

Structure–Activity Relationships (SARs) of the Competence Stimulating Peptide (CSP) in *Streptococcus mutans* Reveal Motifs Critical for Membrane Protease SepM Recognition and ComD Receptor Activation

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Additional experimental details

Resin loading. 0.1 g of Wang resin (0.94 mmol/g) was placed in a clean, dry round bottom flask with a minimal amount of dimethylformamide (DMF) to fully cover the resin, and the resin was allowed to swell at room temperature for 30 min. 0.94 mmol (10 equiv. relative to the overall loading of the resin) of Fmoc-protected amino acid were dissolved in dry dichloromethane (DCM) and placed on an ice bath. A few drops of DMF were added to aid complete dissolution when the amino acid had low solubility in DCM. A solution of 0.47 mmol (5 equiv. relative to the overall loading of the resin) diisopropylcarbodiimide (DIC) in dry DCM was then added dropwise to the amino acid solution. The mixture was stirred for 20 min on an ice bath and the DCM was removed from the mixture using a rotary evaporator. The residue was then dissolved with a minimal amount of DMF and the solution was added to the resin. 0.0094 mmol (0.1 equiv. relative to the overall loading of the resin) of dimethylaminopyridine (DMAP) were dissolved in DMF and the solution was added to the reaction mixture. The mixture was sealed and allowed to stand at room temperature for 2 h with occasional swirling. The resin was washed with DCM, then diethyl ether, followed by overnight drying under vacuum.

Loading test. Two small samples of the amino acid-loaded resin (approximately 5 mg each) were weighed and mixed with 3 mL of fresh 20% piperidine in DMF solution. This solution was stirred on a shaker for 2 h. The absorbance (290 nm) of the supernatants of the two piperidine-resin mixtures was measured by using a UV-Vis spectrophotometer to estimate the level of Fmoc removal, which correlates to first residue attachment. The first residue attachment was estimated from the following equations:

$$\text{Equation 1} \quad A \text{ (mmol/g)} = B \times 1000 / [1000 + (B \times (M - X))]$$

A is the theoretical substitution (mmol/g), B is the substitution of starting resin (mmol/g), and M is the molecular weight of target peptide with all protecting groups. X is 18 for hydroxymethyl-based resin.

$$\text{Equation 2} \quad B \text{ (mmol/g)} = (Abs_{\text{sample}} \times N) / (\text{mg of sample} \times 1.75)$$

B is the experimental first residue attachment and the equation is based on $\epsilon = 5253 \text{ M}^{-1} \text{ cm}^{-1}$. N is the fold of dilution of the supernatant. $\frac{B}{A} \times 100\%$ is the percent yield of the loading. If the average of the percent yield of the two samples was greater than 70%, resin loading was deemed successful and the resin would be used for the synthesis of CSP analogs.

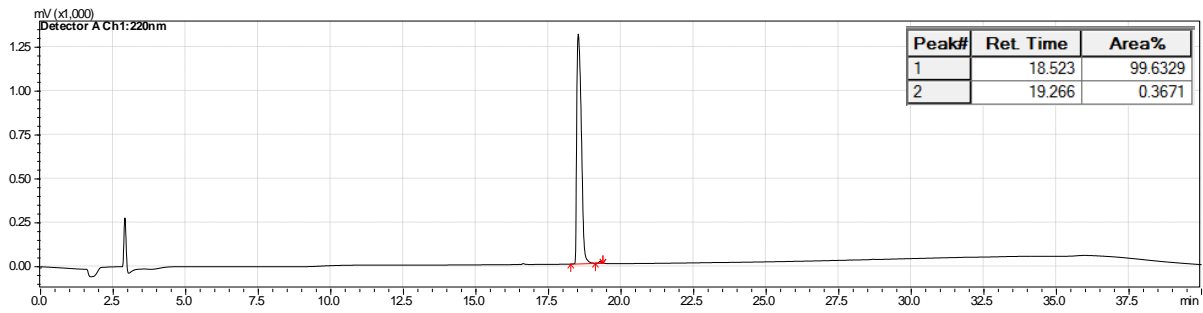
Synthesis. The resin (0.1 g) was first swelled by suspension in DCM for 30 min at room temperature and then drained. The resin was then washed with DMF (3 × 2 mL). To remove the Fmoc-protecting group, the resin was treated with piperidine (2 mL of 20% piperidine in DMF, 2 × 7 min) and washed with DMF (3 × 2 mL). To couple each amino acid, Fmoc-protected amino acids (2 equiv. relative to the overall loading of the resin) were dissolved in DMF (2 mL) and mixed with 2-(1H-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate (HBTU; 2 equiv.) and diisopropylethylamine (DIPEA; 2 equiv.). The solution was allowed to pre-activate for 1 min prior to being added to the resin and agitated for 1 h at room temperature. After each coupling step, the resin was drained and washed with DMF (2 × 2 mL). This process was repeated until the desired peptide sequence was obtained. CSP and analogs were synthesized either manually or with the AAPPTec Focus-XC peptide synthesizer.

Cleavage. Upon completion of peptide synthesis, the resin was washed with diethyl ether (2 mL) and dried under nitrogen stream for 3 min before it was transferred into a 15 mL falcon tube. The peptide was cleaved from the resin, along with all of the protecting groups, by mixing the resin with 3 mL cleavage cocktail of

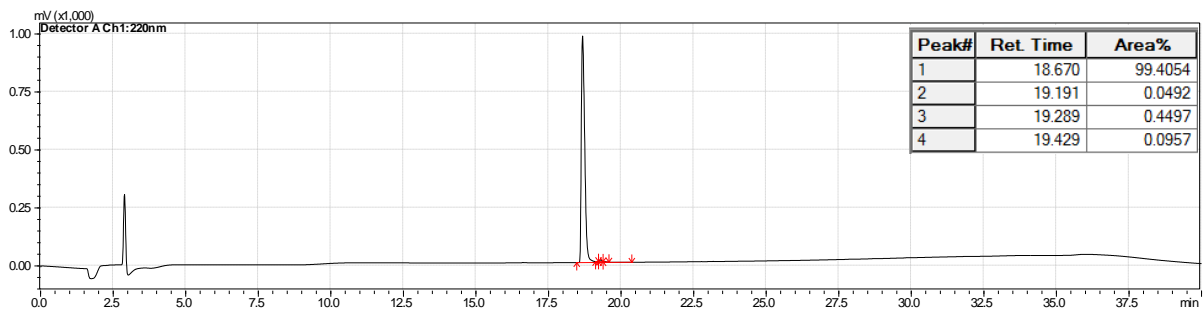
95% trifluoroacetic acid (TFA), 2.5% triisopropylsilane (TIPS), and 2.5% 18 MΩ water for 3 h with agitation. The resulting cleavage product solution was separated from the resin by filtration and the filtrate was transferred into a new 50 mL falcon tube. A cooled solution of diethyl ether:hexane (1:1, 45 mL, 0 °C) was added to the filtrate, and the peptide was allowed to precipitate for 10 min in a freezer at -20 °C. The mixture was centrifuged for 5 min at 3000 RPM and the supernatant was removed to yield crude peptide that was dissolved in 10 mL acetonitrile (ACN):water (1:1) and lyophilized before HPLC purification.

