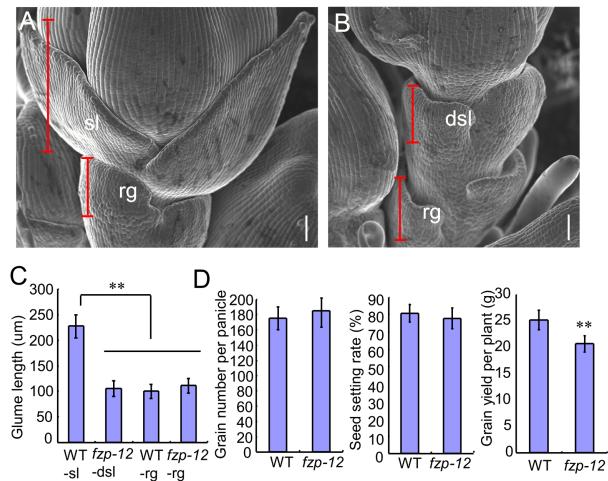


SUPPLEMENTAL FIGURES

Supplemental Figure 1



Supplemental Figure 1. A, wild type spikelet at Sp8 stage. B, *fzp-12* spikelet at Sp8 stage. C, length of the sterile lemma, degenerated sterile lemma, rudimentary glume in the wild type and *fzp-12* mutant. D, grain number per panicle, seed setting rate, and grain yield per plant in the wild type and *fzp-12* mutant. rg, rudimentary glume; sl, sterile lemma; dsl, degenerated sterile lemma. Red lines indicate the organ length. Bars = 50 μm. **Significant difference at $P < 0.01$ compared with the wild type by Student's *t*-test.

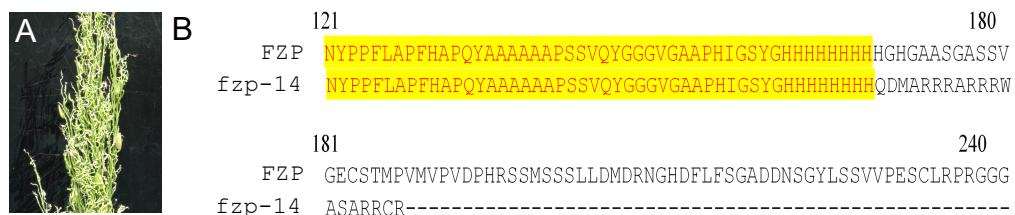
Supplemental Figure 2



Supplemental Figure 2. Protein sequence alignment of FZP, fzp-12, and fzp-13. Red

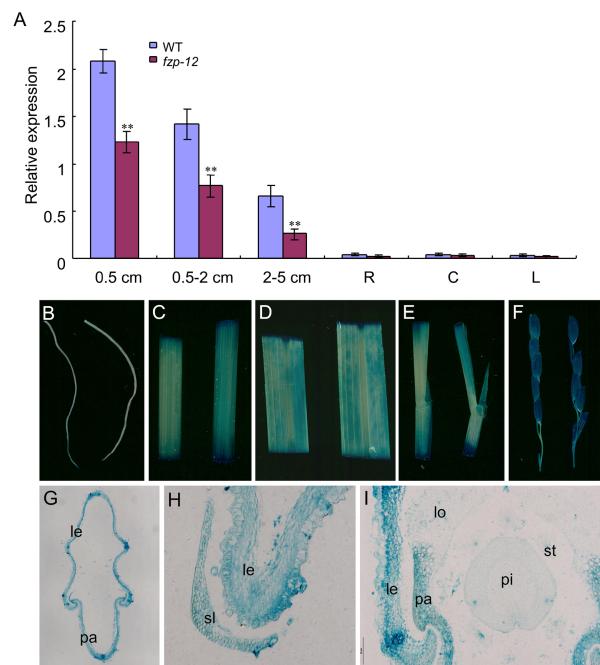
arrows indicate the mutated sites in the *fzp-12* and *fzp-13* mutants. *fzp-13* indicates the Cas9 knock-out plant.

Supplemental Figure 3



Supplemental Figure 3. *fzp-14* mutant and protein sequence alignment of FZP and *fzp-14*. A, *fzp-14* mutant. B, protein sequence alignment of FZP and *fzp-14*. *fzp-14* indicates the Cas9 knock-out plant.

