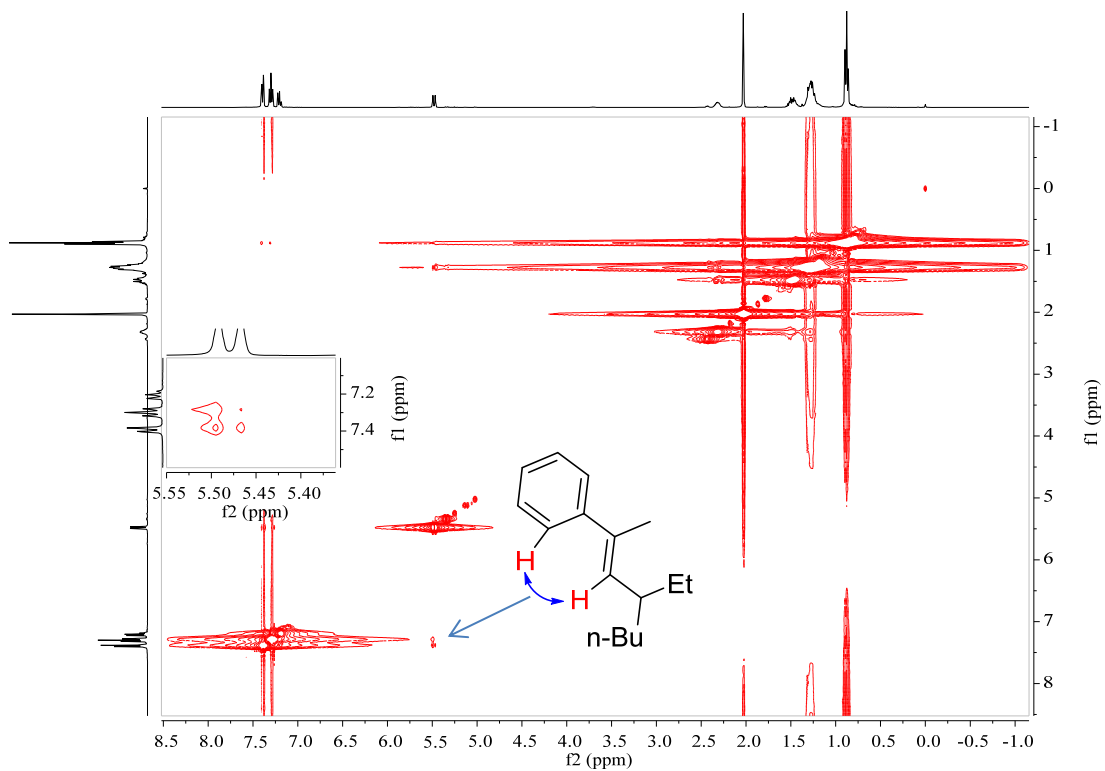


Figure S89. NOE spectrum of compound **24**, related to **Figure 1**.

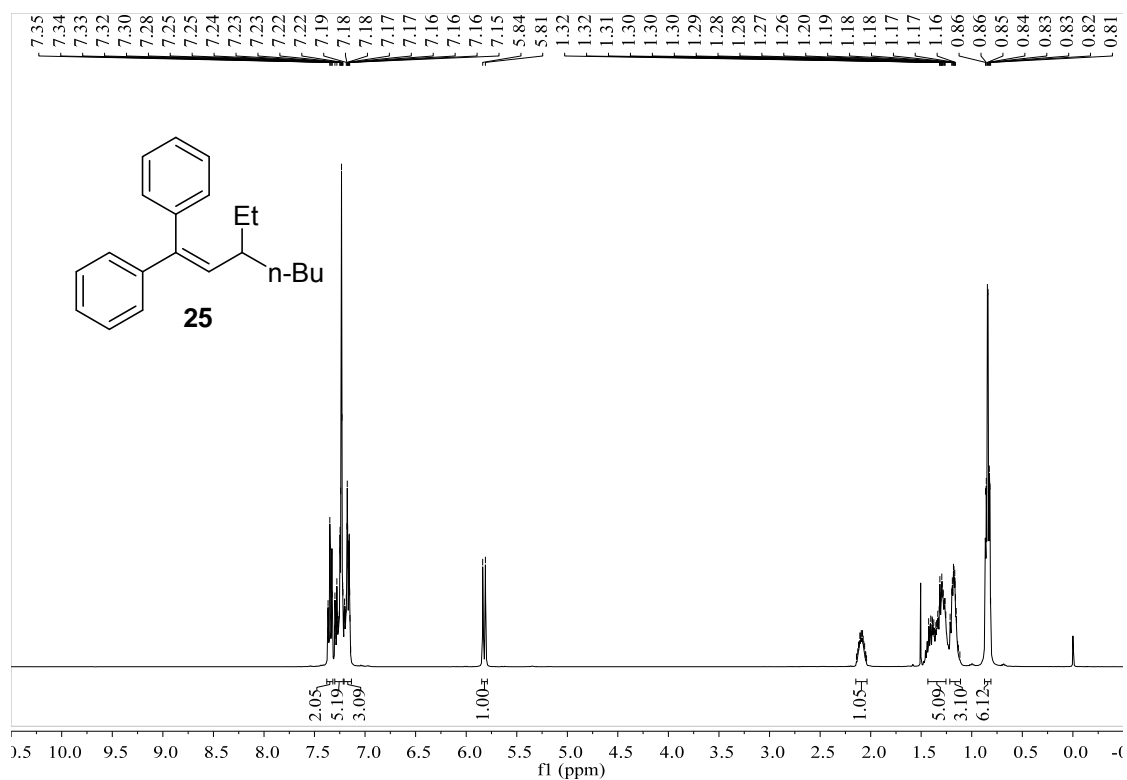


Figure S90. ¹H NMR spectrum of compound **25**, related to **Figure 1**.

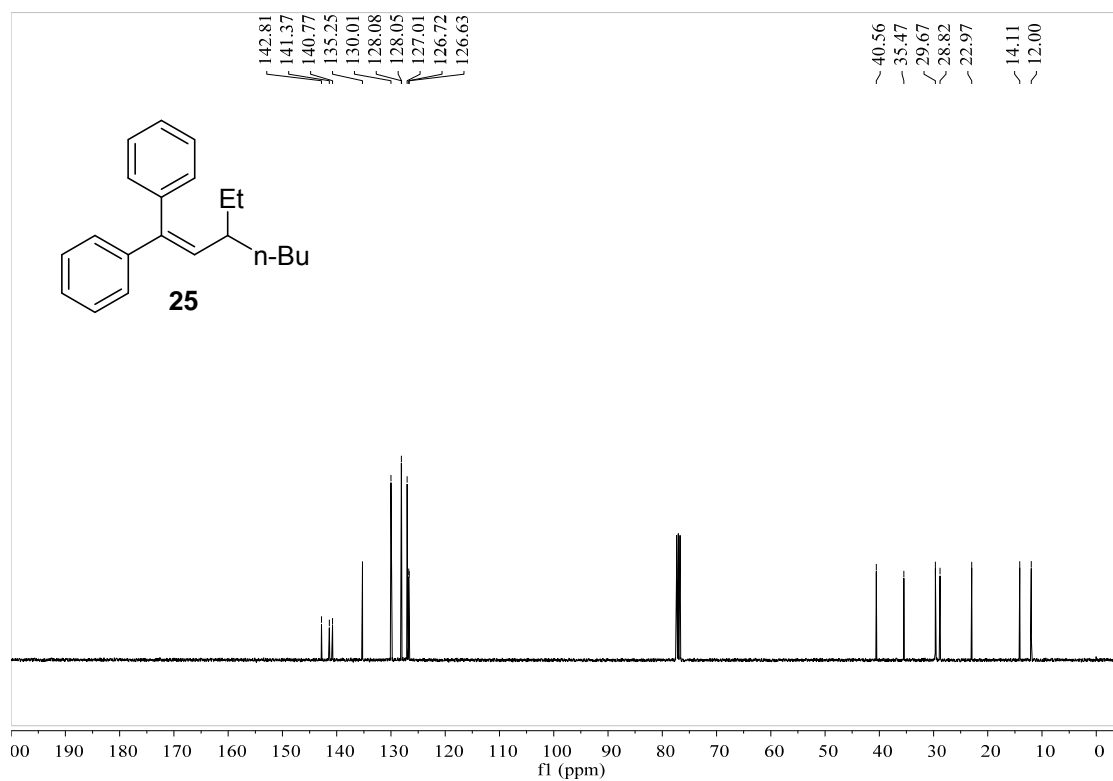


Figure S91. ¹³C NMR spectrum of compound **25**, related to **Figure 1**.

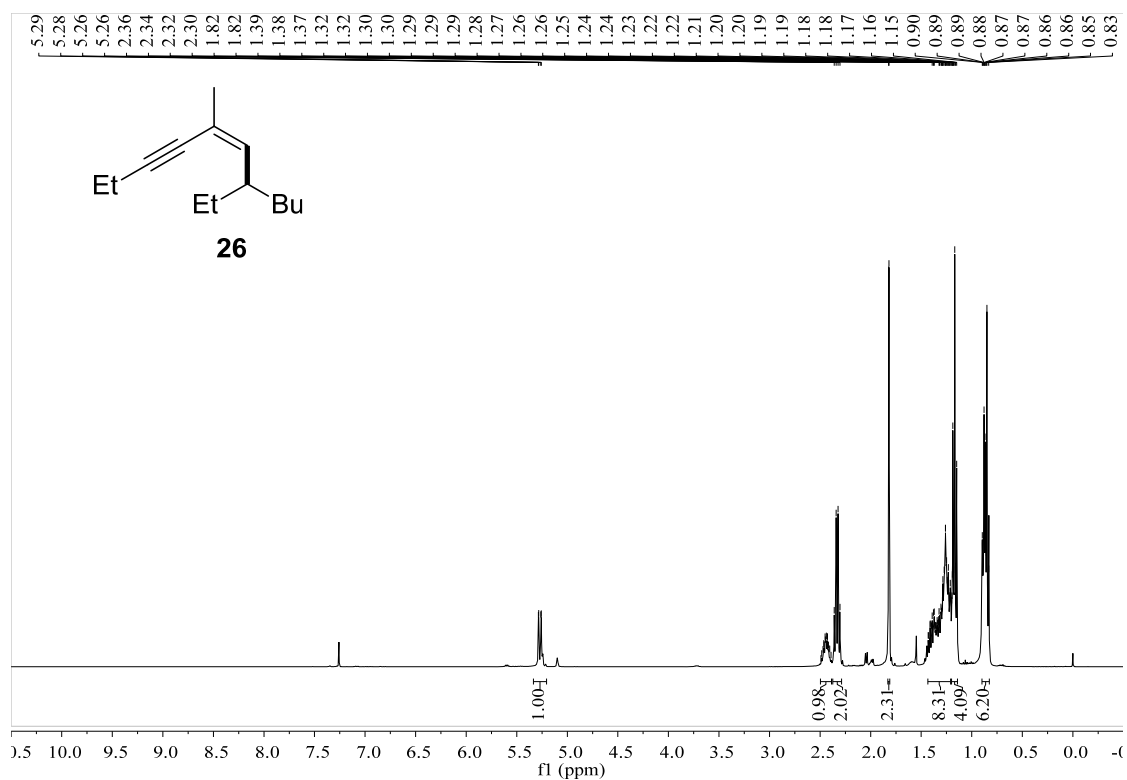


Figure S92. ¹H NMR spectrum of compound **26**, related to **Figure 1**.

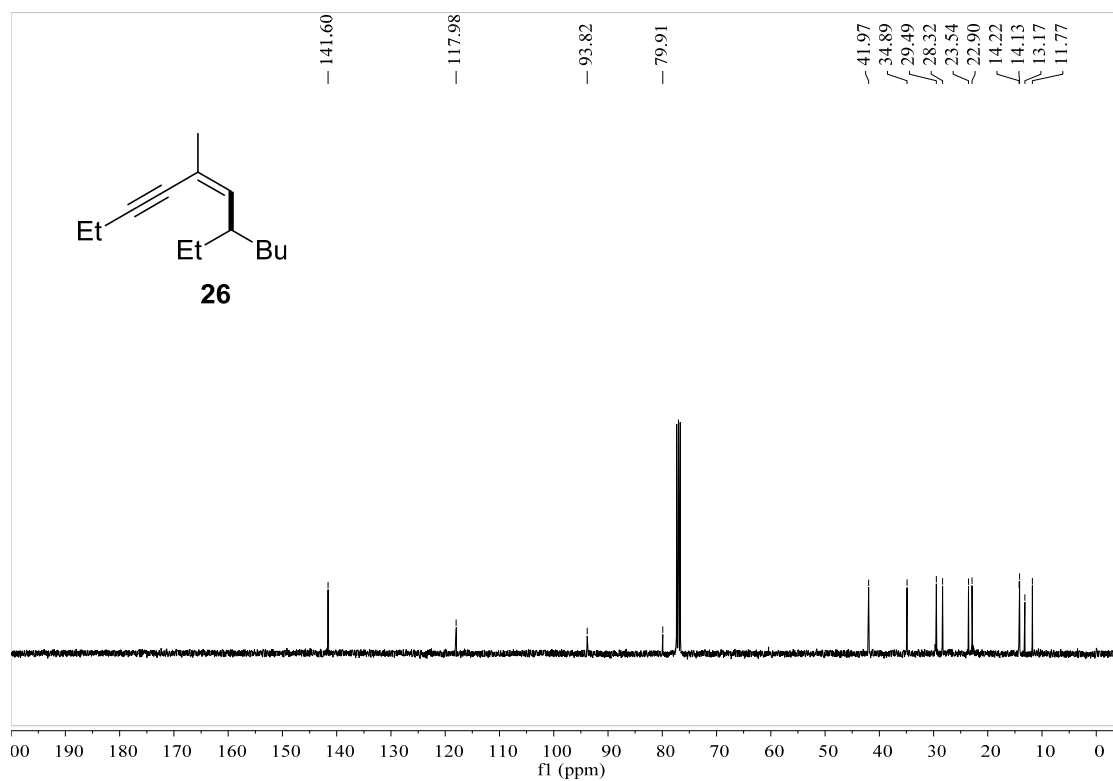


Figure S93. ¹³C NMR spectrum of compound **26**, related to **Figure 1**.

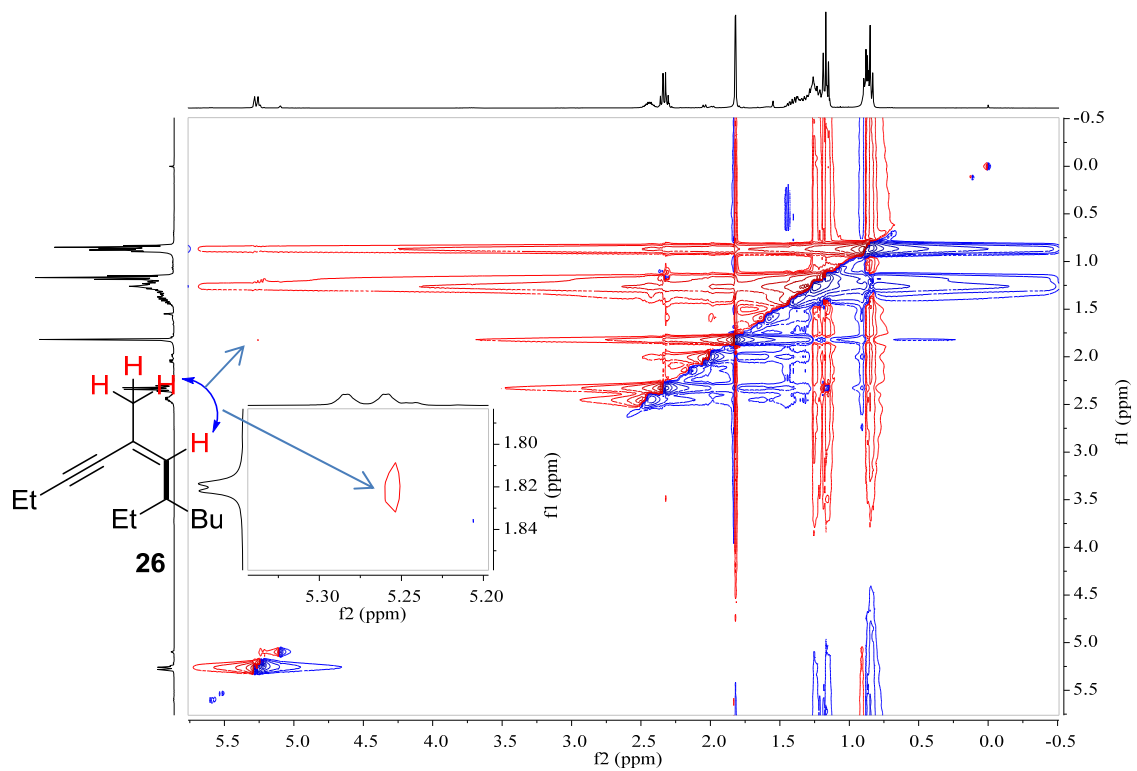


Figure S94. NOE spectrum of compound 26, related to Figure 1.

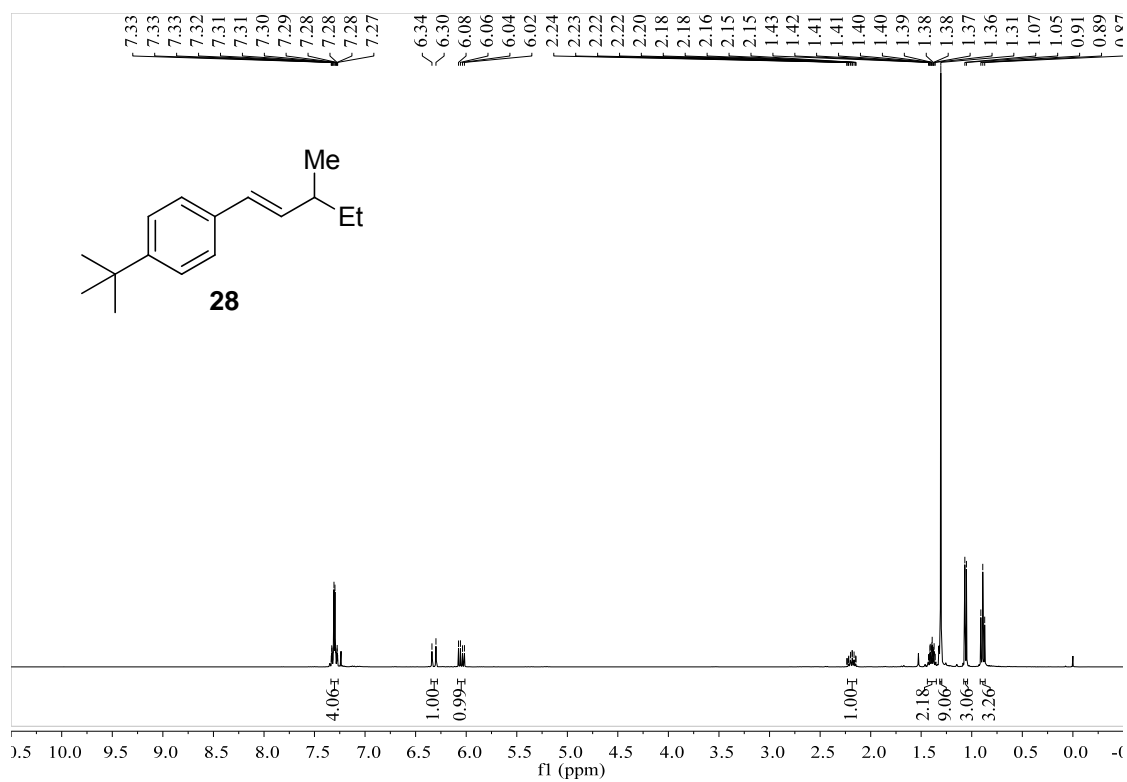


Figure S95. ¹H NMR spectrum of compound **28**, related to Figure 2.

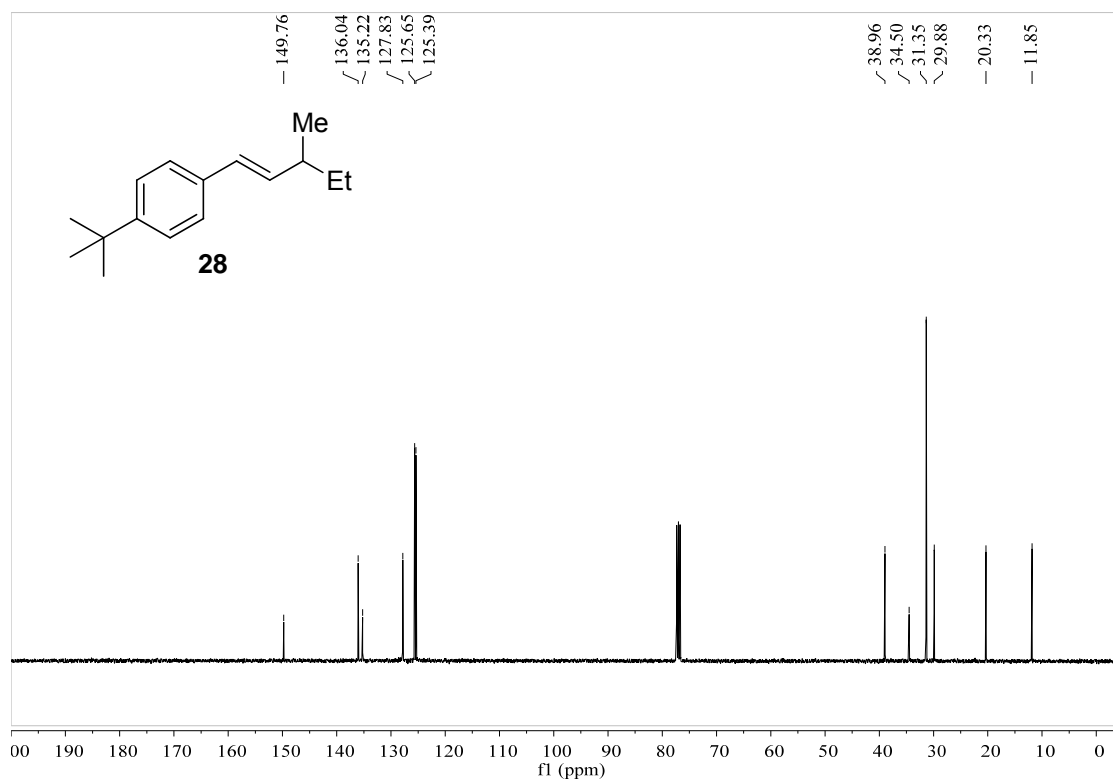


Figure S96. ¹³C NMR spectrum of compound **28**, related to Figure 2.

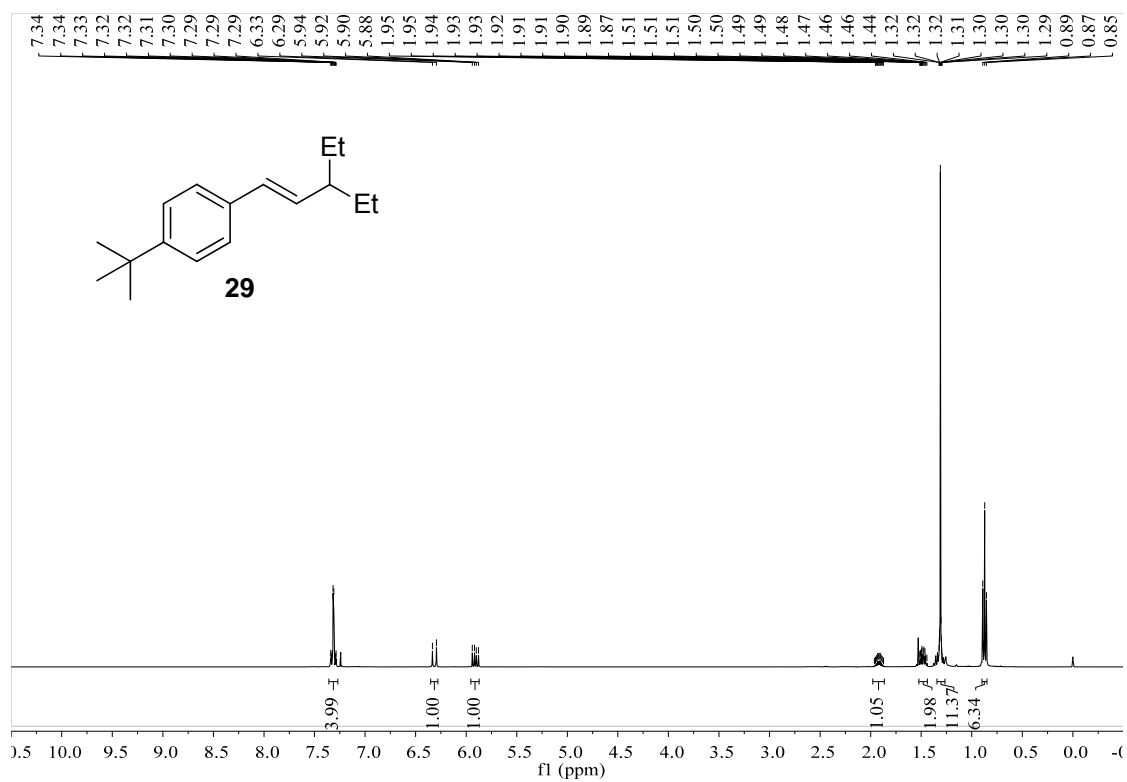


Figure S97. ¹H NMR spectrum of compound **29**, related to Figure 2.

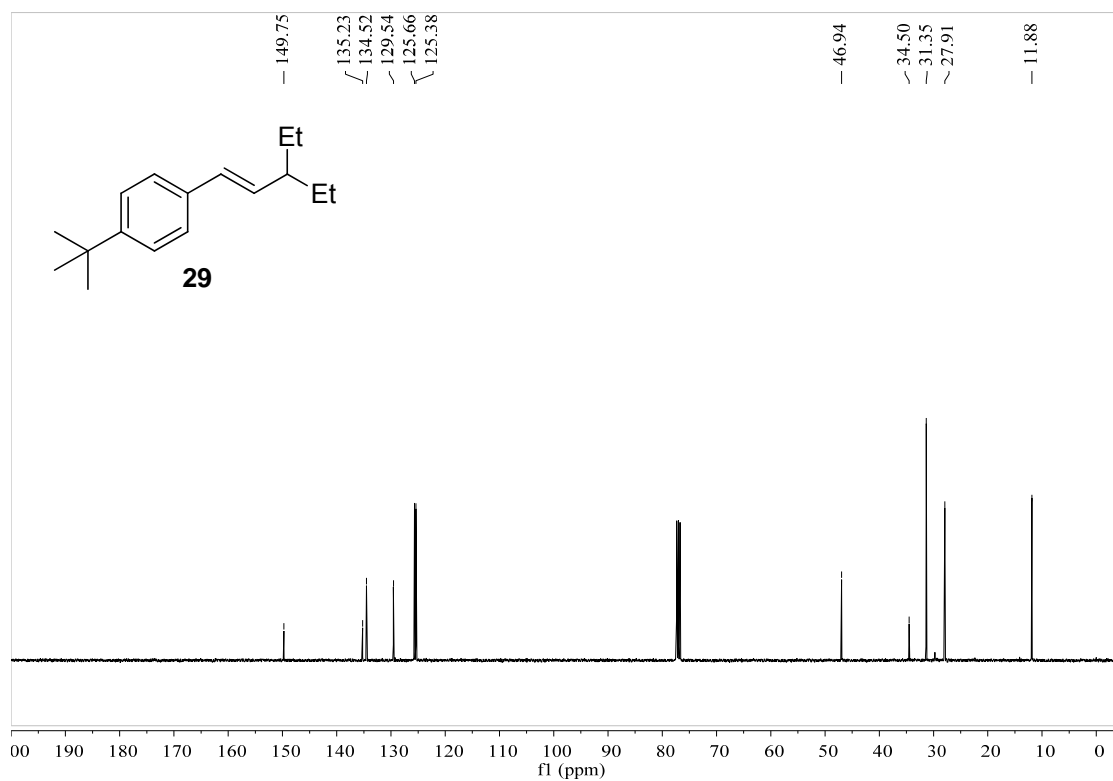


Figure S98. ¹³C NMR spectrum of compound **29**, related to Figure 2.

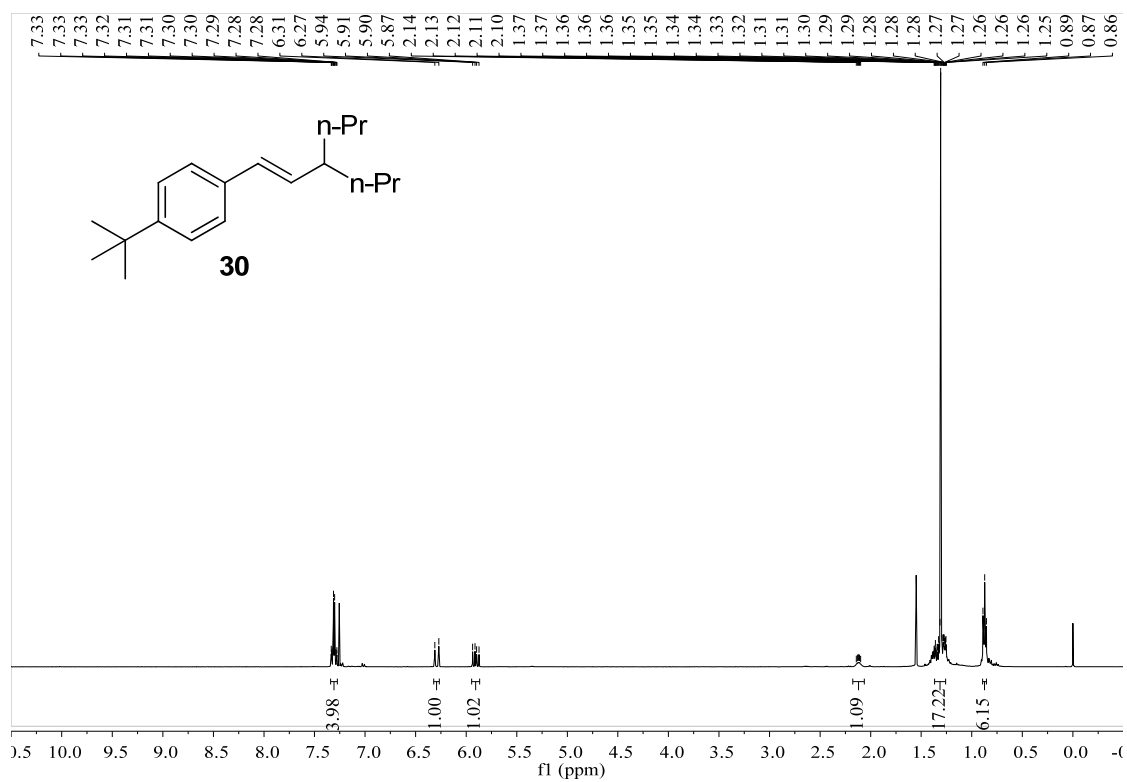


Figure S99. ¹H NMR spectrum of compound **30**, related to **Figure 2**.

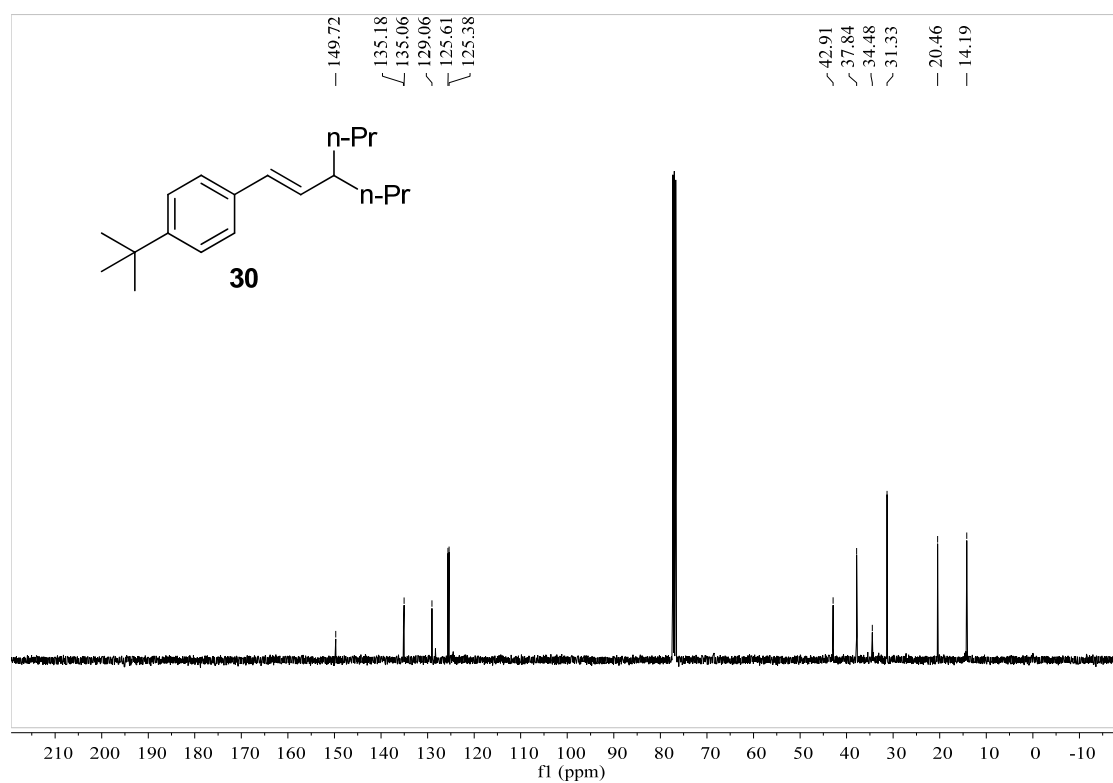


Figure S100. ¹³C NMR spectrum of compound **30**, related to **Figure 2**.

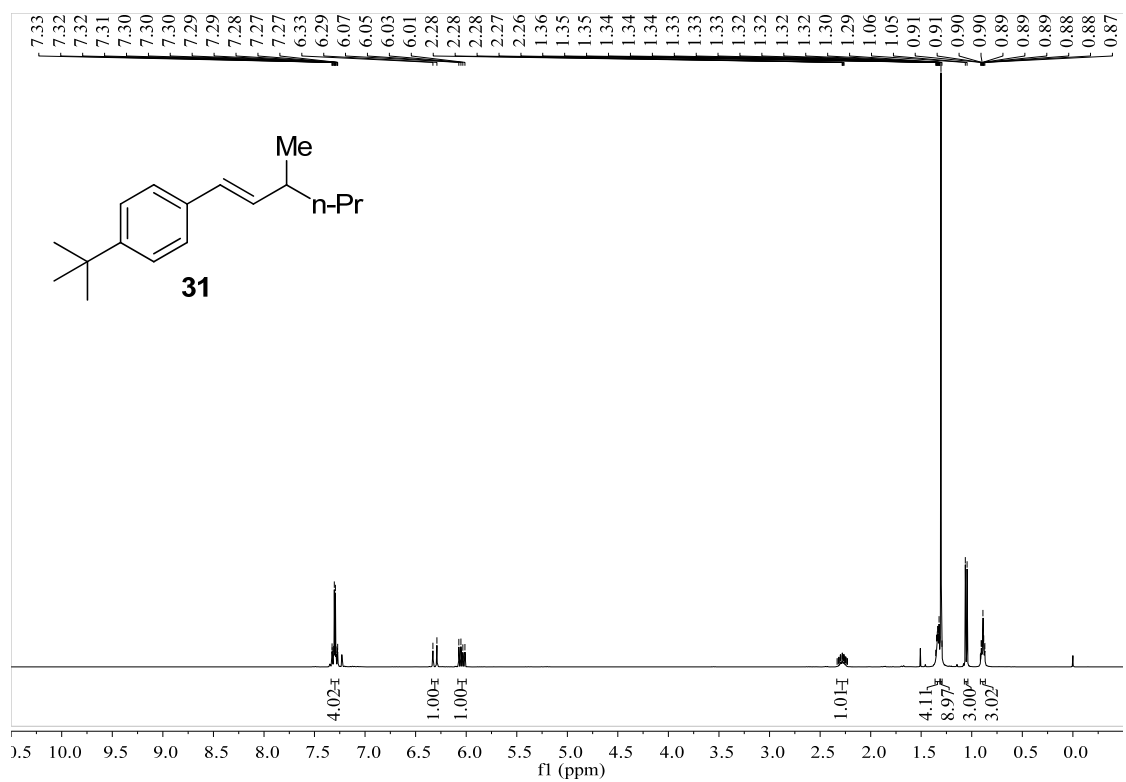


Figure S101. ¹H NMR spectrum of compound **31**, related to Figure 2.

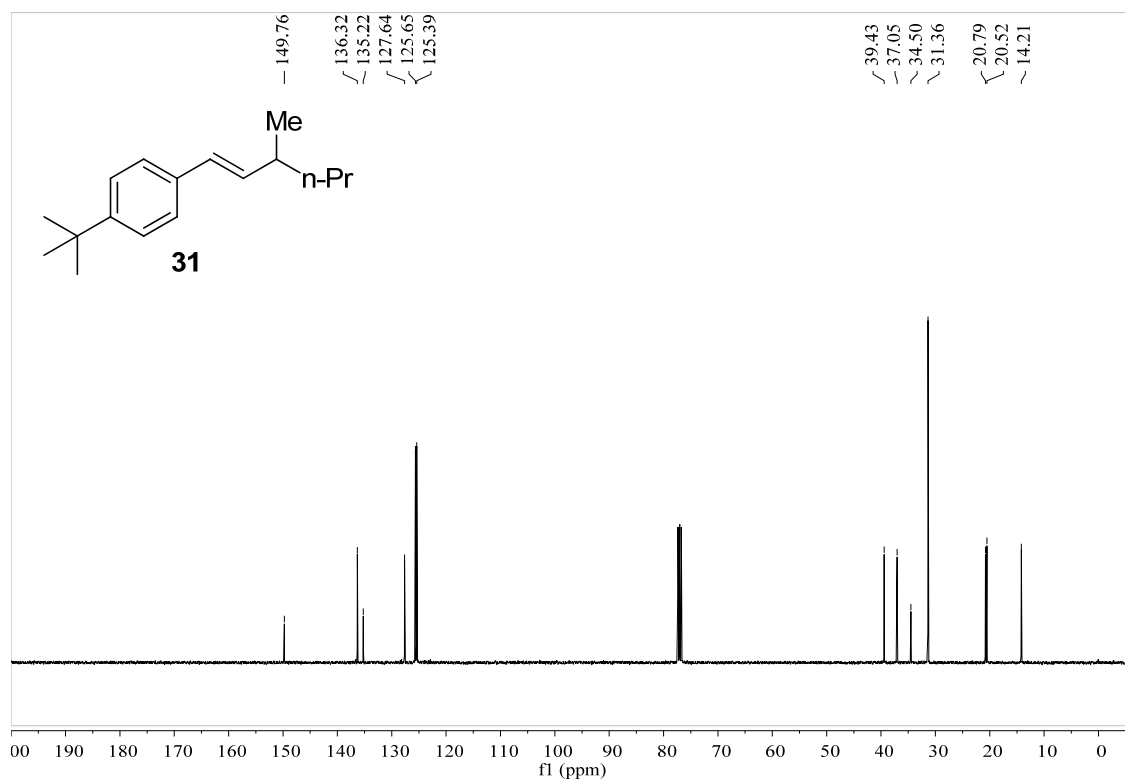


Figure S102. ¹³C NMR spectrum of compound **31**, related to Figure 2.

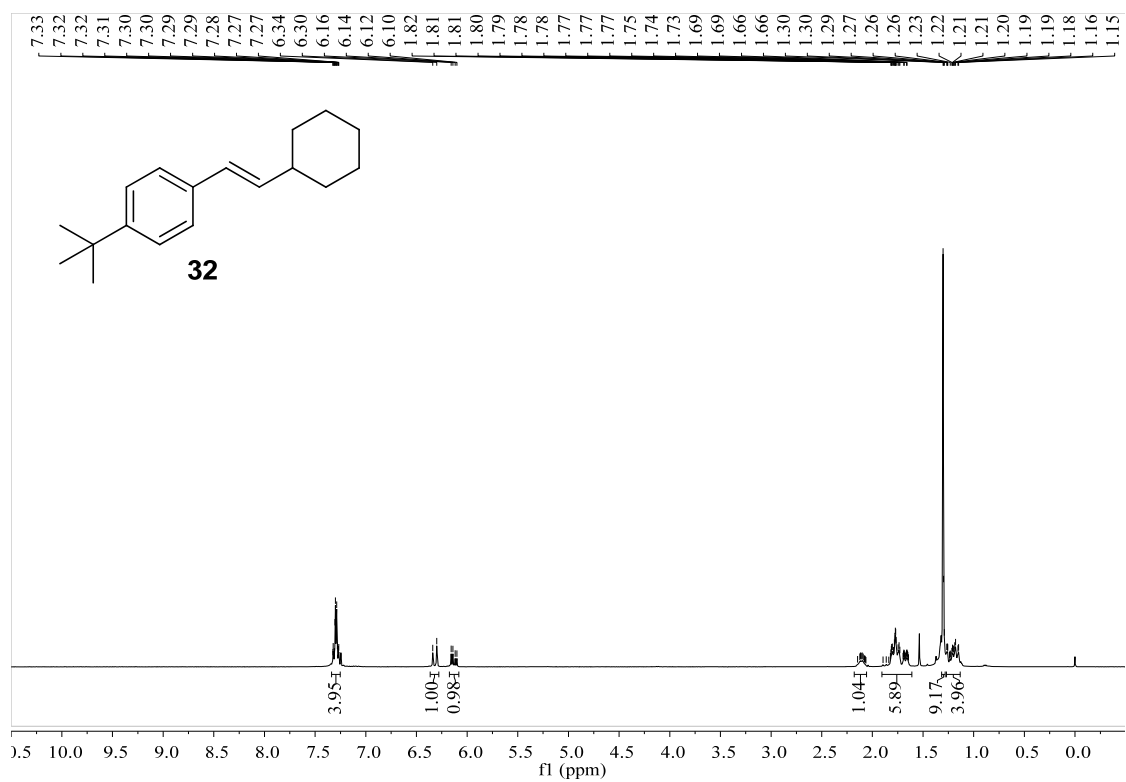


Figure S103. ¹H NMR spectrum of compound **32**, related to **Figure 2**.