HPLC traces for CSP analogs

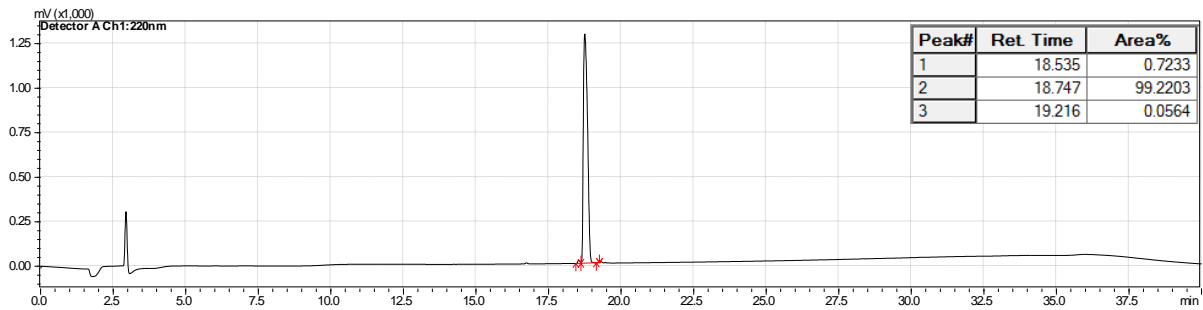
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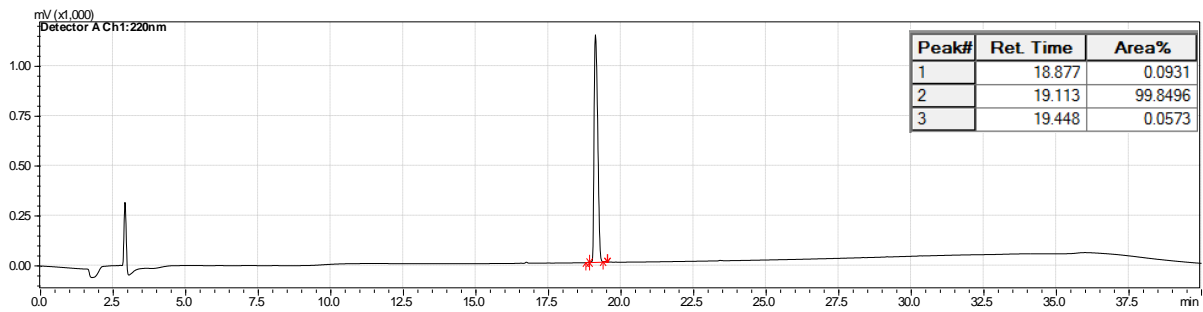
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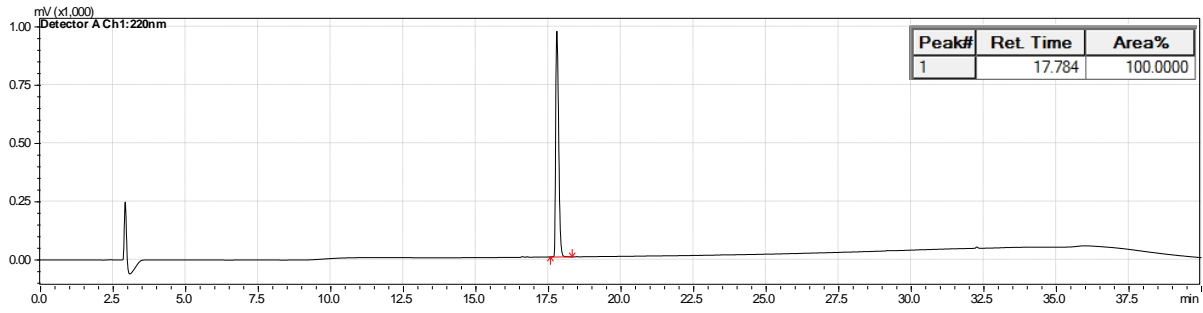
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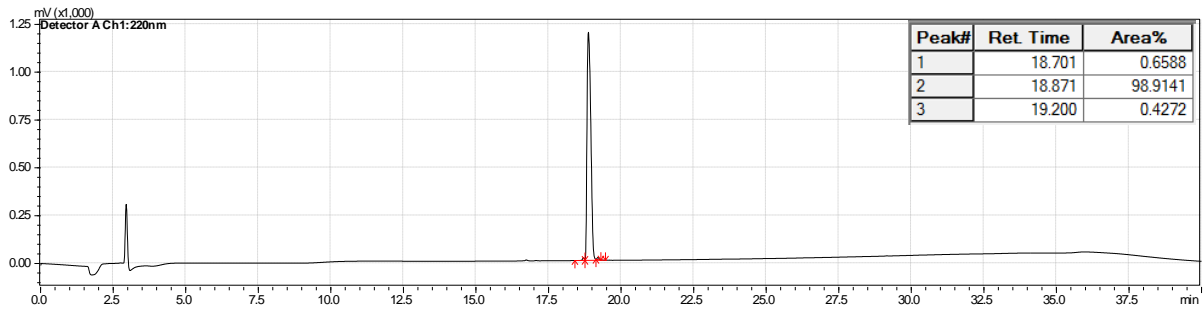
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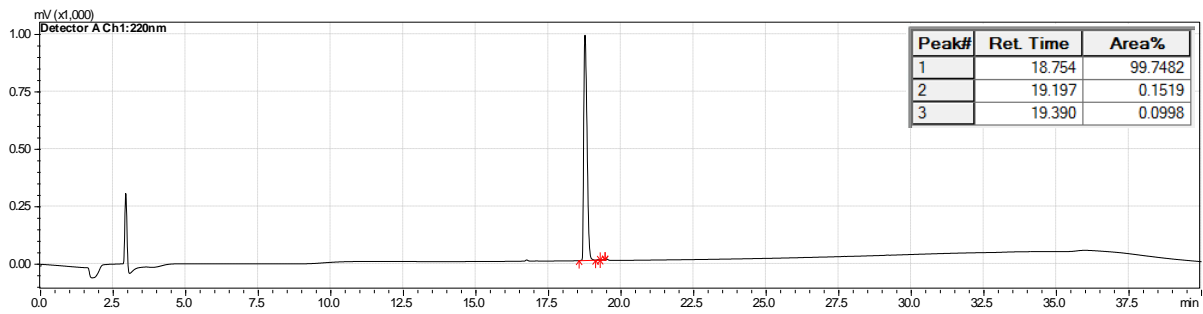
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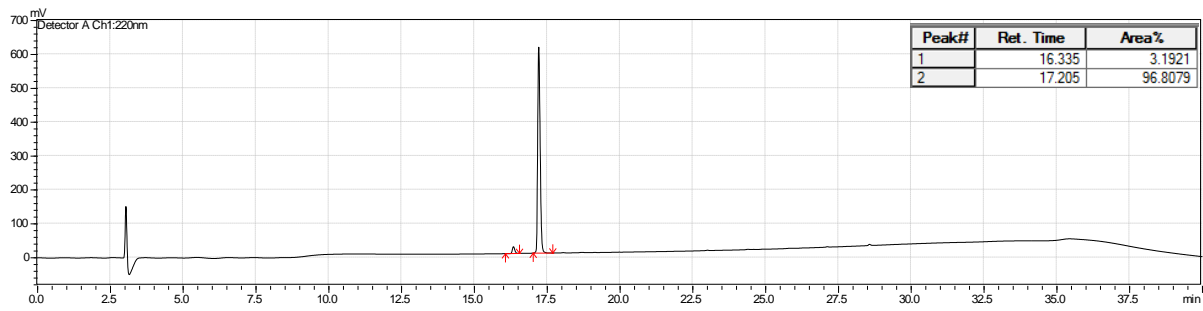
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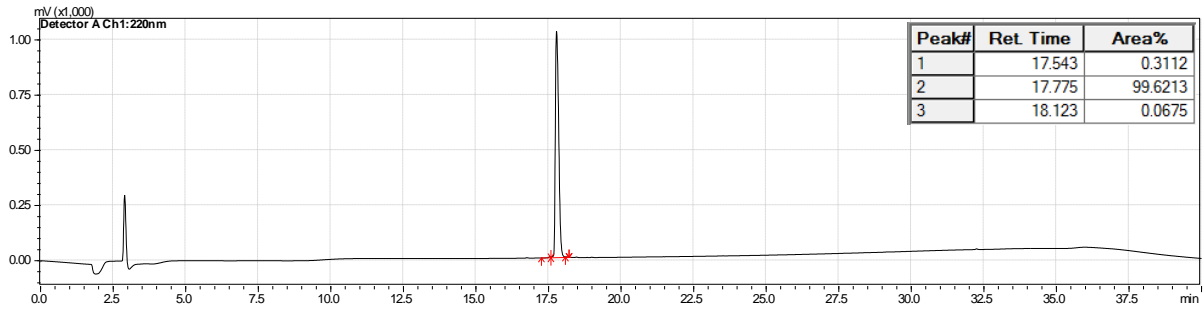
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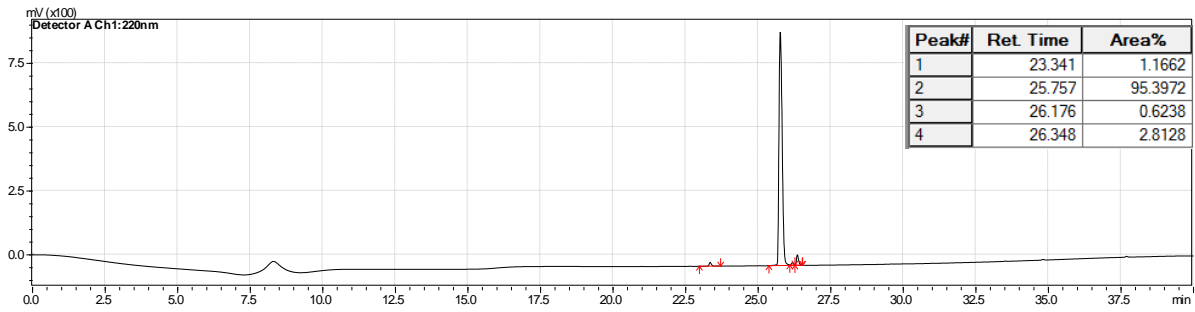