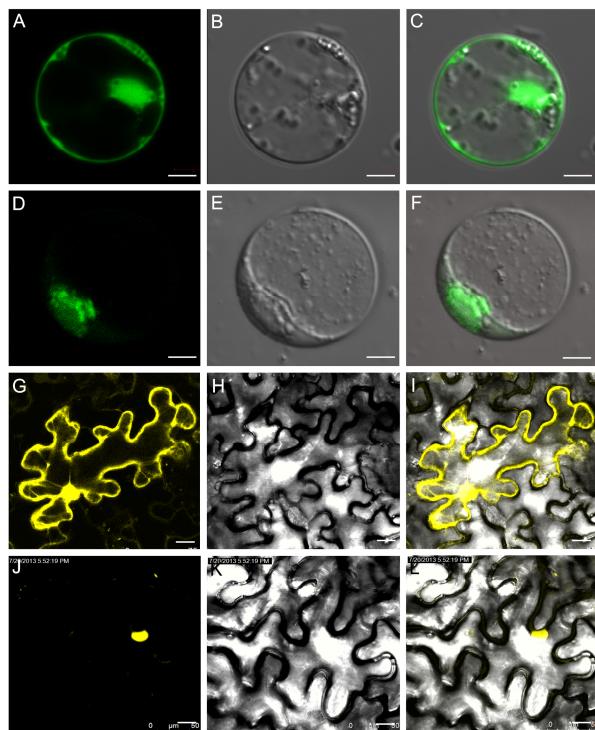
Supplemental Figure 4



Supplemental Figure 4. A, *FZP* expression in different tissues as detected by qPCR. B, GUS staining of root. C, GUS staining of culm. D, GUS staining of leaf. E, GUS staining of leaf sheath. F, GUS staining of spikelet. R, root; C, culm; L, leaf. G-I, Cross-section of the stained spikelets. G, Cross-section of hull. H, Cross-section of sterile lemma. I, Cross-section of inner floral organs. 0.5, young panicles (≤ 0.5 cm);

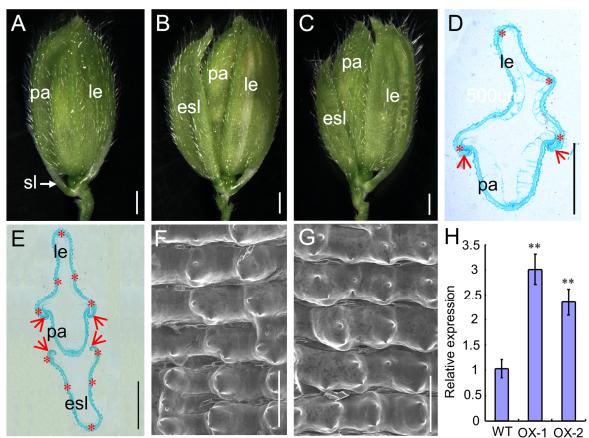
0.5-2, young panicles (0.5-2 cm); 2-5, young panicles (2-5 cm). le, lemma; pa, palea; sl, sterile lemma; st, stamen; pi, pistil. Error bars indicate SD. **Significant difference at $P < 0.01$ compared with the wild type by Student's t -test.

Supplemental Figure 5



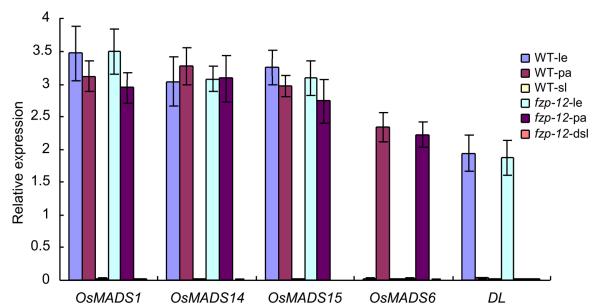
Supplemental Figure 5. Subcellular localization of the FZP protein. A-C, single GFP protein in rice protoplasts. D-F, FZP-GFP in rice protoplasts. G-I, single YFP protein in tobacco (*Nicotiana tabacum*) epidermal cells. J-L, FZP-YFP in tobacco (*Nicotiana tabacum*) epidermal cells. Bars = 5 μm in A-F; 10 μm in G-L.