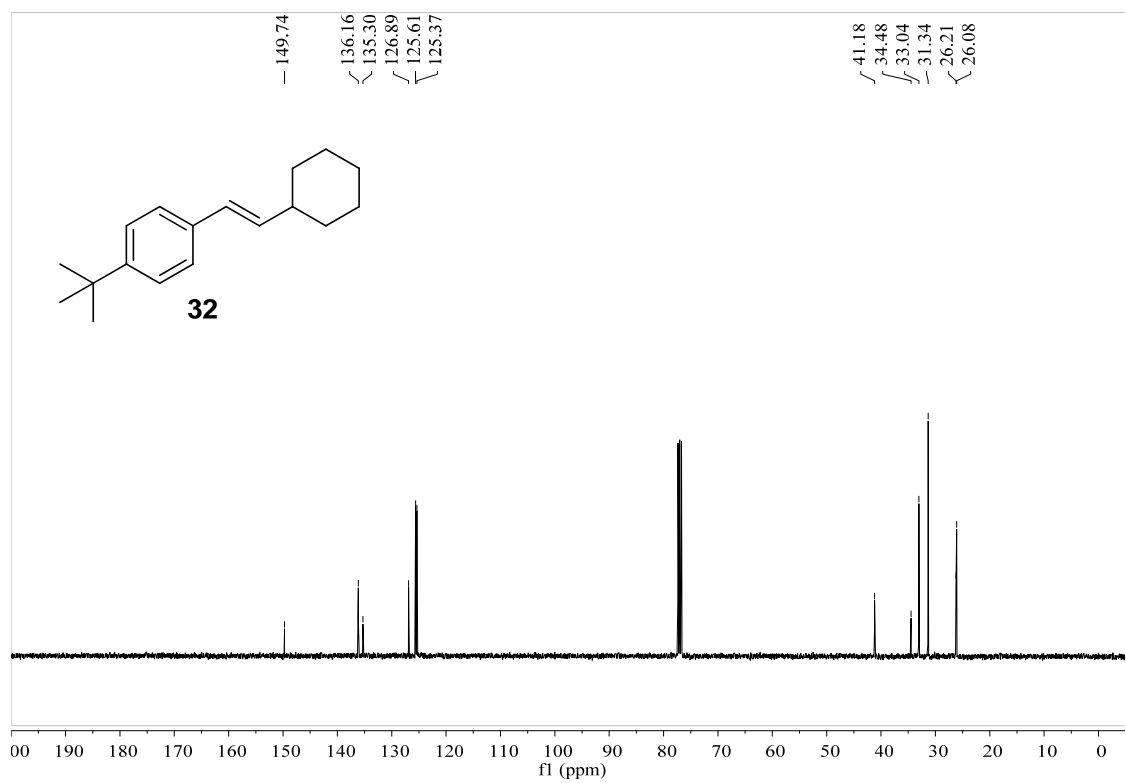


Figure S104. ¹³C NMR spectrum of compound **32**, related to **Figure 2**.

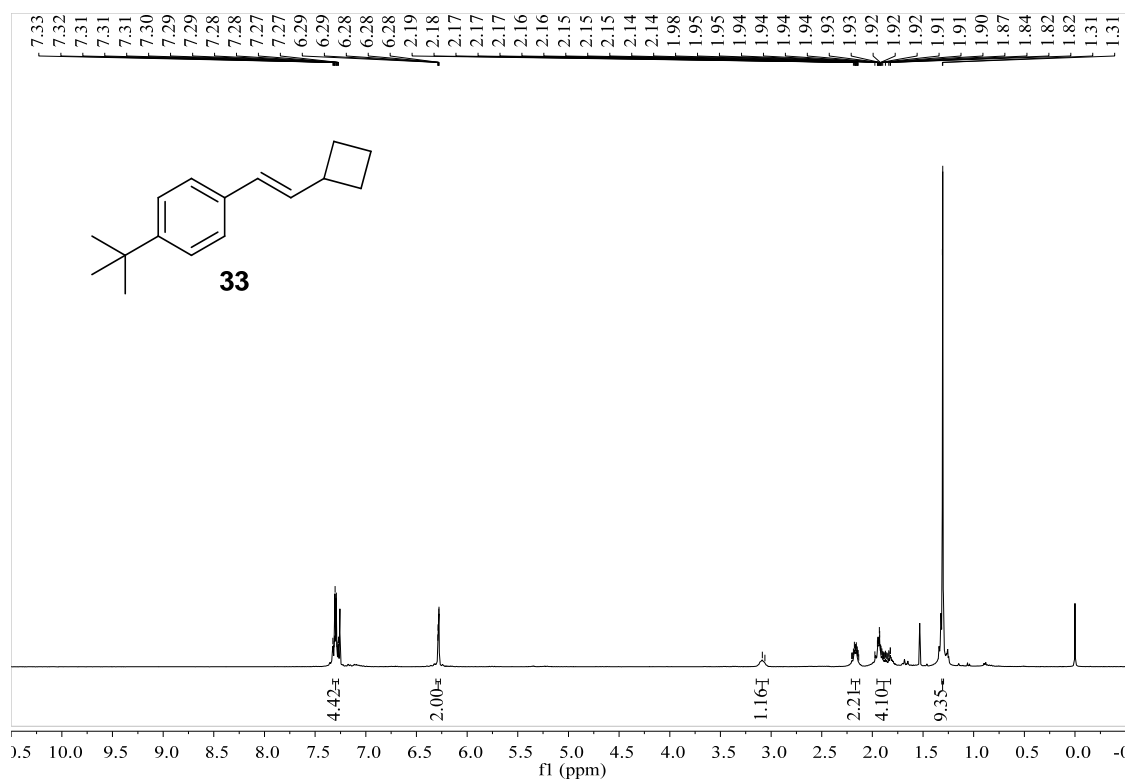


Figure S105. ¹H NMR spectrum of compound **33**, related to **Figure 2**.

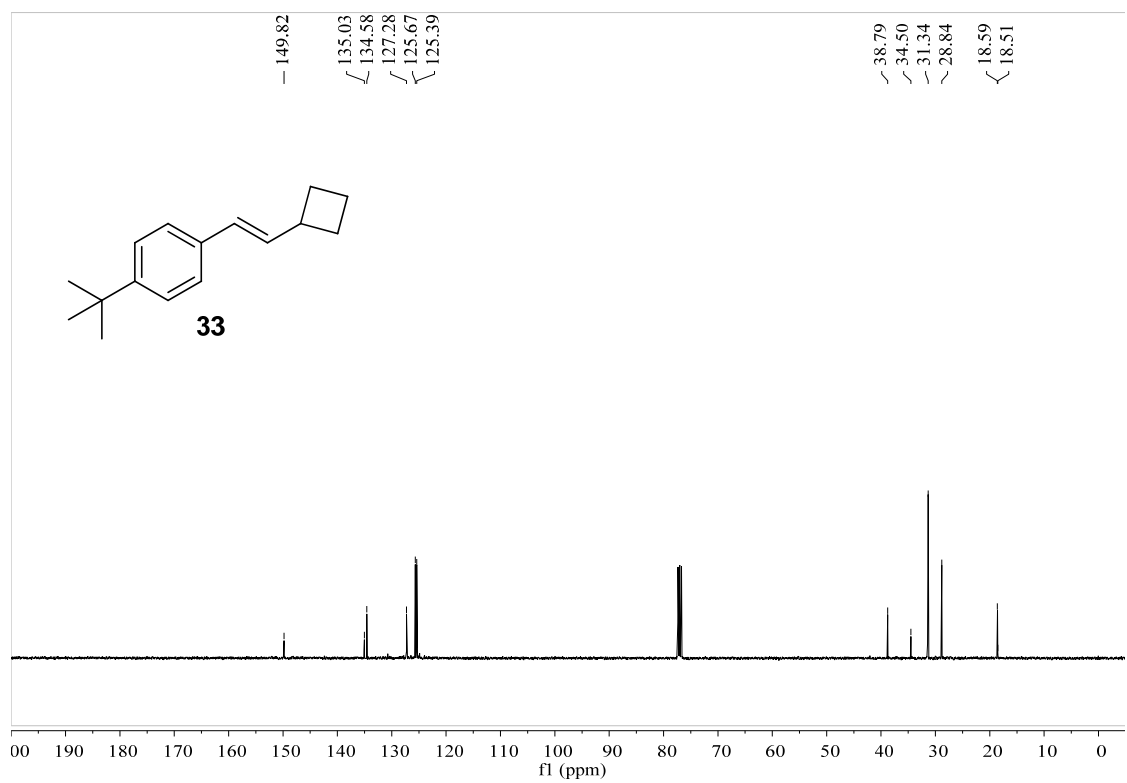


Figure S106. ¹³C NMR spectrum of compound **33**, related to **Figure 2**.

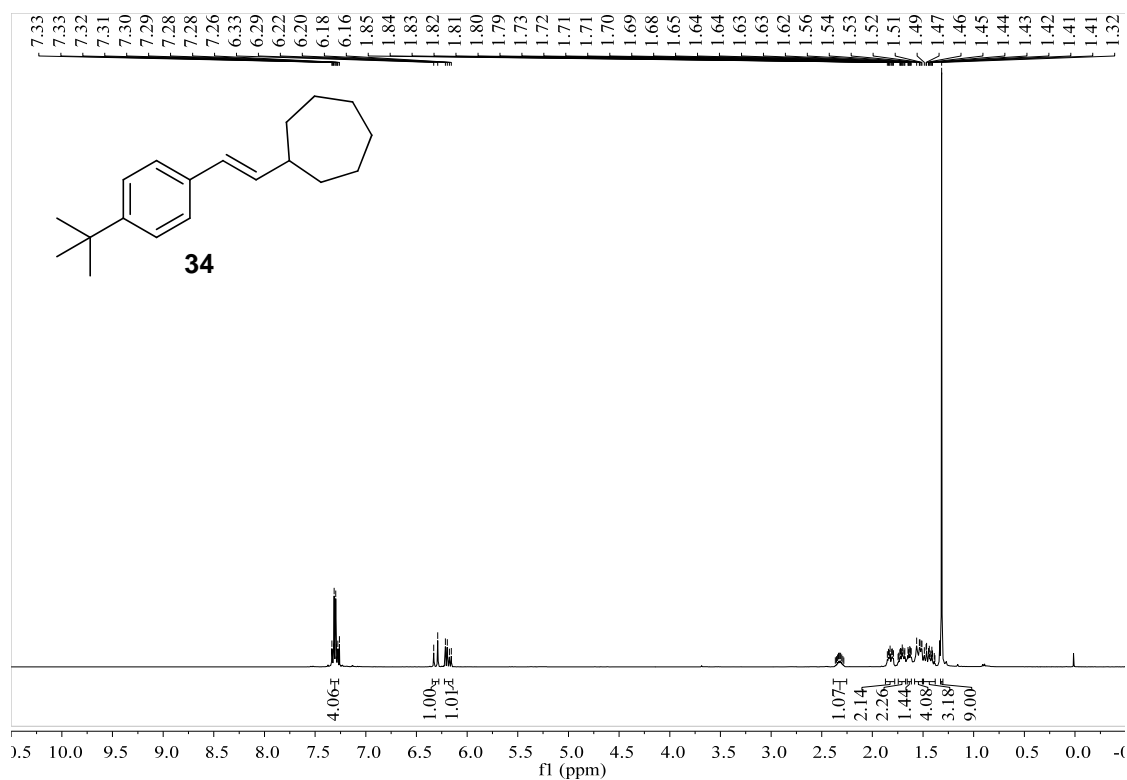


Figure S107. ¹H NMR spectrum of compound **34**, related to Figure 2.

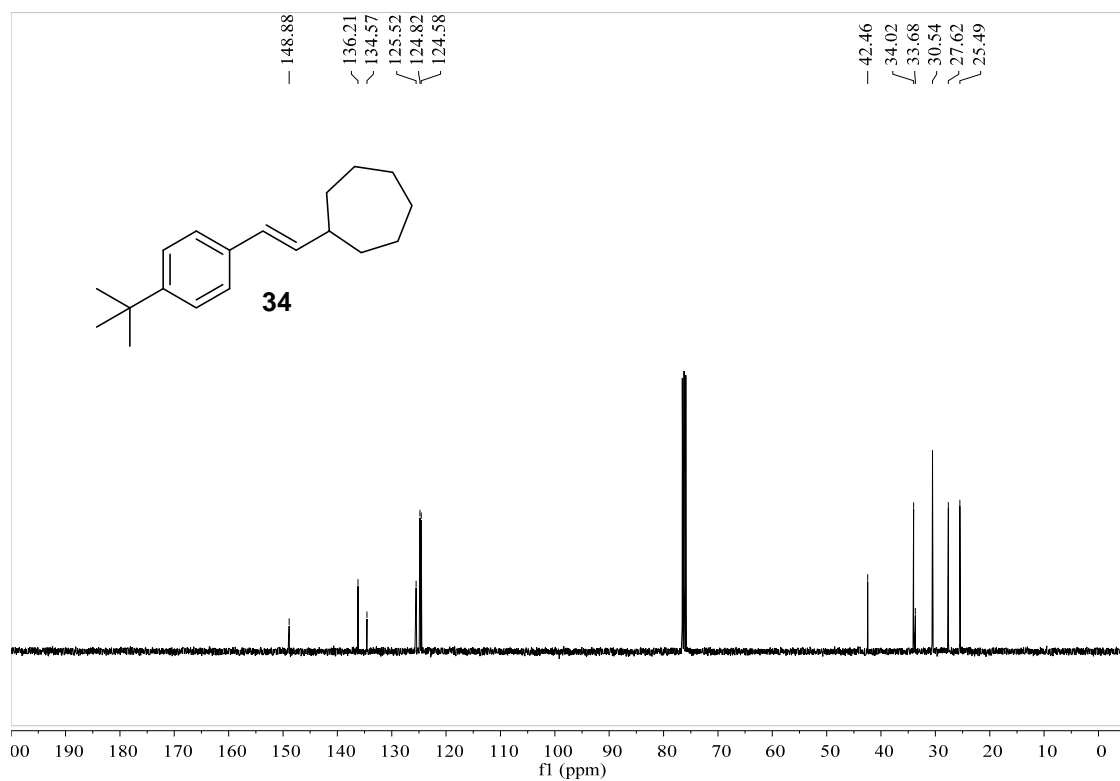
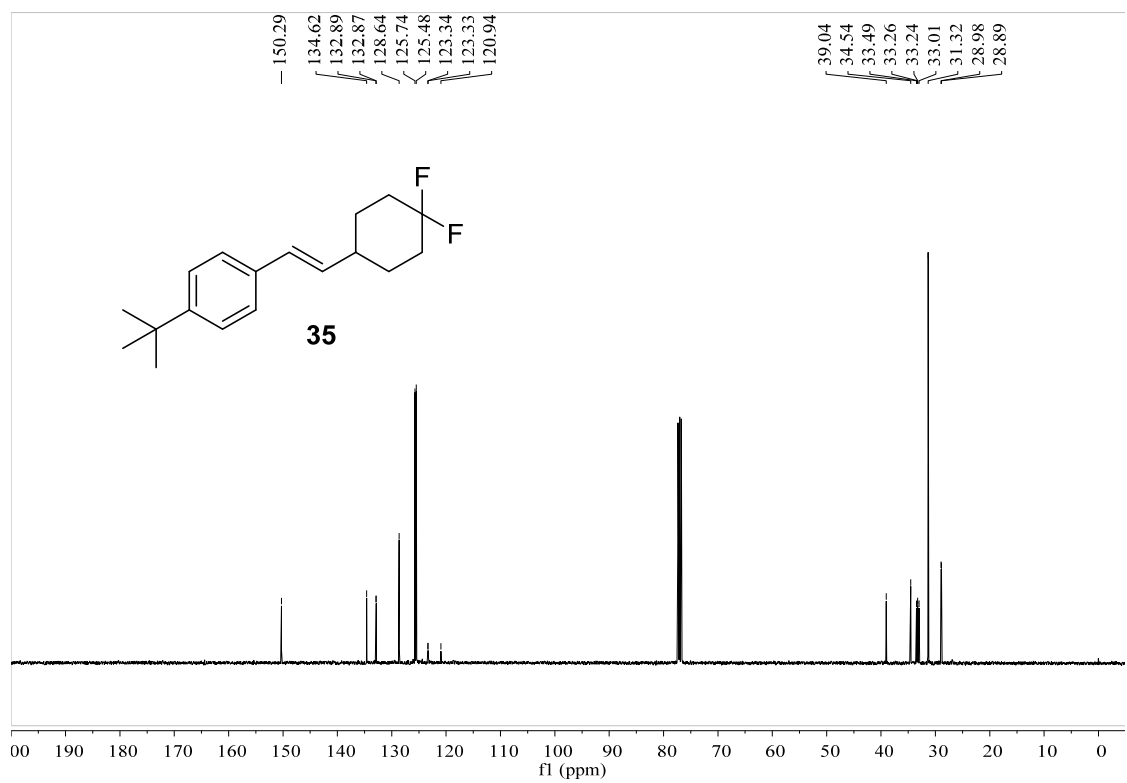
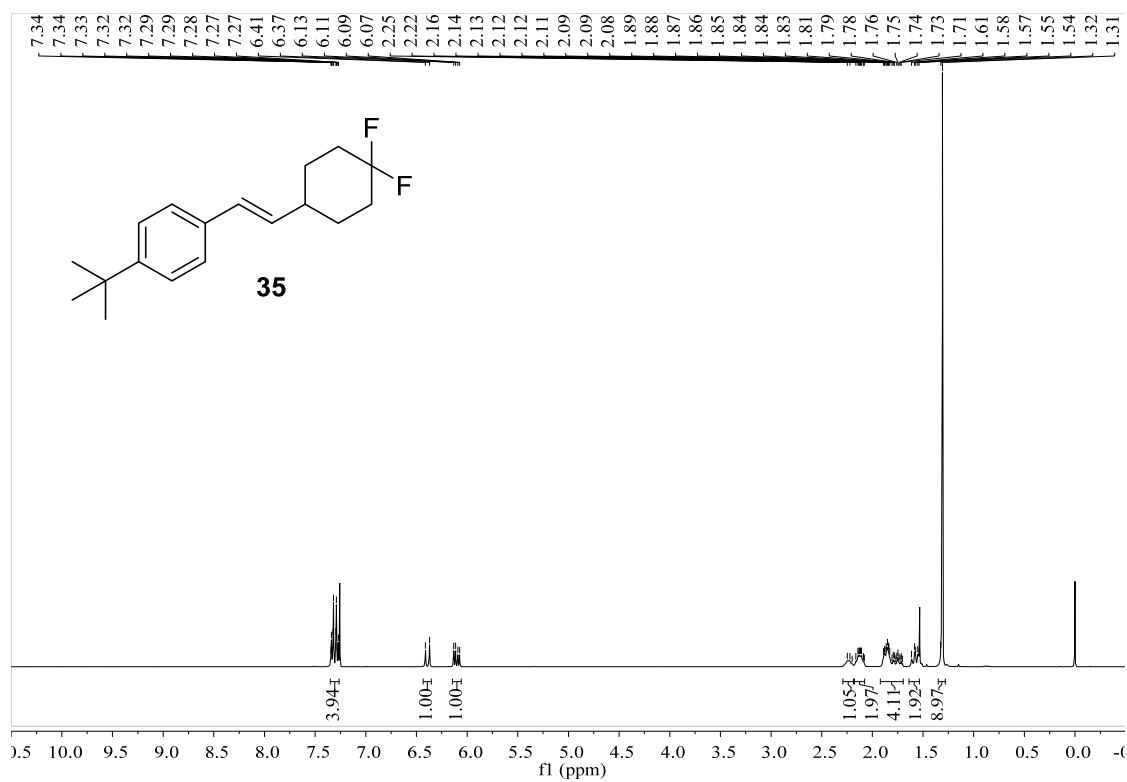


Figure S108. ¹³C NMR spectrum of compound **34**, related to Figure 2.



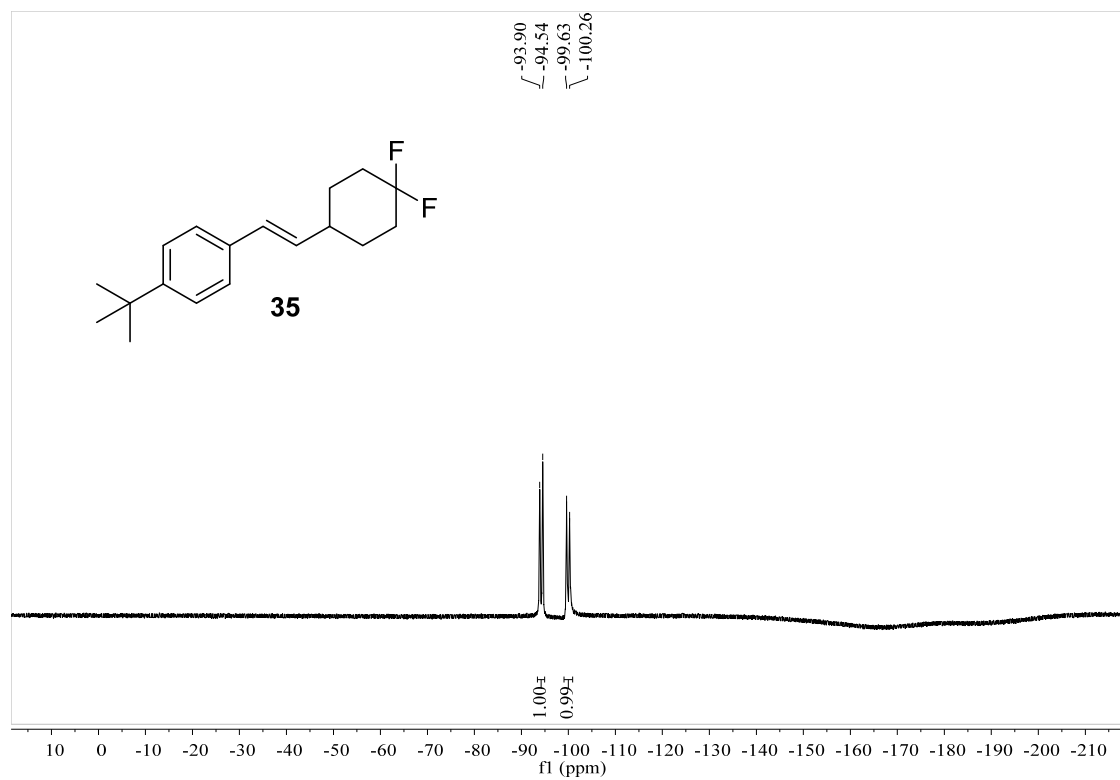


Figure S111. ^{19}F NMR spectrum of compound **35**, related to **Figure 2**.

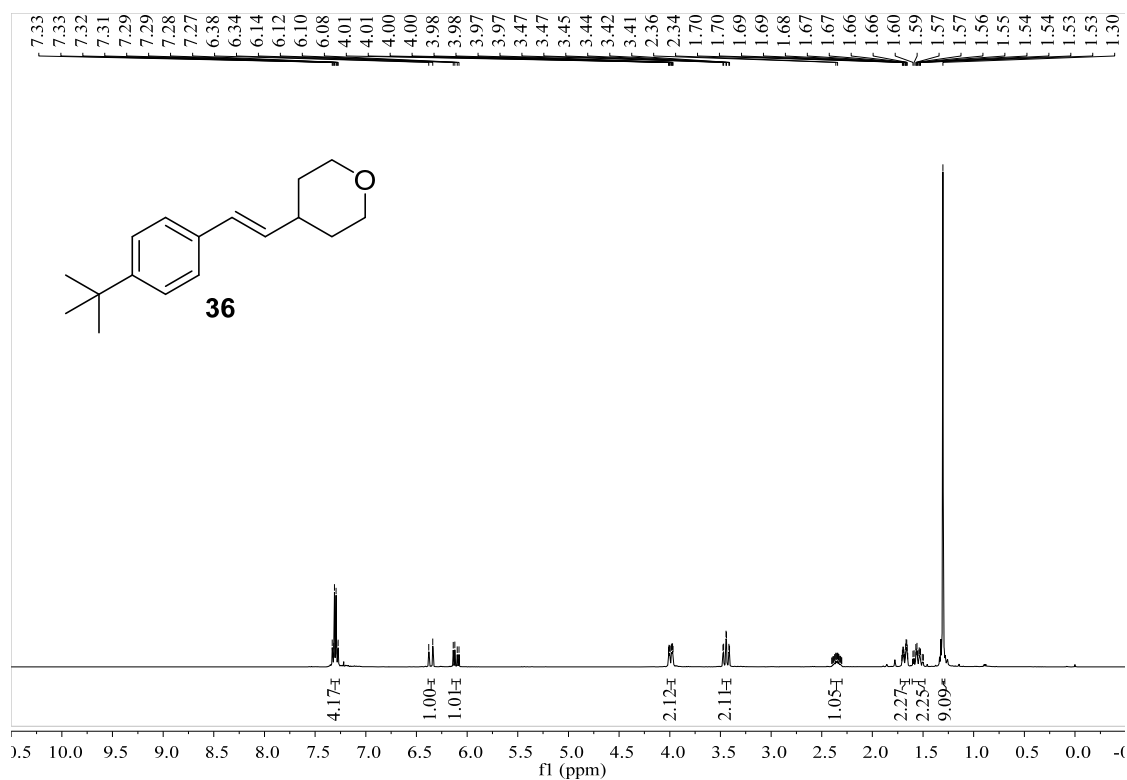


Figure S112. ¹H NMR spectrum of compound **36**, related to Figure 2.

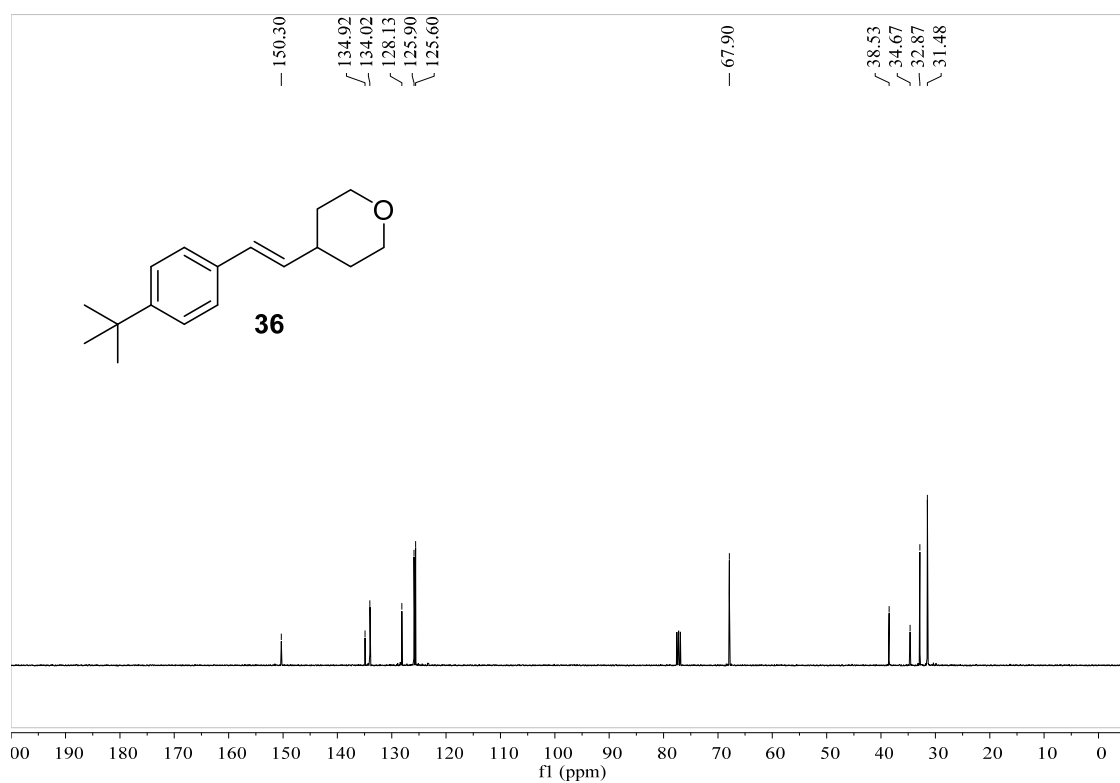


Figure S113. ¹³C NMR spectrum of compound **36**, related to Figure 2.

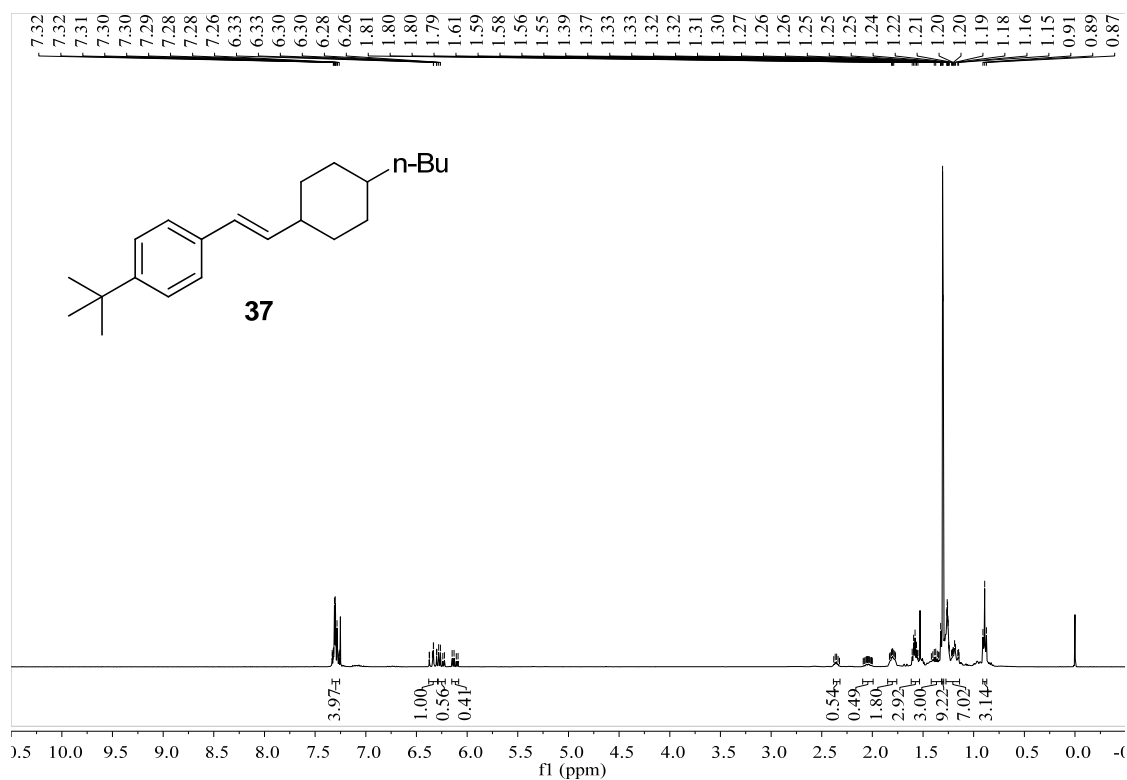


Figure S114. ¹H NMR spectrum of compound **37**, related to Figure 2.

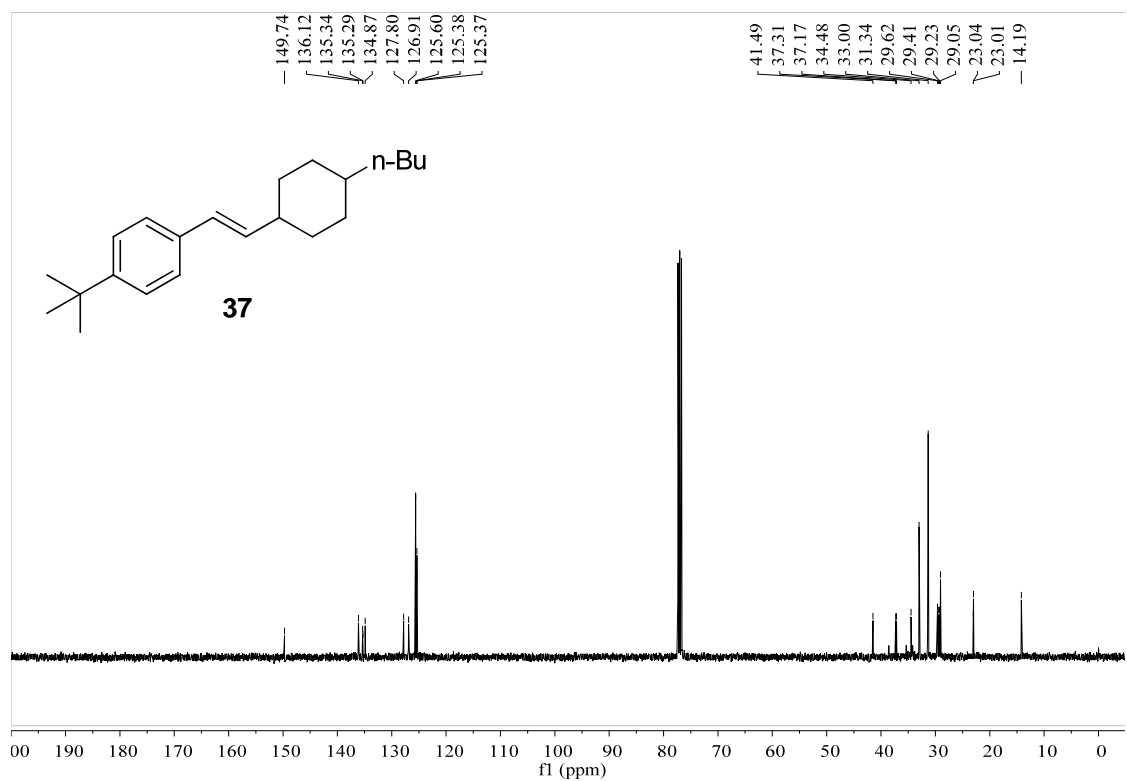


Figure S115. ¹³C NMR spectrum of compound **37**, related to Figure 2.

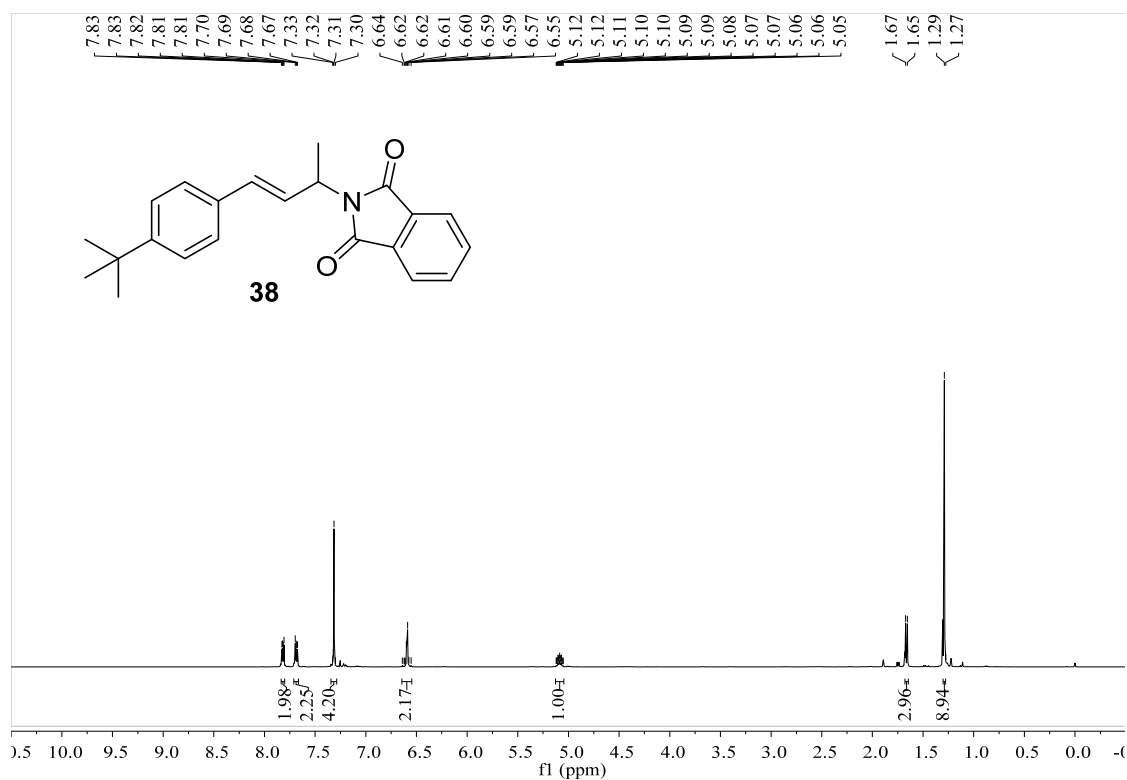


Figure S116. ^1H NMR spectrum of compound **38**, related to **Figure 2**.

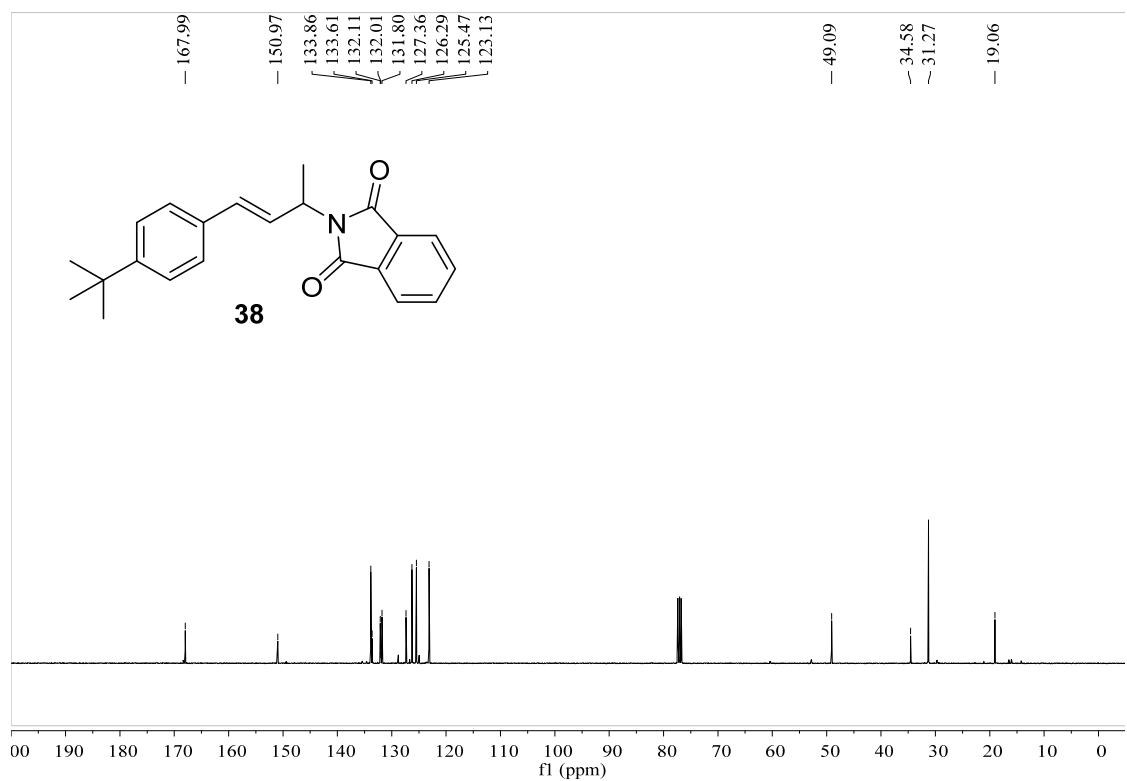


Figure S117. ^{13}C NMR spectrum of compound **38**, related to **Figure 2**.