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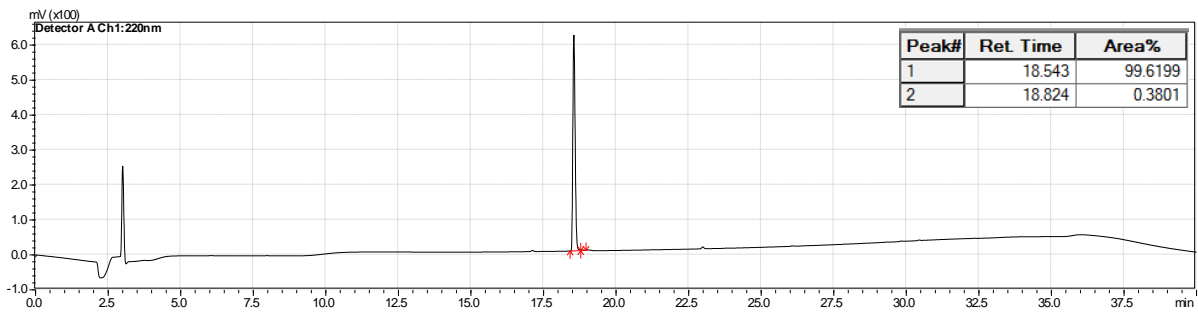
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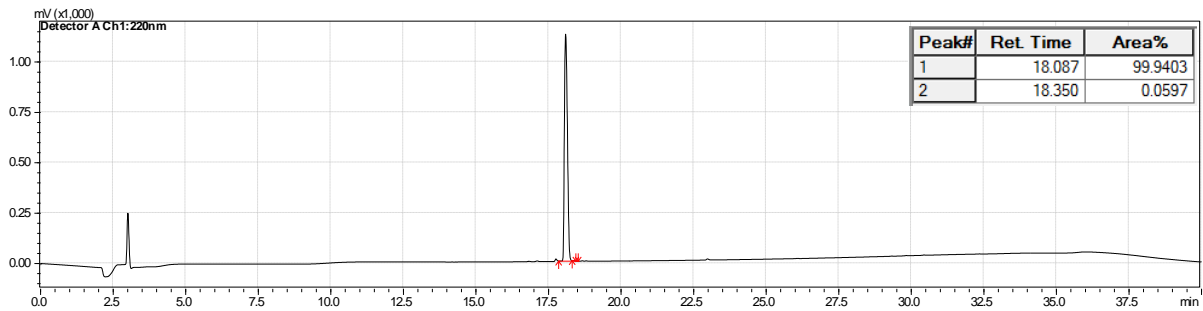
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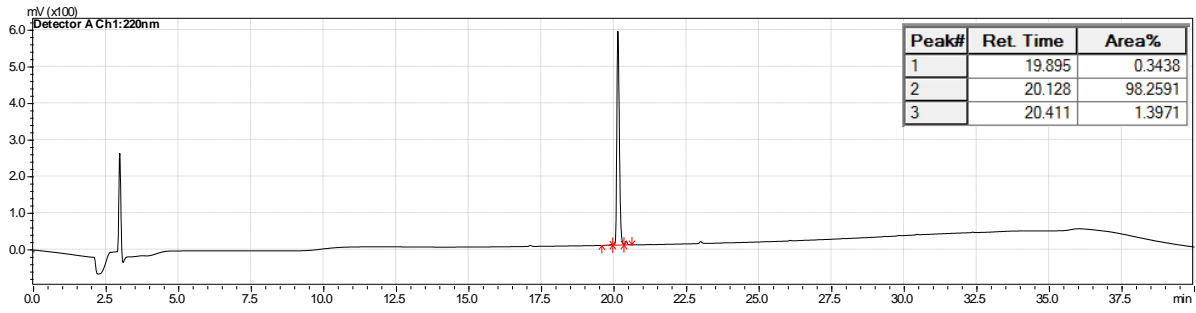
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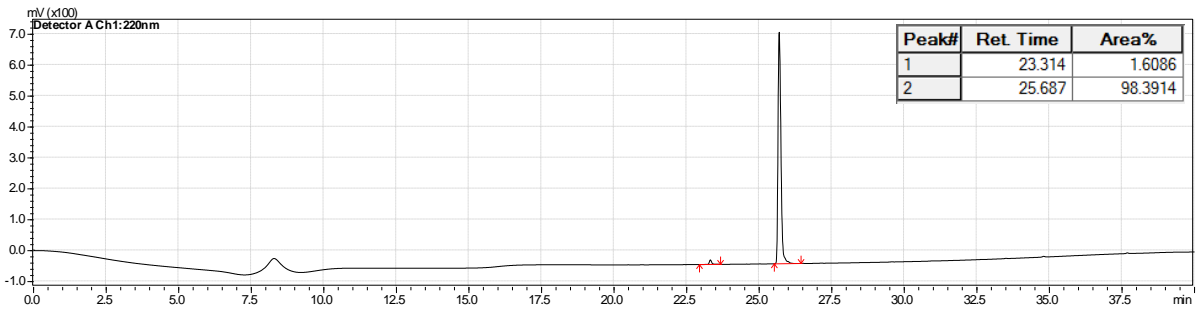
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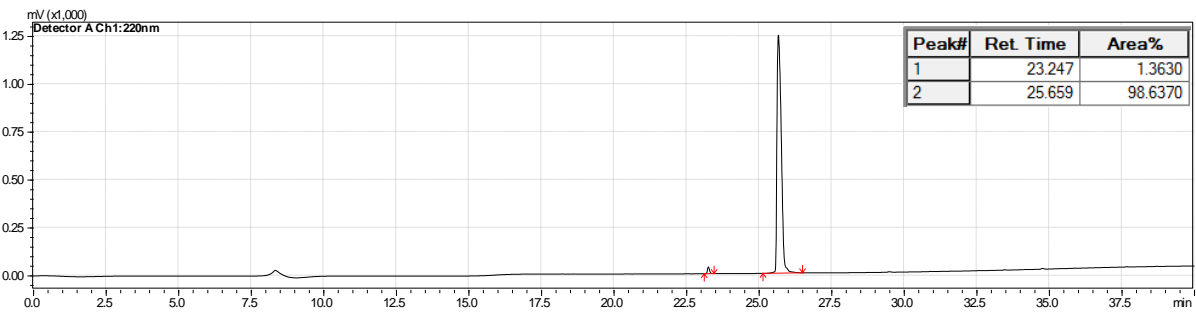
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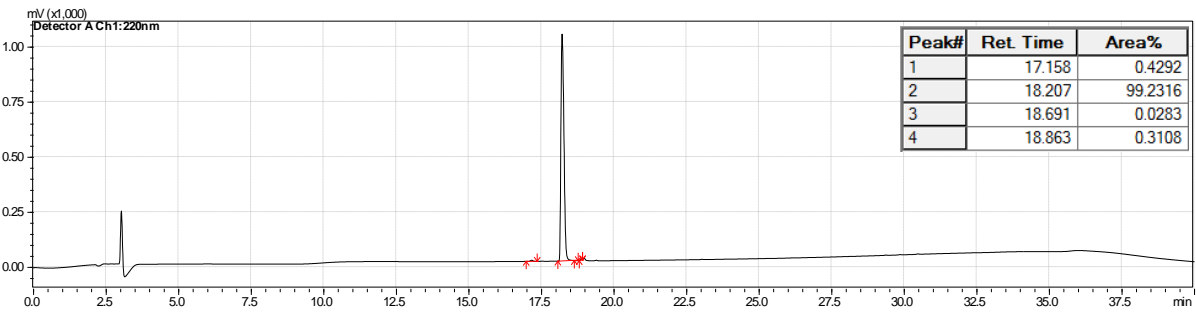
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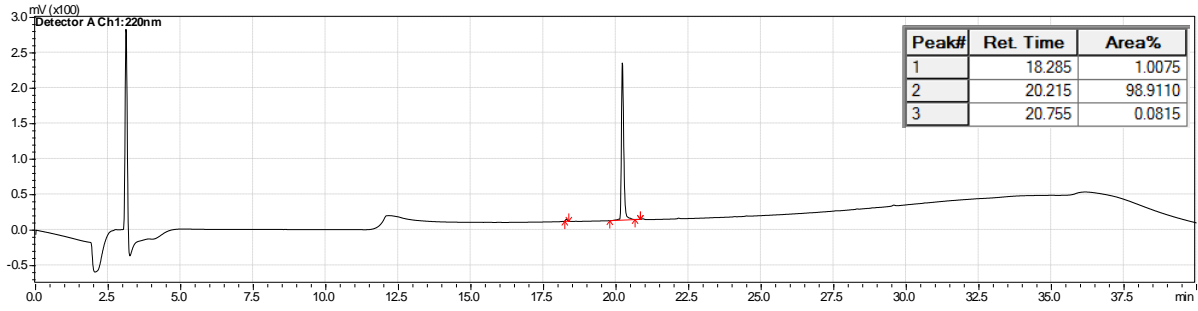
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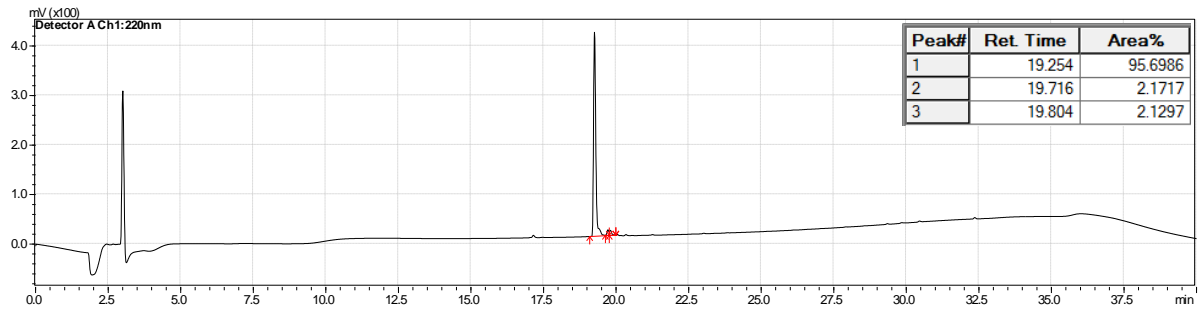