Supplemental Figure 6



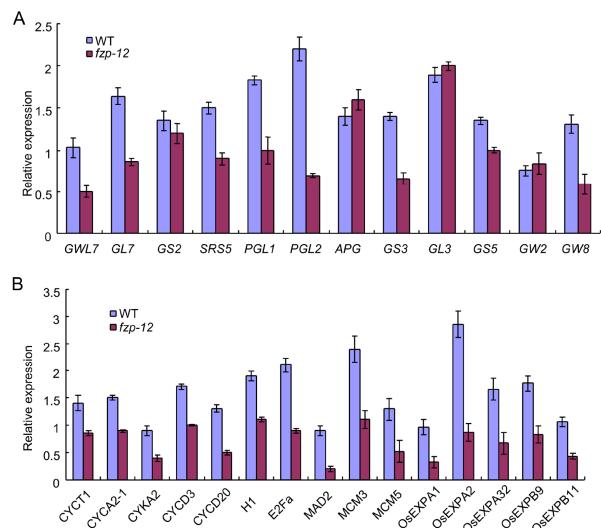
Supplemental Figure 6. A, wild type spikelet. B, spikelet in the *FZP* over-expressing line 1. C, spikelet in the *FZP* over-expressing line 2. D, histological analysis of wild type spikelet. E, histological analysis of spikelet in the *FZP* over-expressing plant. F, epidermal surface of the lemma in the wild type. G, epidermal surface of the sterile lemma in the *FZP* over-expressing plant. H, *FZP* expression analysis in the wild type and *FZP* over-expressing plants. esl, enlarged sterile lemma; sl, sterile lemma; le, lemma; pa, palea. Red stars indicate the vascular bundle. Red arrows indicate the inward hook-like structure. Error bars indicate SD. **Significant difference at $P < 0.01$ compared with the wild type by Student's *t*-test. Bars = 1000 μm in A-C; 500um in D and E; 100um in F and G.

Supplemental Figure 7



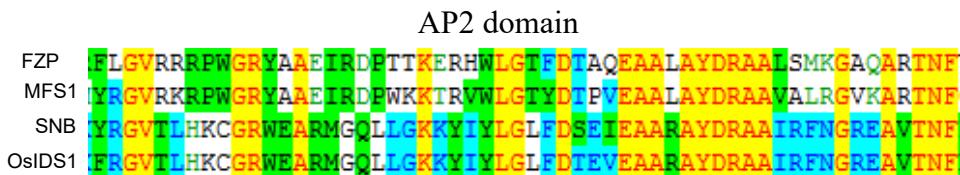
Supplemental Figure 7. Expression of floral organ identity genes in the wild type and *fzp-12* mutant. le, lemma; pa, palea; sl, sterile lemma; dsl, degenerated sterile lemma.

Supplemental Figure 8



Supplemental Figure 8. Expression levels of related genes in wild-type and *fzp-12* mutant. A, Expression analysis of related genes influencing grain size in young panicles. B, Expression analysis of cell proliferation and expansion-related genes in young panicles. Error bars indicate SD. **Significant difference at $P < 0.01$ compared with the wild type by Student's *t*-test.

Supplemental Figure 9. Protein sequence alignment of AP2/ERF genes.



Supplemental Tables

Supplemental Table 1

Cross combination	F1		F2		χ^2 (3:1)	P-value
	Wild type	<i>fzp-12</i>	Wild type	<i>fzp-12</i>		
<i>fzp-12</i> /N6	57	0	3612	1189	0.1406	0.7707
N6/ <i>fzp-12</i>	48	0	3025	1043	0.8863	0.3465

Supplemental Table 1. Genetic analysis of the mutant phenotypes of *fzp-12*.

Supplemental Table 2. Primers used in the study.