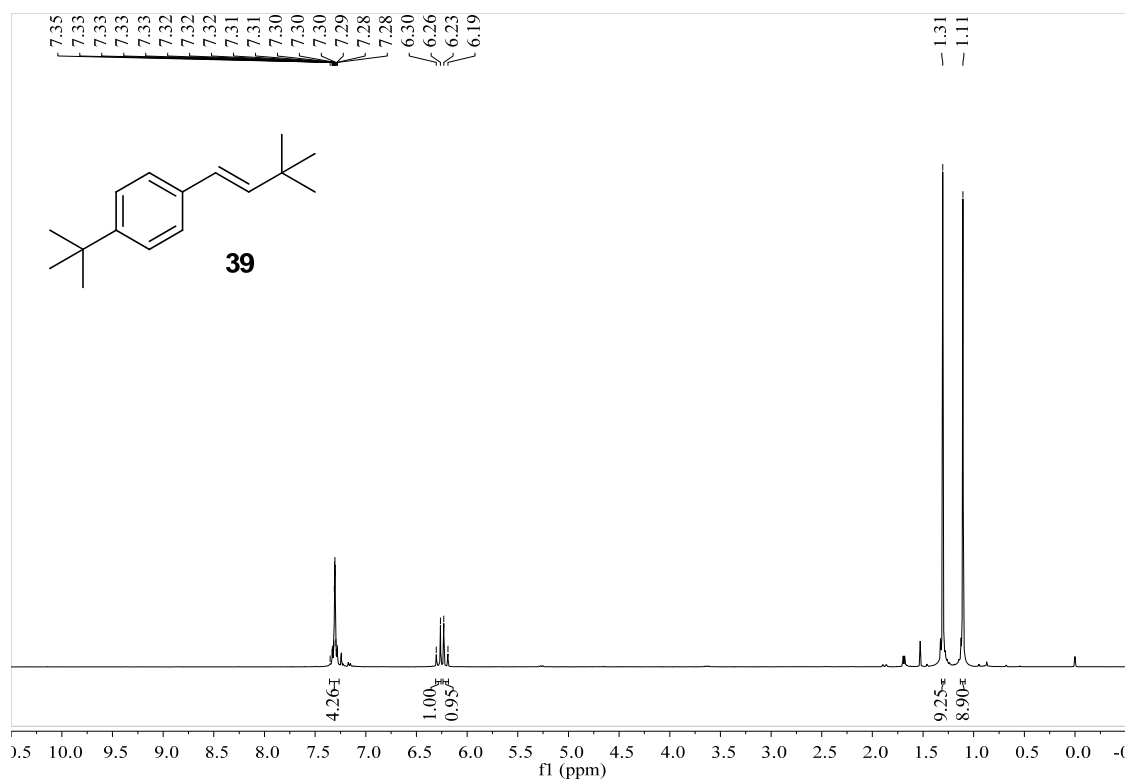


Figure S118. ¹H NMR spectrum of compound **39**, related to **Figure 2**.

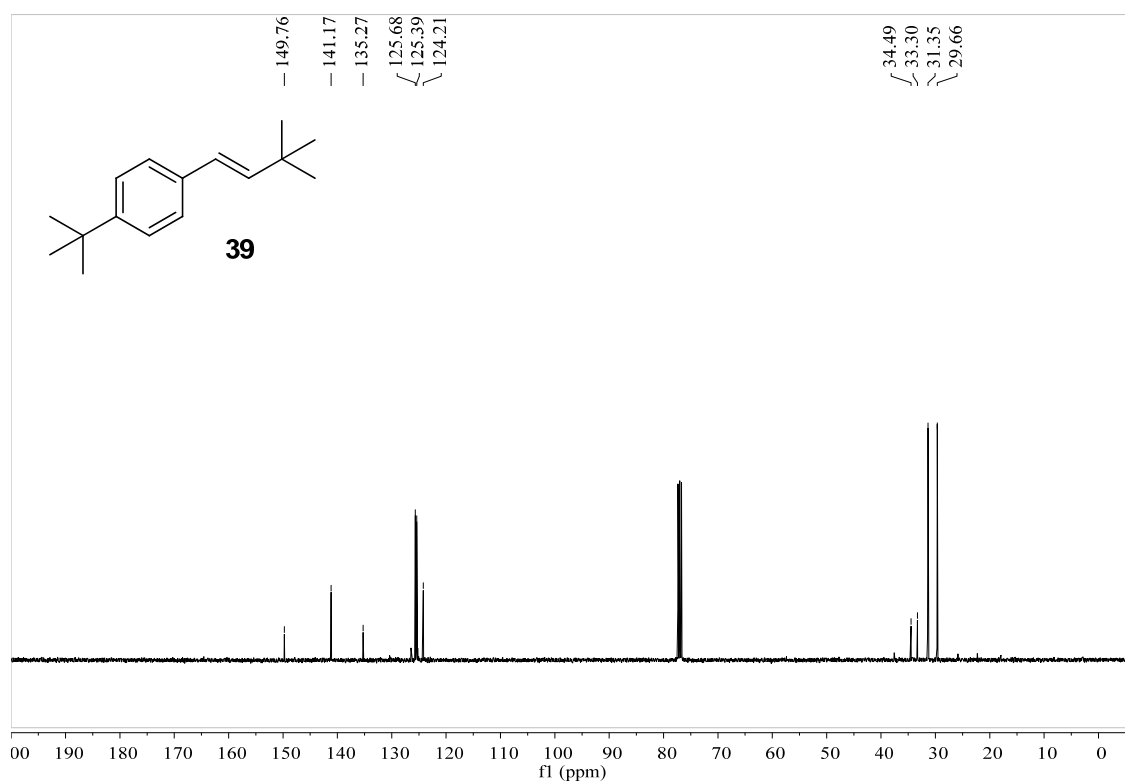
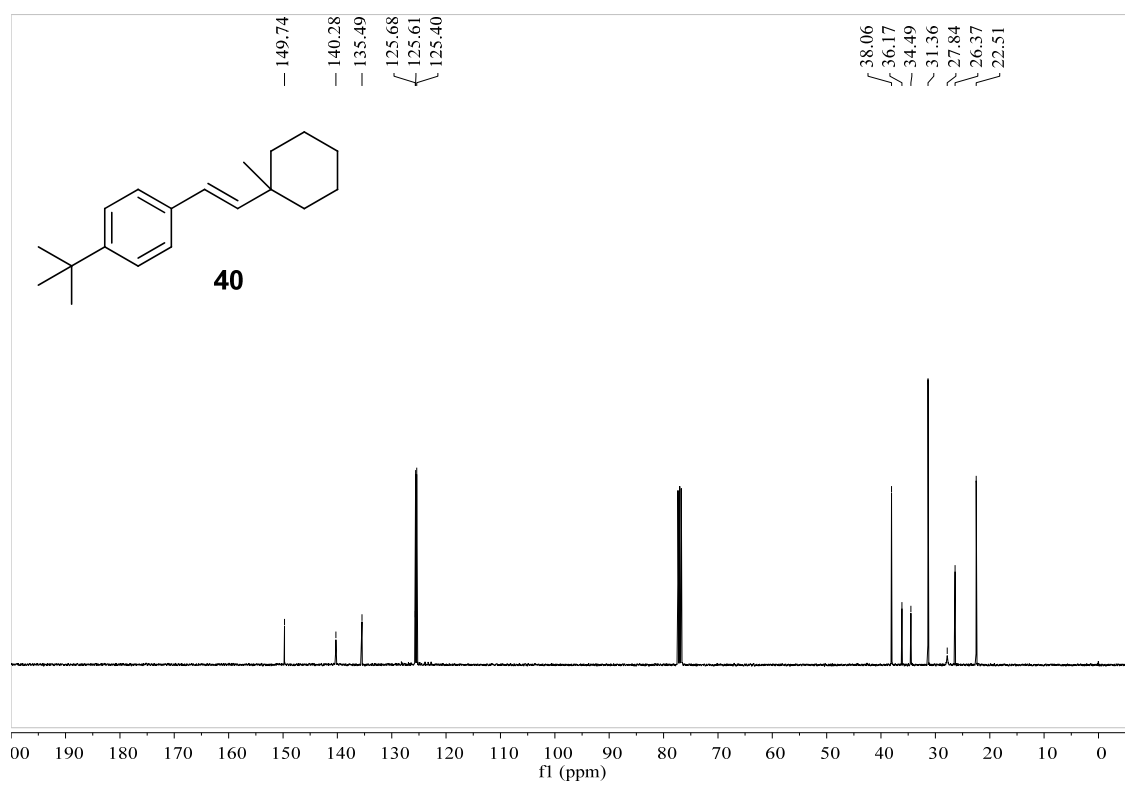
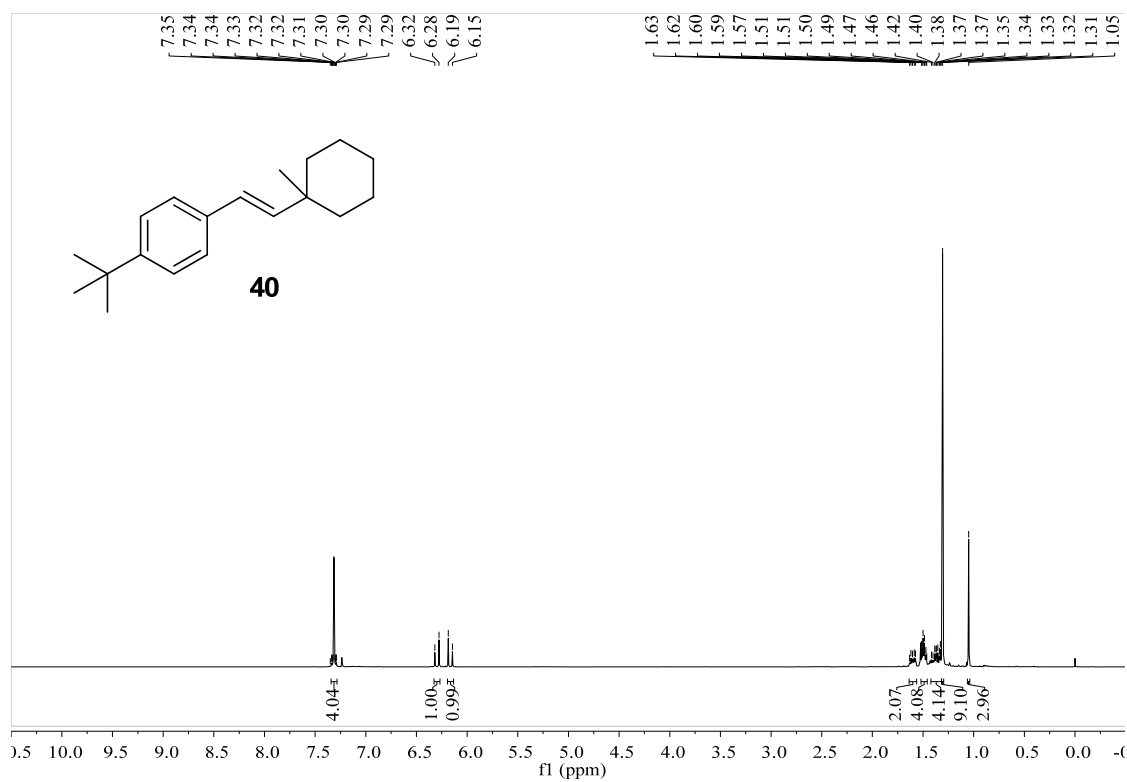


Figure S119. ¹³C NMR spectrum of compound **39**, related to **Figure 2**.



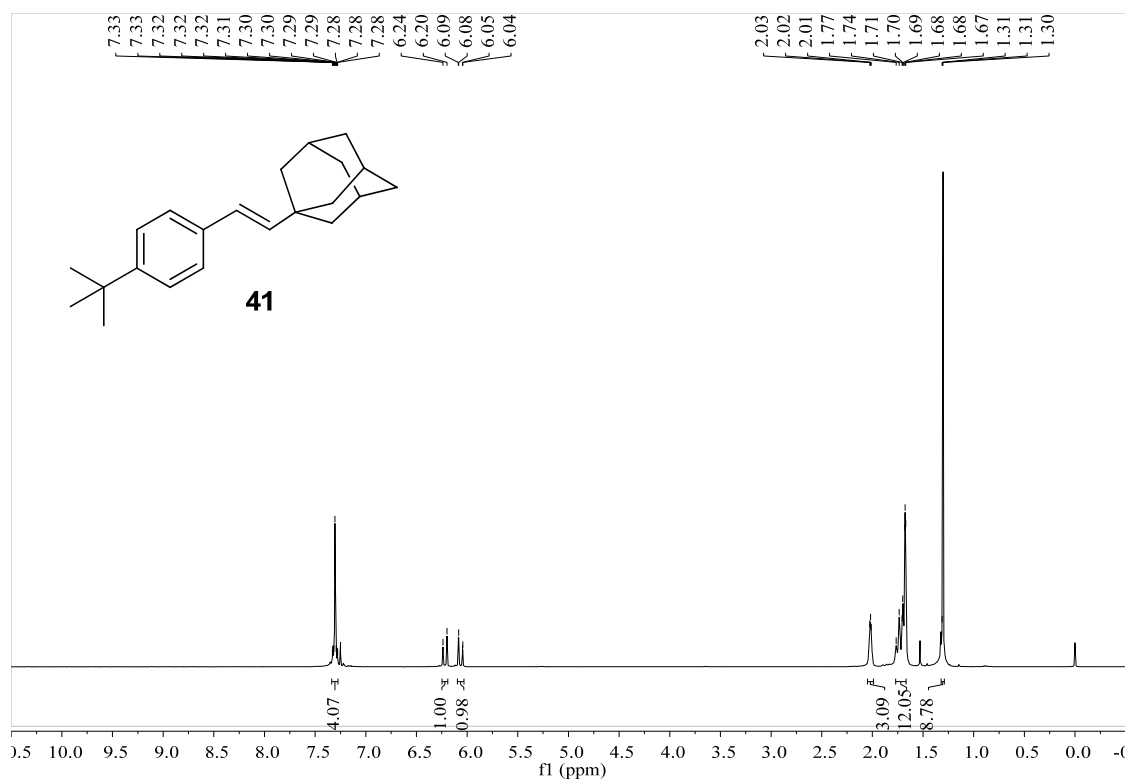


Figure S122. ¹H NMR spectrum of compound **41**, related to **Figure 2**.

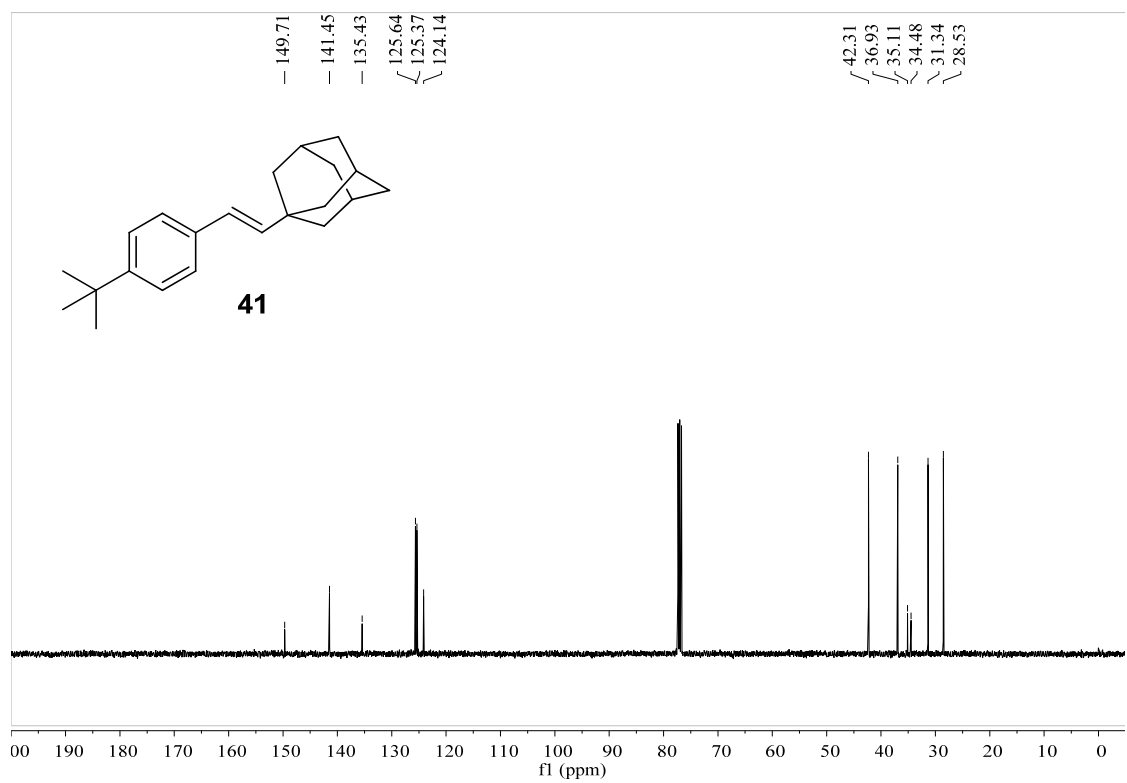


Figure S123. ¹³C NMR spectrum of compound **41**, related to **Figure 2**.

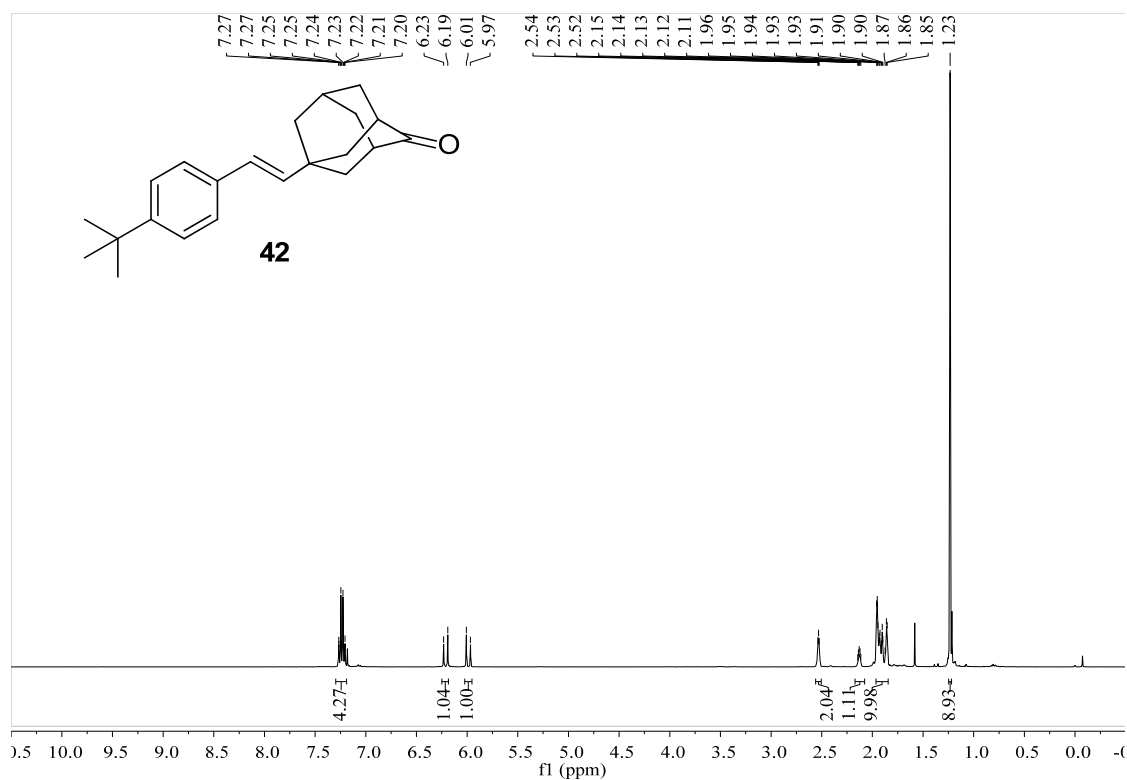


Figure S124. ^1H NMR spectrum of compound **42**, related to **Figure 2**.

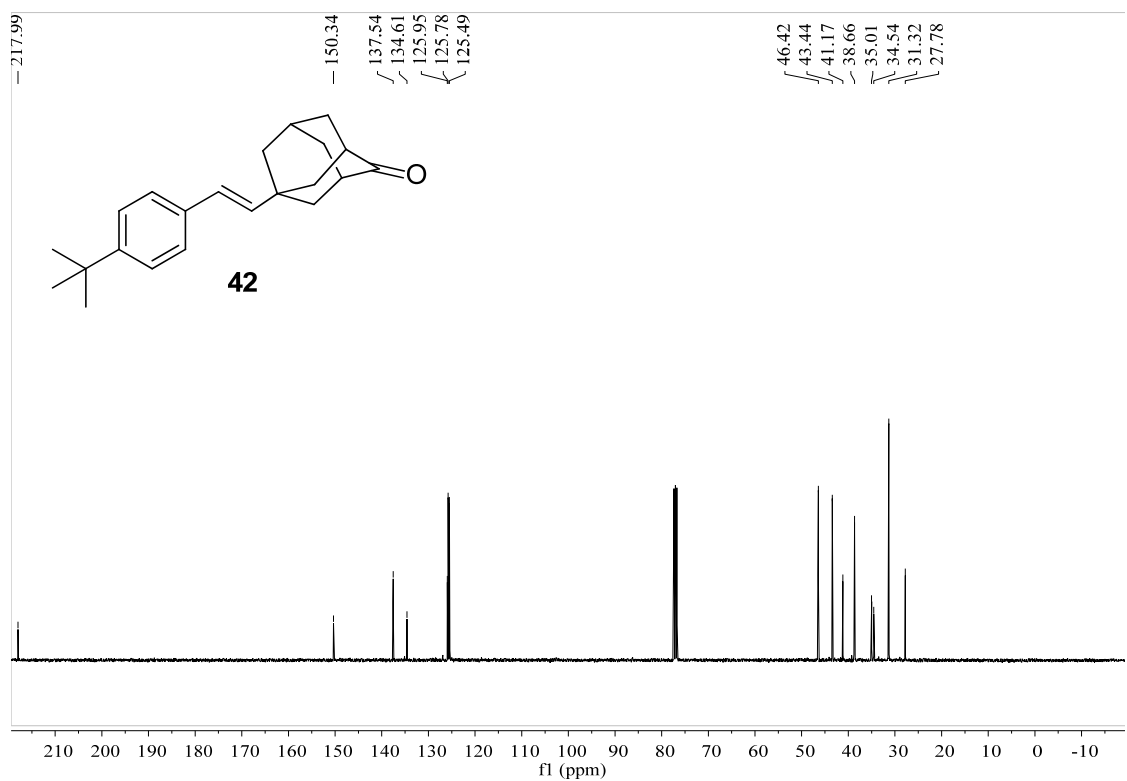


Figure S125. ^{13}C NMR spectrum of compound **42**, related to **Figure 2**.

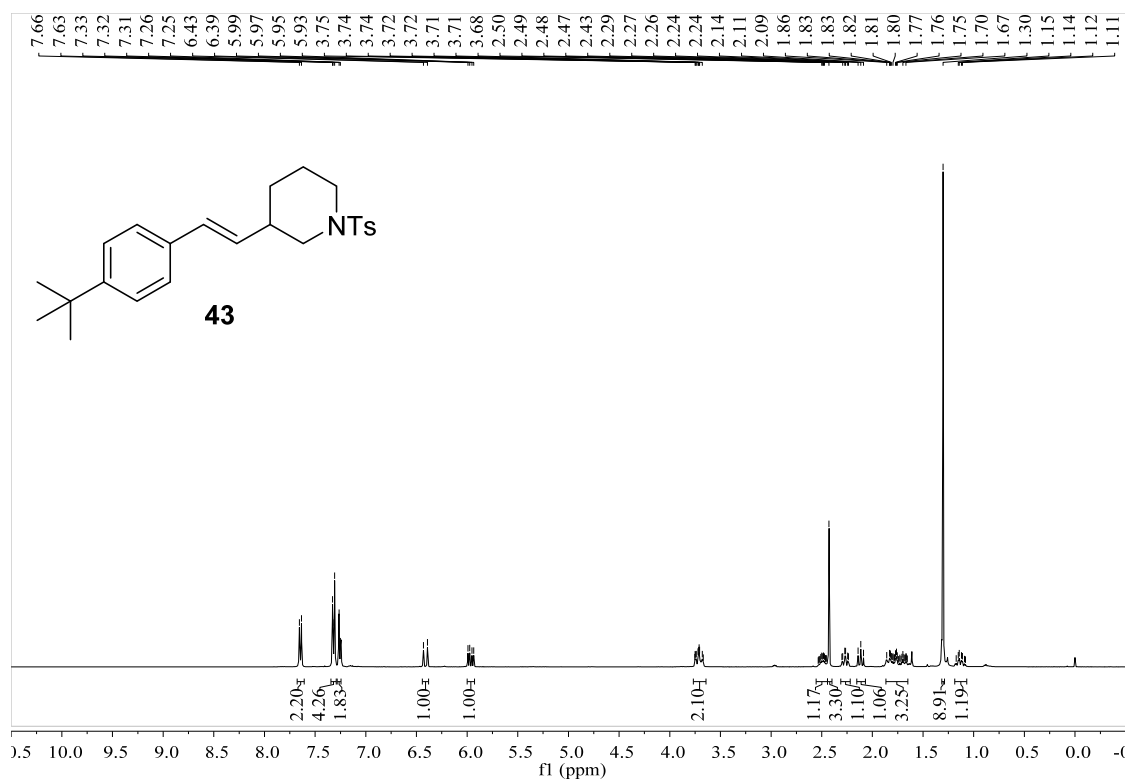


Figure S126. ¹H NMR spectrum of compound 43, related to Figure 2.

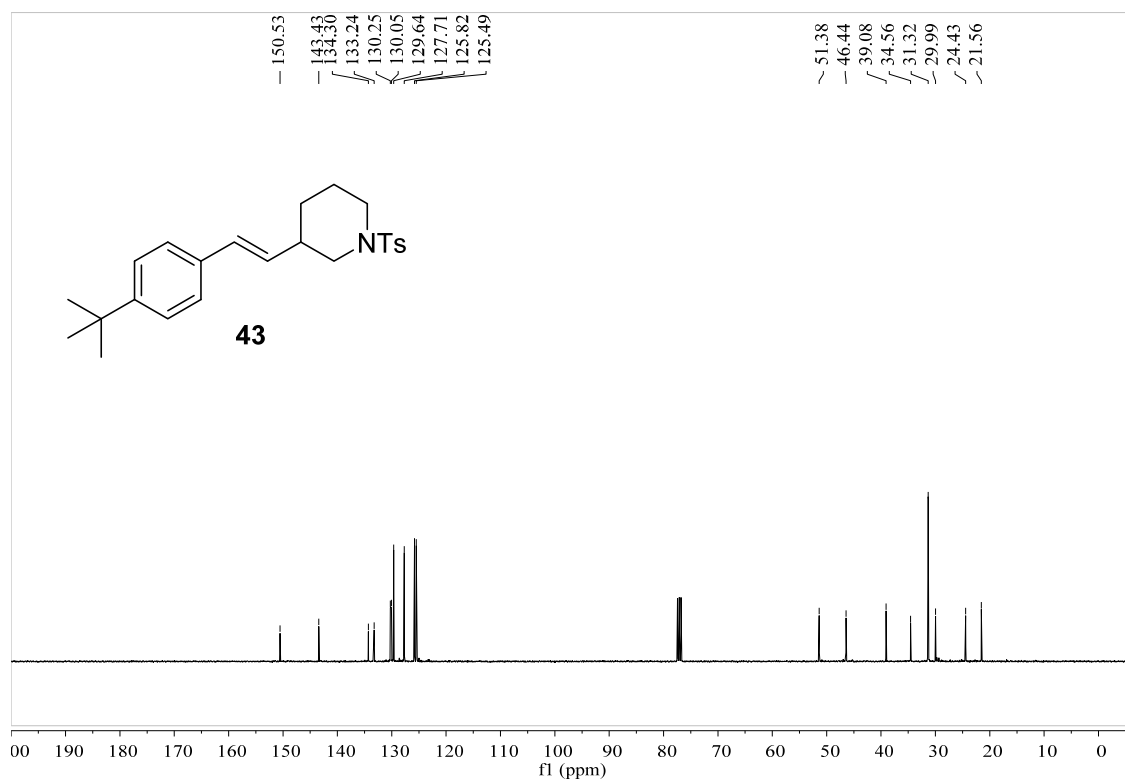


Figure S127. ¹³C NMR spectrum of compound 43, related to Figure 2.

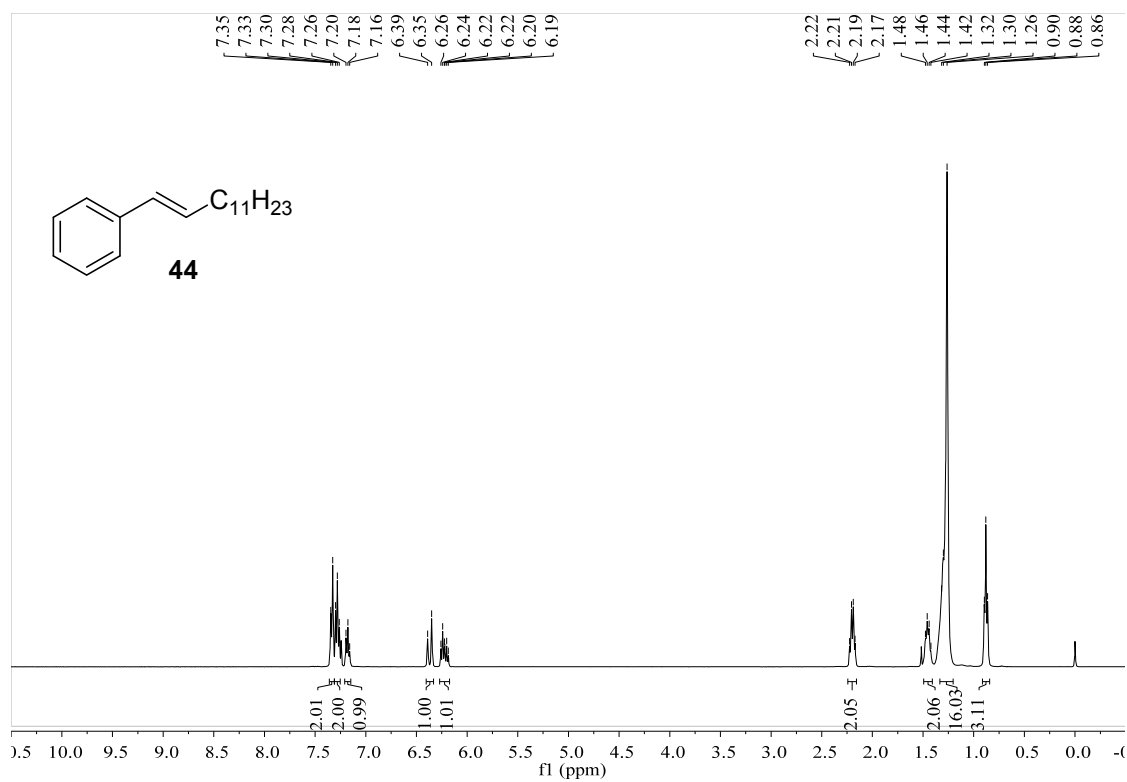


Figure S128. ¹H NMR spectrum of compound 44, related to Figure 3.

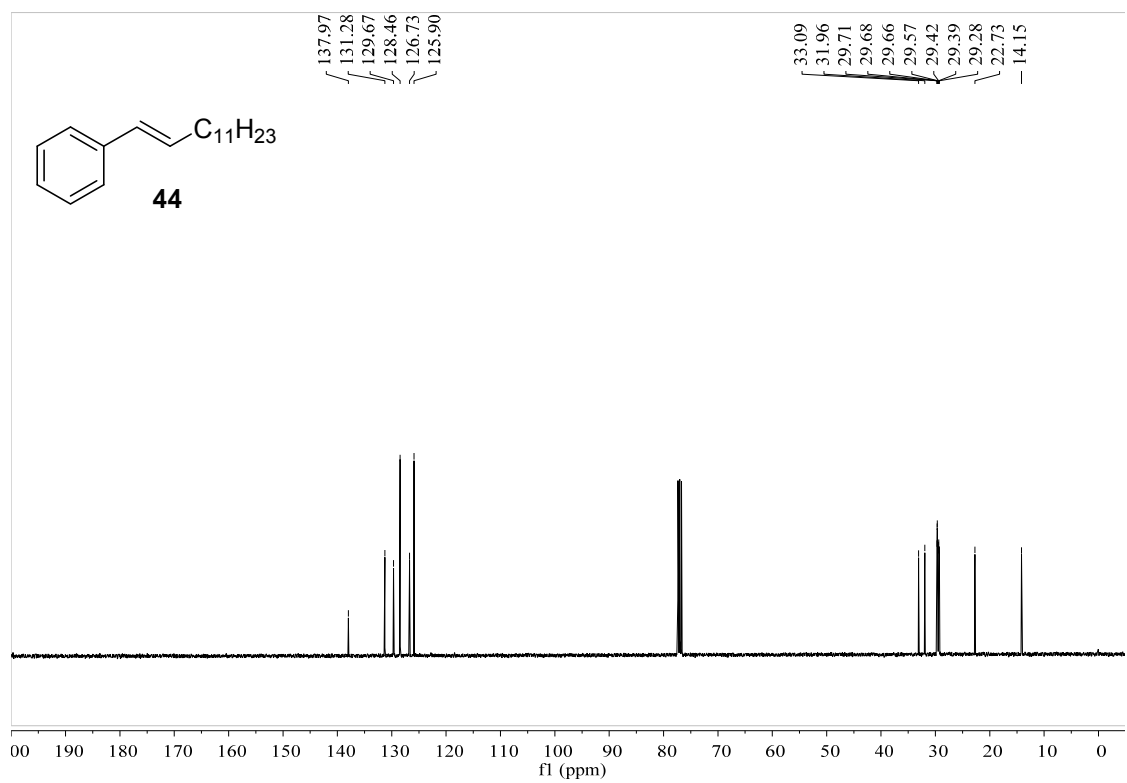


Figure S129. ¹³C NMR spectrum of compound 44, related to Figure 3.

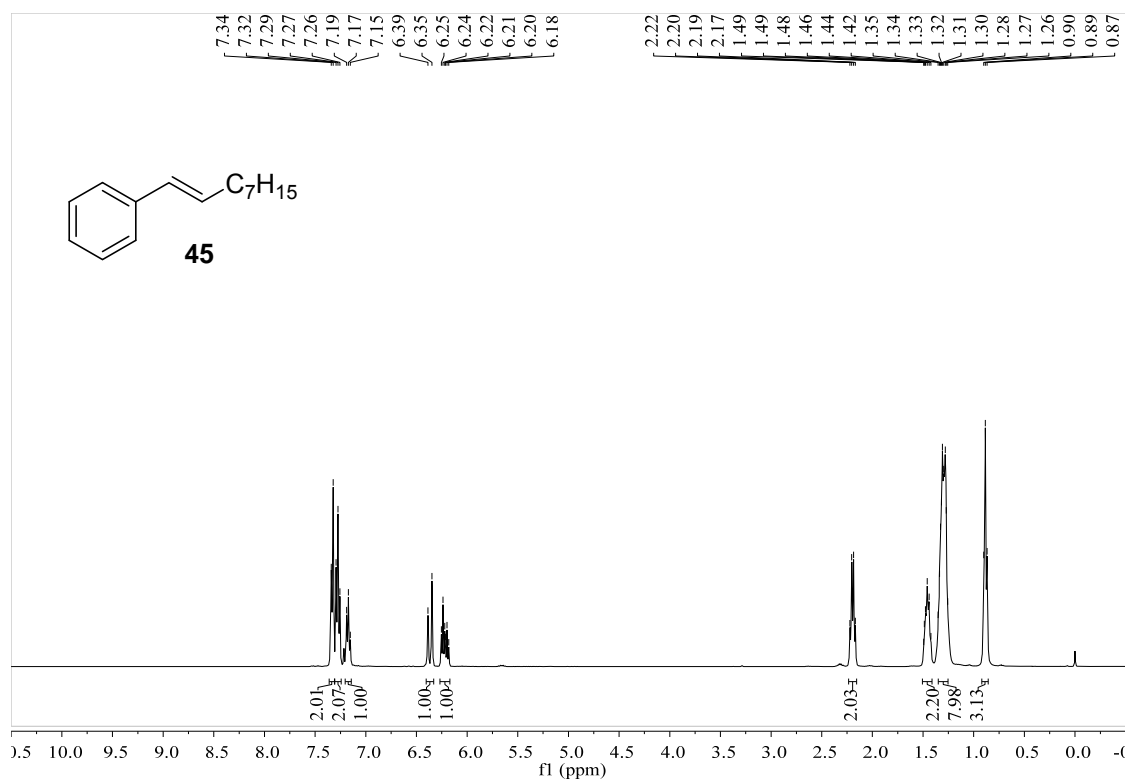


Figure S130. ¹H NMR spectrum of compound 45, related to Figure 3.

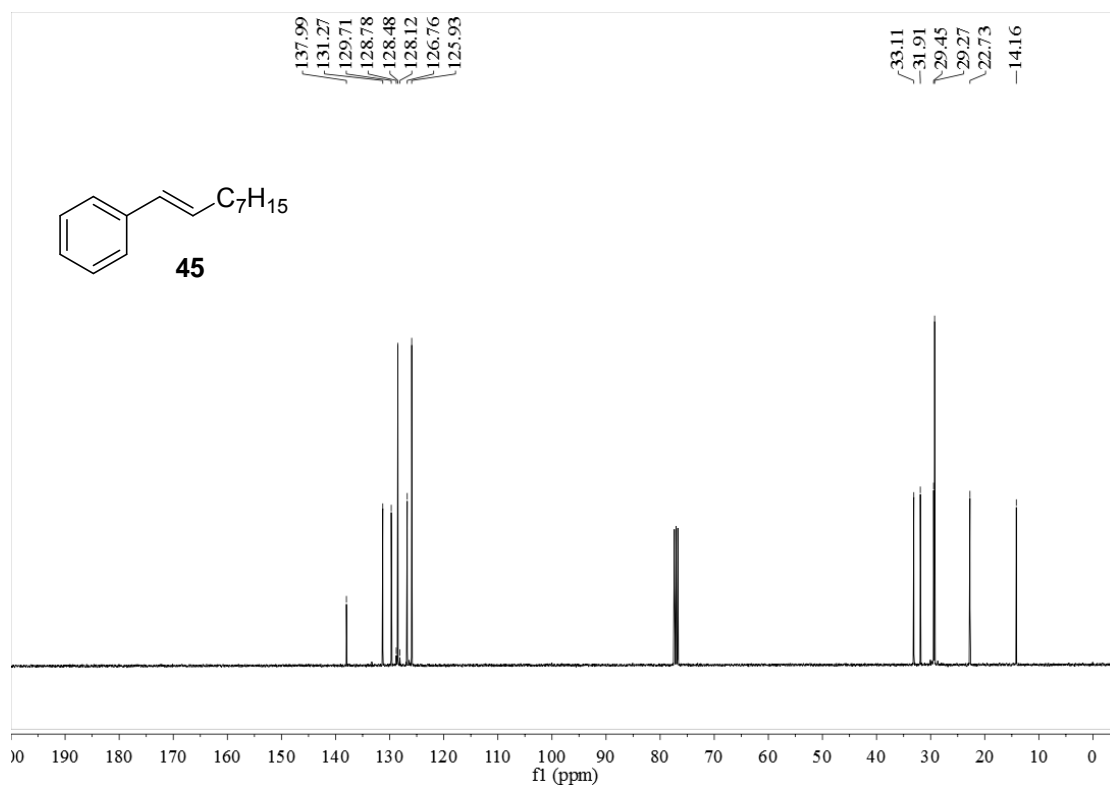


Figure S131. ¹³C NMR spectrum of compound 45, related to Figure 3.