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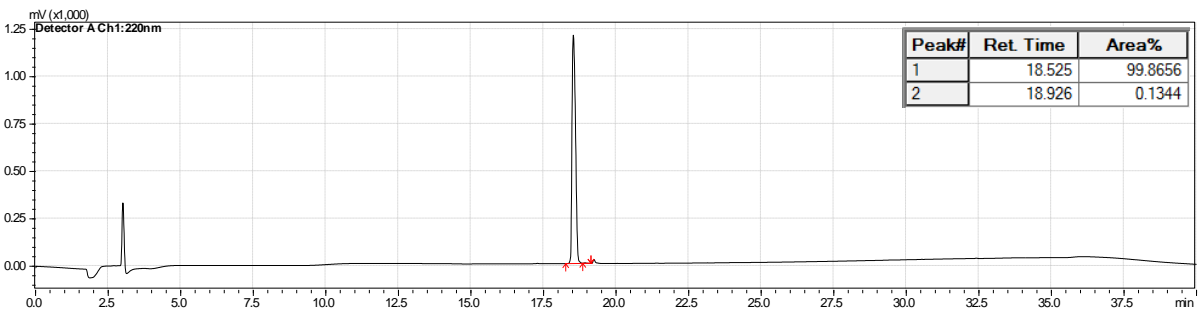
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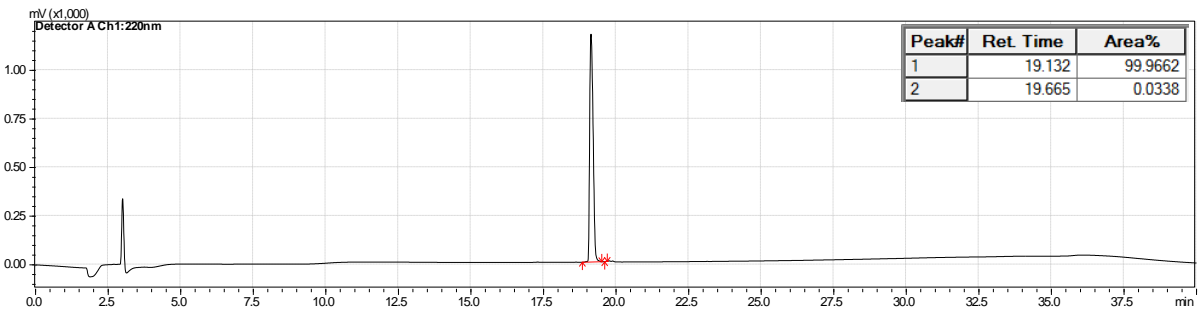
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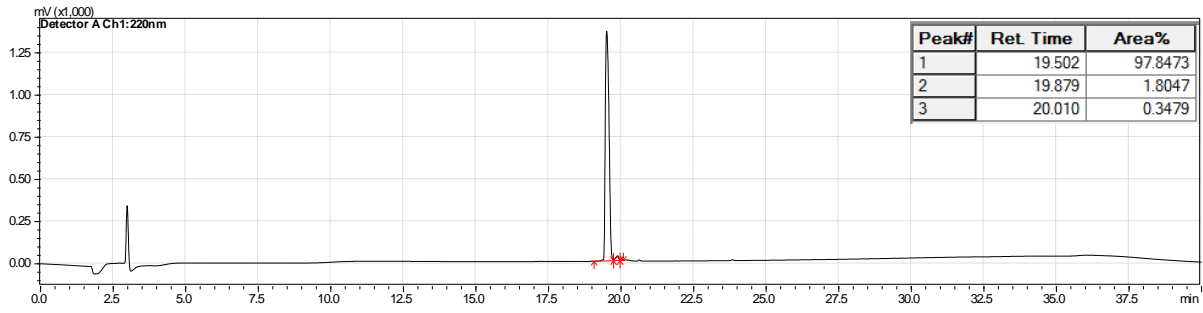
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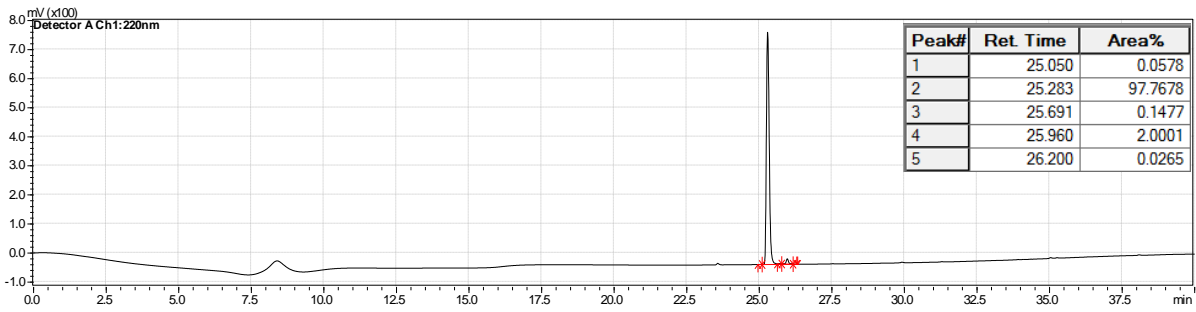
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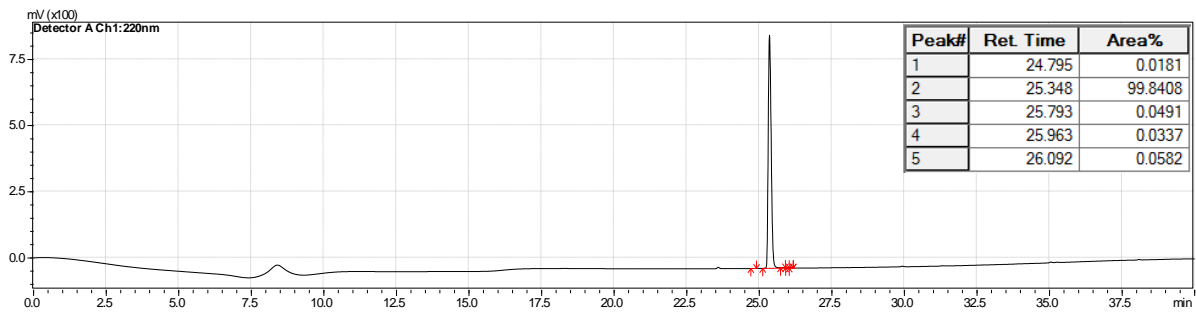
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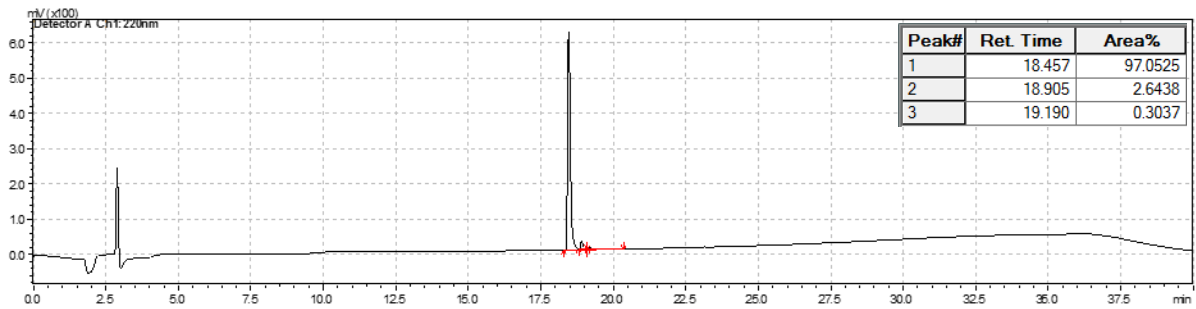
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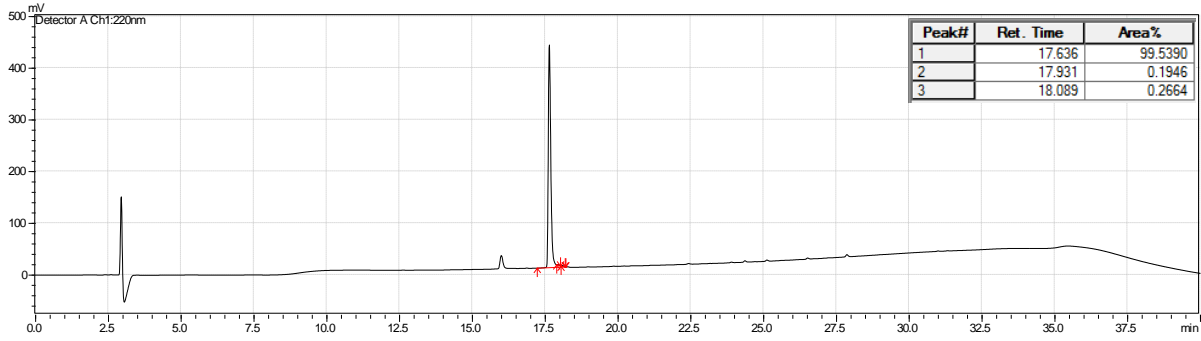
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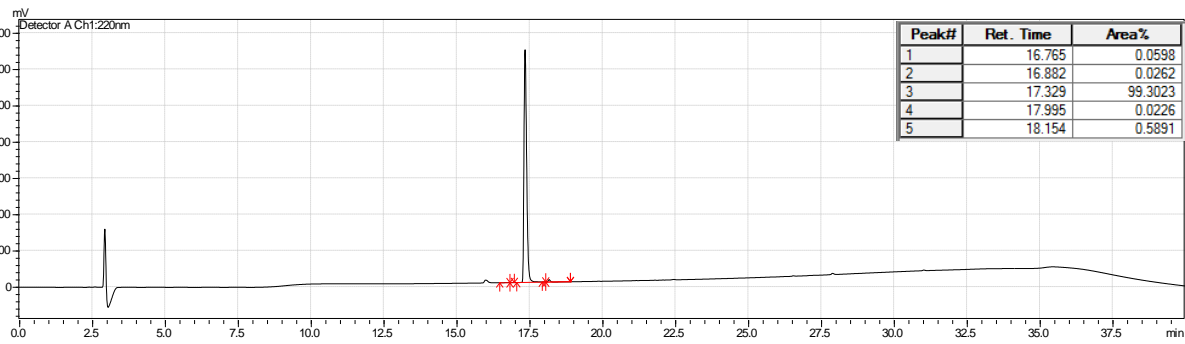