Purpose	Primer name	Sequence
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Mapping	B7-13-1F	ACAGTATCCAAGGCCCTGG
	B7-13-1R	CACGTGAGACAAAGACGGAG
	A7-29-1F	AGTTTCCTTCTTTCTAC
	A7-29-1R	CCTCAGCATGATAACCCAC
	A7-33-1F	GCCCCAAGATTATGAGA
	A7-33-1R	AAGCTCCGACAATACA
	A7-3-1F	GAGTTCAAGGAAGCACAAAA
	A7-3-1R	CAATGAGACTGTTGCTAACCC
	A7-9-1F	TGAGGAGGAGGAGATGGTCG
	A7-9-1R	AGGTGCCCTTGCAGTTGA
	A7-13-1F	TGCTACAAGTACGTGCTTT
	A7-13-1R	GTGGTTGGAGGCGATTTTAT
	A7-25-1F	TGCGTGGTGCAGCATTGTA
	A7-25-1R	GGCATCACATTGAGCAAGAAA
	A7-26-1F	ACGTGCTTTCTTGTACCTT
	A7-26-1R	TTTGTGGTTGGAGGCGATT
	S2-1F	CCTTAAGCTTGCATGGAG
	S2-1R	GGTGGCCGGTGAAAAGT
	S14-1F	GAGCTTGCCCTACAAGG
	S14-1R	CGGGGAACAATGTAACAC
qRT-PCR	<i>FZP</i> -1F	GCATGGCTAATCACGCACTT
	<i>FZP</i> -1R	CCAAGCCACTCTTCTTGTTCG
	<i>OsMADS1</i> -1F	
	<i>OsMADS1</i> -1R	TGATGGTGAGCATGAGGGTG
	<i>OsMADS14</i> -1F	CCATTAACGAGCTTCAACGG
	<i>OsMADS14</i> -1R	TGGTATGGATCTGAAGCCTCC
	<i>OsMADS15</i> -1F	AGTACGCCACTGACTCCAGG
	<i>OsMADS15</i> -1R	TGCTGGCCCCTCACATTC
	<i>OsMADS6</i> -1F	CCAACAATGCACTTCTGAAAC
	<i>OsMADS6</i> -1R	GGAGGCTTGCTGCATGGC
	<i>DL</i> -1F	CCCATCTGCTTACAACCGCTT
	<i>DL</i> -1R	GTTGGAGGTGGAAACCGTCG
	<i>GI</i> -1F	GGCGTCTACTGCCATTCTG
	<i>GI</i> -1R	TCGATCAGCATCAAAGCACAG
	<i>ACTIN</i> -1F	AGCAACTGGATGATATGGA
	<i>ACTIN</i> -1R	CAGGGCGATGTAGGAAAGC
GUS staining	<i>proFZP-GUS</i> -1F	CAAGAGGAATTAACAGTGCTAATTTC
	<i>proFZPGUS</i> -1R	GTGTTCATGACCAATGCAATATTG
In Situ Hybridization	<i>GIH</i> -1F	CGCCTACGACGAGCGTCA
	<i>GISP6</i> -1R	AGATTAGGTGACACTATAGAAAGCAAC
		CACAGTGAGAAGCACAG
	<i>DLH</i> -1F	GGGCCGCAATGGATCTCGTG
	<i>DL SP6</i> -1R	AGATTAGGTGACACTATAGAATCACAA

		CGAAGGGTGCTCT
Complementation test	<i>FZPcom</i> -1F	CAAGAGGAATTAACAGTGCTAATTTC
	<i>FZPcom</i> -1R	CCACCACCTGCGTGTATATG
	<i>FZPOE</i> -1 F	ATGAACACTCGAGGCAGCGG
	<i>FZPOE</i> -1 R	TCAATGGGAGAGGAAGCTGAATG
Over-expressing	<i>FZPOE</i> -1F	ATGAACACTCGAGGCAGCGG
	<i>FZPOE</i> -1R	TCAATGGGAGAGGAAGCTGAATG
EMSA probe (specific)	GCC-1F	CATAAGAGGCCACTCATAAGAGGCC
	GCC-1R	ACTCATAAAGAGCCGCCACT
		GTATTCTCGGCGGTGAGTATTCTCGGCGG
		TGAGTATTCTCGGCGGTGA
EMSA probe (non-specific)	GCC-2F	CATAAGATACTGACTCATAAGATACTTGA
	GCC-2R	CTCATAAAGATACTTGA
		GTATTCTATGAACTGAGTATTCTATGA
		GAGTATTCTATGA
EMSA probe (specific)	DRE-1F	ATACTACCGACATGAGATACTACCGACAT
	DRE-1R	GAG ATACTACCGACATGAG
		TATGATGGCTGTACTCTATGATGGCTGTAC
		TCTATGATGGCTGTACTC
EMSA probe (non-specific)	DRE-2F	ATACGTCTTGACGAGATACGTCTTGAC
	DRE-2R	GAGATACGTCTTGACGAG
		TATGCAGAAACTGCTATGCAGAACTG
		CTC TATGCAGAAACTGCTC