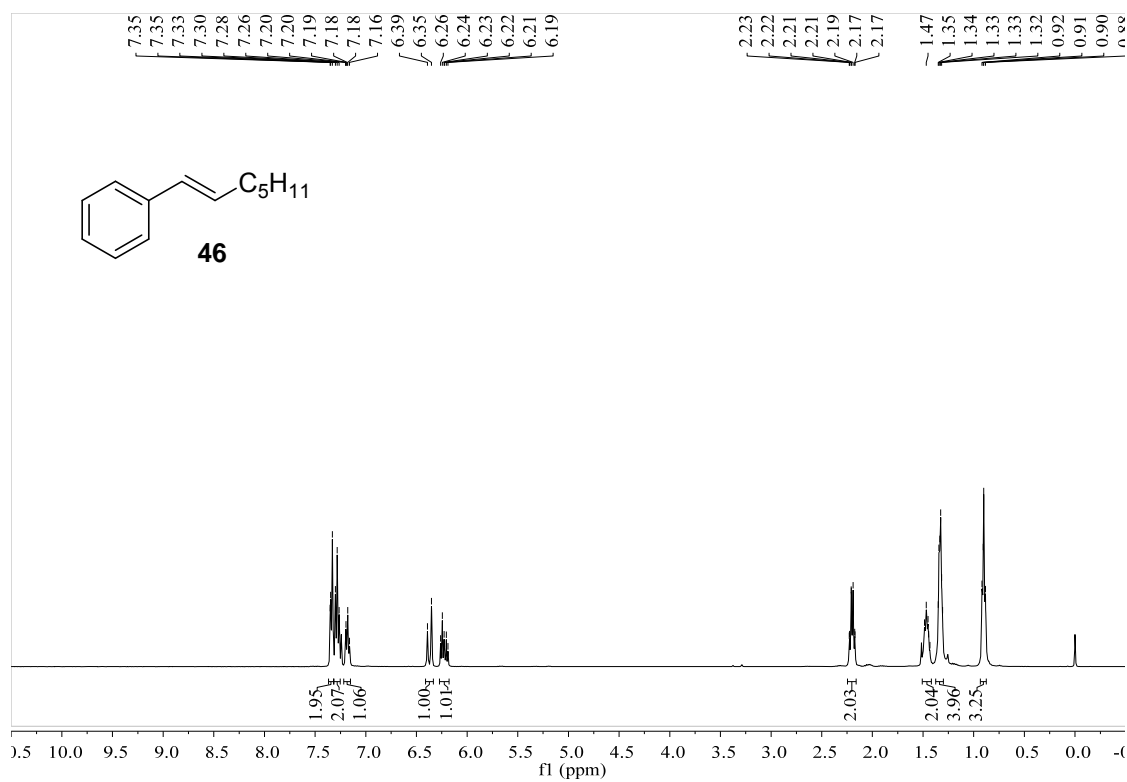


Figure S132. ¹H NMR spectrum of compound **46**, related to **Figure 3**.

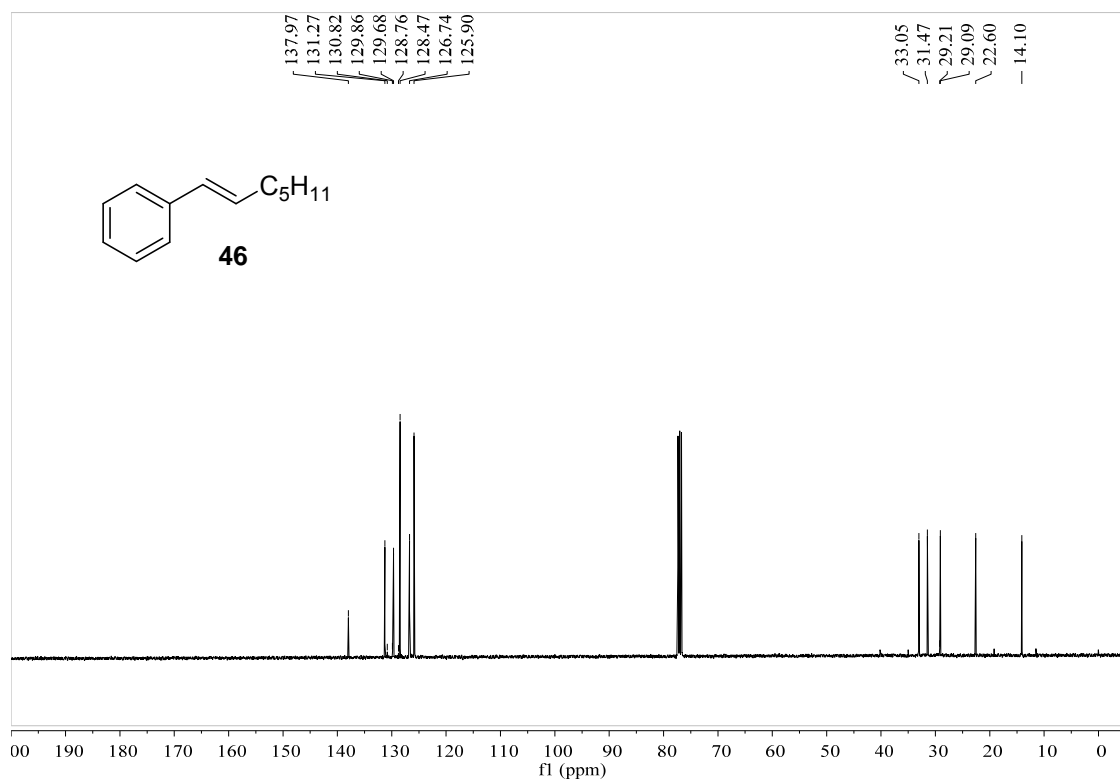


Figure S133. ¹³C NMR spectrum of compound **46**, related to **Figure 3**.

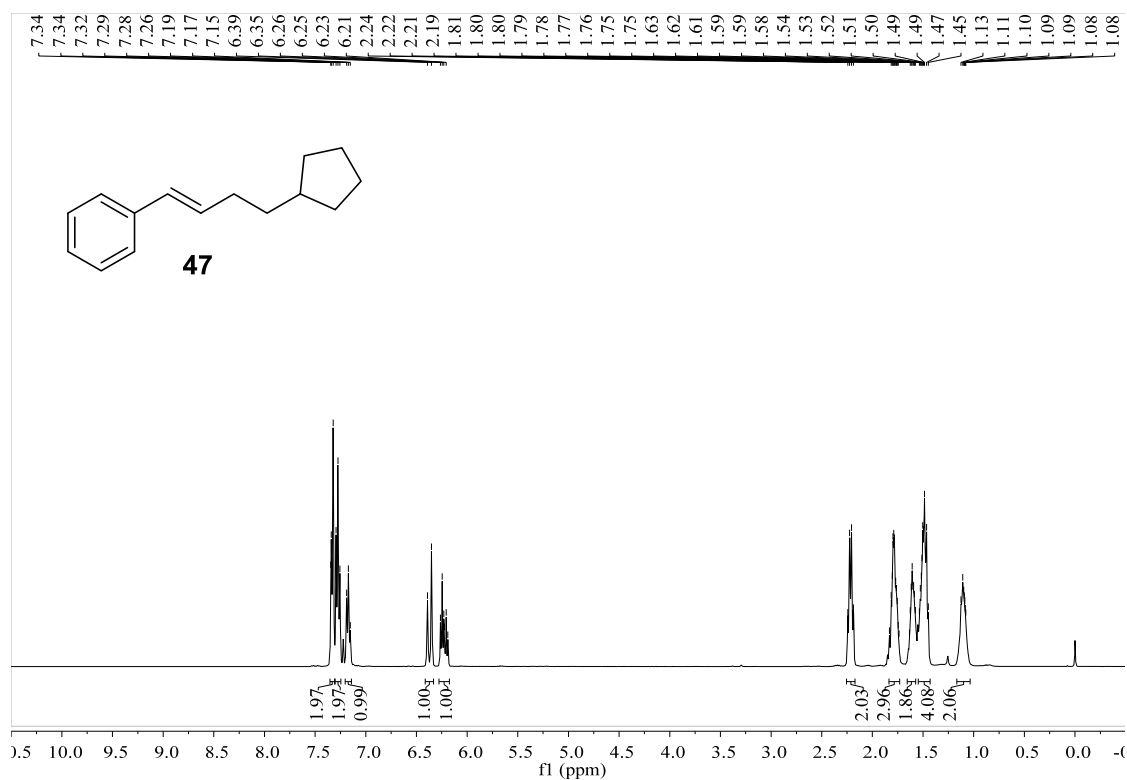


Figure S134. ¹H NMR spectrum of compound 47, related to Figure 3.

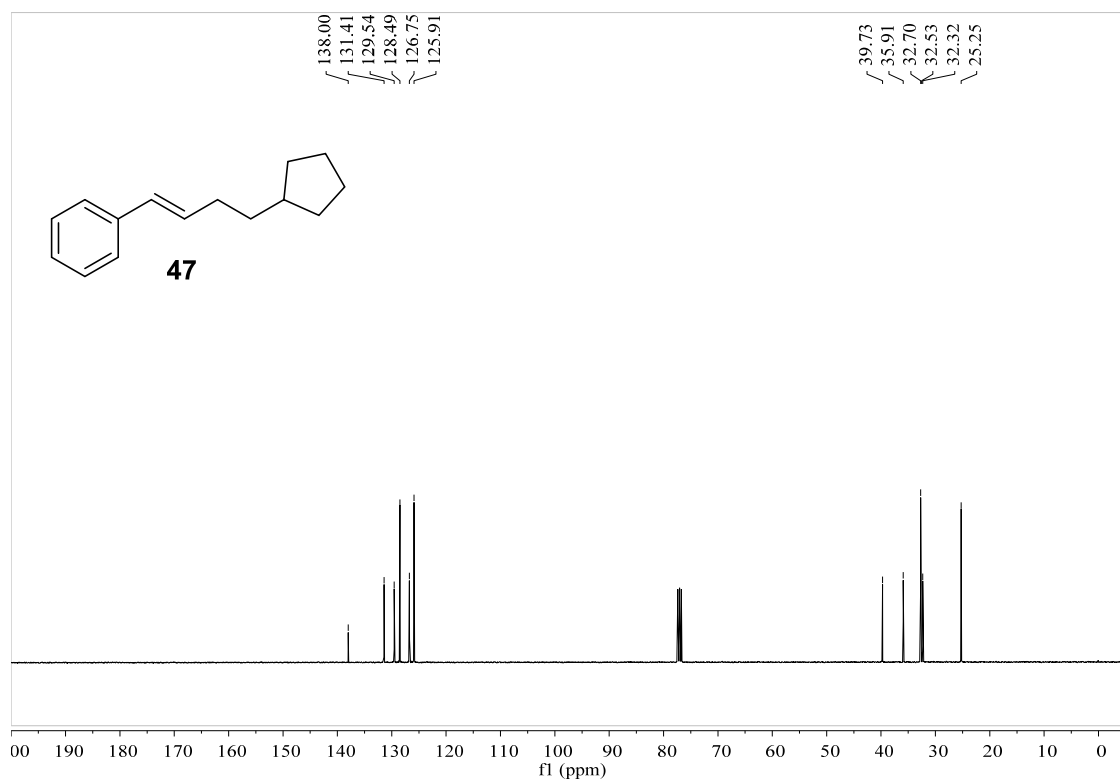


Figure S135. ¹³C NMR spectrum of compound 47, related to Figure 3.

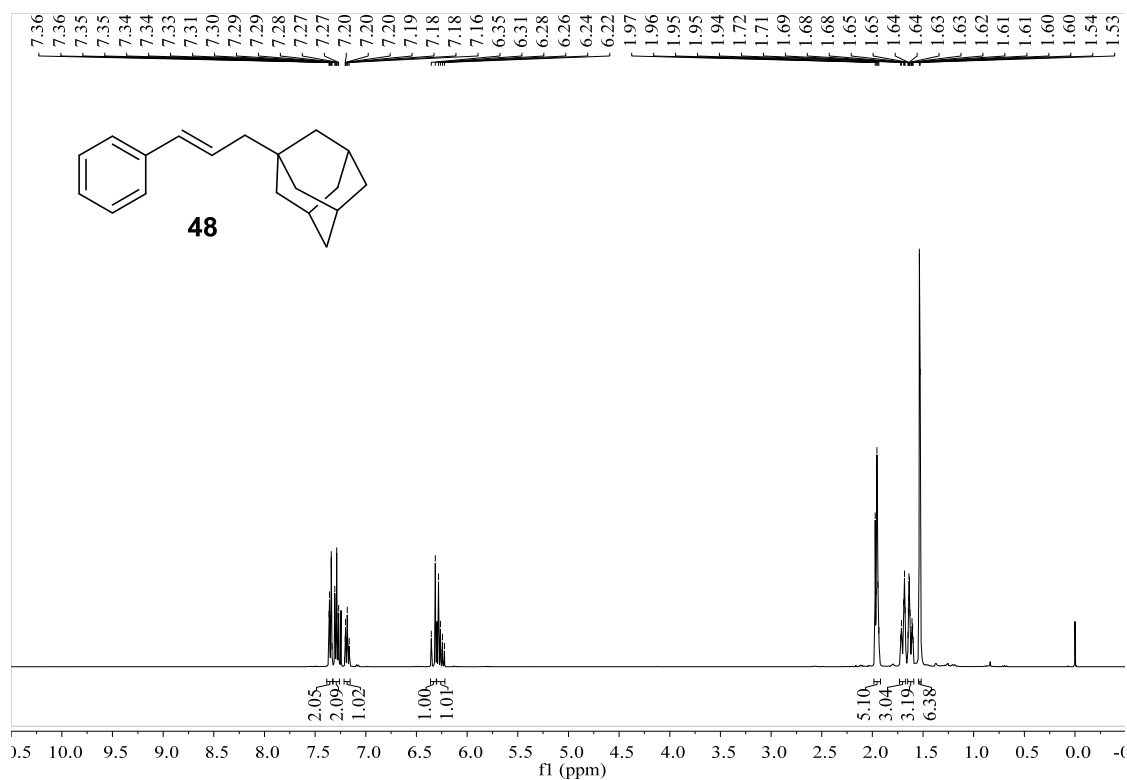


Figure S136. ^1H NMR spectrum of compound **48**, related to **Figure 3**.

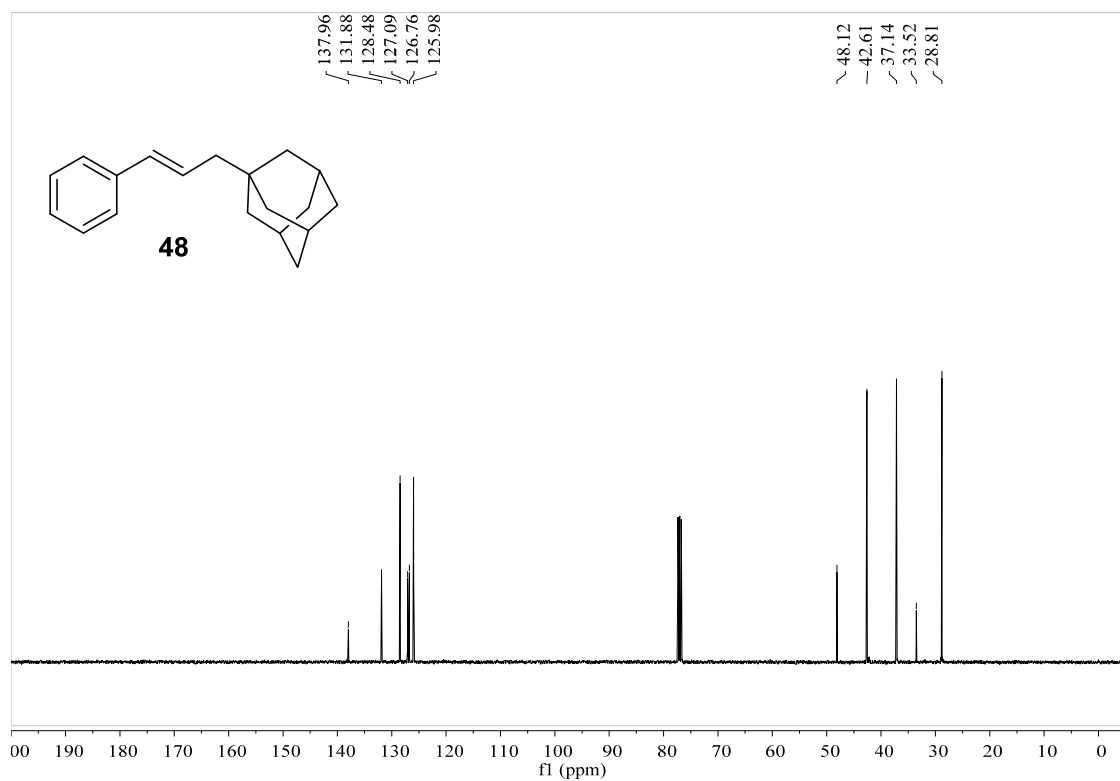


Figure S137. ^{13}C NMR spectrum of compound **48**, related to **Figure 3**.

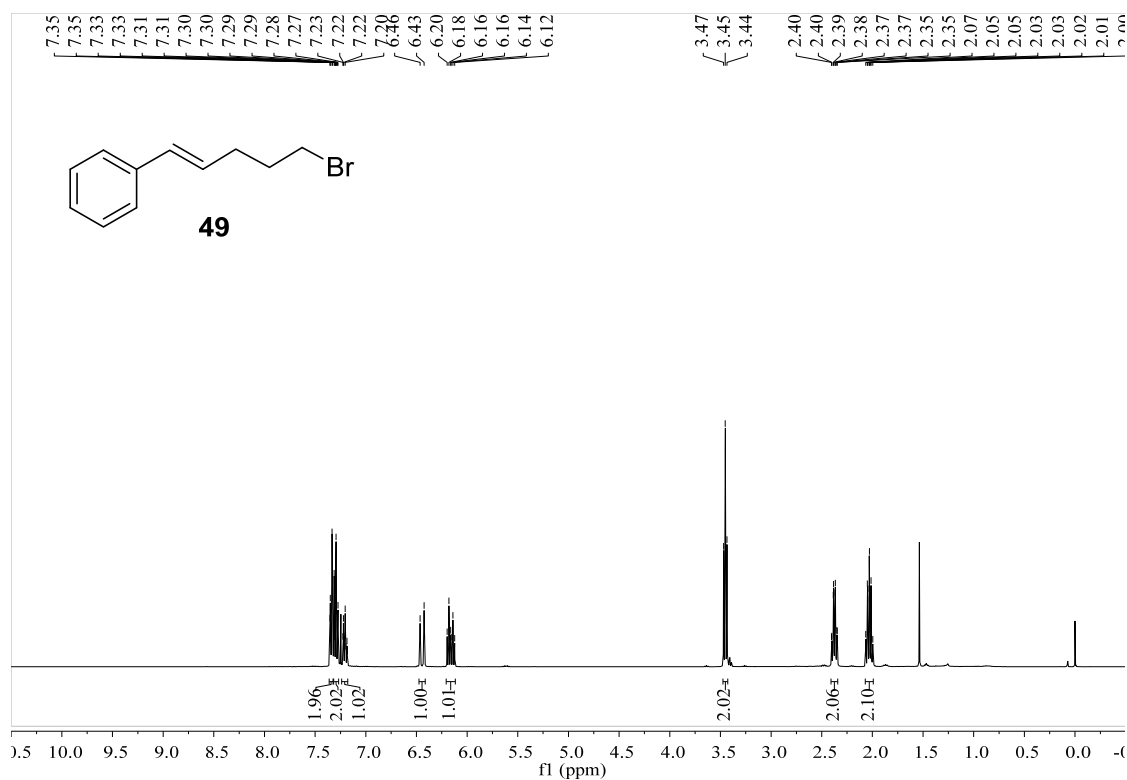


Figure S138. ¹H NMR spectrum of compound 49, related to Figure 3.

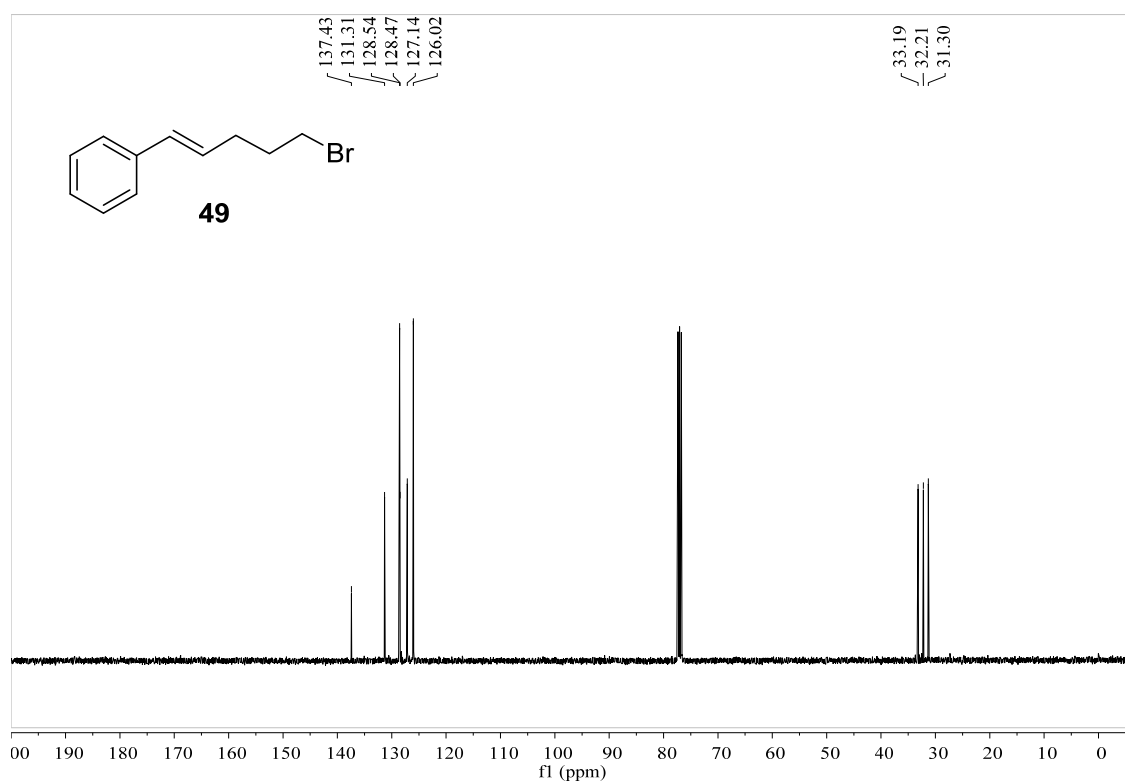


Figure S139. ¹³C NMR spectrum of compound 49, related to Figure 3.

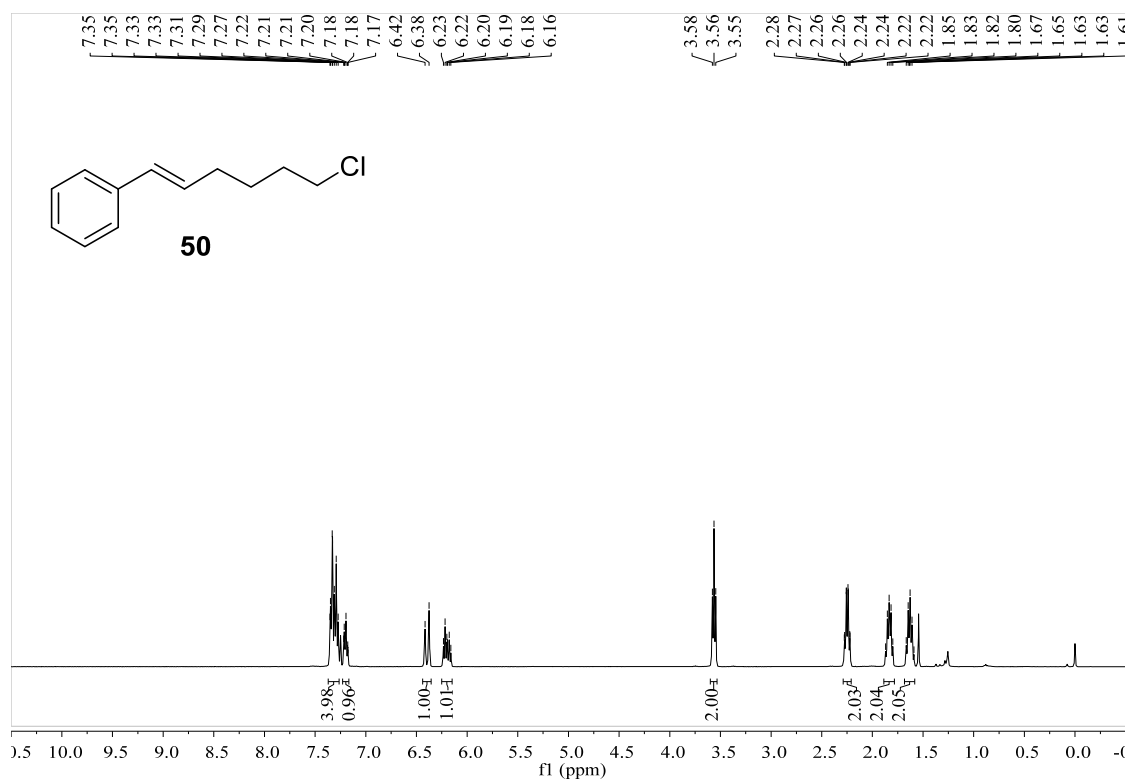


Figure S140. ¹H NMR spectrum of compound **50**, related to **Figure 3**.

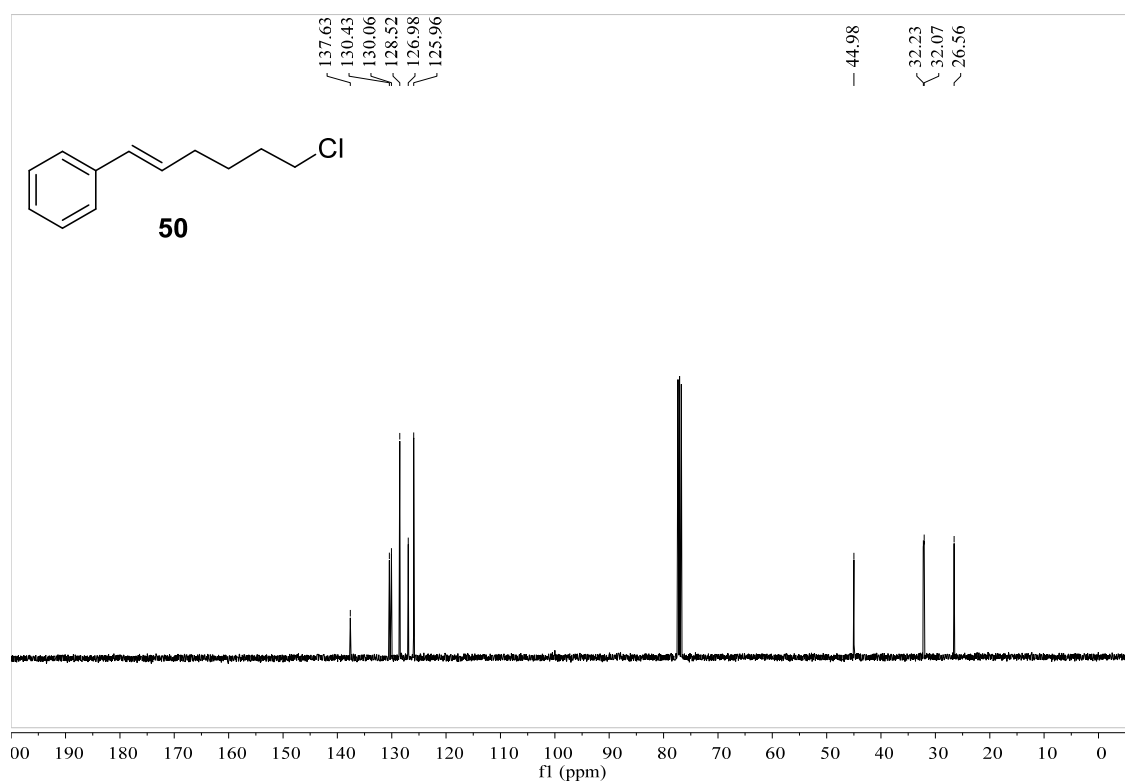


Figure S141. ¹³C NMR spectrum of compound **50**, related to **Figure 3**.

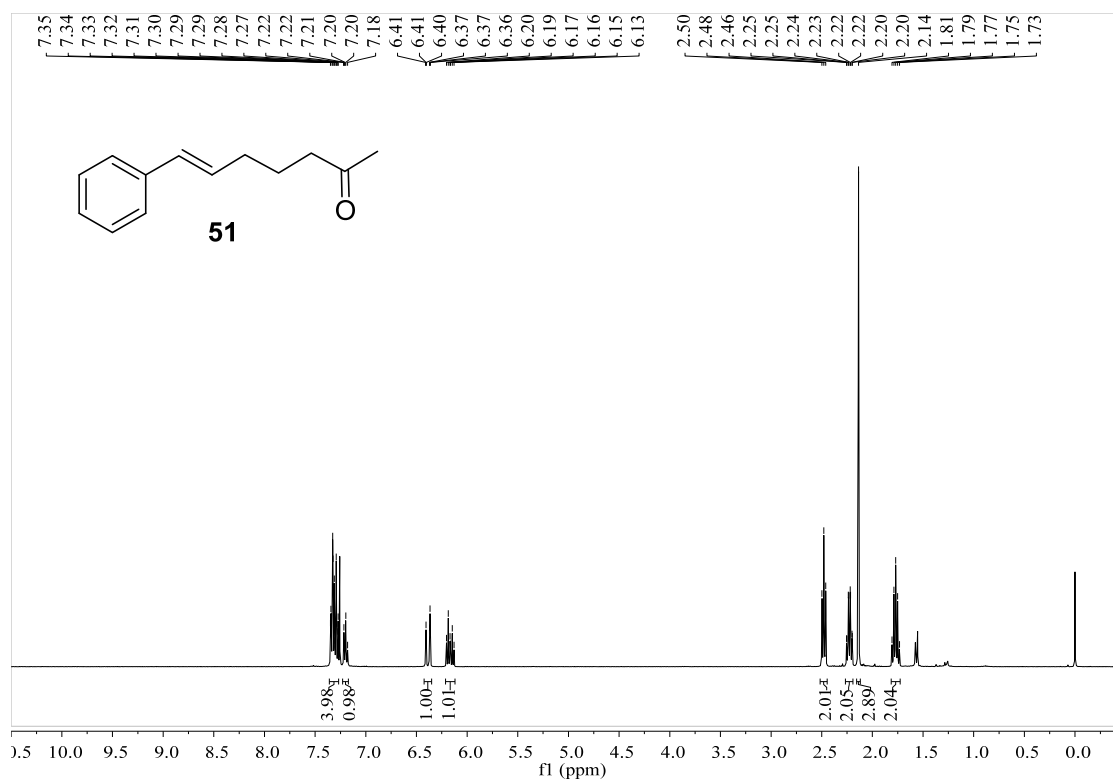


Figure S142. ¹H NMR spectrum of compound **51**, related to **Figure 3**.

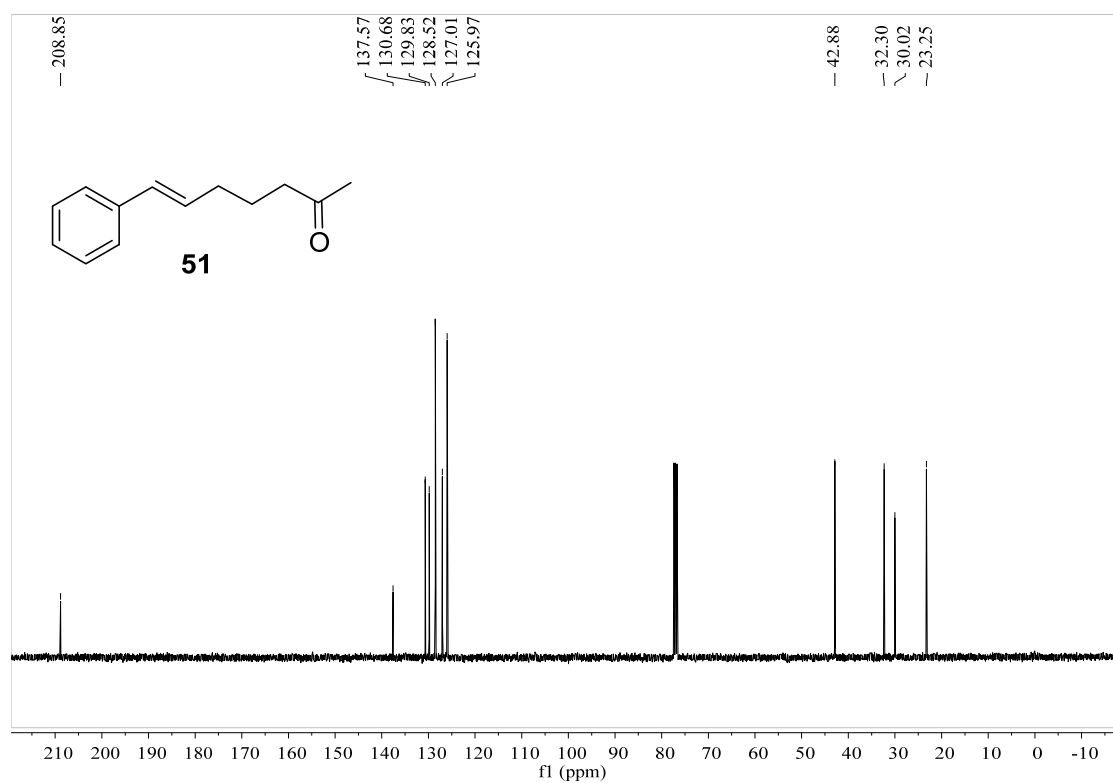


Figure S143. ¹³C NMR spectrum of compound **51**, related to **Figure 3**.

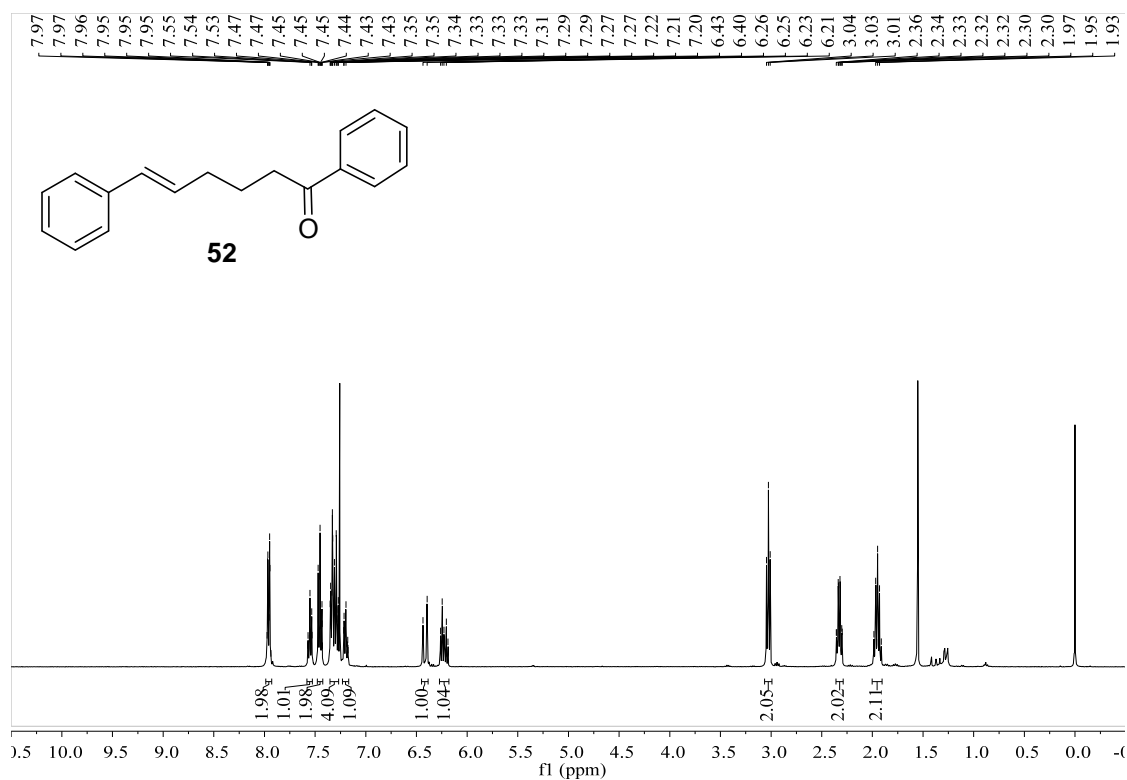


Figure S144. ¹H NMR spectrum of compound **52**, related to Figure 3.

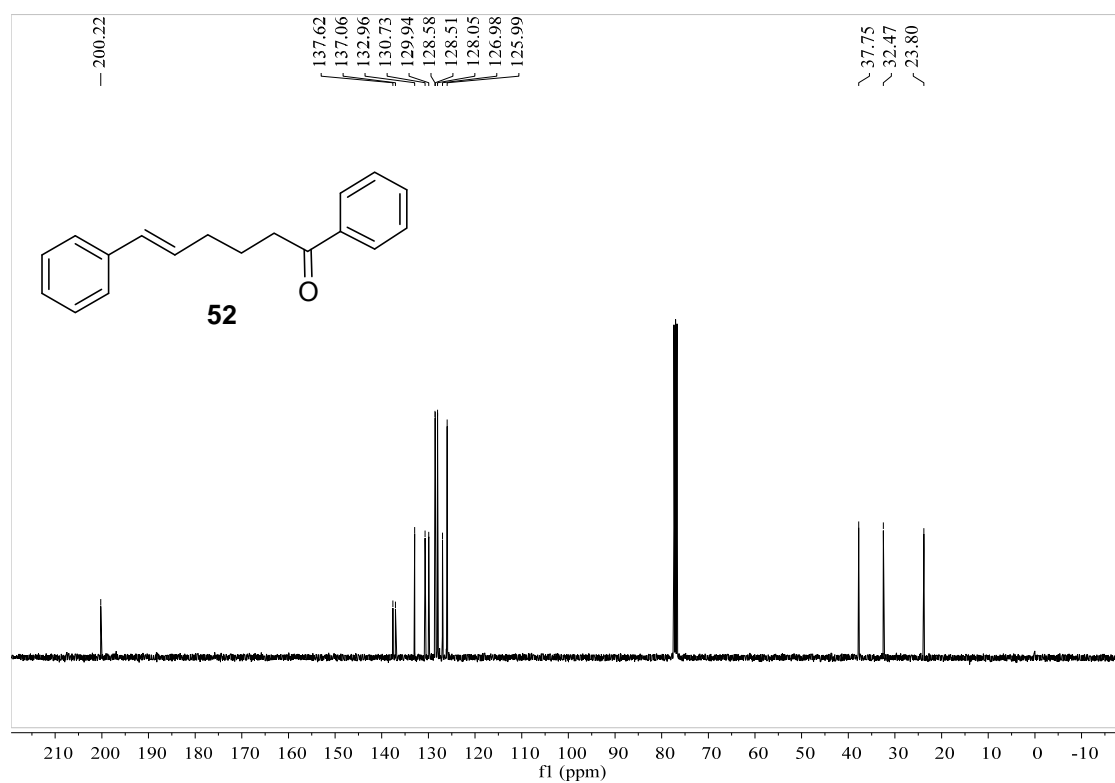


Figure S145. ¹³C NMR spectrum of compound **52**, related to Figure 3.

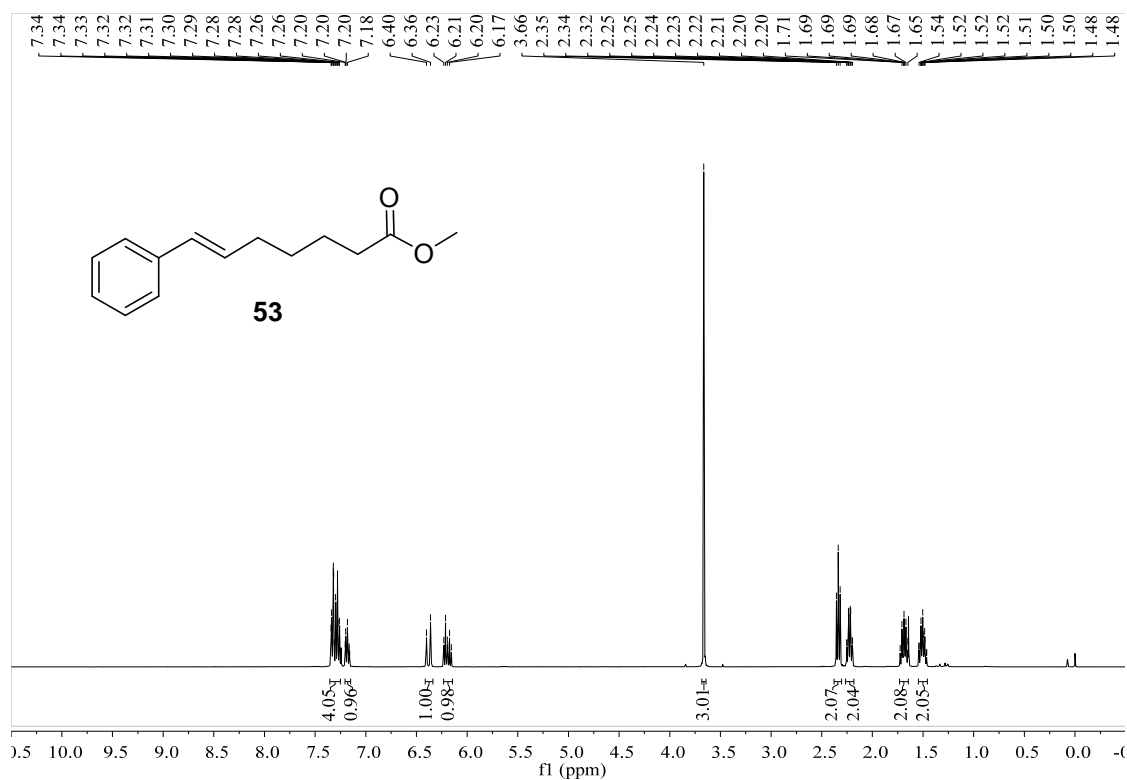


Figure S146. ¹H NMR spectrum of compound **53**, related to **Figure 3**.