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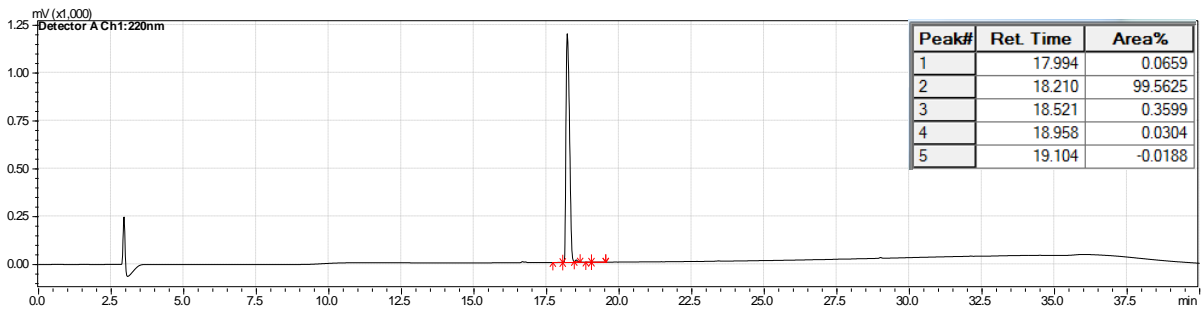
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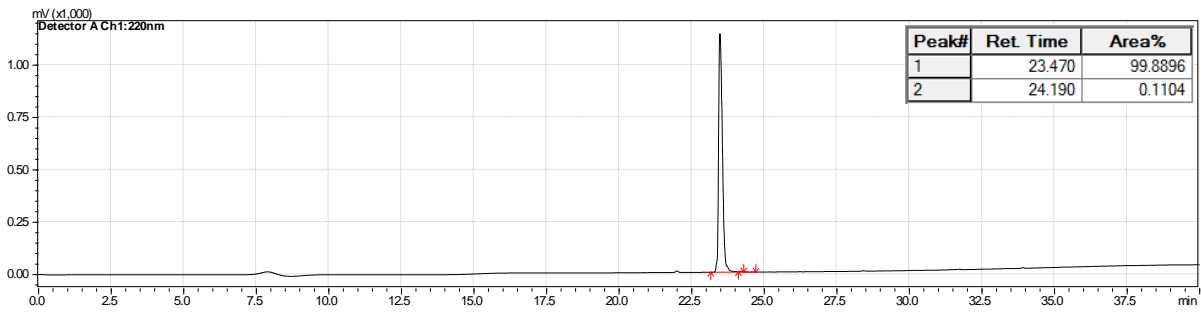
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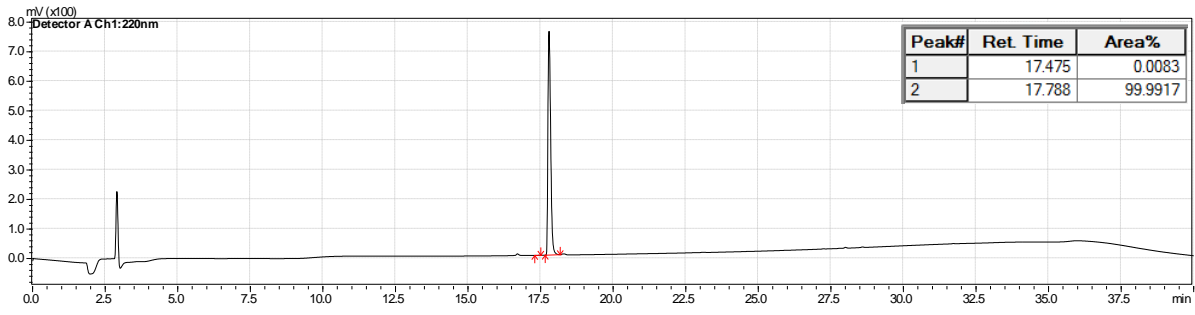
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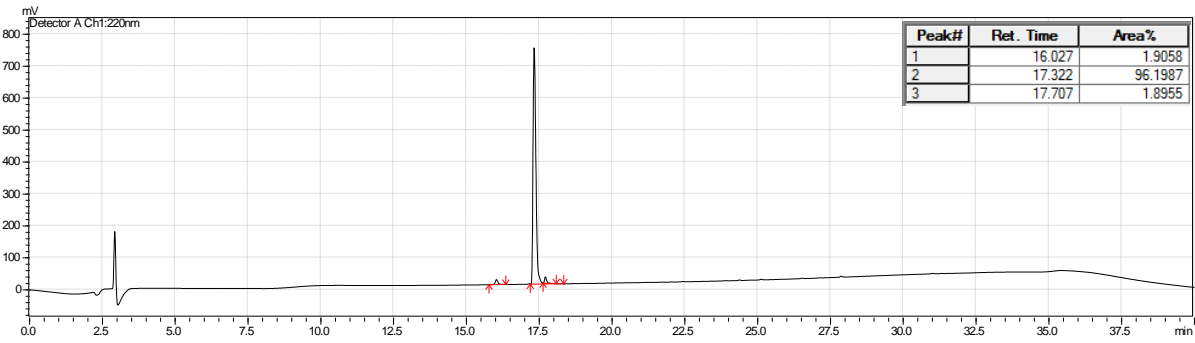
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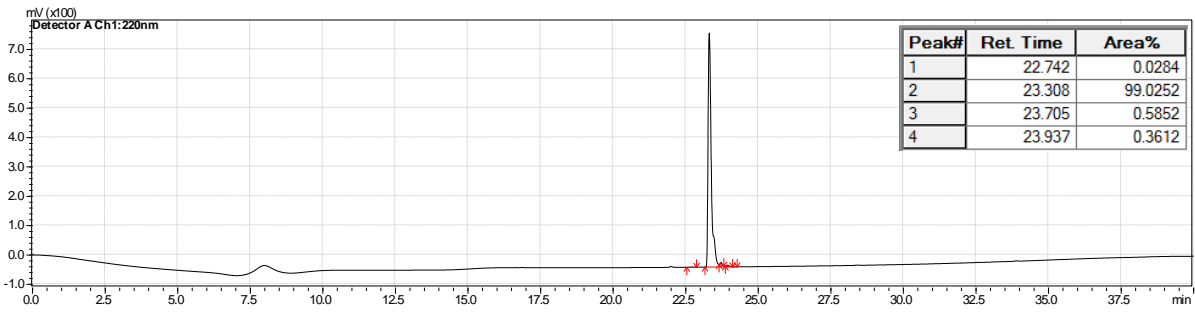
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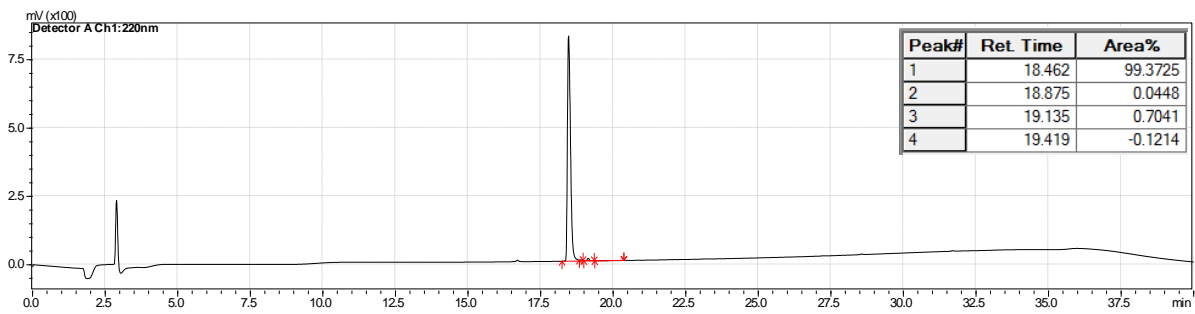
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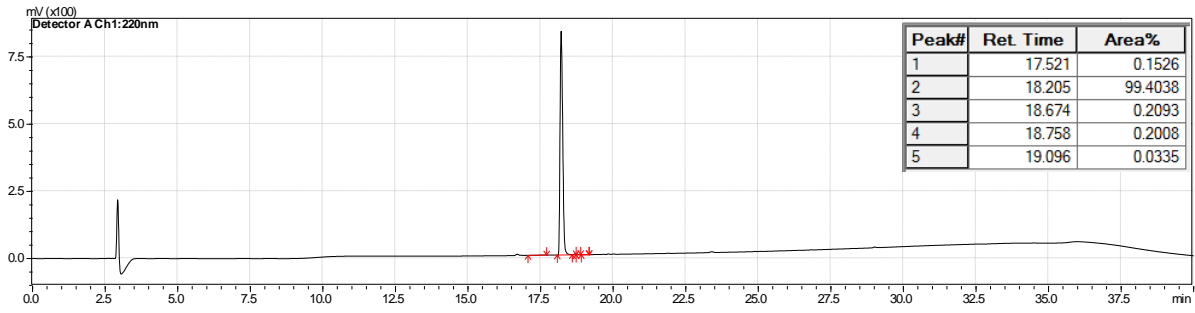