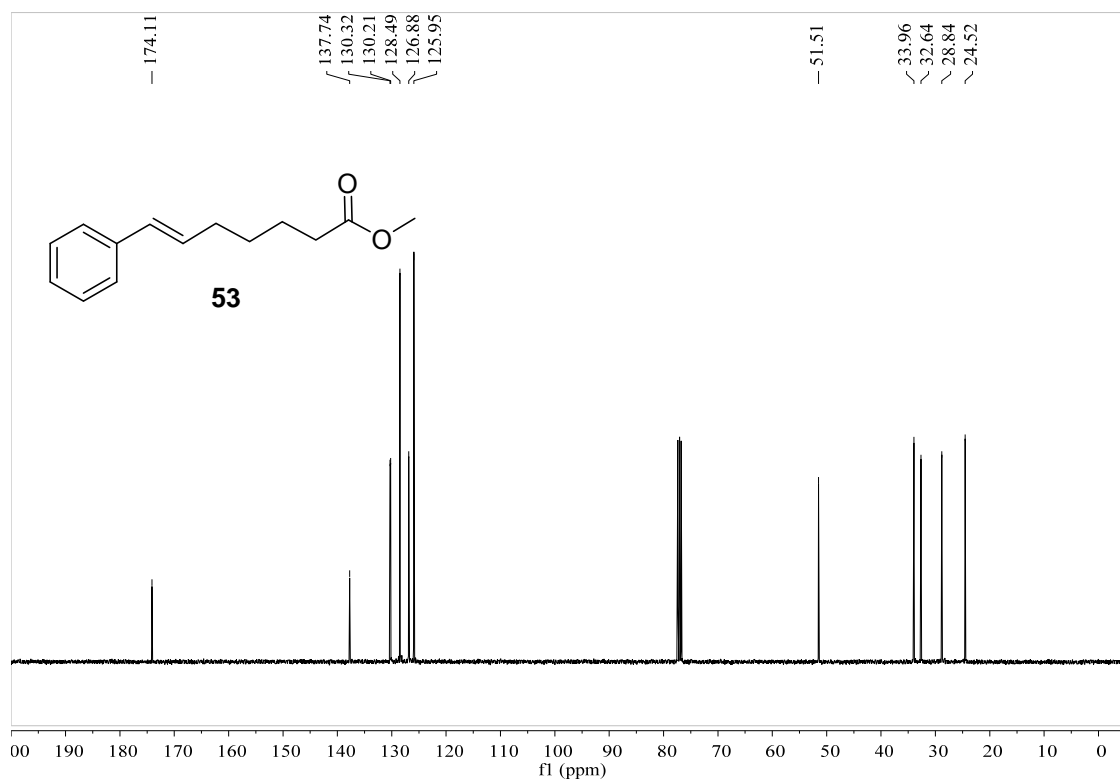


Figure S147. ¹³C NMR spectrum of compound **53**, related to **Figure 3**.

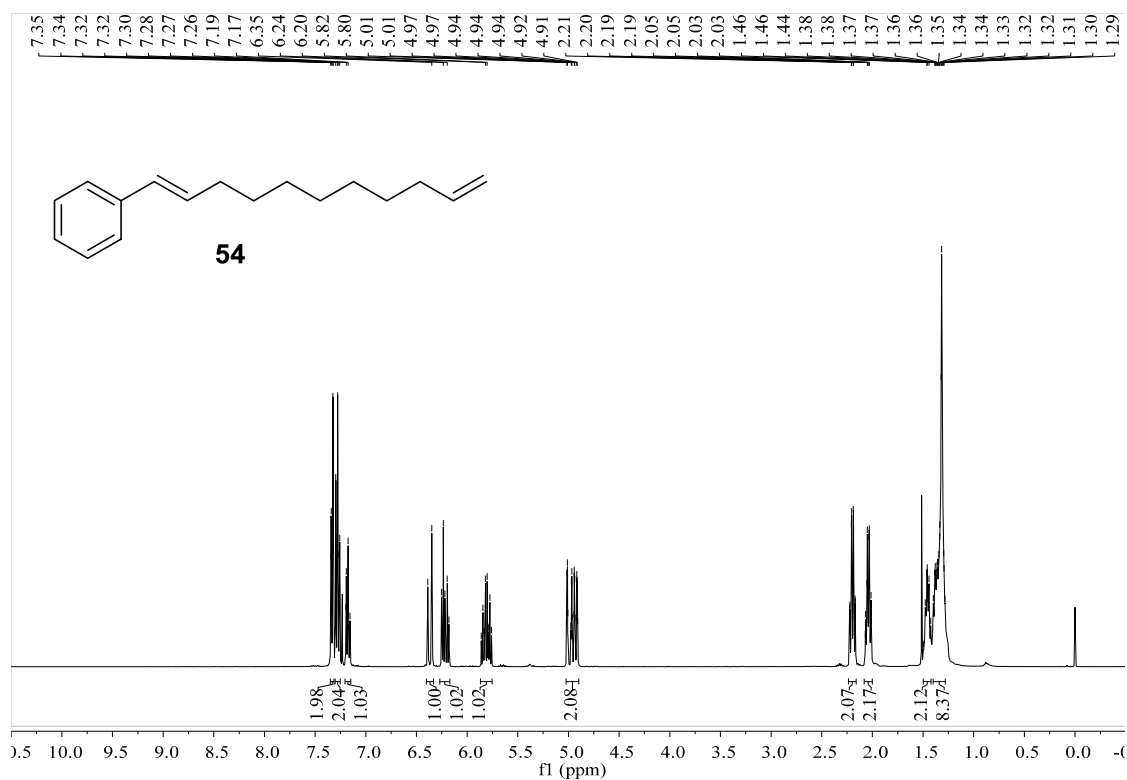


Figure S148. ¹H NMR spectrum of compound **54**, related to Figure 3.

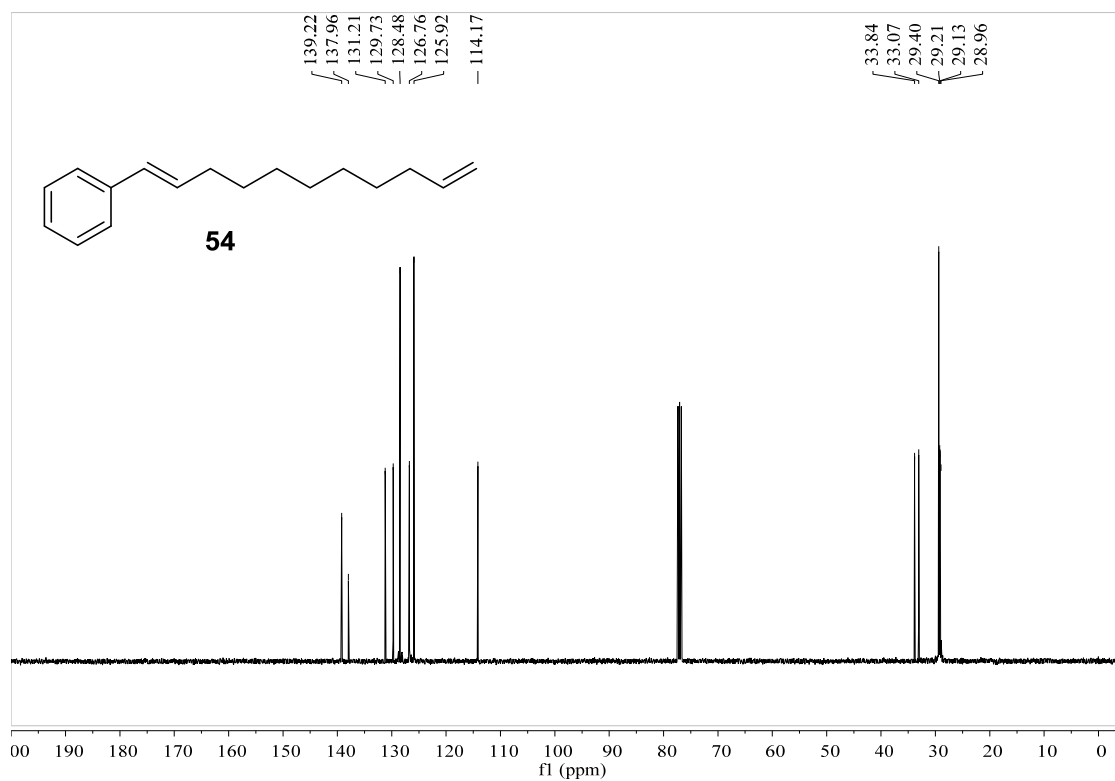


Figure S149. ¹³C NMR spectrum of compound **54**, related to Figure 3.

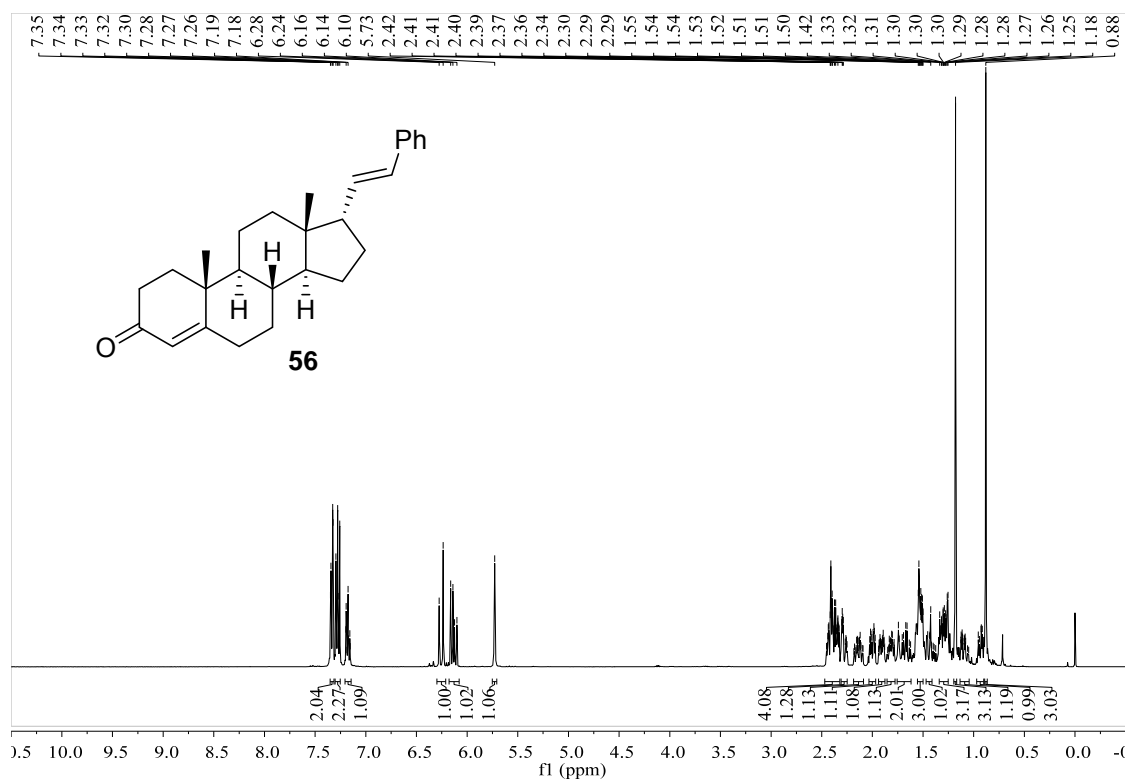


Figure S150. ¹H NMR spectrum of compound **56**, related to **Scheme 2**.

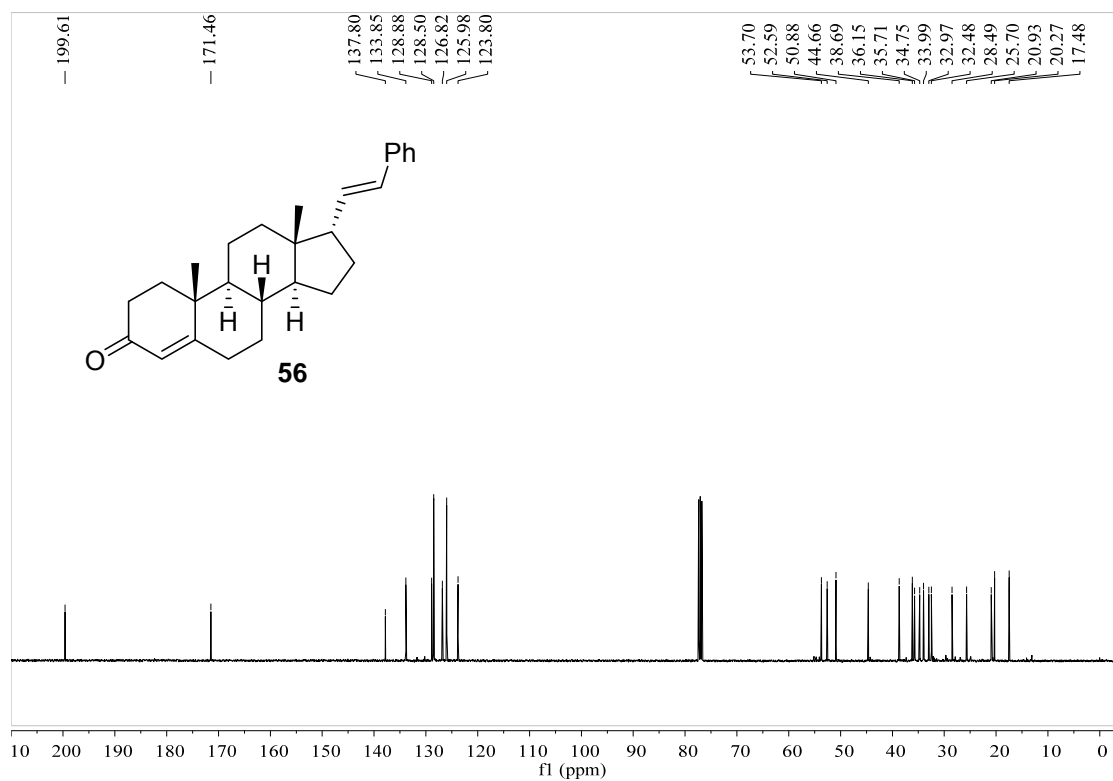
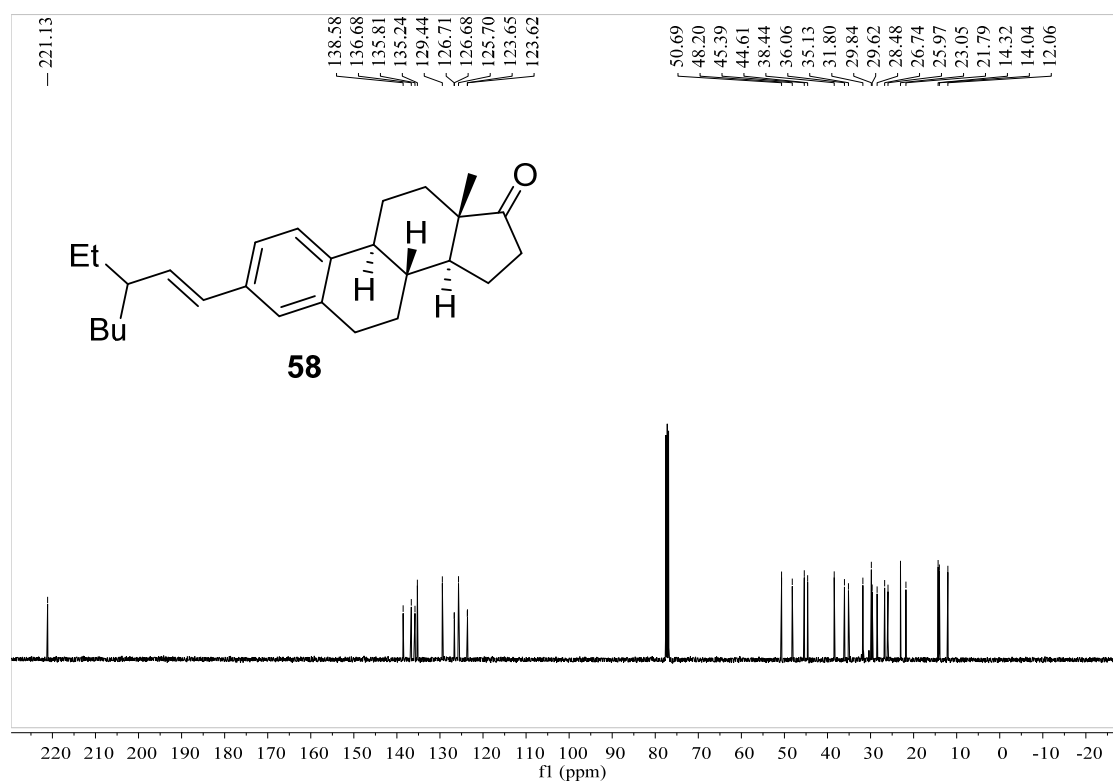
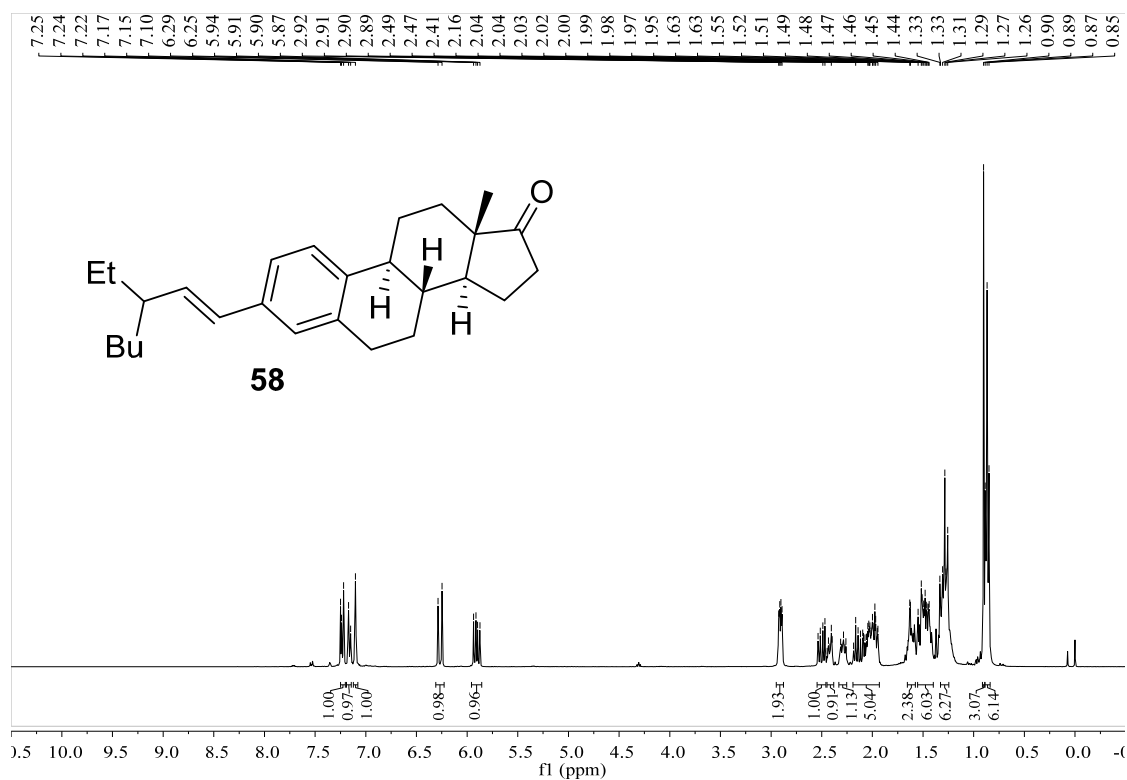


Figure S151. ¹³C NMR spectrum of compound **56**, related to **Scheme 2**.



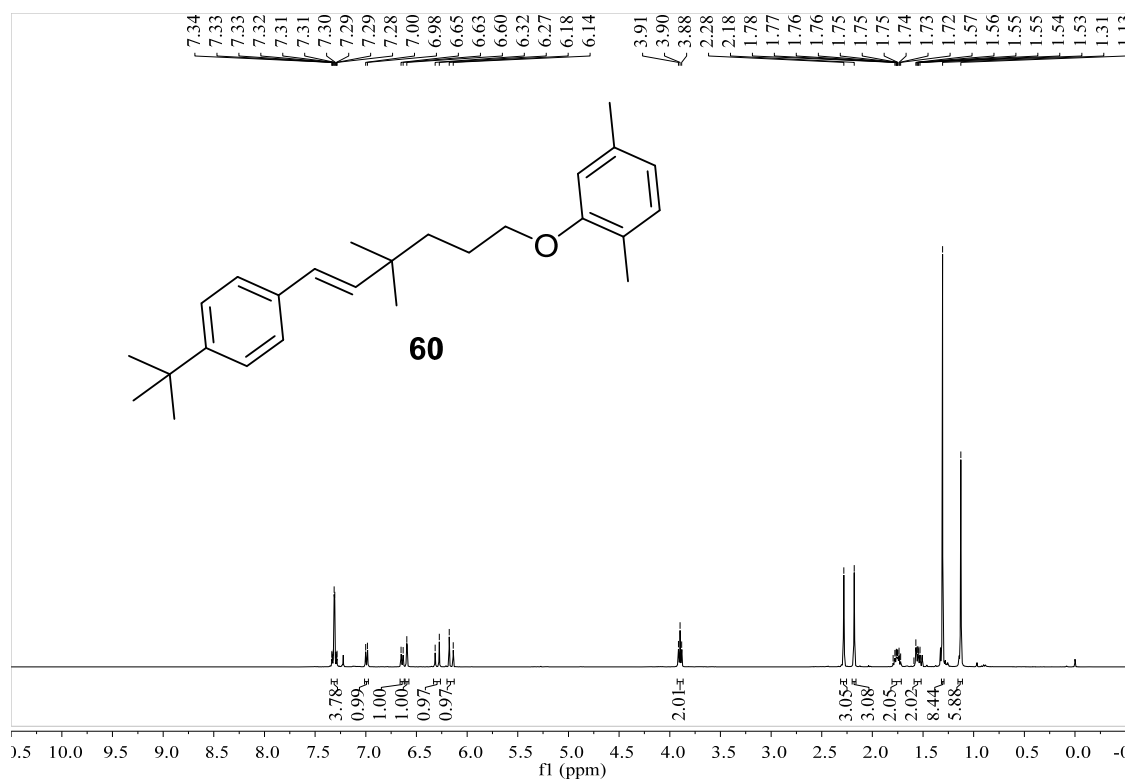


Figure S154. ^1H NMR spectrum of compound **60**, related to Scheme 2.

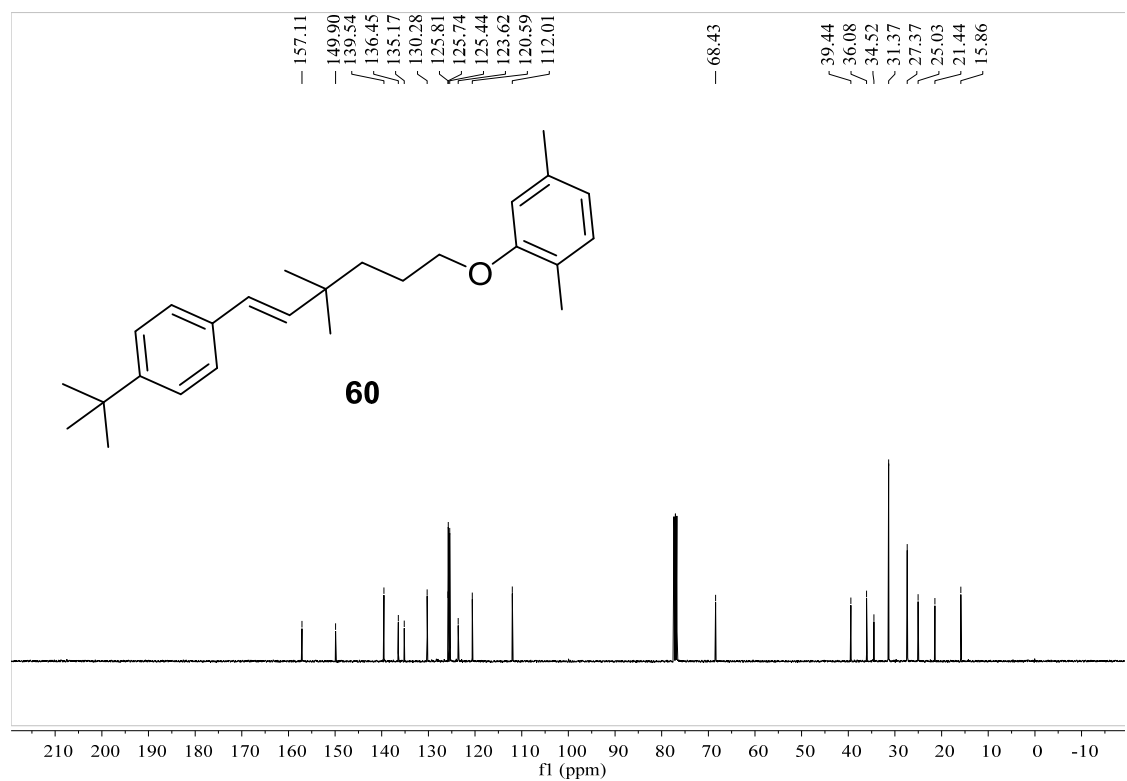


Figure S155. ^{13}C NMR spectrum of compound **60**, related to Scheme 2.

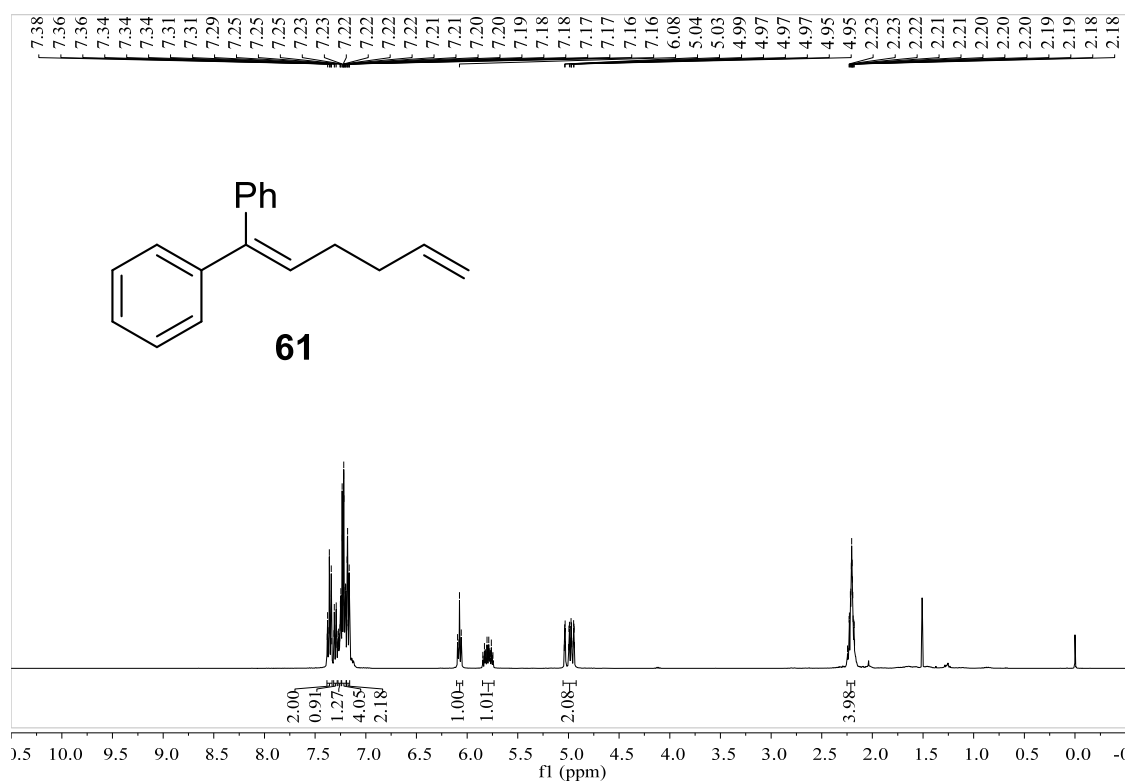


Figure S156. ¹H NMR spectrum of compound **61**, related to Scheme 3A.

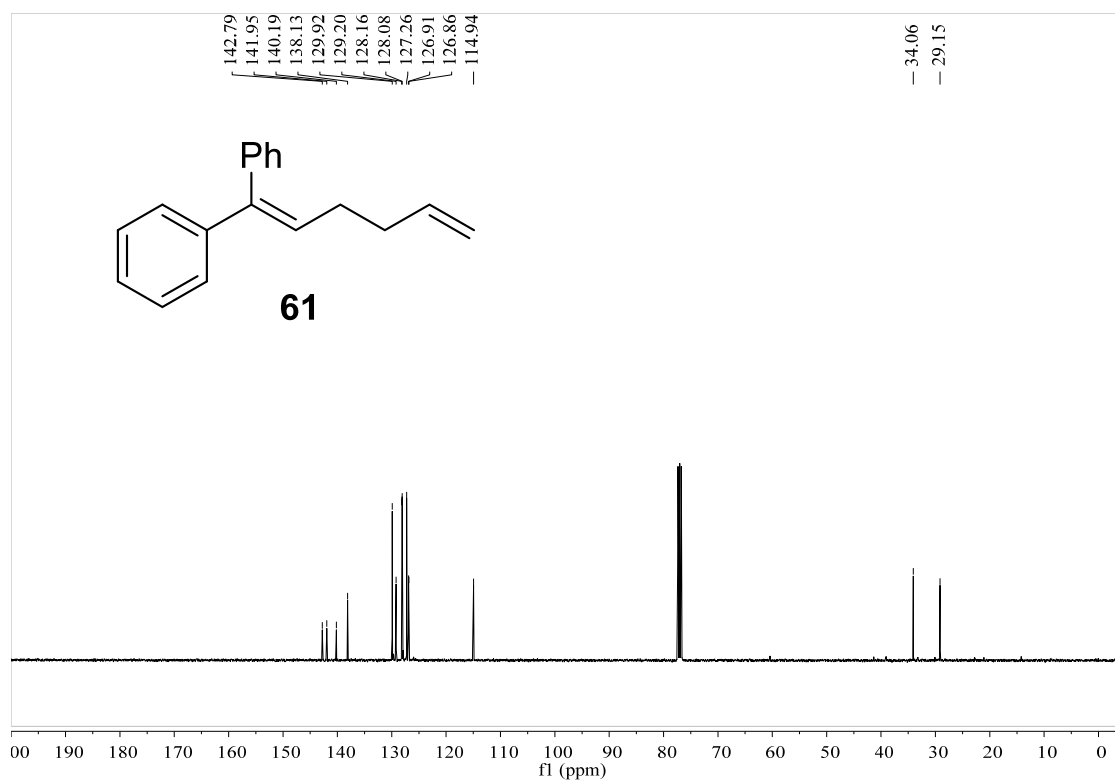


Figure S157. ¹³C NMR spectrum of compound **61**, related to Scheme 3A.

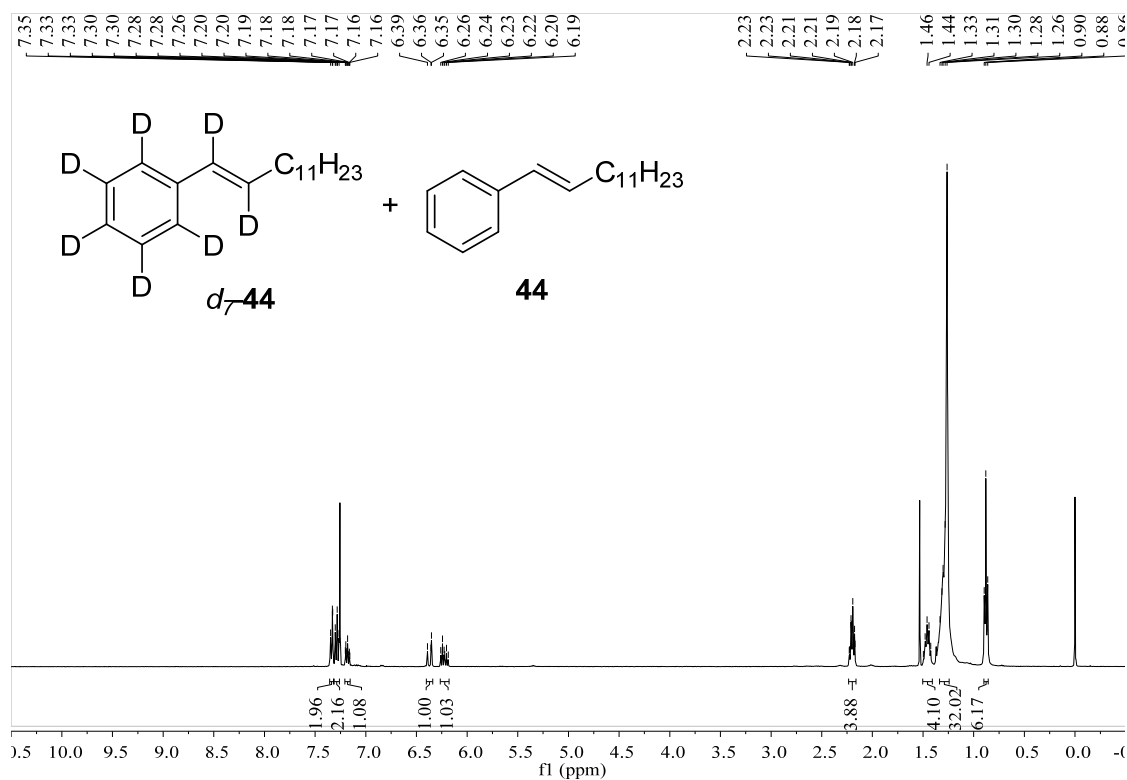


Figure S158. 1H NMR spectrum of compounds d_7-44 and 44 , related to **Scheme 3B**.

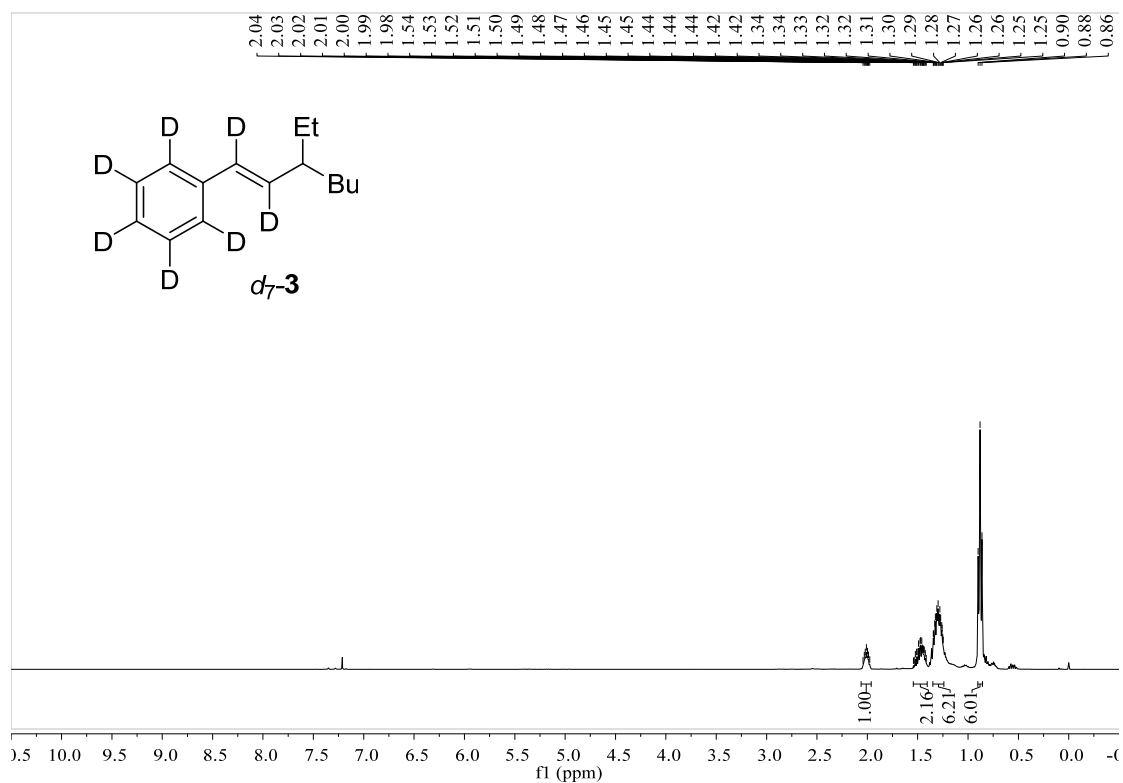


Figure S159. 1H NMR spectrum of compound d_7-3 , related to **Scheme 3B**.

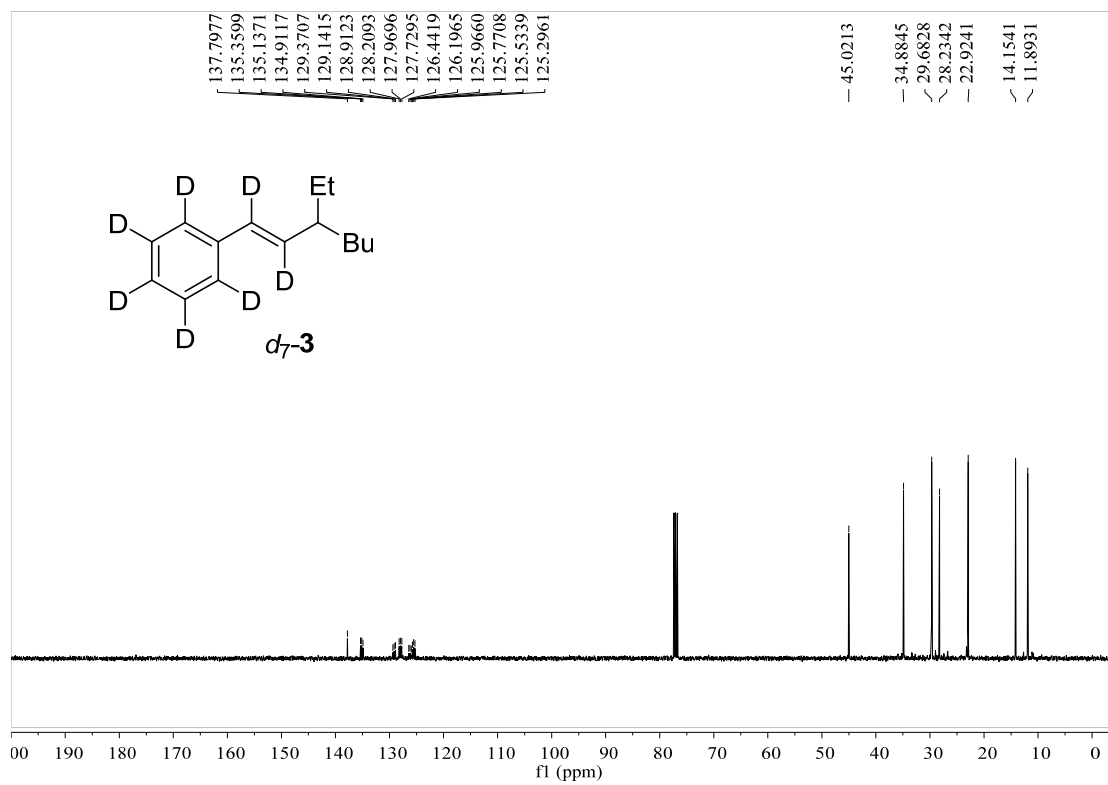


Figure S160. ^{13}C NMR spectrum of compound $d_7\text{-3}$, related to Scheme 3B.

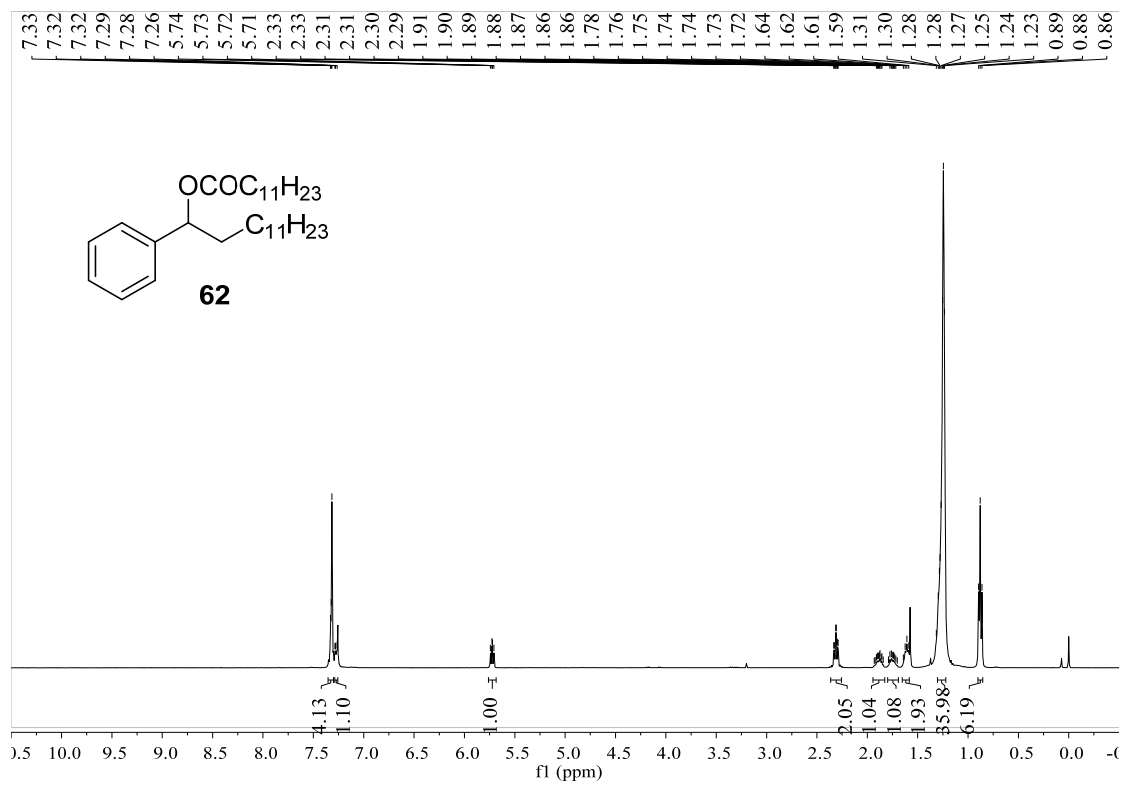


Figure S161. ¹H NMR spectrum of compound **62**, related to **Scheme 3C**.

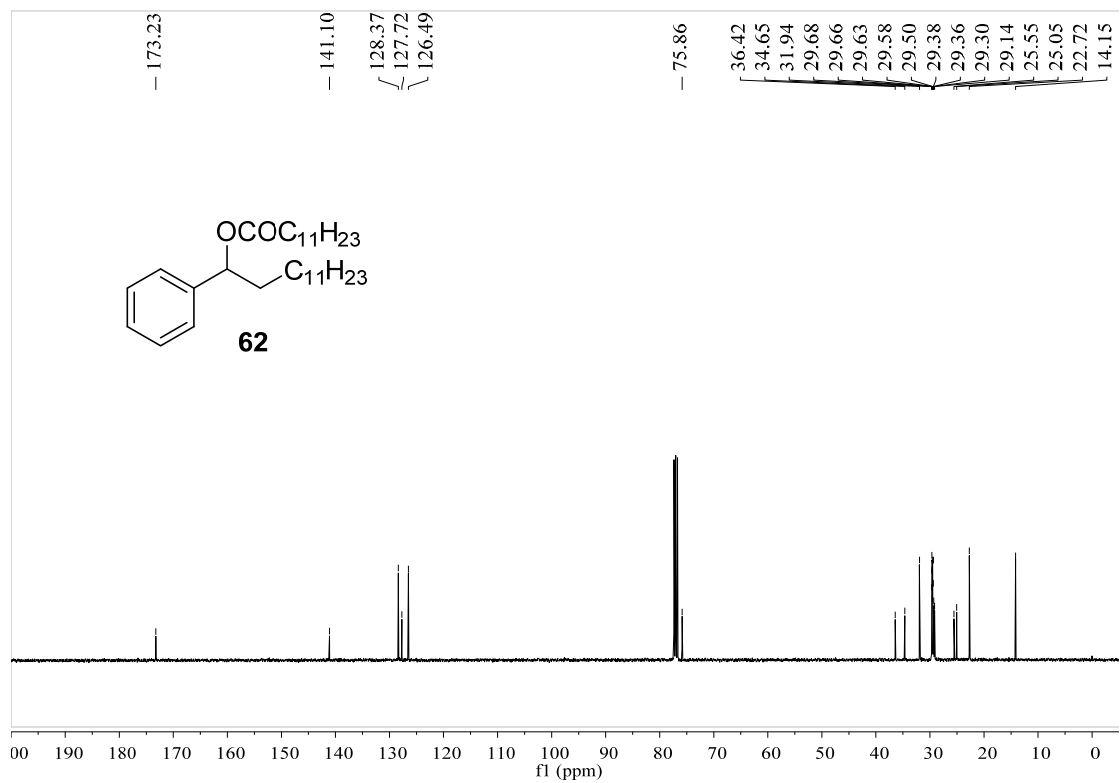


Figure S162. ¹³C NMR spectrum of compound **62**, related to **Scheme 3C**.

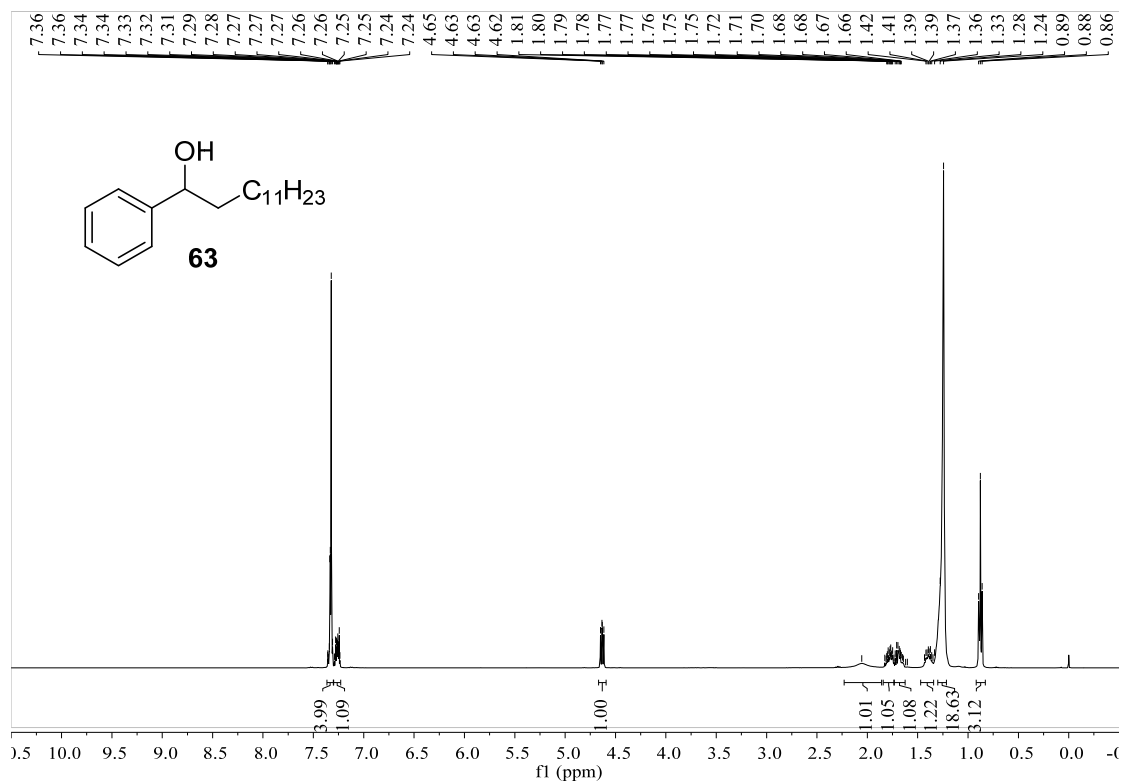


Figure S163. ¹H NMR spectrum of compound **63**, related to **Scheme 3C**.

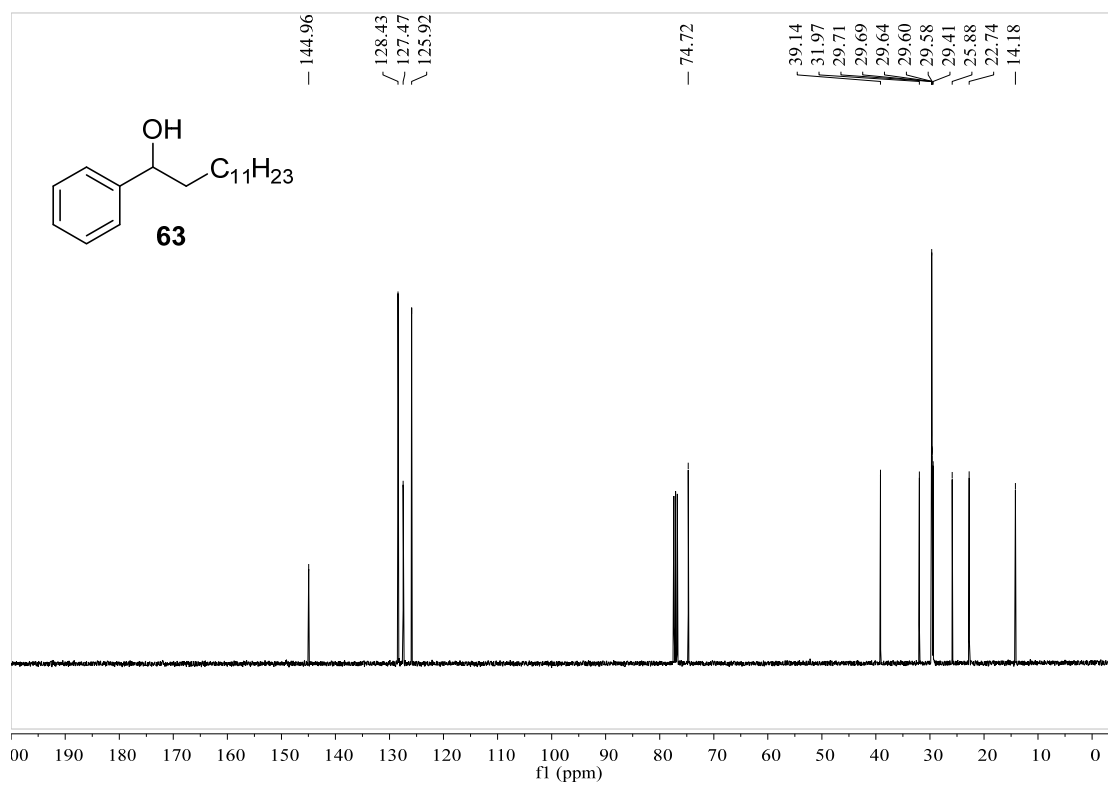
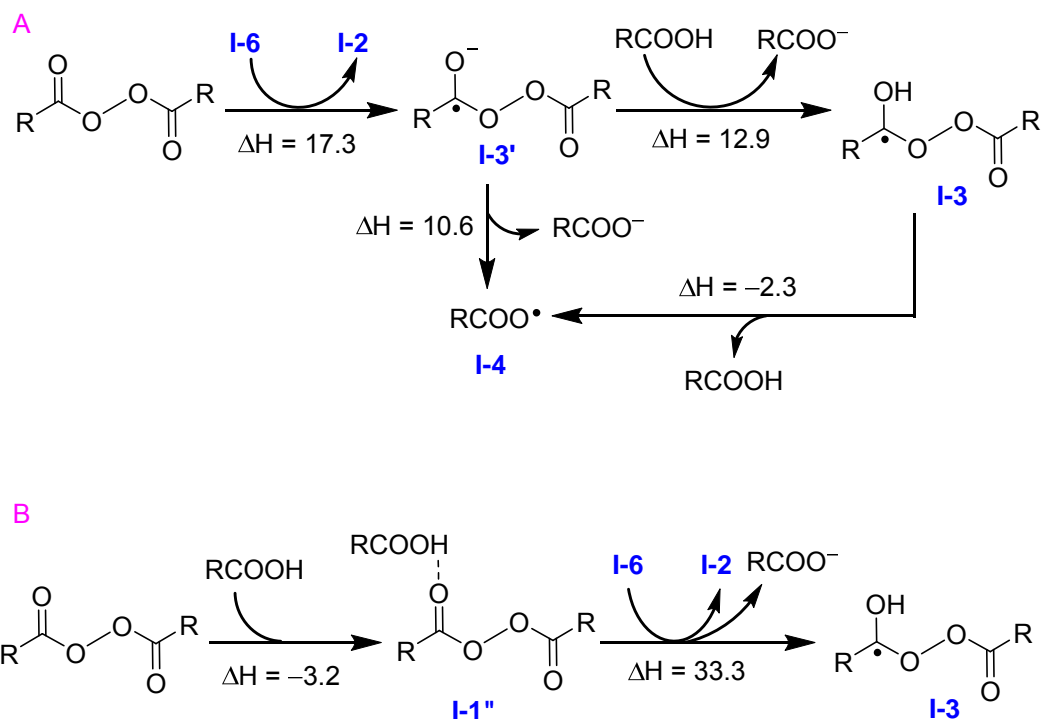


Figure S164. ¹³C NMR spectrum of compound **63**, related to **Scheme 3C**.

Supplemental Schemes

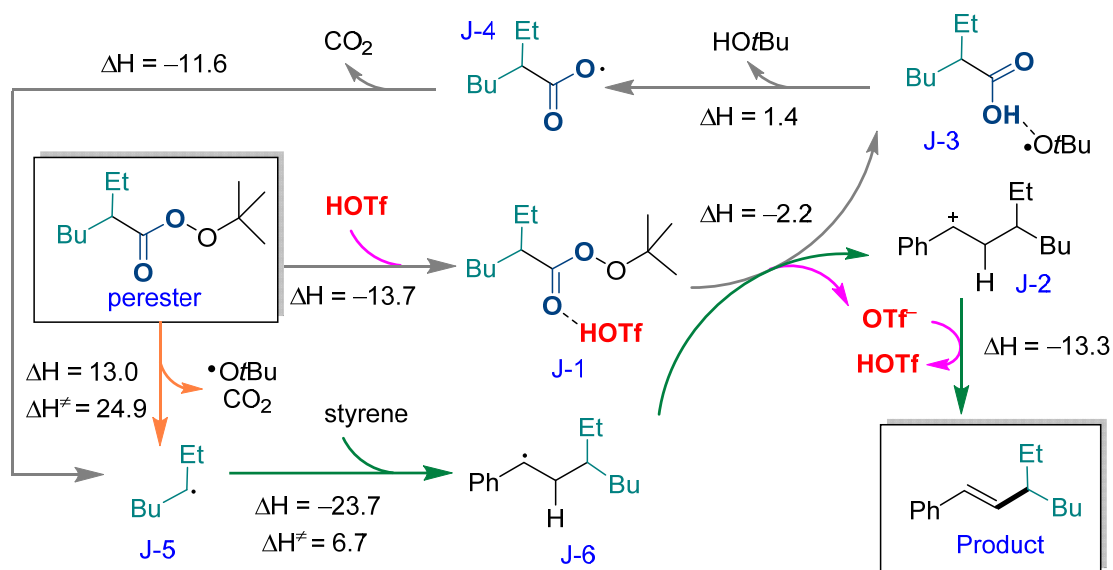
Note: R = *n*-C₅H₁₁ was used for the calculation



Scheme S1. The reaction profiles without HOTf. Related to **Scheme 4**.

(A) Direct electron transfer from **I-6** to peroxide requires a high energy of 17.3 kcal/mol. Protonation of **I-2** by RCOOH to form **I-3** is endothermic of 12.9 kcal/mol. Without acid direct O-O cleavage to form **I-4** is also endothermic of 10.6 kcal/mol. Combined with previous energy requirement of electron transfer the overall reaction energies are more than 27 kcal/mol.

(B) Binding a carboxylic acid RCOOH to peroxide forms a weak hydrogen bond of 3.2 kcal/mol. However, electron transfer process requires high energy of 33.3 kcal/mol. As such, both paths are disfavored compared to the HOTf involved reactions.



Scheme S2. The reaction profile of perester, Related to **Scheme 4**.

The perester species has a similar mechanism catalyzed by HOTf. Before the catalytic cycle the R• radical **J-5** can be formed by homolytic dissociation of the alkyl diacyl peroxide (perester), which is a very slow step with a high barrier of 24.9 kcal/mol. However, this is considered as the *trigger* to invoke the following catalytic cycle. Attack on the styrene substrate by the active species R• radical **J-5** to form a benzyl radical (**J-6**) leads to energies lower by 23.7 kcal/mol with a small barrier of 6.7 kcal/mol, indicating that such reaction is both thermodynamically and kinetically favourable. In the beginning of the catalytic cycle, LPO binding a molecule of HOTf forms a complex **J-1** with a strong hydrogen bonding of 13.7 kcal/mol. This complex oxidizes benzyl radical (**J-6**) to yield a benzyl cation species (**J-2**), a radical (**J-3**) and an OTf⁻ anion, which is exothermic by 2.2 kcal/mol. Meanwhile, the generated OTf⁻ deprotonates **J-1** to yield the product and regenerate acid HOTf with reaction energy of -13.3 kcal/mol. Thus, from the reactions of LPO and **J-6** to the product and **J-3** stepwise electron and proton transfers are promoted by HOTf, which serves as the driving force and proton source for the reaction. Thereby hydrogen transfer of **J-3** leads to RCOO• radical (**J-4**) and tBuOH, which is nearly thermal neutral of 1.4 kcal/mol without any barrier. Subsequently, C-C cleavage of **J-4** is exothermic by 11.6 kcal/mol in energy which releases the active species R• radical (**J-5**) and CO₂ to close the catalytic cycle.

Supplemental Tables

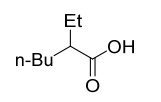
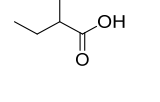
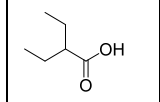
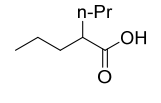
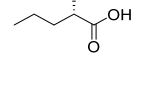
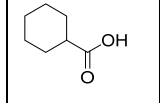
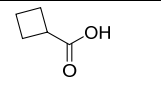
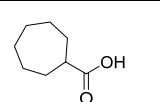
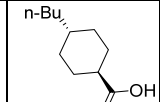
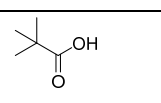
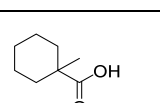
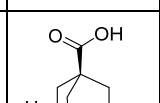
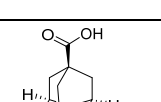
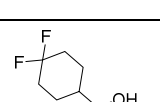
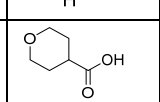
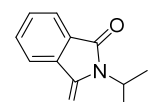
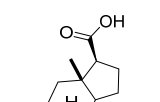

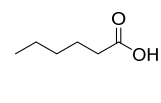
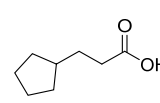
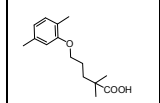
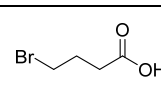
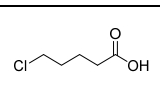
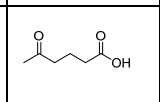
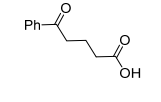
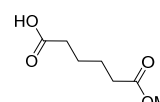
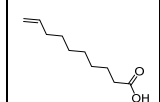
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	Energy-chemical		TCI-chemicals		Energy-chemical
	Bide-pharmatech		Bide-pharmatech		Bide-pharmatech
	Aladdin		Heowns		Energy-chemical
	Energy-chemical		Energy-chemical		Energy-chemical
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Table S1. Sources of acids, related to Figure 2 and Figure 3.


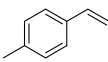



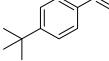
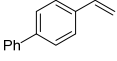
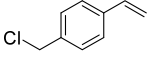
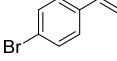
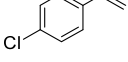
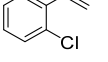
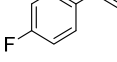
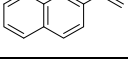
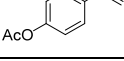
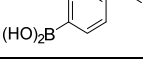
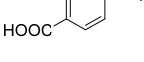
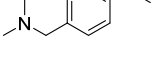
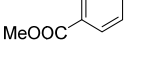
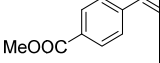


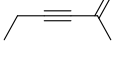
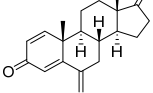
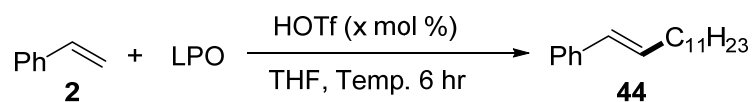
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	Meryer		Energy-chemical		Energy-chemical
	Energy-chemical		Jkchemical		Energy-chemical
	Energy-chemical		Energy-chemical		Energy-chemical
	Sigma-aldrich		Energy-chemical		

Table S2. Sources of alkenes, related to Figure 1, Figure 2 and Figure 3.

structure	yield	structure	yield
	90%		85%
	84%		65%
	84%		73%
	71%		76%
	55%		76%
	75%		70%
	78%		70%
	64%		80%

Table S3. The synthesis of peroxides, related to Figure 3.



Entry	HOTf (x mol %)	THF (y mL)	Temp.	Yield (%) ^b
1	5 mol %	2 mL	90°C	5%
2	10 mol %	2 mL	90°C	10%
3	15 mol %	2 mL	90°C	36%
4	20 mol %	2 mL	90°C	65%
5	40 mol %	2 mL	90°C	60%
6	50 mol %	2 mL	90°C	76% ^c
7	20 mol %	2 mL	100°C	62%
8 ^d	20 mol %	2 mL	80°C	43%
9 ^d	20 mol %	2 mL	70°C	6%
10	20 mol %	1 mL	90°C	76% (73% ^c)
11 ^e	20 mol %	1 mL	90°C	67%
12 ^f	20 mol %	1 mL	90°C	54%
13 ^g	20 mol %	1 mL	90°C	70%
14 ^h	20 mol %	1 mL	90°C	40%

Table S4. Optimizations of reaction conditions with primary aliphatic acid, Related to **Figure 3**.^a

^a**2** (0.5 mmol), LPO (1.0 mmol).

^bYield detected by GC.

^cIsolated product.

^dReaction with 8 hr.

^e**2** (0.5 mmol), LPO (0.75 mmol).

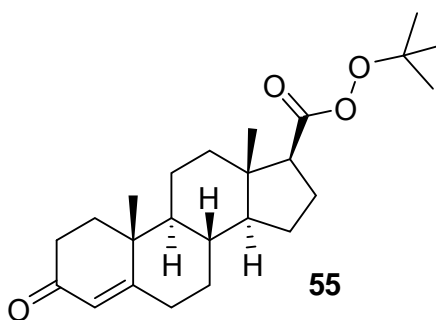
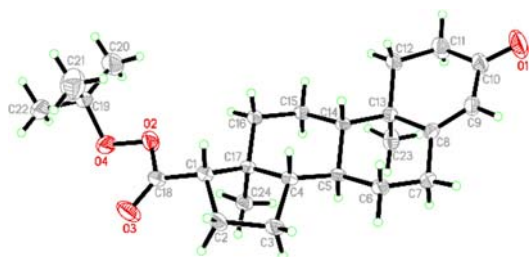
^f**2** (0.5 mmol), LPO (0.5 mmol).

^g**2** (0.6 mmol), LPO (0.5 mmol).

^h**2** (0.75 mmol), LPO (0.5 mmol).

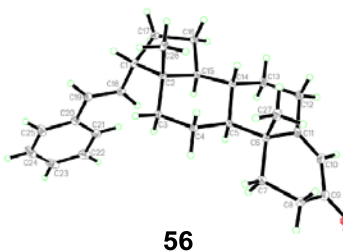
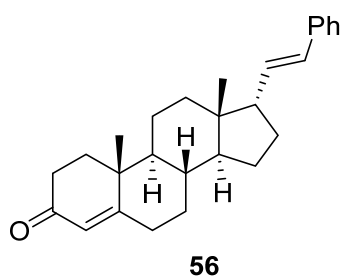
Single Crystal Data of **55** and **56**

Single crystal of **55** and **56** suitable for X-ray diffraction was mounted in Paratone oil onto a glass fiber and frozen under a nitrogen cold stream. The data was collected at 220.0(1) K using a Agilent SuperNova, Dual, Cu at zero, Atlas fitted with Cu K α radiation ($\lambda = 1.54184 \text{ \AA}$). Data collection and unit cell refinement were executed by using CrysAlisPro software. Data processing and absorption correction, giving minimum and maximum transmission factors, were accomplished with CrysAlisPro. The structure was solved with the SHELXT-2014 and refined with the SHELXL-2014 using Least Squares minimisation. All non-hydrogen atoms were refined with anisotropic displacement parameters. All carbon bound hydrogen atom positions were determined by geometry and refined by a riding model. CCDC 1477011 and CCDC 1476738 for **55** and **56** contain the supplementary crystallographic data. Crystal data and structure refinements of **55** and **56** are listed in Table S5 and Table S6. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.



Identification code	55
Empirical formula	C ₂₄ H ₃₆ O ₄
Formula weight	388.53
Temperature	220.0(1) K
Wavelength	1.54184 Å
Crystal system	Orthorhombic
Space group	P 21 21 21
Unit cell dimensions	a = 6.16790(10) Å b = 12.5681(3) Å c = 29.0822(7) Å
Volume	2254.42(8) Å ³
Z	4
Density (calculated)	1.145 Mg/m ³
Absorption coefficient	0.603 mm ⁻¹
F(000)	848
Crystal size	0.220 x 0.200 x 0.170 mm ³
Theta range for data collection	3.831 to 73.663°.
Index ranges	-5 ≤ h ≤ 7, -9 ≤ k ≤ 15, -23 ≤ l ≤ 35
Reflections collected	7382
Independent reflections	3937 [R(int) = 0.0387]
Completeness to theta = 67.684°	99.3 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.00000 and 0.60854
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	3937 / 6 / 258
Goodness-of-fit on F ²	1.023
Final R indices [I > 2σ(I)]	R1 = 0.0531, wR2 = 0.1363
R indices (all data)	R1 = 0.0649, wR2 = 0.1499
Absolute structure parameter	0.0(3)
Extinction coefficient	n/a
Largest diff. peak and hole	0.276 and -0.212 e.Å ⁻³

Table S5. Crystal data and structure refinement for **55**, Related to **Scheme 2**.



Identification code	56
Empirical formula	C ₂₇ H ₃₄ O
Formula weight	374.54
Temperature	100.0(2) K
Wavelength	1.54184 Å
Crystal system	Orthorhombic
Space group	P 21 21 21
Unit cell dimensions	a = 6.23490(10) Å b = 28.7978(4) Å c = 11.7959(2) Å
Volume	2117.97(6) Å ³
Z	4
Density (calculated)	1.175 Mg/m ³
Absorption coefficient	0.520 mm ⁻¹
F(000)	816
Crystal size	0.200 x 0.180 x 0.150 mm ³
Theta range for data collection	4.050 to 73.331°.
Index ranges	-7<=h<=2, -31<=k<=35, -14<=l<=7
Reflections collected	5630
Independent reflections	3761 [R(int) = 0.0141]
Completeness to theta = 67.684°	99.9 %
Absorption correction	Semi-empirical from equivalents
Max. and min. transmission	1.00000 and 0.88083
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	3761 / 0 / 256
Goodness-of-fit on F ²	1.025
Final R indices [I>2sigma(I)]	R1 = 0.0291, wR2 = 0.0733
R indices (all data)	R1 = 0.0302, wR2 = 0.0742
Absolute structure parameter	-0.29(15)
Extinction coefficient	0.0041(3)
Largest diff. peak and hole	0.252 and -0.140 e.Å ⁻³

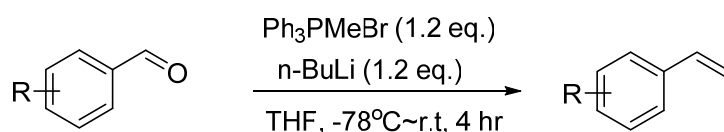
Table S6. Crystal data and structure refinement for **56**, Related to **Scheme 2**.

Transparent Methods

All reactions were carried out under an atmosphere of nitrogen in flame-dried glassware with magnetic stirring unless otherwise indicated. Commercially obtained reagents were used as received. Solvents were dried by Innovative Technology Solvent Purification System. Liquids and solutions were transferred via syringe. All reactions were monitored by thin-layer chromatography. GC-MS data were recorded on Thermo ISQ QD. ^1H , ^{13}C and ^{19}F NMR spectra were recorded on Bruker-BioSpin AVANCE III HD. Data for ^1H NMR spectra are reported relative to chloroform as an internal standard (7.26 ppm) and are reported as follows: chemical shift (ppm), multiplicity, coupling constant (Hz), and integration. Data for ^{13}C NMR spectra are reported relative to chloroform as an internal standard (77.23 ppm) and are reported in terms of chemical shift (ppm). HRMS data were recorded on Waters Micromass GCT Premier or Thermo Fisher Scientific LTQ FTICR-MS.

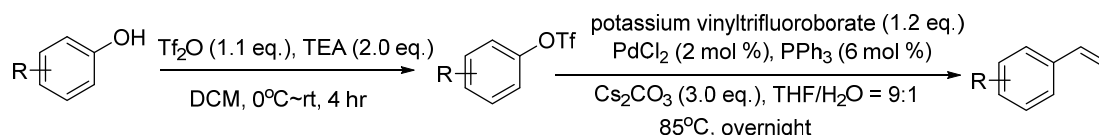
Experimental procedures for synthesis of materials

Procedure A for the synthesis of alkenes



General procedure (Haubenreisser et al., 2016): The reaction vessel was charged with phosphonium salt (1.2 equiv) in dry THF. To the stirred mixture, *n*-butyl lithium (1.2 equiv) was added under N_2 atmosphere at -78°C . The mixture was stirred at 0°C for 5 mins and then substituted aldehyde (1.0 equiv.) in dry THF was added dropwise in over 15 min. After stirring at rt for 4 hr, the mixture was quenched with saturated NH_4Cl , then extracted three times with dichloromethane and water. The combined organic layers were dried over anhydrous sodium sulfate, concentrated and purified by flash column chromatography afford the desired product.

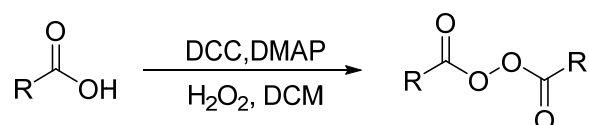
Procedure B for the synthesis of alkenes



General procedure (Huang and Doyle, 2012): A flask was flame dried and charged with phenol (1.0 equiv), dichloromethane, and Et_3N (2.0 equiv). The mixture was cooled in a 0°C ice-water bath, and Tf_2O (1.1 equiv) was added dropwise. The mixture was allowed to warm up to room temperature and stirred at room temperature under argon for 5 hr. The resulting brown mixture was diluted with dichloromethane, washed with sat. NH_4Cl , and the aqueous layer was

extracted with dichloromethane. The combined organic layers were dried over MgSO₄, and the filtrate was concentrated. The crude was purified with column chromatography to afford triflate. A threaded tube was charged with triflate (1.0 equiv), potassium vinyltrifluoroborate (1.2 equiv), PdCl₂ (2 mol %), PPh₃ (6 mol %), Cs₂CO₃ (3.0 equiv) were added, then THF and water were added under N₂ atmosphere. The mixture was stirred at 85°C for overnight. The resulting dark brown mixture was allowed to cool to room temperature, diluted with dichloromethane, and washed with water. The aqueous layer was extracted with dichloromethane. The combined organic layers were dried over MgSO₄, and the filtrate was concentrated. The crude was purified with column chromatography to afford the desired product.

Procedure C for the synthesis of diacyl peroxides



General procedure (Jian et al., 2017): A solution of DMAP (10 mol %), 30% hydrogen peroxide (1.2 equiv), and acid in CH₂Cl₂ was cooled to -5°C for about 10 min. Then DCC (1.2 equiv) was added. Then the mixture was stirred at -5°C to room temperature for 1.5 ~4 hr. The solution was concentrated on a rotary evaporator under vacuum at 10~15°C and the residue was chromatographed on silica gel to give the diacyl peroxide.

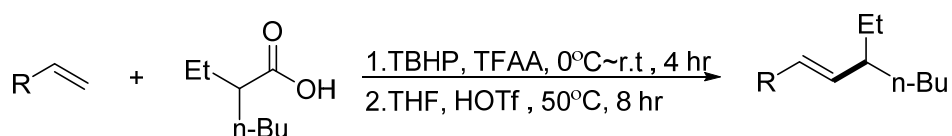
Procedure D for the synthesis of peresters



General procedure (Zhu et al., 2017): A solution of DMAP (10 mol %), TBHP (aqueous solution, 1.2 equiv), and acid in CH₂Cl₂ was cooled to -5°C for about 10 min. Then DCC (1.2 equiv) was added. Then the mixture was stirred at -5°C to room temperature for 4 hr. The solution was concentrated on a rotary evaporator under vacuum at 20~25°C and the residue was chromatographed on silica gel to give the perester.

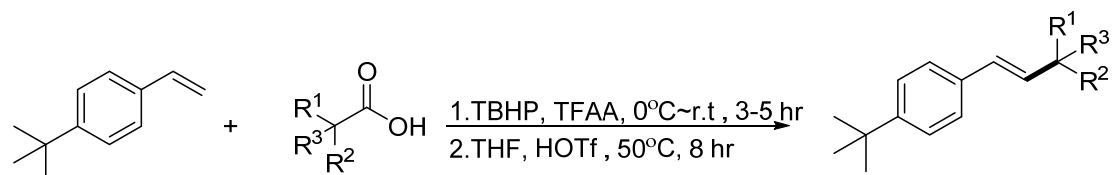
General procedure for the alkyl-Heck-type reaction

Procedure E for the alkyl-Heck-type reaction of alkenes



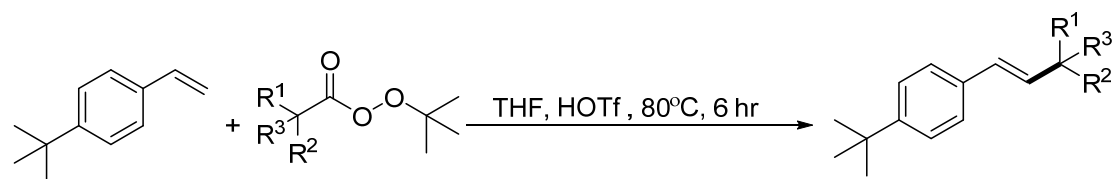
To a flame-dried Schlenk tube were added 2-ethylhexanoic acid (1.5 mmol, 3.0 equiv), TBHP (1.5 mmol, 3.0 equiv, in decane) and TFAA (2.0 mmol, 4.0 equiv) at 0°C under the atmosphere of nitrogen. The mixture was then stirred at rt for 4 hours. After completion, THF (2 mL), alkene (0.5 mmol, 1.0 equiv) and HOTf (0.05 mmol, 10 mol %) were added into the Schlenk tube under the atmosphere of nitrogen. The mixture was then stirred at 50°C for 8 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product.

Procedure F for the alkyl-Heck-type reaction of secondary and tertiary aliphatic acids



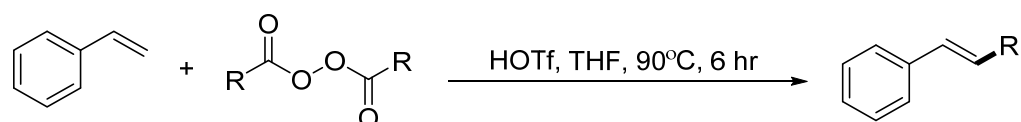
General procedure: To a flame-dried Schlenk tube were added acid (1.5 mmol, 3.0 equiv), TBHP (1.5 mmol, 3.0 equiv, in decane) and TFAA (2.0 mmol, 4.0 equiv) at 0°C under the atmosphere of nitrogen. The mixture was then stirred at rt for 3-5 hours. After completion, THF (2 mL), 4-*tert*-butylstyrene (0.5 mmol, 1.0 equiv) and HOTf (0.05 mmol, 10 mol %) were added into the Schlenk tube under the atmosphere of nitrogen. The mixture was then stirred at 50°C for 8 hours. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product.

Procedure G for the alkyl-Heck-type reaction with peresters



To a flame-dried Schlenk tube were added THF (2 mL), 4-*tert*-butylstyrene (0.5 mmol, 1.0 equiv), perester (1.25 mmol, 2.5 equiv) and HOTf (0.1 mmol, 20 mol %) under the atmosphere of nitrogen. The mixture was then stirred at 80°C for 6 hours. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product.

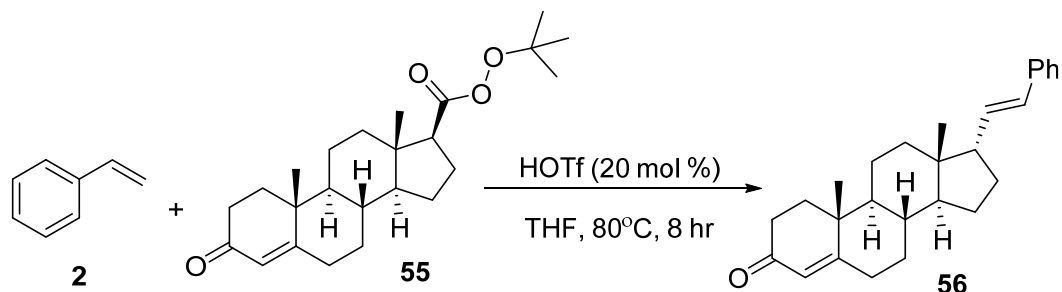
Procedure H for the alkyl-Heck-type reaction with diacyl peroxides



To a flame-dried Schlenk tube were added THF (1 mL), styrene (0.5 mmol, 1.0 equiv) diacyl peroxides (1.0 mmol, 2.0 equiv) and HOTf (0.1 mmol, 20 mol %) under the atmosphere of nitrogen. The mixture was then stirred at 90°C for 6 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product.

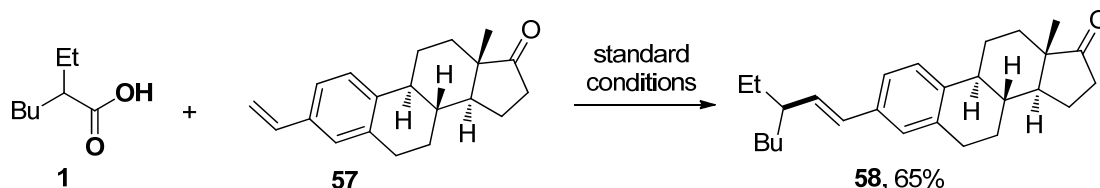
Experimental procedures for synthetic applications

(i)



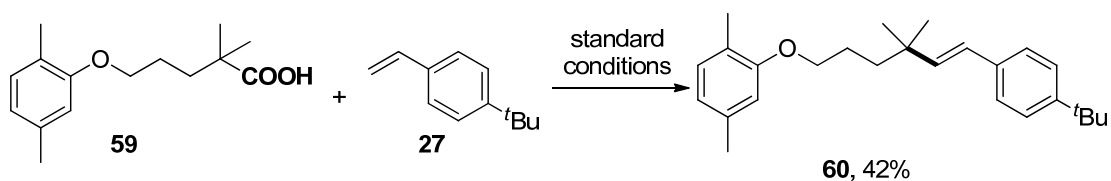
To a flame-dried Schlenk tube were added THF (2 mL), styrene **2** (0.5 mmol, 1.0 equiv) perester **55** (1.25 mmol, 2.5 equiv) and HOTf (0.1 mmol, 20 mol %) under the atmosphere of nitrogen. The mixture was then stirred at 80°C for 8 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product **56**.

(ii)



To a flame-dried Schlenk tube were added acid **1** (1.5 mmol, 3.0 equiv), TBHP (1.5 mmol, 3.0 equiv) and TFAA (2.0 mmol, 4.0 equiv) at 0°C under the atmosphere of nitrogen. The mixture was then stirred at rt for 5 hr. After completion, THF (2 mL), alkene **57** (0.5 mmol, 1.0 equiv) and HOTf (0.05 mmol, 10 mol %) were added into the Schlenk tube under the atmosphere of nitrogen. The mixture was then stirred at 50°C for 8 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product **58**.

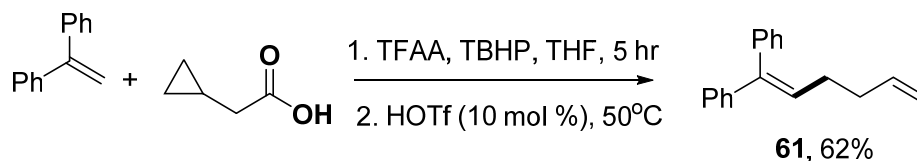
(iii)



To a flame-dried Schlenk tube were added acid **59** (1.5 mmol, 3.0 equiv), TBHP (1.5 mmol, 3.0 equiv) and TFAA (2.0 mmol, 4.0 equiv) at 0°C under the atmosphere of nitrogen. The mixture was then stirred at rt for 5 hr. After completion THF (2 mL), 4-*tert*-butylstyrene **27** (0.5 mmol, 1.0 equiv) and HOTf (0.05 mmol, 10 mol %) were added into the Schlenk tube under the atmosphere of nitrogen. The mixture was then stirred at 50°C for 8 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product **60**.