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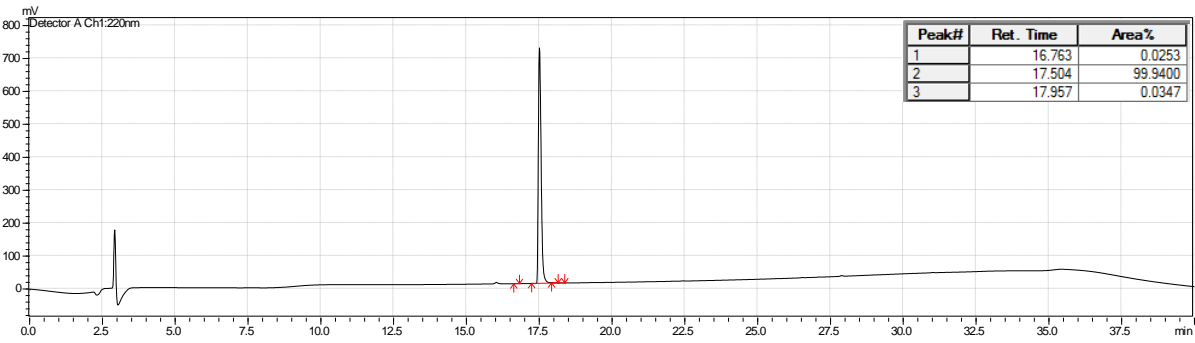
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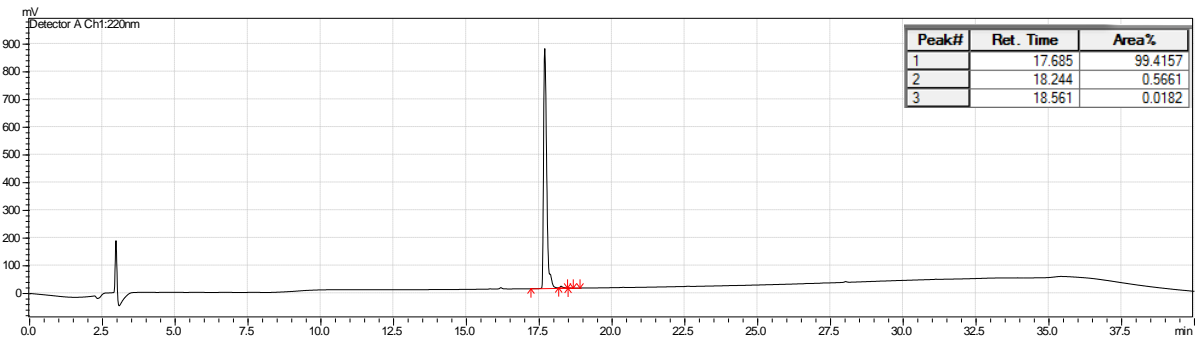
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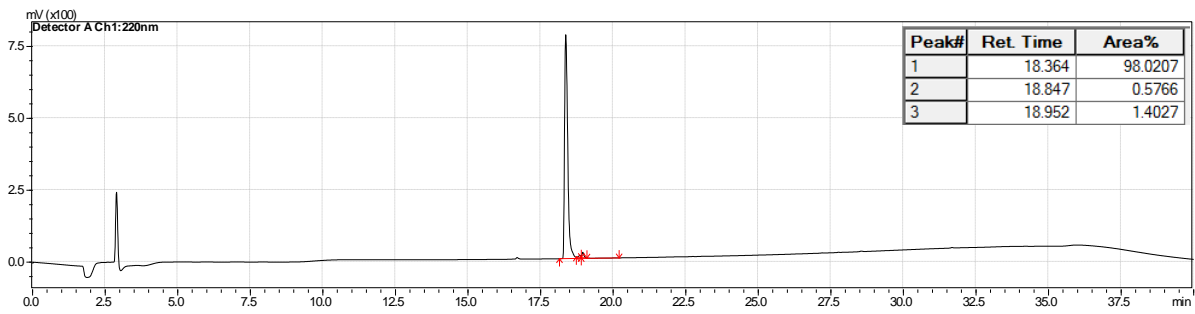
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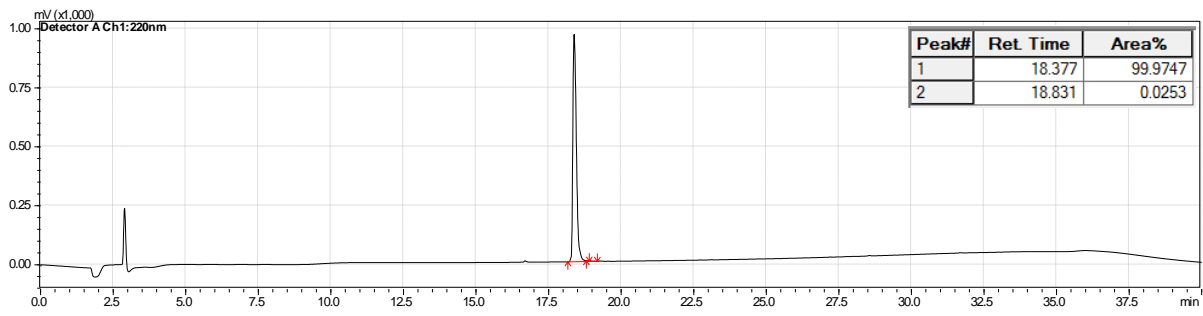
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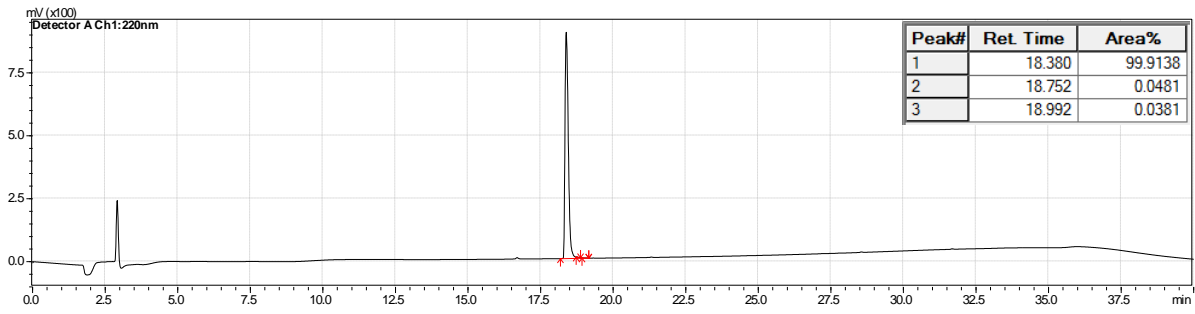
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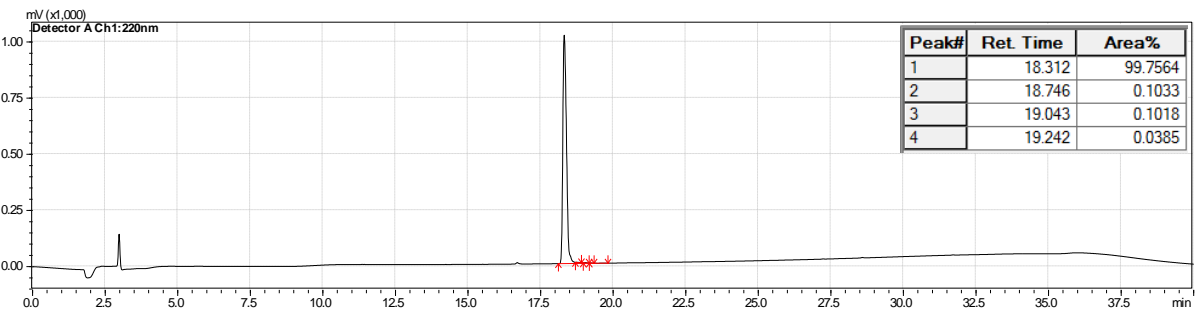
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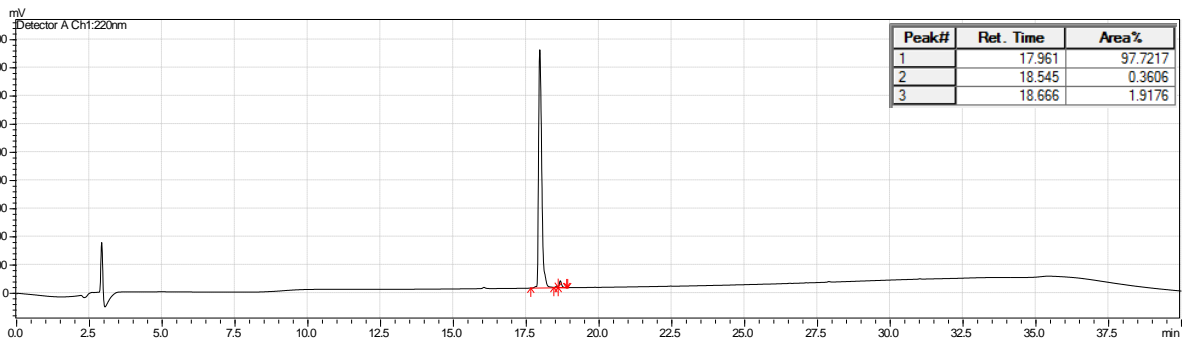
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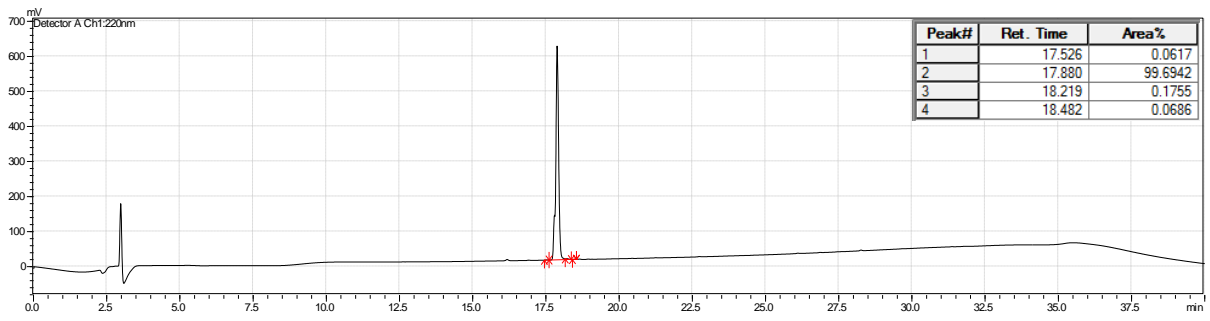