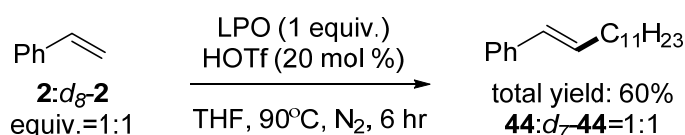
Experimental procedures for preliminary mechanistic studies

Radical clock experiment

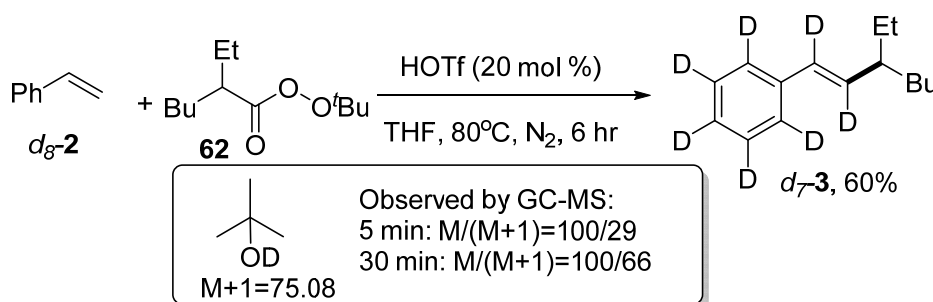


To a flame-dried Schlenk tube were added acid (1.5 mmol, 3.0 equiv), TBHP (1.5 mmol, 3.0 equiv) and TFAA (2.0 mmol, 4.0 equiv) at 0°C under the atmosphere of nitrogen. The mixture was then stirred at rt for 5 hr. After completion, THF (2 mL), styrene (0.5 mmol, 1.0 equiv) and HOTf (0.05 mmol, 10 mol %) were added into the Schlenk tube under the atmosphere of nitrogen. The mixture was then stirred at 50°C for 8 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product **61**.

Deuterium labeling experiment

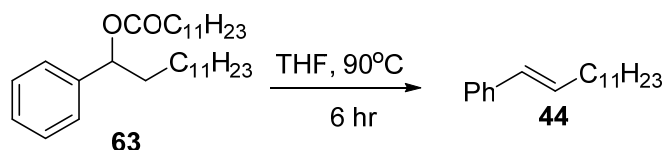


To a flame-dried Schlenk tube were added THF (1 mL), **2** (1.0 mmol, 1.0 equiv), d_8 -**2** (1.0 mmol, 1.0 equiv), LPO (0.5 mmol, 0.5 equiv) and HOTf (0.1 mmol, 20 mol %) under the atmosphere of nitrogen. The mixture was then stirred at 90°C for 6 hr. After completion detected by TLC, the solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired product **44** and d_7 -**44**.



To a flame-dried Schlenk tube were added THF (2 mL), d_8 -**2** (0.50 mmol, 1.0 equiv), **62** (1.25 mmol, 2.5 equiv) and HOTf (0.1 mmol, 20 mol %) under the atmosphere of nitrogen. The mixture was then stirred at 80°C. The reaction mixture was tested by GC-MS after 5 minutes and 30 minutes. The deuterated side-products d (OD)-butanol was detected by GC-MS. The mixture was then stirred at 80°C for 6 hr. After the reaction mixture was cooled to ambient temperature. The solvent was removed by rotary evaporation under vacuum, and the residue was chromatographed on silica gel to give the desired d_7 -**3**.

Exclusion of possible intermediates



0%, **63** was recovered in 92% yield

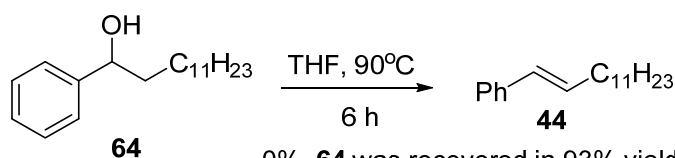
0%, C₁₁H₂₃COOH (1 equiv), **63** was recovered in 93% yield

91%, HOTf (20 mol %).

Ester **63** (Ge et al., 2017): (0.2 mmol) was charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6 hr. No reaction was observed and the ester **63** was recovered in 92% yield.

Ester **63** (0.2 mmol) and C₁₁H₂₃COOH (1 equiv) were charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6 hr. No reaction was observed and the ester **63** was recovered in 93% yield.

Ester **63** (0.2 mmol) and HOTf (20 mol %) were charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6hr. The reaction mixture was diluted with DCM (30 mL) and washed with saturated aq. NaHCO₃. The organic layer was dried (anhydrous Na₂SO₄), filtered, and concentrated in vacuo. The residue was chromatographed on silica gel affording product **44** in 91% yield.



0%, **64** was recovered in 93% yield

0%, C₁₁H₂₃COOH (1 equiv), **64** was recovered in 93% yield

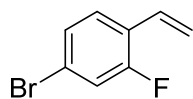
87%, HOTf(20 mol %, 6 hr)

Benzyl alcohol **64** (Jian et al., 2017) (0.2 mmol) was charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6 hr. No reaction was observed and the alcohol **64** was recovered in 93% yield.

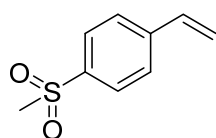
Benzyl alcohol **64** (0.2 mmol) and C₁₁H₂₃COOH (1 equiv) were charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6hr. No reaction was observed and the alcohol **64** was recovered in 93% yield.

Benzyl alcohol **64** (0.2 mmol) and HOTf (20 mol %) were charged in anhydrous THF (0.5 mL) at 90°C and then stirred for 6hr. The reaction mixture was diluted with DCM (30 mL) and washed with saturated aq. NaHCO₃. The organic layer was dried (anhydrous Na₂SO₄), filtered, and concentrated in vacuo. The residue was chromatographed on silica gel affording product **44** in 87% yield.

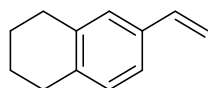
Characterization of all compounds



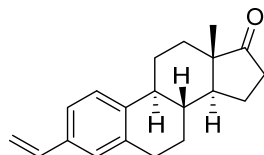
Following procedure A for the synthesis of alkenes, 4-bromo-2-fluoro-1-vinylbenzene (related to **Figure 1**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 (t, J = 8.3 Hz, 1H), 7.26 – 7.19 (m, 2H), 6.79 (dd, J = 17.7, 11.2 Hz, 1H), 5.81 (d, J = 17.7 Hz, 1H), 5.40 (d, J = 11.2 Hz, 1H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 159.92 (d, J = 254.0 Hz), 128.45 (d, J = 3.6 Hz), 128.11 (d, J = 4.4 Hz), 127.43 (d, J = 3.7 Hz), 124.49 (d, J = 12.4 Hz), 121.34 (d, J = 9.7 Hz), 119.34 (d, J = 25.5 Hz), 117.10 (d, J = 4.7 Hz). ^{19}F NMR (376 MHz, Chloroform-*d*) δ -115.94.



Following procedure A for synthesis of alkenes, 1-(methylsulfonyl)-4-vinylbenzene (related to **Figure 1**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.90 (d, J = 8.4 Hz, 2H), 7.58 (d, J = 8.3 Hz, 2H), 6.77 (dd, J = 17.6, 10.9 Hz, 1H), 5.91 (d, J = 17.6 Hz, 1H), 5.47 (d, J = 10.9 Hz, 1H), 3.05 (s, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 142.88, 139.34, 135.21, 127.74, 126.95, 118.02, 44.57.

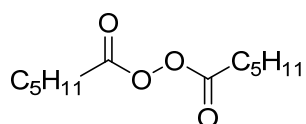


Following procedure B for synthesis of alkenes, 6-vinyl-1,2,3,4-tetrahydronaphthalene (related to **Figure 1**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.14 (d, J = 7.8 Hz, 1H), 7.09 (s, 1H), 7.01 (d, J = 7.9 Hz, 1H), 6.65 (dd, J = 17.6, 10.9 Hz, 1H), 5.67 (d, J = 17.6 Hz, 1H), 5.15 (d, J = 10.9 Hz, 1H), 2.79 – 2.70 (m, 4H), 1.84 – 1.74 (m, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.19, 137.03, 136.91, 134.91, 129.32, 127.05, 123.27, 112.64, 29.46, 29.28, 23.26.

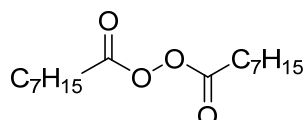


Following procedure B for synthesis of alkenes, (8*R*,9*S*,13*S*,14*S*)-13-Methyl-3-vinyl-6,7,8,9,11,12,13,14,15,16-decahydro-17*H*-cyclopenta[*a*]phenanthren-17-one (compound **57**, related to **scheme 2**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.28 – 7.20 (m, 2H), 7.14 (s, 1H), 6.66 (dd, J = 17.6, 10.9 Hz, 1H), 5.70 (dd, J = 17.6, 1.0 Hz, 1H), 5.19 (dd, J = 10.9, 0.9 Hz, 1H), 2.91 (dd, J = 9.0, 4.2 Hz, 2H), 2.50 (dd, J = 18.8, 8.7 Hz, 1H), 2.45 – 2.39 (m, 1H), 2.34 – 2.25 (m, 1H), 2.19 – 1.93 (m, 4H), 1.66 – 1.41 (m, 6H), 0.91 (s, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 220.89, 139.55, 136.59, 135.22, 126.89, 125.56, 123.62, 113.20, 50.52, 48.00, 44.46, 38.18, 35.88, 31.61, 29.40, 26.52, 25.74, 21.61, 13.87. The spectrum data matches previously reported values

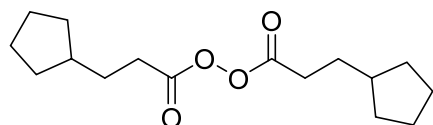
(Huang and Doyle, 2012)



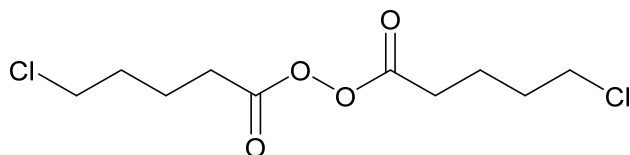
Following procedure C for synthesis of diacyl peroxides, hexanoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. 1H NMR (400 MHz, Chloroform-*d*) δ 2.42 (t, $J = 7.5$ Hz, 4H), 1.78 – 1.66 (m, 4H), 1.41 – 1.29 (m, 8H), 0.91 (t, $J = 7.1$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 169.25, 31.03, 29.95, 24.49, 22.15, 13.78.



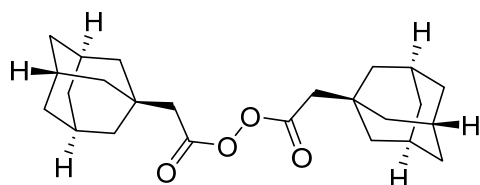
Following procedure C for synthesis of diacyl peroxides, octanoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. 1H NMR (400 MHz, Chloroform-*d*) δ 2.42 (t, $J = 7.5$ Hz, 4H), 1.71 (p, $J = 7.4$ Hz, 4H), 1.42 – 1.22 (m, 16H), 0.87 (t, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 169.25, 31.54, 30.00, 28.87, 28.75, 24.81, 22.55, 14.00.



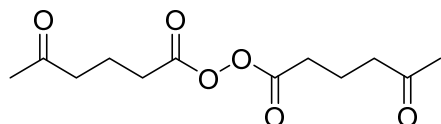
Following procedure C for synthesis of diacyl peroxides, 3-cyclopentylpropanoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. 1H NMR (400 MHz, Chloroform-*d*) δ 2.44 (t, $J = 7.8$ Hz, 4H), 1.86 – 1.69 (m, 10H), 1.68 – 1.49 (m, 8H), 1.17 – 1.03 (m, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 169.39, 39.46, 32.29, 30.94, 29.35, 25.10.



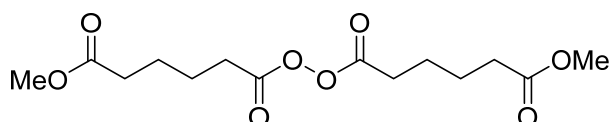
Following procedure C for synthesis of diacyl peroxides, 5-chloropentanoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. 1H NMR (400 MHz, Chloroform-*d*) δ 3.61 – 3.54 (m, 4H), 2.53 – 2.45 (m, 4H), 1.97 – 1.82 (m, 8H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 168.70, 44.17, 31.36, 29.14, 22.09.



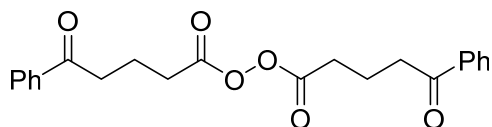
Following procedure C for synthesis of diacyl peroxides, 2-((3*r*,5*r*,7*r*)-adamantan-1-yl)acetic peroxyanhydride (related to **Figure 3**) was obtained as a white solid. 1H NMR (400 MHz, Chloroform-*d*) δ 2.19 (s, 4H), 2.00 (s, 6H), 1.74 – 1.62 (m, 24H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 166.72, 44.46, 42.11, 36.53, 32.99, 28.55.



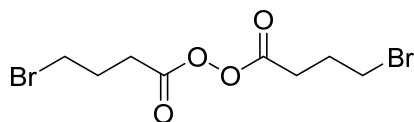
Following procedure C for synthesis of diacyl peroxides, 5-oxohexanoic peroxyanhydride (related to **Figure 3**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 2.60 (t, $J = 7.0$ Hz, 4H), 2.49 (t, $J = 7.1$ Hz, 4H), 2.16 (s, 6H), 1.97 (p, $J = 6.9$ Hz, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 207.42, 168.74, 41.59, 29.94, 28.88, 18.63.



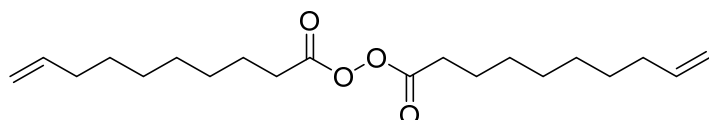
Following procedure C for synthesis of diacyl peroxides, 6-methoxy-6-oxohexanoic peroxyanhydride (related to **Figure 3**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 3.67 (s, 6H), 2.46 (t, $J = 7.0$ Hz, 4H), 2.35 (t, $J = 6.9$ Hz, 4H), 1.81 – 1.68 (m, 8H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 173.46, 168.75, 51.59, 33.42, 29.65, 24.21, 24.09.



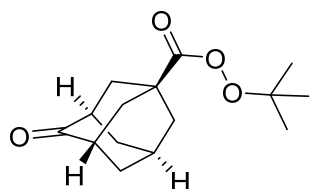
Following procedure C for synthesis of diacyl peroxides, 5-oxo-5-phenylpentanoic peroxyanhydride (related to **Figure 3**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.00 – 7.93 (m, 4H), 7.63 – 7.54 (m, 2H), 7.53 – 7.42 (m, 4H), 3.14 (t, $J = 7.0$ Hz, 4H), 2.60 (t, $J = 7.0$ Hz, 4H), 2.18 (p, $J = 7.0$ Hz, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 198.85, 168.92, 136.68, 133.21, 128.64, 128.03, 36.85, 29.23, 19.20.



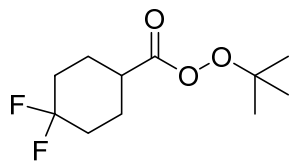
Following procedure C for synthesis of diacyl peroxides, 4-bromobutanoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 3.55 – 3.40 (m, 4H), 2.69 – 2.56 (m, 4H), 2.31 – 2.14 (m, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 168.19, 31.77, 28.39, 27.60.



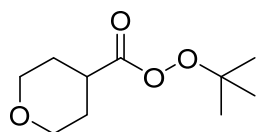
Following procedure C for synthesis of diacyl peroxides, dec-9-enoic peroxyanhydride (related to **Figure 3**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 5.88 – 5.72 (m, 2H), 5.17 – 4.77 (m, 4H), 2.42 (t, $J = 7.4$ Hz, 4H), 2.11 – 1.94 (m, 4H), 1.80 – 1.64 (m, 4H), 1.52 – 1.16 (m, 16H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 169.20, 138.99, 114.23, 33.71, 29.97, 28.92, 28.84, 28.80, 28.78, 24.78.



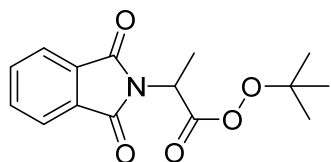
Following procedure D for synthesis of peresters, *tert*-butyl (1*s*,3*r*,5*s*,7*s*)-4-oxoadamantane-1-carboperoxoate (related to **Figure 2**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 2.62 (t, $J = 3.0$ Hz, 2H), 2.27 (d, $J = 2.9$ Hz, 4H), 2.22 (q, $J = 3.1$ Hz, 1H), 2.19 (d, $J = 3.2$ Hz, 2H), 2.11 – 1.99 (m, 4H), 1.32 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 215.64, 172.31, 83.74, 45.66, 40.59, 39.99, 38.12, 37.81, 27.15, 26.07.



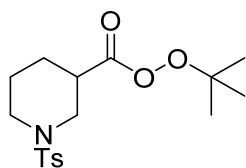
Following procedure D for synthesis of peresters, *tert*-butyl 4,4-difluorocyclohexane-1-carboperoxoate (related to **Figure 2**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 2.54 – 2.44 (m, 1H), 2.21 – 2.08 (m, 2H), 2.06 – 1.71 (m, 6H), 1.33 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 171.35, 122.25 (t, $J = 241.2$ Hz), 83.65, 38.63, 32.45 (t, $J = 24.7$ Hz), 26.11, 25.22 (dd, $J = 7.8$ Hz, $J = 2.5$ Hz). ^{19}F NMR (376 MHz, Chloroform-*d*) δ -94.22 (d, $J = 238.5$ Hz), -99.94 (d, $J = 237.9$ Hz).



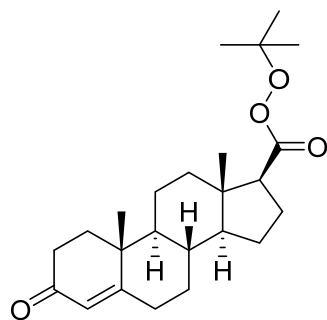
Following procedure D for synthesis of peresters, *tert*-butyl tetrahydro-2*H*-pyran-4-carboperoxoate (related to **Figure 2**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 4.00 (t, $J = 3.6$ Hz, 1H), 3.97 (t, $J = 3.6$ Hz, 1H), 3.48 – 3.41 (m, 2H), 2.68 – 2.59 (m, 1H), 1.93 – 1.82 (m, 4H), 1.33 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 171.51, 83.58, 66.87, 38.30, 28.58, 26.11.



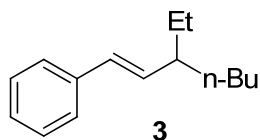
Following procedure D for synthesis of peresters, *tert*-butyl 2-(1,3-dioxoisindolin-2-yl)propaneperoxoate (related to **Figure 2**) was obtained as a clear liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.83 – 7.78 (m, 2H), 7.71 – 7.66 (m, 2H), 5.00 (q, J = 7.3 Hz, 1H), 1.68 (d, J = 7.3 Hz, 3H), 1.21 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 167.08, 166.96, 134.35, 131.73, 123.59, 84.49, 46.38, 26.04, 15.28.



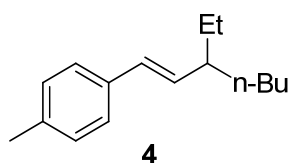
Following procedure D for synthesis of peresters, *tert*-butyl 1-tosylpiperidine-3-carboperoxoate (related to **Figure 2**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 7.64 (d, J = 8.1 Hz, 2H), 7.33 (d, J = 8.0 Hz, 2H), 3.80 (dd, J = 11.6, 3.7 Hz, 1H), 3.66 – 3.56 (m, 1H), 2.77 – 2.65 (m, 1H), 2.56 (t, J = 10.9 Hz, 1H), 2.44 (s, 3H), 2.40 – 2.29 (m, 1H), 2.03 – 1.88 (m, 1H), 1.83 – 1.76 (m, 1H), 1.73 – 1.61 (m, 1H), 1.56 – 1.42 (m, 1H), 1.33 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 170.12, 143.81, 132.89, 129.78, 127.66, 83.94, 47.55, 46.19, 39.23, 26.67, 26.12, 23.84, 21.54.



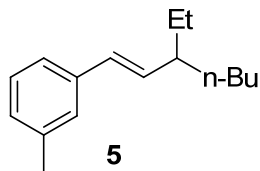
Following procedure D for synthesis of peresters, *tert*-butyl(8*S*,9*S*,10*R*,13*S*,14*S*,17*S*)-10,13-dimethyl-3-oxo-2,3,6,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthrene-17-carboperoxoate (compound **55**, related to **scheme 2**) was obtained as a white solid. ^1H NMR (400 MHz, Chloroform-*d*) δ 5.74 (s, 1H), 2.50 – 2.25 (m, 5H), 2.24 – 2.13 (m, 1H), 2.09 – 2.00 (m, 2H), 1.95 – 1.82 (m, 2H), 1.80 – 1.66 (m, 3H), 1.64 – 1.56 (m, 3H), 1.51 – 1.39 (m, 1H), 1.33 (s, 9H), 1.19 (s, 3H), 1.15 – 1.04 (m, 2H), 1.02 – 0.92 (m, 1H), 0.80 (s, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 199.42, 170.94, 170.84, 123.96, 82.94, 55.30, 53.66, 52.67, 44.14, 38.59, 37.89, 35.72, 35.67, 33.95, 32.76, 31.88, 26.32, 24.45, 23.88, 20.90, 17.38, 13.46.



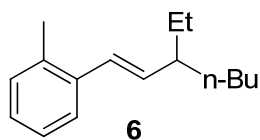
Following the procedure E, product **3** (related to **Figure 1**) was obtained as a clear liquid (75.8 mg, 75% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 (d, *J* = 7.5 Hz, 2H), 7.29 (t, *J* = 7.5 Hz, 2H), 7.18 (t, *J* = 7.2 Hz, 1H), 6.32 (d, *J* = 15.8 Hz, 1H), 5.95 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.05 – 1.95 (m, 1H), 1.52 – 1.40 (m, 2H), 1.34 – 1.22 (m, 6H), 0.88 (t, *J* = 7.5 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 138.09, 135.64, 129.78, 128.55, 126.82, 126.08, 45.30, 35.02, 29.78, 28.36, 23.02, 14.24, 11.98. The spectrum data matches previously reported values (Xu et al., 2017).



Following the procedure E, product **4** (related to **Figure 1**) was obtained as a clear liquid (87.5 mg, 81% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.25 (d, *J* = 7.8 Hz, 2H), 7.09 (d, *J* = 7.7 Hz, 2H), 6.29 (d, *J* = 15.8 Hz, 1H), 5.89 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.32 (s, 3H), 2.03 – 1.94 (m, 1H), 1.52 – 1.39 (m, 2H), 1.37 – 1.20 (m, 6H), 0.87 (t, *J* = 7.2 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 136.41, 135.24, 134.61, 129.44, 129.17, 125.87, 45.19, 34.97, 29.69, 28.31, 22.93, 21.16, 14.17, 11.91. HRMS (EI+) calcd for [C₁₆H₂₄]⁺ ([M]⁺): 216.1878, found: 216.1887.

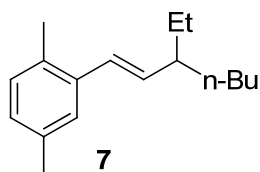


Following the procedure E, product **5** (related to **Figure 1**) was obtained as a clear liquid (78.8 mg, 73% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.21 – 7.13 (m, 3H), 7.00 (d, *J* = 7.2 Hz, 1H), 6.29 (d, *J* = 15.8 Hz, 1H), 5.94 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.33 (s, 3H), 2.05 – 1.95 (m, 1H), 1.52 – 1.41 (m, 2H), 1.35 – 1.24 (m, 6H), 0.88 (t, *J* = 7.4 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 138.01, 135.40, 129.81, 128.45, 127.59, 126.74, 123.24, 45.31, 35.03, 29.76, 28.37, 22.99, 21.49, 14.21, 11.96. HRMS (EI+) calcd for [C₁₆H₂₄]⁺ ([M]⁺): 216.1878, found: 216.1880.

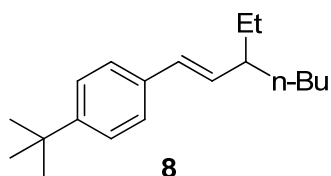


Following the procedure E, product **6** (related to **Figure 1**) was obtained as a clear liquid (79.9 mg, 74% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.41 (d, *J* = 7.0 Hz, 1H), 7.16 – 7.08 (m, 3H), 6.50 (d, *J* = 15.6 Hz, 1H), 5.79 (dd, *J* = 15.6, 9.0 Hz, 1H), 2.33 (s, 3H), 2.07 – 1.98 (m, 1H), 1.52 – 1.45 (m, 2H), 1.36 – 1.27 (m, 6H), 0.90 (t, *J* = 7.4 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 137.33, 137.09, 134.90, 130.12, 127.66, 126.72, 126.00, 125.61, 45.45, 34.91,

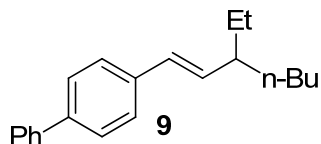
29.70, 28.29, 22.88, 19.91, 14.17, 11.93. HRMS (EI+) calcd for [C₁₆H₂₄]⁺ ([M]⁺): 216.1878, found: 216.1883.



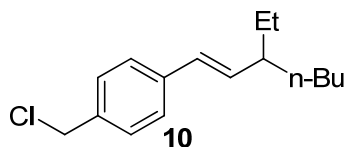
Following the procedure E, product **7** (related to **Figure 1**) was obtained as a clear liquid (90.7 mg, 79% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.24 (s, 1H), 7.01 (d, *J* = 7.7 Hz, 1H), 6.93 (dd, *J* = 7.8, 1.9 Hz, 1H), 6.48 (d, *J* = 15.6 Hz, 1H), 5.77 (dd, *J* = 15.7, 9.1 Hz, 1H), 2.31 (s, 3H), 2.29 (s, 3H), 2.07 – 1.98 (m, 1H), 1.54 – 1.42 (m, 2H), 1.37 – 1.25 (m, 6H), 0.93 – 0.86 (m, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 137.01, 136.75, 135.32, 131.84, 130.07, 127.70, 127.48, 126.16, 45.48, 34.95, 29.70, 28.32, 22.88, 21.07, 19.43, 14.17, 11.94. HRMS (EI+) calcd for [C₁₇H₂₆]⁺ ([M]⁺): 230.2035, found: 230.2028.



Following the procedure E, product **8** (related to **Figure 1**) was obtained as a clear liquid (108 mg, 84% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.43 – 7.26 (m, 4H), 6.30 (d, *J* = 15.8 Hz, 1H), 5.91 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.06 – 1.95 (m, 1H), 1.53 – 1.41 (m, 2H), 1.35 – 1.24 (m, 15H), 0.87 (t, *J* = 7.3 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 149.75, 135.25, 134.86, 129.36, 125.68, 125.42, 45.27, 35.02, 34.52, 31.38, 29.71, 28.37, 22.92, 14.18, 11.93. HRMS (EI+) calcd for [C₁₉H₃₀]⁺ ([M]⁺): 258.2348, found: 258.2343.

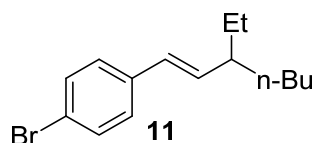


Following the procedure E, product **9** (related to **Figure 1**) was obtained as a white solid (94.5 mg, 68% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.59 – 7.55 (m, 2H), 7.54 – 7.50 (m, 2H), 7.43 – 7.38 (m, 4H), 7.33 – 7.26 (m, 1H), 6.36 (d, *J* = 15.8 Hz, 1H), 6.00 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.08 – 1.97 (m, 1H), 1.55 – 1.44 (m, 2H), 1.37 – 1.24 (m, 6H), 0.92 – 0.85 (m, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 141.01, 139.61, 137.12, 135.90, 129.26, 128.82, 127.26, 127.19, 126.98, 126.45, 45.34, 34.99, 29.76, 28.34, 22.99, 14.23, 11.98. HRMS (EI+) calcd for [C₂₁H₂₆]⁺ ([M]⁺): 278.2035, found: 278.2040.

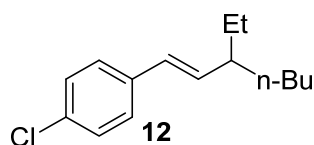


Following the procedure E, product **10** (related to **Figure 1**) was obtained as a clear liquid (87.6 mg, 70% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.28 (m, 4H), 6.32 (d, *J* = 15.8 Hz, 1H), 5.97 (dd, *J* = 15.8, 9.0 Hz, 1H), 4.57 (s, 2H), 2.07 – 1.96 (m, 1H), 1.53 – 1.41 (m, 2H), 1.35 – 1.23 (m, 6H), 0.87 (t, *J* = 7.3 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 138.25,

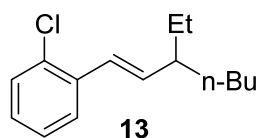
136.53, 135.80, 128.98, 128.83, 126.27, 46.26, 45.20, 34.81, 29.65, 28.17, 22.87, 14.12, 11.86. HRMS (EI+) calcd for $[C_{16}H_{24}Cl]^+$ ($[M]^+$): 250.1488, found: 250.1485.



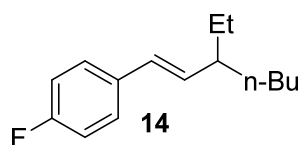
Following the procedure E product **11** (related to **Figure 1**) was obtained as a clear liquid (105.4 mg, 75% yield). 1H NMR (400 MHz, Chloroform-*d*) δ 7.40 (d, $J = 8.5$ Hz, 2H), 7.21 (d, $J = 8.5$ Hz, 2H), 6.26 (d, $J = 15.8$ Hz, 1H), 5.94 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.05 – 1.95 (m, 1H), 1.51 – 1.42 (m, 2H), 1.35 – 1.22 (m, 6H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 136.89, 136.53, 131.49, 128.46, 127.51, 120.30, 45.17, 34.74, 29.64, 28.10, 22.87, 14.12, 11.86. HRMS (EI+) calcd for $[C_{15}H_{21}Br]^+$ ($[M]^+$): 280.0827, found: 280.0829.



Following the procedure E, product **12** (related to **Figure 1**) was obtained as a clear liquid (93.9 mg, 80% yield). 1H NMR (400 MHz, Chloroform-*d*) δ 7.29 – 7.22 (m, 4H), 6.27 (d, $J = 15.8$ Hz, 1H), 5.93 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.06 – 1.95 (m, 1H), 1.54 – 1.41 (m, 2H), 1.35 – 1.24 (m, 6H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 136.48, 136.39, 132.25, 128.57, 128.44, 127.17, 45.16, 34.78, 29.65, 28.13, 22.88, 14.12, 11.85. The spectrum data matches previously reported values (Mai et al., 2016)

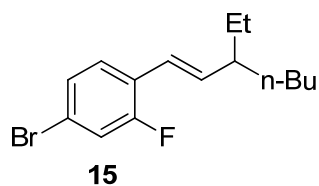


Following the procedure E, product **13** (related to **Figure 1**) was obtained as a clear liquid (81.3 mg, 69% yield). 1H NMR (400 MHz, Chloroform-*d*) δ 7.51 (dd, $J = 7.8, 1.8$ Hz, 1H), 7.32 (d, $J = 7.9$ Hz, 1H), 7.19 (t, $J = 7.4$ Hz, 1H), 7.15 – 7.09 (m, 1H), 6.70 (d, $J = 15.8$ Hz, 1H), 5.93 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.13 – 2.04 (m, 1H), 1.52 – 1.43 (m, 2H), 1.35 – 1.27 (m, 6H), 0.90 (t, $J = 7.8$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 138.53, 136.11, 132.55, 129.57, 127.74, 126.68, 125.97, 45.21, 34.70, 29.59, 28.09, 22.86, 14.12, 11.84. HRMS (EI+) calcd for $[C_{15}H_{21}Cl]^+$ ($[M]^+$): 236.1332, found: 236.1329.

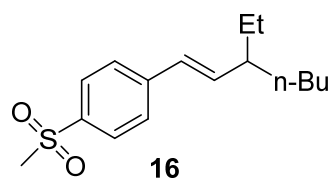


Following the procedure E, product **14** (related to **Figure 1**) was obtained as a clear liquid (76.8 mg, 70% yield). 1H NMR (400 MHz, Chloroform-*d*) δ 7.30 (dd, $J = 8.6, 5.4$ Hz, 2H), 6.97 (t, $J = 8.6$ Hz, 2H), 6.28 (d, $J = 15.8$ Hz, 1H), 5.86 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.04 – 1.94 (m, 1H), 1.52 – 1.41 (m, 2H), 1.35 – 1.22 (m, 6H), 0.88 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 161.85 (d, $J = 245.4$ Hz), 135.34 (d, $J = 2.2$ Hz), 134.13 (d, $J = 3.3$ Hz), 128.43, 127.33 (d, $J = 7.7$ Hz), 115.26 (d, $J = 21.5$ Hz), 45.13, 34.85, 29.66, 28.20, 22.89, 14.12, 11.86. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -115.97. HRMS (EI+) calcd for $[C_{15}H_{21}F]^+$ ($[M]^+$):

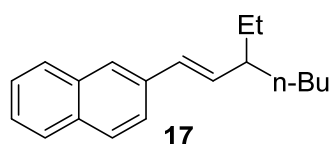
220.1627, found: 220.1621.



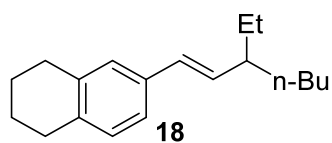
Following the procedure E, product **15** (related to **Figure 1**) was obtained as a clear liquid (80.5 mg, 54% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.31 (t, $J = 8.3$ Hz, 1H), 7.23 – 7.17 (m, 2H), 6.40 (d, $J = 16.0$ Hz, 1H), 6.03 (dd, $J = 16.0, 9.0$ Hz, 1H), 2.09 – 1.99 (m, 1H), 1.53 – 1.42 (m, 2H), 1.36 – 1.25 (m, 6H), 0.88 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 159.57 (d, $J = 252.6$ Hz), 139.09 (d, $J = 4.3$ Hz), 128.01 (d, $J = 4.8$ Hz), 127.26 (d, $J = 3.6$ Hz), 124.87 (d, $J = 12.5$ Hz), 120.99 (d, $J = 3.3$ Hz), 119.90 (d, $J = 9.7$ Hz), 119.19 (d, $J = 25.7$ Hz), 45.57, 34.63, 29.60, 28.00, 22.84, 14.09, 11.81. ^{19}F NMR (376 MHz, Chloroform-*d*) δ -116.27. HRMS (EI+) calcd for $[\text{C}_{15}\text{H}_{20}\text{BrF}]^+$ ($[\text{M}]^+$): 298.0732, found: 298.0739.



Following the procedure E, product **16** (related to **Figure 1**) was obtained as a white solid (33.6 mg, 24% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.78 (d, $J = 8.4$ Hz, 2H), 7.44 (d, $J = 8.4$ Hz, 2H), 6.31 (d, $J = 15.8$ Hz, 1H), 6.08 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.96 (s, 3H), 2.05 – 1.93 (m, 1H), 1.50 – 1.36 (m, 2H), 1.29 – 1.18 (m, 6H), 0.81 (t, $J = 7.5$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 143.44, 140.29, 138.16, 128.11, 127.69, 126.59, 45.32, 44.64, 34.57, 29.62, 27.96, 22.82, 14.08, 11.83. HRMS (EI+) calcd for $[\text{C}_{16}\text{H}_{24}\text{O}_2\text{S}]^+$ ($[\text{M}]^+$): 280.1497, found: 280.1501.

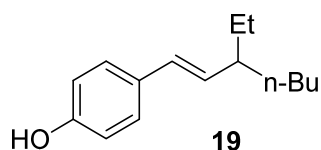


Following the procedure E, product **17** (related to **Figure 2**) was obtained as a clear liquid (91.9 mg, 73% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.84-7.78 (m, 3H), 7.72 (s, 1H), 7.63 (dd, $J = 8.6, 1.8$ Hz, 1H), 7.50 – 7.41 (m, 2H), 6.53 (d, $J = 15.8$ Hz, 1H), 6.13 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.16 – 2.06 (m, 1H), 1.60 – 1.52 (m, 2H), 1.43 – 1.30 (m, 6H), 0.97 – 0.91 (m, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 136.16, 135.44, 133.75, 132.64, 129.72, 128.00, 127.79, 127.62, 126.11, 125.39, 125.27, 123.68, 45.30, 34.91, 29.69, 28.26, 22.91, 14.14, 11.92. HRMS (EI+) calcd for $[\text{C}_{19}\text{H}_{24}]^+$ ($[\text{M}]^+$): 252.1878, found: 252.1873.

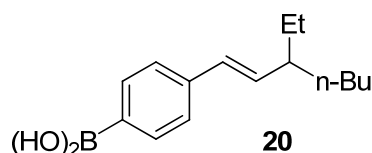


Following the procedure E, product **18** (related to **Figure 1**) was obtained as a clear liquid (79.8 mg, 63% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.10 (dd, $J = 7.9, 1.4$ Hz, 1H), 7.05 (s, 1H), 6.98 (d, $J = 7.9$ Hz, 1H), 6.25 (d, $J = 15.8$ Hz, 1H), 5.87 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.74

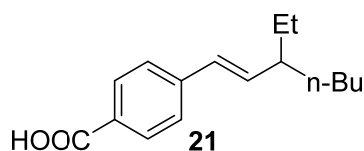
(d, $J = 6.1$ Hz, 4H), 2.04 – 1.93 (m, 1H), 1.82 – 1.74 (m, 4H), 1.52 – 1.40 (m, 2H), 1.33 – 1.23 (m, 6H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform- d) δ 137.11, 135.86, 135.31, 134.47, 129.59, 129.27, 126.62, 123.16, 45.22, 35.00, 29.69, 29.49, 29.21, 28.35, 23.34, 23.32, 22.91, 14.16, 11.90. HRMS (EI+) calcd for $[\text{C}_{19}\text{H}_{28}]^+$ ($[\text{M}]^+$): 256.2191, found: 256.2197.



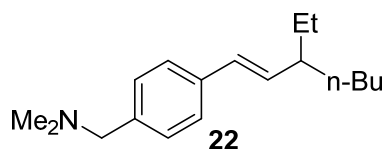
Following the procedure E, product **19** (related to **Figure 1**) was obtained as a clear liquid (89.3 mg, 82% yield). ^1H NMR (400 MHz, Chloroform- d) δ 7.24 (d, $J = 8.7$ Hz, 2H), 6.76 (d, $J = 8.5$ Hz, 2H), 6.25 (d, $J = 15.8$ Hz, 1H), 5.79 (dd, $J = 15.8, 9.0$ Hz, 1H), 4.88 (s, 1H), 2.02 – 1.94 (m, 1H), 1.52 – 1.38 (m, 2H), 1.32 – 1.25 (m, 6H), 0.87 (t, $J = 7.4$ Hz, 6H). ^{13}C NMR (100 MHz, Chloroform- d) δ 154.48, 133.55, 131.06, 128.82, 127.21, 115.33, 45.08, 34.94, 29.64, 28.28, 22.89, 14.12, 11.86. HRMS (EI+) calcd for $[\text{C}_{15}\text{H}_{22}\text{O}]^+$ ($[\text{M}]^+$): 218.1671, found: 218.1673.



Following the procedure E, product **20** (related to **Figure 1**) was obtained as a light yellow oil (43.9 mg, 36% yield). ^1H NMR (400 MHz, Chloroform- d) δ 8.17 (d, $J = 7.8$ Hz, 2H), 7.50 (d, $J = 7.9$ Hz, 2H), 6.43 (d, $J = 15.8$ Hz, 1H), 6.15 (dd, $J = 15.8, 8.9$ Hz, 1H), 2.10 (m, 1H), 1.61 – 1.47 (m, 2H), 1.44 – 1.30 (m, 6H), 0.96 – 0.90 (m, 6H). ^{13}C NMR (100 MHz, Chloroform- d) δ 142.23, 137.77, 136.14, 129.90, 125.73, 45.50, 34.99, 29.88, 28.35, 23.11, 14.34, 12.11. HRMS (DART-) calcd for $[\text{C}_{15}\text{H}_{22}\text{O}_2^{10}\text{B}]^-$ ($[\text{M}-\text{H}]^-$): 244.1755, found: 244.1757.

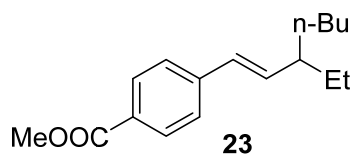


Following the procedure E, product **21** (related to **Figure 1**) was obtained as a clear liquid (64 mg, 52% yield). ^1H NMR (400 MHz, Chloroform- d) δ 8.04 (d, $J = 8.3$ Hz, 2H), 7.44 (d, $J = 8.3$ Hz, 2H), 6.38 (d, $J = 15.8$ Hz, 1H), 6.13 (dd, $J = 15.8, 9.0$ Hz, 1H), 2.13 – 2.00 (m, 1H), 1.56 – 1.44 (m, 2H), 1.32 – 1.25 (m, 6H), 0.91 – 0.86 (m, 6H). ^{13}C NMR (100 MHz, Chloroform- d) δ 171.83, 143.37, 139.21, 130.55, 128.87, 127.33, 125.91, 45.32, 34.67, 29.63, 28.04, 22.85, 14.09, 11.85. HRMS (EI+) calcd for $[\text{C}_{16}\text{H}_{22}\text{O}_2]^+$ ($[\text{M}]^+$): 246.1620, found: 246.1621.

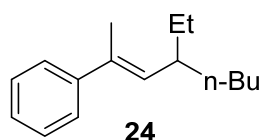


Following the procedure E, product **22** (related to **Figure 1**) was obtained as a clear liquid (65 mg, 50% yield). ^1H NMR (400 MHz, Chloroform- d) δ 7.32 (d, $J = 8.1$ Hz, 2H), 7.23 (d, $J = 7.9$ Hz, 2H), 6.32 (d, $J = 15.8$ Hz, 1H), 5.95 (dd, $J = 15.8, 9.0$ Hz, 1H), 3.40 (s, 2H), 2.24 (s, 6H), 2.07 – 1.95 (m, 1H), 1.54 – 1.43 (m, 2H), 1.31 – 1.26 (m, 6H), 0.92 – 0.85 (m, 6H). ^{13}C NMR (100 MHz, Chloroform- d) δ 137.27, 136.89, 135.33, 129.33, 129.28, 125.84, 64.11, 45.31, 45.16, 34.88, 29.64, 28.23, 22.88, 14.12, 11.87. HRMS (EI+) calcd for $[\text{C}_{18}\text{H}_{29}\text{N}]^+$ ($[\text{M}]^+$):

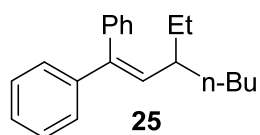
259.2300, found: 259.2305.



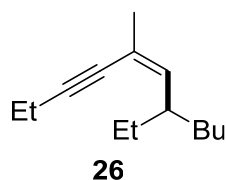
Following the procedure E, product **23** (related to **Figure 1**) was obtained as a clear liquid (66.3 mg, 51% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.88 (d, J = 8.4 Hz, 2H), 7.32 (d, J = 8.4 Hz, 2H), 6.28 (d, J = 15.8 Hz, 1H), 6.02 (dd, J = 15.8, 9.0 Hz, 1H), 3.82 (s, 3H), 2.01 – 1.92 (m, 1H), 1.50 – 1.33 (m, 2H), 1.32 – 1.14 (m, 6H), 0.81 (t, J = 7.5 Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 167.00, 142.49, 138.63, 129.87, 128.90, 128.19, 125.79, 51.97, 45.29, 34.68, 29.63, 28.05, 22.85, 14.09, 11.84. HRMS (EI+) calcd for $[\text{C}_{17}\text{H}_{24}\text{O}_2]^+$ ($[\text{M}]^+$): 260.1776, found: 260.1778.



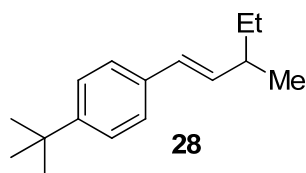
Following the procedure E, product **24** (related to **Figure 1**) was obtained as a clear liquid (88.5 mg, 82% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.42 – 7.36 (m, 2H), 7.30 (t, J = 7.7 Hz, 2H), 7.23 – 7.19 (m, 1H), 5.48 (d, J = 9.9 Hz, 1H), 2.36 – 2.27 (m, 1H), 2.03 (d, J = 1.4 Hz, 3H), 1.54 – 1.44 (m, 2H), 1.33 – 1.22 (m, 6H), 0.88 (t, J = 7.4 Hz, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 144.52, 134.52, 134.42, 128.32, 126.60, 125.90, 40.63, 35.78, 29.98, 29.08, 23.21, 16.57, 14.37, 12.16. HRMS (EI+) calcd for $[\text{C}_{16}\text{H}_{24}]^+$ ($[\text{M}]^+$): 216.1878, found: 216.1875.



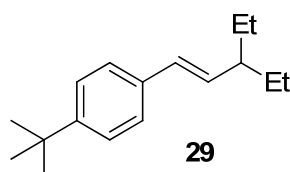
Following the procedure E, product **25** (related to **Figure 1**) was obtained as a clear liquid (129.7 mg, 93% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.37 – 7.32 (m, 2H), 7.30 – 7.21 (m, 5H), 7.21 – 7.13 (m, 3H), 5.82 (d, J = 10.5 Hz, 1H), 2.15 – 2.03 (m, 1H), 1.47 – 1.23 (m, 5H), 1.22 – 1.11 (m, 3H), 0.87 – 0.81 (m, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 142.81, 141.37, 140.77, 135.25, 130.01, 128.08, 128.05, 127.01, 126.72, 126.63, 40.56, 35.47, 29.67, 28.82, 22.97, 14.11, 12.00. HRMS (EI+) calcd for $[\text{C}_{21}\text{H}_{26}]^+$ ($[\text{M}]^+$): 278.2035, found: 278.2038.



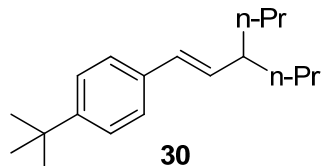
Following the procedure E, product **26** (related to **Figure 1**) was obtained as a clear liquid (68.2 mg, 71% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 5.27 (dd, J = 9.8, 1.6 Hz, 1H), 2.49 – 2.39 (m, 1H), 2.33 (q, J = 7.5 Hz, 2H), 1.82 (d, J = 1.4 Hz, 2H), 1.43 – 1.21 (m, 8H), 1.20 – 1.14 (m, 4H), 0.90 – 0.83 (m, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 141.60, 117.98, 93.82, 79.91, 41.97, 34.89, 29.49, 28.32, 23.54, 22.90, 14.22, 14.13, 13.17, 11.77. HRMS (EI+) calcd for $[\text{C}_{14}\text{H}_{24}]^+$ ($[\text{M}]^+$): 192.1878, found: 192.1887.



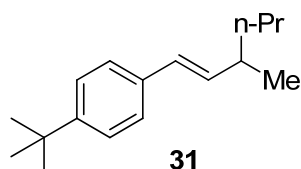
Following the procedure F, product **28** (related to **Figure 2**) was obtained as a clear liquid (97.2 mg, 90% yield). $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.34 – 7.27 (m, 4H), 6.32 (d, J = 15.8 Hz, 1H), 6.05 (dd, J = 15.8, 7.9 Hz, 1H), 2.24 – 2.14 (m, 1H), 1.45 – 1.35 (m, 2H), 1.31 (s, 9H), 1.06 (d, J = 6.7 Hz, 3H), 0.89 (t, J = 7.4 Hz, 3H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 149.76, 136.04, 135.22, 127.83, 125.65, 125.39, 38.96, 34.50, 31.35, 29.88, 20.33, 11.85. HRMS (EI+) calcd for $[\text{C}_{16}\text{H}_{24}]^+$ ($[\text{M}]^+$): 216.1878, found: 216.1877.



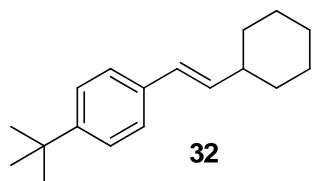
Following the procedure F, product **29** (related to **Figure 2**) was obtained as a clear liquid (93.4 mg, 81% yield). $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.36 – 7.27 (m, 4H), 6.31 (d, J = 15.8 Hz, 1H), 5.91 (dd, J = 15.8, 8.9 Hz, 1H), 1.97 – 1.87 (m, 1H), 1.52 – 1.44 (m, 2H), 1.33 – 1.28 (m, 11H), 0.87 (t, J = 7.4 Hz, 6H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 149.75, 135.23, 134.52, 129.54, 125.66, 125.38, 46.94, 34.50, 31.35, 27.91, 11.88. HRMS (EI+) calcd for $[\text{C}_{17}\text{H}_{26}]^+$ ($[\text{M}]^+$): 230.2035, found: 230.2030.



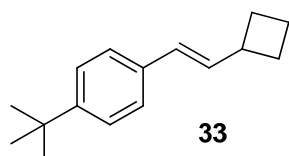
Following the procedure F, product **30** (related to **Figure 2**) was obtained as a clear liquid (105.8 mg, 82% yield). $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.35 – 7.27 (m, 4H), 6.29 (d, J = 15.8 Hz, 1H), 5.90 (dd, J = 15.8, 9.1 Hz, 1H), 2.18 – 2.08 (m, 1H), 1.37 – 1.25 (m, 17H), 0.87 (t, J = 6.9 Hz, 6H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 149.72, 135.18, 135.06, 129.06, 125.61, 125.38, 42.91, 37.84, 34.48, 31.33, 20.46, 14.19. HRMS (EI+) calcd for $[\text{C}_{19}\text{H}_{30}]^+$ ($[\text{M}]^+$): 258.2348, found: 258.2350.



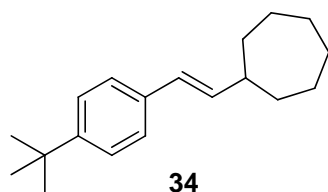
Following the procedure F, product **31** (related to **Figure 2**) was obtained as a clear liquid (90.9 mg, 79% yield). $^1\text{H NMR}$ (400 MHz, Chloroform-*d*) δ 7.33 – 7.26 (m, 4H), 6.31 (d, J = 15.8 Hz, 1H), 6.04 (dd, J = 15.8, 8.0 Hz, 1H), 2.33 – 2.23 (m, 1H), 1.36 – 1.31 (m, 4H), 1.30 (s, 9H), 1.05 (d, J = 6.7 Hz, 3H), 0.91 – 0.87 (m, 3H). $^{13}\text{C NMR}$ (100 MHz, Chloroform-*d*) δ 149.76, 136.32, 135.22, 127.64, 125.65, 125.39, 39.43, 37.05, 34.50, 31.36, 20.79, 20.52, 14.21. HRMS (EI+) calcd for $[\text{C}_{17}\text{H}_{26}]^+$ ($[\text{M}]^+$): 230.2035, found: 230.2038.



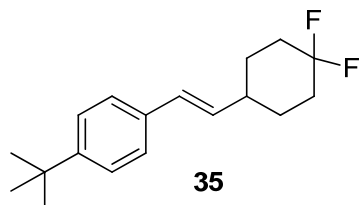
Following the procedure F, product **32** (related to **Figure 2**) was obtained as a clear liquid (115.0 mg, 95% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 – 7.25 (m, 4H), 6.32 (d, J = 16.0 Hz, 1H), 6.13 (dd, J = 15.9, 7.0 Hz, 1H), 2.17 – 2.06 (m, 1H), 1.91 – 1.62 (m, 6H), 1.30 (s, 9H), 1.27 – 1.13 (m, 4H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.74, 136.16, 135.30, 126.89, 125.61, 125.37, 41.18, 34.48, 33.04, 31.34, 26.21, 26.08. The spectrum data matches previously reported values (Zhu and Wei, 2014)



Following the procedure F, product **33** (related to **Figure 2**) was obtained as a clear liquid (80.1 mg, 75% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 – 7.27 (m, 4H), 6.31 – 6.26 (m, 2H), 3.14 – 3.03 (m, 1H), 2.21 – 2.13 (m, 2H), 2.00 – 1.78 (m, 4H), 1.31 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.82, 135.03, 134.58, 127.28, 125.67, 125.39, 38.79, 34.50, 31.34, 28.84, 18.59. HRMS (EI+) calcd for $[\text{C}_{16}\text{H}_{22}]^+$ ($[\text{M}]^+$): 214.1722, found: 214.1725.

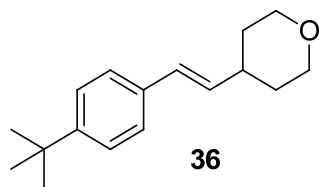


Following the procedure F, product **34** (related to **Figure 2**) was obtained as a clear liquid (117.8 mg, 92% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.27 (m, 4H), 6.31 (d, J = 15.9 Hz, 1H), 6.19 (dd, J = 15.9, 7.5 Hz, 1H), 2.38 – 2.26 (m, 1H), 1.87 – 1.78 (m, 2H), 1.74 – 1.67 (m, 2H), 1.66 – 1.61 (m, 1H), 1.58 – 1.51 (m, 4H), 1.50 – 1.38 (m, 3H), 1.32 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 148.88, 136.21, 134.57, 125.52, 124.82, 124.58, 42.46, 34.02, 33.68, 30.54, 27.62, 25.49. HRMS (EI+) calcd for $[\text{C}_{19}\text{H}_{28}]^+$ ($[\text{M}]^+$): 256.2191, found: 256.2190.

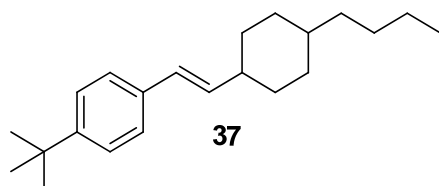


Following the procedure G for the reaction with perester, product **35** (related to **Figure 2**) was obtained as a white solid (73.7 mg, 53% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.26 (m, 4H), 6.39 (d, J = 16.0 Hz, 1H), 6.10 (dd, J = 16.0, 7.0 Hz, 1H), 2.29 – 2.19 (m, 1H), 2.18 – 2.07 (m, 2H), 1.92 – 1.70 (m, 4H), 1.63 – 1.54 (m, 2H), 1.31 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 150.29, 134.62, 132.88 (d, J = 2.5 Hz), 128.64, 125.74, 125.48, 122.14 (d, J = 240.2 Hz), 39.04, 34.54, 33.25 (dd, J = 25.2, 22.9 Hz), 31.32, 28.93 (d, J = 9.2 Hz). ^{19}F NMR

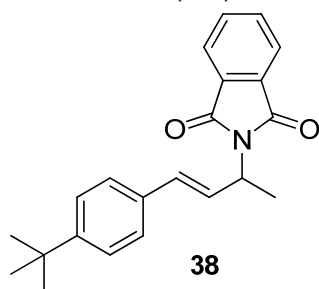
(376 MHz, Chloroform-*d*) δ -94.22 (d, J = 238.6 Hz), -99.94 (d, J = 238.3 Hz). HRMS (EI+) calcd for $[\text{C}_{18}\text{H}_{24}\text{F}_2]^+$ ($[\text{M}]^+$): 278.1846, found: 278.1848.



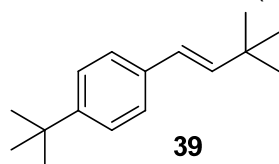
Following the procedure G for the reaction with perester, product **36** (related to **Figure 2**) was obtained as a white solid (90.3 mg, 74% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.27 (m, 4H), 6.36 (d, J = 16.0 Hz, 1H), 6.11 (dd, J = 15.9, 6.8 Hz, 1H), 4.02 – 3.96 (m, 2H), 3.48 – 3.37 (m, 2H), 2.40 – 2.30 (m, 1H), 1.71 – 1.65 (m, 2H), 1.61 – 1.49 (m, 2H), 1.30 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 150.30, 134.92, 134.02, 128.13, 125.90, 125.60, 67.90, 38.53, 34.67, 32.87, 31.48. HRMS (EI+) calcd for $[\text{C}_{17}\text{H}_{24}\text{O}]^+$ ($[\text{M}]^+$): 244.1827, found: 244.1835.



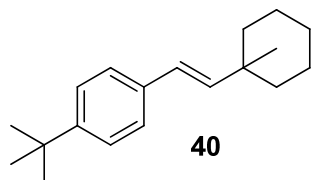
Following the procedure F, product **37** (related to **Figure 2**) was obtained as a clear liquid (104.5 mg, 70% yield, dr = 1:1). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 – 7.26 (m, 4H), 6.38 – 6.29 (m, 1H), 6.25 (dd, J = 16.0, 6.4 Hz, 0.56H), 6.12 (dd, J = 15.9, 7.0 Hz, 0.41H), 2.39 – 2.32 (m, 0.54H), 2.09 – 1.99 (m, 0.49H), 1.85 – 1.76 (m, 2H), 1.63 – 1.54 (m, 3H), 1.42 – 1.32 (m, 3H), 1.31 – 1.30 (m, 9H), 1.27 – 1.14 (m, 7H), 0.89 (t, J = 6.9 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.74, 136.12, 135.34, 135.29, 134.87, 127.80, 126.91, 125.60, 125.38, 125.37, 41.49, 37.31, 37.17, 34.48, 33.00, 31.34, 29.62, 29.41, 29.23, 29.05, 23.04, 23.01, 14.19. HRMS (EI+) calcd for $[\text{C}_{22}\text{H}_{34}]^+$ ($[\text{M}]^+$): 298.2661, found: 298.2668.



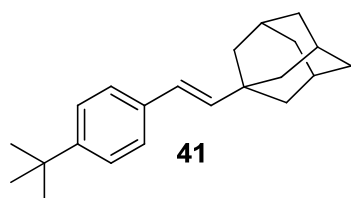
Following the procedure G for the reaction with perester, product **38** (related to **Figure 2**) was obtained as a clear liquid (95 mg, 57% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.82 (dd, J = 5.4, 3.1 Hz, 2H), 7.69 (dd, J = 5.5, 3.0 Hz, 2H), 7.33 – 7.30 (m, 4H), 6.64 – 6.55 (m, 2H), 5.13 – 5.04 (m, 1H), 1.66 (d, J = 7.1 Hz, 3H), 1.29 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 167.99, 150.97, 133.86, 133.61, 132.11, 131.80, 127.36, 126.29, 125.47, 123.13, 49.09, 34.58, 31.27, 19.06. HRMS (EI+) calcd for $[\text{C}_{22}\text{H}_{23}\text{NO}_2]^+$ ($[\text{M}]^+$): 333.1729, found: 333.1725.



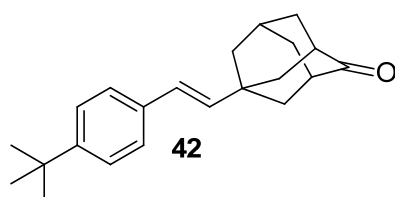
Following the procedure F, product **39** (related to **Figure 2**) was obtained as clear liquid (82.1 mg, 76 % yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.26 (m, 4H), 6.28 (d, J = 16.2 Hz, 1H), 6.21 (d, J = 16.2, 1H), 1.31 (s, 9H), 1.11 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.76, 141.17, 135.27, 125.68, 125.39, 124.21, 34.49, 33.30, 31.35, 29.66. The spectrum data matches previously reported values (Aydin et al., 2009).



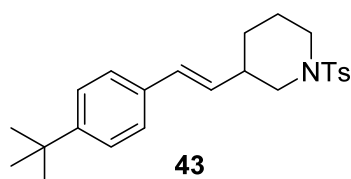
Following the procedure F, product **40** (related to **Figure 2**) was obtained as a clear liquid (102.4 mg, 80% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.28 (m, 4H), 6.30 (d, J = 16.4 Hz, 1H), 6.17 (d, J = 16.4 Hz, 1H), 1.64 – 1.56 (m, 2H), 1.52 – 1.46 (m, 4H), 1.42 – 1.32 (m, 4H), 1.31 (s, 9H), 1.05 (s, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.74, 140.28, 135.49, 125.68, 125.61, 125.40, 38.06, 36.17, 34.49, 31.36, 27.84, 26.37, 22.51. HRMS (EI+) calcd for $[\text{C}_{19}\text{H}_{28}]^+$ ($[\text{M}]^+$): 256.2191, found: 256.2189.



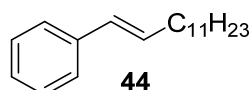
Following the procedure F, product **41** (related to **Figure 2**) was obtained as a white solid (128 mg, 87% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.27 (m, 4H), 6.22 (d, J = 16.2 Hz, 1H), 6.06 (d, J = 16.3, 1H), 2.05 – 1.99 (m, 3H), 1.77 – 1.67 (m, 12H), 1.32 – 1.29 (m, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 149.71, 141.45, 135.43, 125.64, 125.37, 124.14, 42.31, 36.93, 35.11, 34.48, 31.34, 28.53. HRMS (EI+) calcd for $[\text{C}_{22}\text{H}_{30}]^+$ ($[\text{M}]^+$): 294.2348, found: 294.2352.



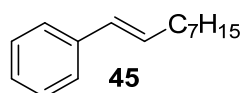
Following the procedure G for the reaction with perester, product **42** (related to **Figure 2**) was obtained as a white solid (127 mg, 82% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.28 – 7.20 (m, 4H), 6.21 (d, J = 16.2 Hz, 1H), 5.99 (d, J = 16.3 Hz, 1H), 2.56 – 2.52 (m, 2H), 2.17 – 2.11 (m, 1H), 1.98 – 1.84 (m, 10H), 1.23 (s, 9H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 217.99, 150.34, 137.54, 134.61, 125.95, 125.78, 125.49, 46.42, 43.44, 41.17, 38.66, 35.01, 34.54, 31.32, 27.78. HRMS (EI+) calcd for $[\text{C}_{22}\text{H}_{28}\text{O}]^+$ ($[\text{M}]^+$): 308.2140, found: 308.2134.



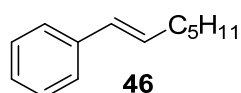
Following the procedure G for the reaction with perester, product **43** (related to **Figure 2**) was obtained as white solid (79 mg, 40% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.65 (d, J = 8.3 Hz, 2H), 7.34 – 7.29 (m, 4H), 7.26 (d, J = 5.2 Hz, 2H), 6.41 (d, J = 16.0 Hz, 1H), 5.96 (dd, J = 16.0, 7.2 Hz, 1H), 3.79 – 3.57 (m, 2H), 2.54 – 2.46 (m, 1H), 2.43 (s, 3H), 2.30 – 2.23 (m, 1H), 2.11 (t, J = 10.9 Hz, 1H), 1.86 – 1.65 (m, 3H), 1.30 (s, 9H), 1.13 (m, 1H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 150.53, 143.43, 134.30, 133.24, 130.25, 130.05, 129.64, 127.71, 125.82, 125.49, 51.38, 46.44, 39.08, 34.56, 31.32, 29.99, 24.43, 21.56. HRMS (EI+) calcd for $[\text{C}_{24}\text{H}_{31}\text{NO}_2\text{S}]^+$ ($[\text{M}]^+$): 397.2075, found: 397.2079.



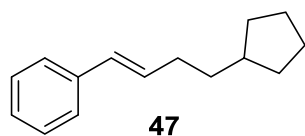
Following the procedure H for the reaction with diacyl peroxide, product **44** (related to **Figure 3**) was obtained as a clear liquid (94 mg, 73% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 (d, J = 7.7 Hz, 2H), 7.28 (t, J = 7.5 Hz, 2H), 7.18 (t, J = 7.2 Hz, 1H), 6.37 (d, J = 15.8 Hz, 1H), 6.27 – 6.18 (m, 1H), 2.24 – 2.16 (m, 2H), 1.49 – 1.40 (m, 2H), 1.34 – 1.22 (m, 16H), 0.88 (t, J = 6.6 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.97, 131.28, 129.67, 128.46, 126.73, 125.90, 33.09, 31.96, 29.71, 29.68, 29.66, 29.57, 29.42, 29.39, 29.28, 22.73, 14.15. The spectrum data matches previously reported values (Habrant et al., 2007).



Following the procedure H for the reaction with diacyl peroxide, product **45** (related to **Figure 3**) was obtained as a clear liquid (61 mg, 60% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 (d, J = 7.6 Hz, 2H), 7.27 (t, J = 7.5 Hz, 2H), 7.17 (t, J = 7.2 Hz, 1H), 6.37 (d, J = 15.8 Hz, 1H), 6.27 – 6.15 (m, 1H), 2.25 – 2.14 (m, 2H), 1.52 – 1.41 (m, 2H), 1.36 – 1.25 (m, 8H), 0.88 (t, J = 6.5 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.99, 131.27, 129.71, 128.48, 126.76, 125.93, 33.11, 31.91, 29.45, 29.27, 22.73, 14.16. The spectrum data matches previously reported values (Thiot et al., 2007).

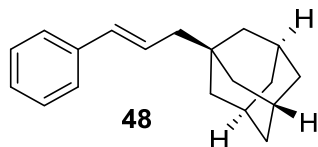


Following the procedure H for the reaction with diacyl peroxide, product **46** (related to **Figure 3**) was obtained as a clear liquid (59 mg, 68% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.34 (d, J = 7.1 Hz, 2H), 7.28 (t, J = 7.5 Hz, 2H), 7.22 – 7.15 (m, 1H), 6.37 (d, J = 15.8 Hz, 1H), 6.27 – 6.18 (m, 1H), 2.24 – 2.17 (m, 2H), 1.50 – 1.43 (m, 2H), 1.37 – 1.29 (m, 4H), 0.90 (t, J = 6.7 Hz, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.97, 131.27, 129.68, 128.47, 126.74, 125.90, 33.05, 31.47, 29.09, 22.60, 14.10. The spectrum data matches previously reported values (Denmark and Wehrli, 2000).

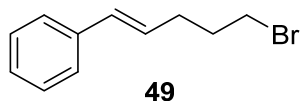


Following the procedure H for the reaction with diacyl peroxide, product **47** (related to **Figure 3**) was obtained as a clear liquid (77 mg, 77% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.33 (d,

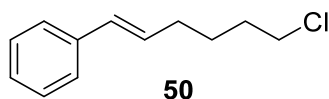
$J = 7.2$ Hz, 2H), 7.28 (t, $J = 7.6$ Hz, 2H), 7.17 (t, $J = 7.2$ Hz, 1H), 6.37 (d, $J = 15.9$ Hz, 1H), 6.27 – 6.18 (m, 1H), 2.25 – 2.18 (m, 2H), 1.84 – 1.73 (m, 3H), 1.66 – 1.57 (m, 2H), 1.54 – 1.44 (m, 4H), 1.17 – 1.04 (m, 2H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 138.00, 131.41, 129.54, 128.49, 126.75, 125.91, 39.73, 35.91, 32.70, 32.32, 25.25. The spectrum data matches previously reported values (Li et al., 2016).



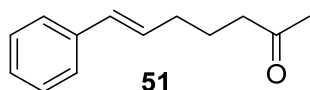
Following the procedure H for the reaction with diacyl peroxide, product **48** (related to **Figure 3**) was obtained as clear liquid (83 mg, 66% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.37 – 7.33 (m, 2H), 7.32 – 7.26 (m, 2H), 7.21 – 7.15 (m, 1H), 6.33 (d, $J = 15.9$ Hz, 1H), 6.29 – 6.22 (m, 1H), 1.98 – 1.93 (m, 5H), 1.73 – 1.67 (m, 3H), 1.66 – 1.59 (m, 3H), 1.55 – 1.52 (m, 6H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.96, 131.88, 128.48, 127.09, 126.76, 125.98, 48.12, 42.61, 37.14, 33.52, 28.81. The spectrum data matches previously reported values (Tanaka et al., 1987).



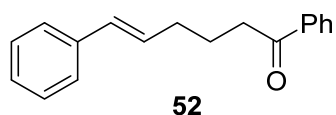
Following the procedure H for the reaction with diacyl peroxide, product **49** (related to **Figure 3**) was obtained as a clear liquid (62 mg, 55% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.32 (m, 2H), 7.32 – 7.27 (m, 2H), 7.23 – 7.18 (m, 1H), 6.44 (d, $J = 15.8$ Hz, 1H), 6.21 – 6.10 (m, 1H), 3.45 (t, $J = 6.7$ Hz, 2H), 2.42 – 2.34 (m, 2H), 2.07 – 1.99 (m, 2H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.43, 131.31, 128.54, 128.47, 127.14, 126.02, 33.19, 32.21, 31.30. The spectrum data matches previously reported values (Matsubara and Jamison, 2010)



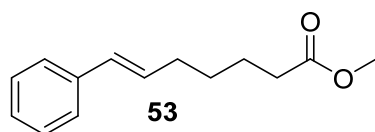
Following the procedure H for the reaction with diacyl peroxide, product **50** (related to **Figure 3**) was obtained as a clear liquid (53 mg, 55% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.38 – 7.26 (m, 4H), 7.23 – 7.17 (m, 1H), 6.40 (d, $J = 15.8$ Hz, 1H), 6.26 – 6.15 (m, 1H), 3.56 (t, $J = 6.7$ Hz, 2H), 2.29 – 2.22 (m, 2H), 1.90 – 1.77 (m, 2H), 1.69 – 1.57 (m, 2H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 137.63, 130.43, 130.06, 128.52, 126.98, 125.96, 44.98, 32.23, 32.0, 26.56. The spectrum data matches previously reported values (Hu et al., 1999).



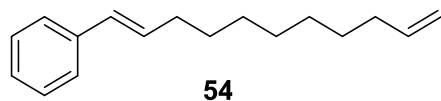
Following the procedure H for the reaction with diacyl peroxide, product **51** (related to **Figure 3**) was obtained as a white solid (50.7 mg, 54% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.26 (m, 4H), 7.24 – 7.17 (m, 1H), 6.39 (d, $J = 15.7$ Hz, 1H), 6.21 – 6.12 (m, 1H), 2.48 (t, $J = 7.4$ Hz, 2H), 2.26 – 2.19 (m, 1H), 2.14 (s, 3H), 1.82 – 1.72 (m, 1H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 208.85, 137.57, 130.68, 129.83, 128.52, 127.01, 125.97, 42.88, 32.30, 30.02, 23.25. The spectrum data matches previously reported values (Musacchio et al., 2014).



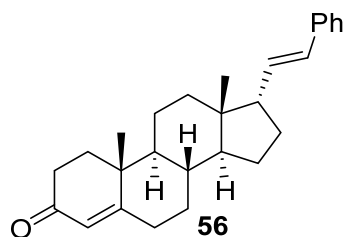
Following the procedure H for the reaction with diacyl peroxide, product **52** (related to **Figure 3**) was obtained as a white solid (70 mg, 56% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.98 – 7.93 (m, 2H), 7.58 – 7.53 (m, 1H), 7.48 – 7.42 (m, 2H), 7.36 – 7.27 (m, 4H), 7.23 – 7.17 (m, 1H), 6.41 (d, $J = 15.9$ Hz, 1H), 6.27 – 6.18 (m, 1H), 3.03 (t, $J = 7.3$ Hz, 2H), 2.36 – 2.29 (m, 2H), 1.99 – 1.91 (m, 2H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 200.22, 137.62, 137.02, 132.96, 130.73, 129.94, 128.58, 128.51, 128.05, 126.98, 125.99, 37.75, 32.47, 23.80. The spectrum data matches previously reported values (Hilt et al., 2016).



Following the procedure H for the reaction with diacyl peroxide, product **53** (related to **Figure 3**) was obtained as a white solid (61 mg, 56% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.25 (m, 4H), 7.21 – 7.15 (m, 1H), 6.38 (d, $J = 15.9$ Hz, 1H), 6.23 – 6.15 (m, 1H), 3.66 (s, 3H), 2.34 (t, $J = 7.5$ Hz, 2H), 2.26 – 2.19 (m, 2H), 1.74 – 1.65 (m, 2H), 1.55 – 1.46 (m, 2H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 174.11, 137.74, 130.32, 130.21, 128.49, 126.88, 125.95, 51.51, 33.96, 32.64, 28.84, 24.52. The spectrum data matches previously reported values (Ramón-Azcón et al., 2006).

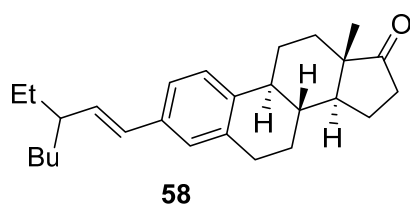


Following the procedure H for the reaction with diacyl peroxide, product **54** (related to **Figure 3**) was obtained as a clear liquid (70 mg, 61% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.32 (m, 2H), 7.30 – 7.25 (m, 2H), 7.21 – 7.15 (m, 1H), 6.37 (d, $J = 15.9$ Hz, 1H), 6.27 – 6.17 (m, 1H), 5.91 – 5.74 (m, 1H), 5.04 – 4.90 (m, 2H), 2.25 – 2.16 (m, 2H), 2.09 – 2.00 (m, 2H), 1.50 – 1.42 (m, 2H), 1.40 – 1.28 (m, 8H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 139.22, 137.96, 131.21, 129.73, 128.48, 126.76, 125.92, 114.17, 33.84, 33.07, 29.40, 29.21, 29.13, 28.96. The spectrum data matches previously reported values (Kulasegaram and Kulawiec, 1997).

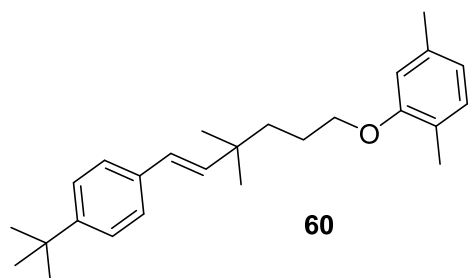


Following the procedure G, product **56** (related to **Scheme 2**) was obtained as a white solid (89.8 mg, 48% yield). ^1H NMR (400 MHz, Chloroform-*d*) δ 7.35 – 7.32 (m, 2H), 7.31 – 7.25 (m, 2H), 7.20 – 7.15 (m, 1H), 6.26 (d, $J = 15.7$ Hz, 1H), 6.13 (dd, $J = 15.7, 9.3$ Hz, 1H), 5.73 (s, 1H), 2.46 – 2.33 (m, 4H), 2.31–2.15 (m, 1H), 2.18 – 2.09 (m, 1H), 2.03 – 1.97 (m, 1H), 1.94 – 1.88 (m, 1H), 1.85 – 1.78 (m, 1H), 1.75 – 1.62 (m, 2H), 1.55 – 1.50 (m, 3H), 1.46 – 1.37 (m, 1H), 1.36 – 1.23 (m, 3H), 1.18 (s, 3H), 1.14 – 1.04 (m, 1H), 0.97 – 0.91 (m, 1H), 0.88 (s, 3H). ^{13}C NMR (100 MHz, Chloroform-*d*) δ 199.61, 171.46, 137.80, 133.85, 128.88, 128.50, 126.82,

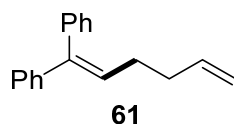
125.98, 123.80, 53.70, 52.59, 50.88, 44.66, 38.69, 36.15, 35.71, 34.75, 33.99, 32.97, 32.48, 28.49, 25.70, 20.93, 20.27, 17.48. HRMS (EI⁺) calcd for [C₂₇H₃₄O]⁺ ([M]⁺): 374.2610, found: 374.2613.



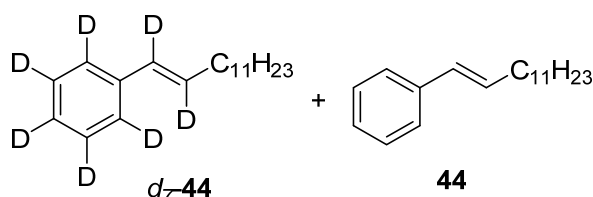
Following the procedure E, product **58** (related to **Scheme 2**) was obtained as a white solid (123 mg, 65% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.23 (d, *J* = 8.1 Hz, 1H), 7.16 (d, *J* = 8.3 Hz, 1H), 7.10 (s, 1H), 6.27 (d, *J* = 15.8 Hz, 1H), 5.90 (dd, *J* = 15.8, 9.0 Hz, 1H), 2.91 (dd, *J* = 8.8, 4.0 Hz, 2H), 2.50 (dd, *J* = 18.7, 8.6 Hz, 1H), 2.45 – 2.39 (m, 1H), 2.33 – 2.26 (m, 1H), 2.19 – 1.93 (m, 5H), 1.65 – 1.58 (m, 2H), 1.55 – 1.39 (m, 6H), 1.33 – 1.24 (m, 6H), 0.90 (s, 3H), 0.87 (t, *J* = 7.4 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 221.13, 138.58, 136.68, 135.81, 135.24, 129.44, 126.71, 126.68, 125.70, 123.65, 123.62, 50.69, 48.20, 45.39, 44.61, 38.44, 36.06, 35.13, 31.80, 29.84, 29.62, 28.48, 26.74, 25.97, 23.05, 21.79, 14.32, 14.04, 12.06. HRMS (EI⁺) calcd for [C₂₇H₃₈O]⁺ ([M]⁺): 378.2923, found: 378.2929.



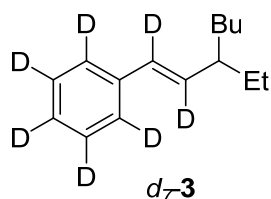
Following the procedure E, product **60** (related to **Scheme 2**) was obtained as a clear liquid (76 mg, 42% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.34 – 7.28 (m, 4H), 6.99 (d, *J* = 7.4 Hz, 1H), 6.64 (d, *J* = 8.1 Hz, 1H), 6.60 (s, 1H), 6.29 (d, *J* = 16.2 Hz, 1H), 6.16 (d, *J* = 16.2 Hz, 1H), 3.90 (t, *J* = 6.4 Hz, 2H), 2.28 (s, 3H), 2.18 (s, 3H), 1.80 – 1.71 (m, 2H), 1.59 – 1.52 (m, 2H), 1.31 (s, 9H), 1.13 (s, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 157.11, 149.90, 139.54, 136.45, 135.17, 130.28, 125.81, 125.74, 125.44, 123.62, 120.59, 112.01, 68.43, 39.44, 36.08, 34.52, 31.37, 27.37, 25.03, 21.44, 15.86. HRMS (EI⁺) calcd for [C₂₆H₃₆O]⁺ ([M]⁺): 364.2766, found: 364.2761.



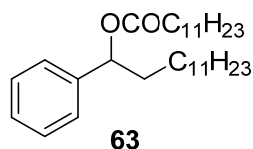
Following the procedure E, product **61** (related to **Scheme 3A**) was obtained as a clear liquid (72.3 mg, 62%). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.38 – 7.33 (m, 2H), 7.32 – 7.28 (m, 1H), 7.27 – 7.24 (m, 1H), 7.24 – 7.19 (m, 4H), 7.19 – 7.16 (m, 2H), 6.10 – 6.05 (m, 1H), 5.86 – 5.73 (m, 1H), 5.08 – 4.80 (m, 2H), 2.25 – 2.17 (m, 4H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 142.79, 141.95, 140.19, 138.13, 129.92, 129.20, 128.16, 128.08, 127.26, 126.91, 126.86, 114.94, 34.06, 29.15. The spectrum data matches previously reported values (Jiménez-Aquino et al., 2011).



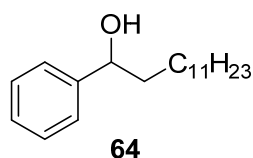
Following the procedure H, product **44** and *d*₇-**44** (related to **Scheme 3B**) were obtained as a clear liquid (60% total yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.37 – 7.31 (m, 2H), 7.31 – 7.26 (m, 2H), 7.21 – 7.15 (m, 1H), 6.37 (d, *J* = 15.9 Hz, 1H), 6.27 – 6.17 (m, 1H), 2.23 – 2.16 (m, 4H), 1.50 – 1.41 (m, 4H), 1.34 – 1.24 (m, 32H), 0.88 (t, *J* = 6.8 Hz, 6H).



Following the procedure G, product *d*₇-**3** (related to **Scheme 3B**) was obtained as a clear liquid (62.2 mg, 60% yield). ¹H NMR (400 MHz, Chloroform-*d*) δ 2.05 – 1.97 (m, 1H), 1.54 – 1.41 (m, 2H), 1.35 – 1.24 (m, 6H), 0.88 (t, *J* = 7.5 Hz, 6H). ¹³C NMR (101 MHz, Chloroform-*d*) δ 137.80, 135.14 (t, *J* = 22.4 Hz), 129.14 (t, *J* = 22.9 Hz), 127.97 (t, *J* = 24.00 Hz), 126.20 (t, *J* = 23.8 Hz), 125.54 (t, *J* = 23.7 Hz), 45.02, 34.88, 29.68, 28.23, 22.92, 14.15, 11.89.



Compound **63** (related to **Scheme 3C**): ¹H NMR (400 MHz, Chloroform-*d*) δ 7.35-7.30 (m, 4H), 7.29-7.26 (m, 1H), 5.72 (dd, *J* = 7.8, 6.1 Hz, 1H), 2.34-2.28 (m, 2H), 1.94-1.83 (m, 1H), 1.80-1.69 (m, 1H), 1.65-1.58 (m, 2H), 1.31-1.21 (m, 36H), 0.88 (t, *J* = 6.7 Hz, 6H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 173.23, 141.10, 128.37, 127.72, 126.49, 75.86, 36.42, 34.65, 31.94, 29.68, 29.66, 29.63, 29.58, 29.50, 29.38, 29.36, 29.30, 29.14, 25.55, 25.05, 22.72, 14.15. Synthesized according to reported method (Ge et al., 2017).



Compound **64** (related to **Scheme 3C**): ¹H NMR (400 MHz, Chloroform-*d*) δ 7.36 – 7.30 (m, 4H), 7.29 – 7.22 (m, 1H), 4.63 (dd, *J* = 7.5, 5.9 Hz, 1H), 2.06 (s, 1H), 1.83 – 1.73 (m, 1H), 1.72 – 1.62 (m, 1H), 1.45 – 1.35 (m, 1H), 1.31 – 1.18 (m, 19H), 0.88 (t, *J* = 6.9 Hz, 3H). ¹³C NMR (100 MHz, Chloroform-*d*) δ 144.96, 128.43, 127.47, 125.92, 74.72, 39.14, 31.97, 29.71, 29.69, 29.64, 29.60, 29.58, 29.41, 25.88, 22.74, 14.18. Synthesized according to reported method (Jian et al., 2017).

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