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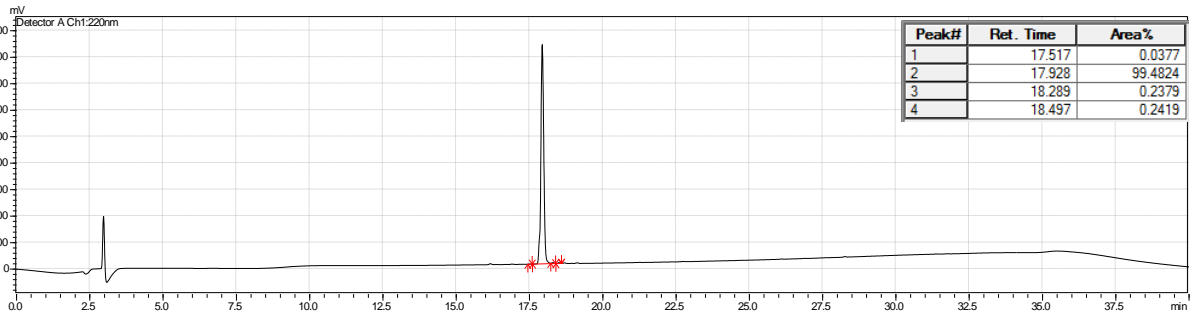
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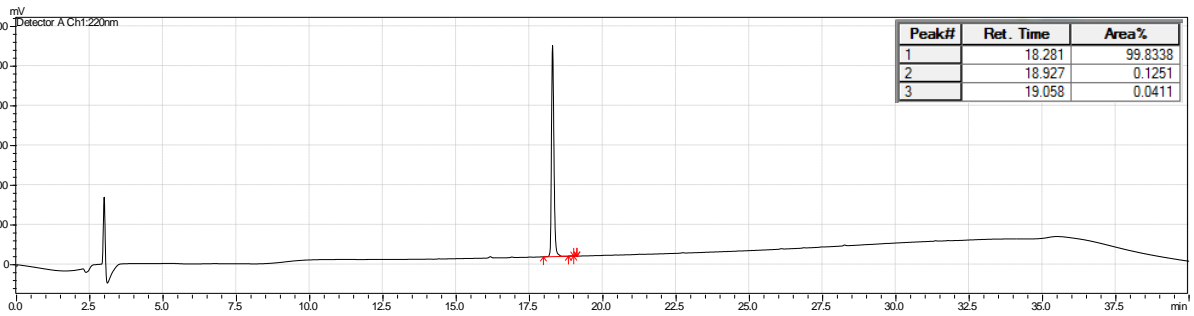
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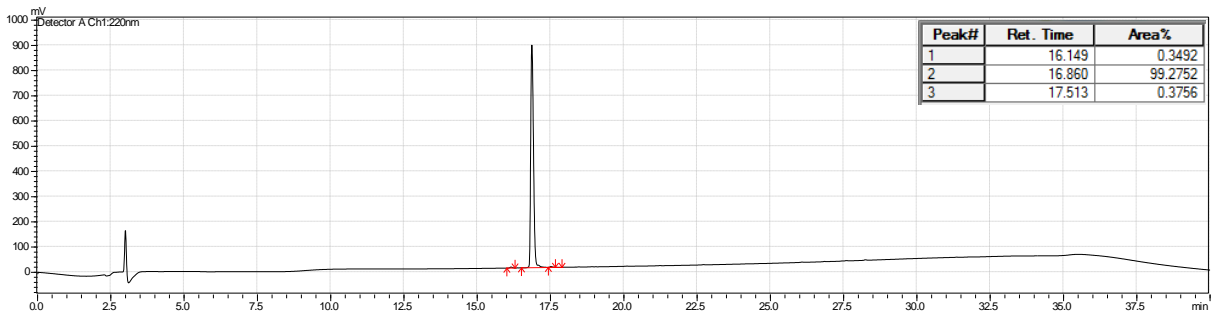
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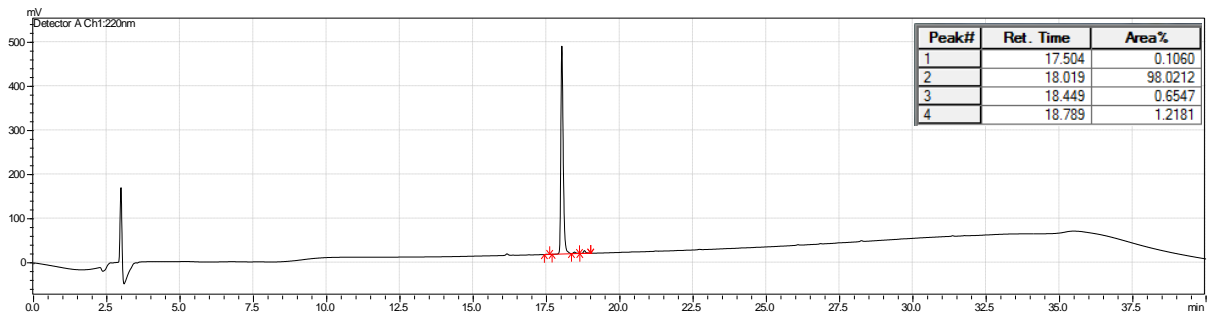
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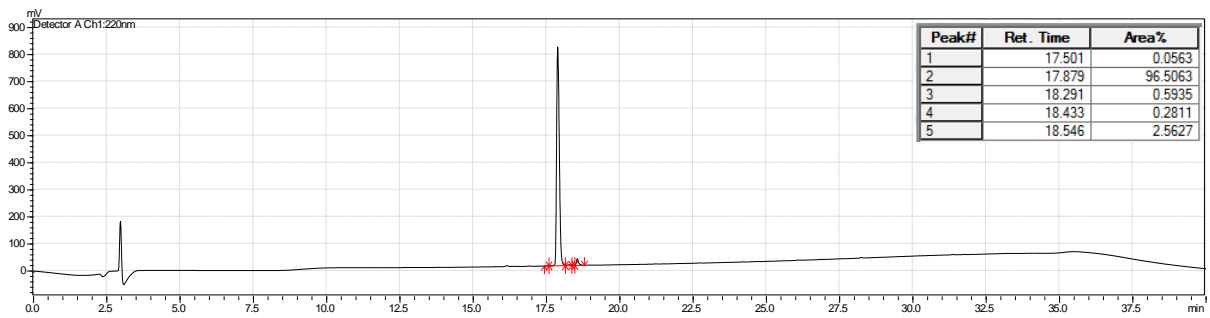
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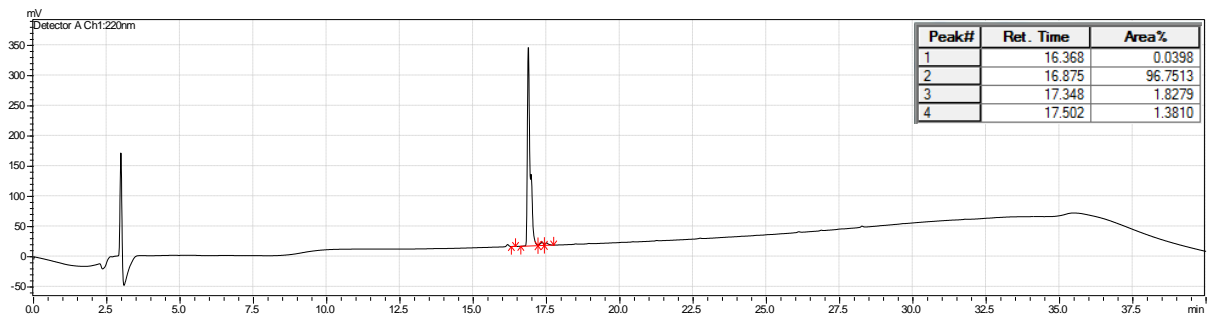
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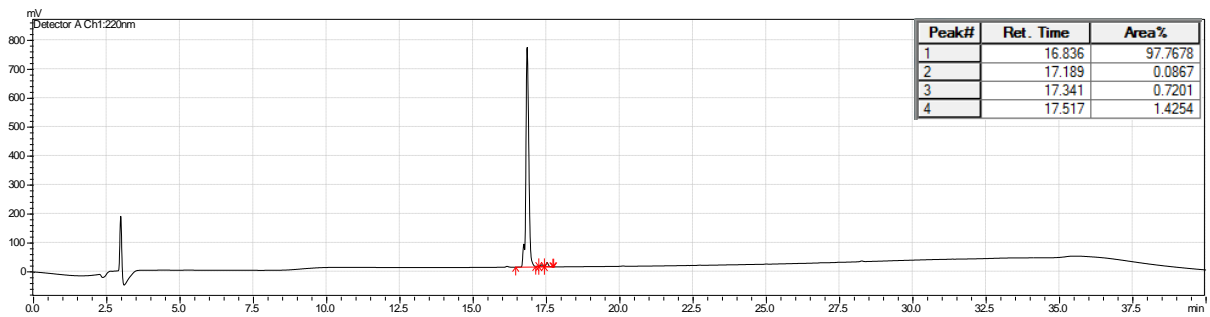
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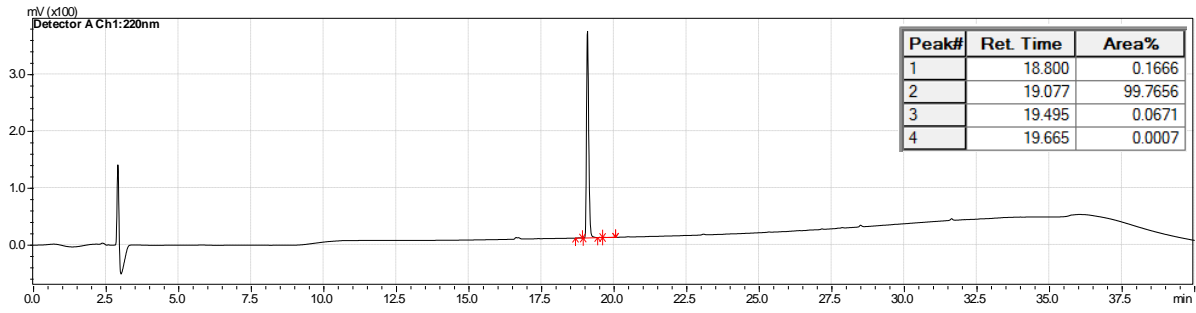
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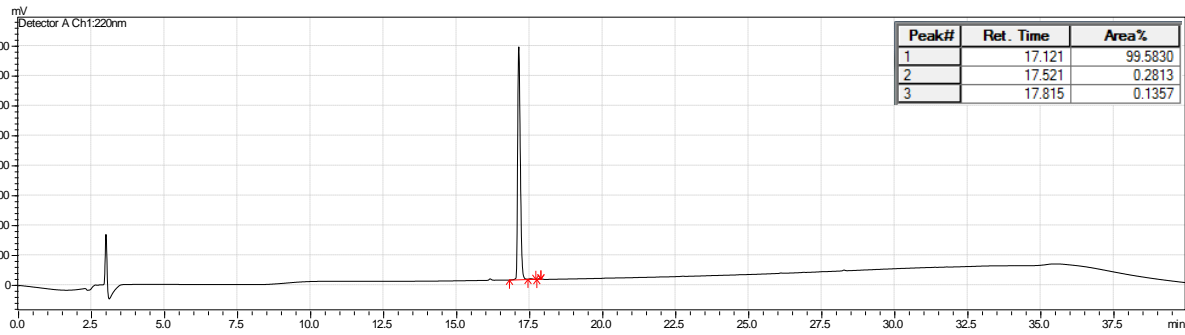
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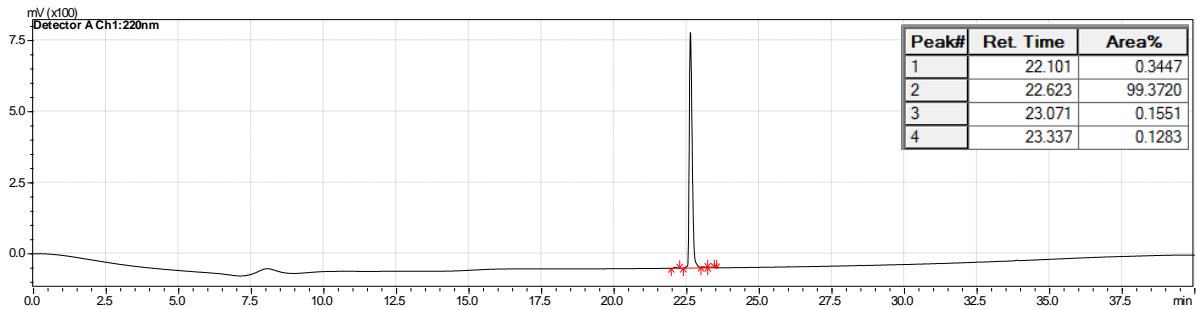
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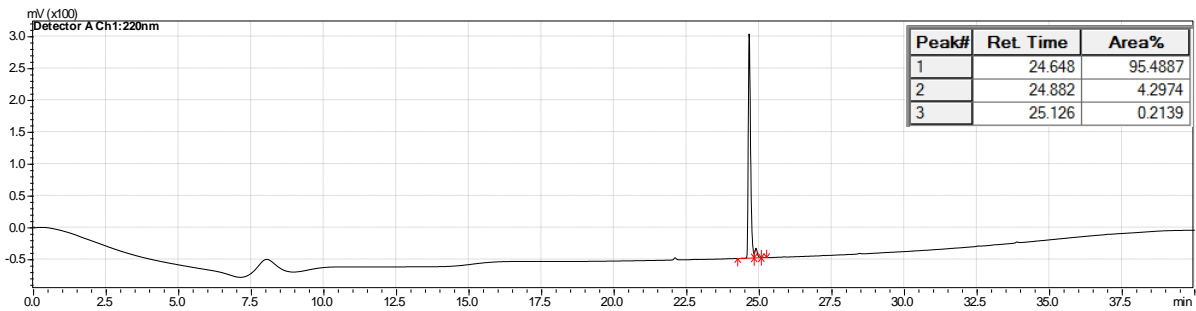
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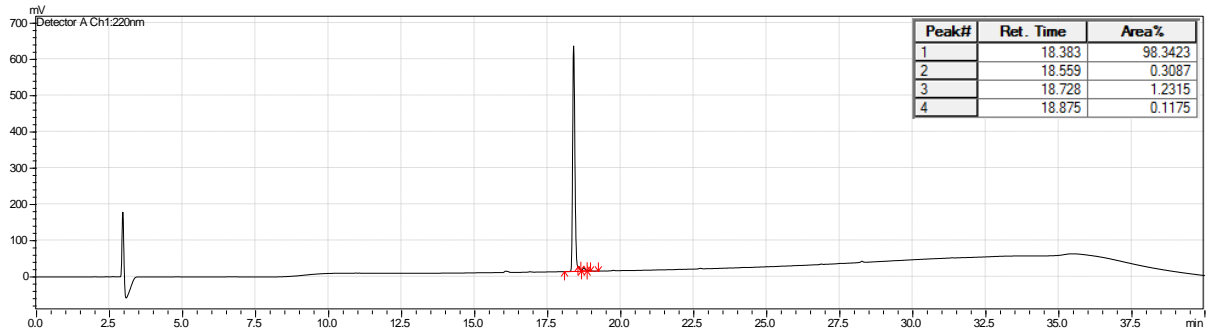
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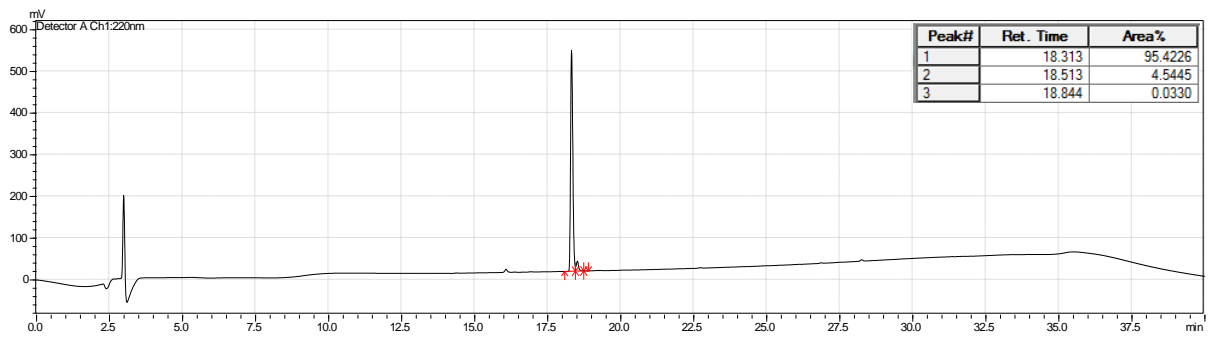
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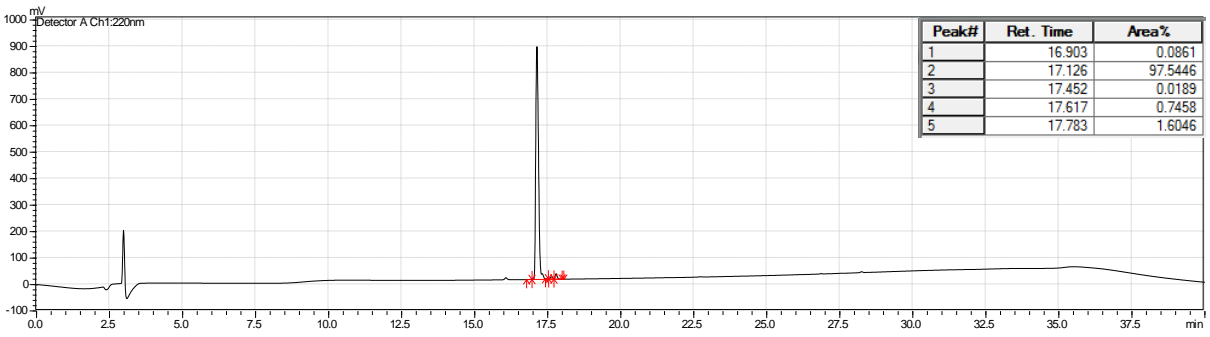
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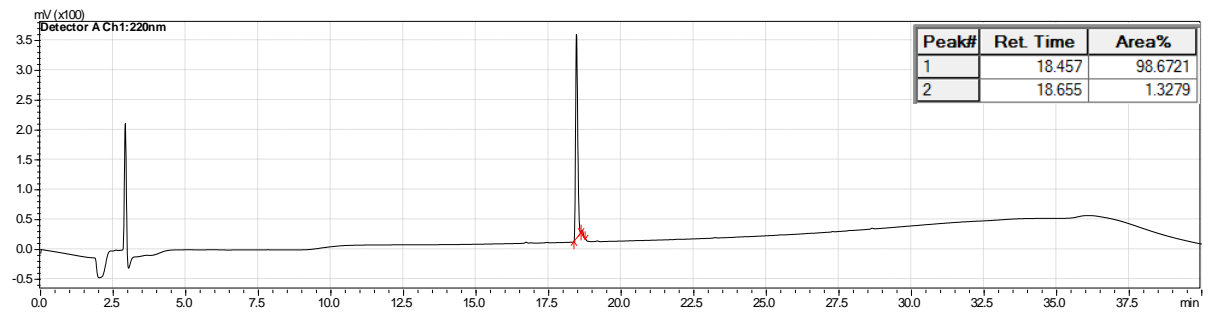
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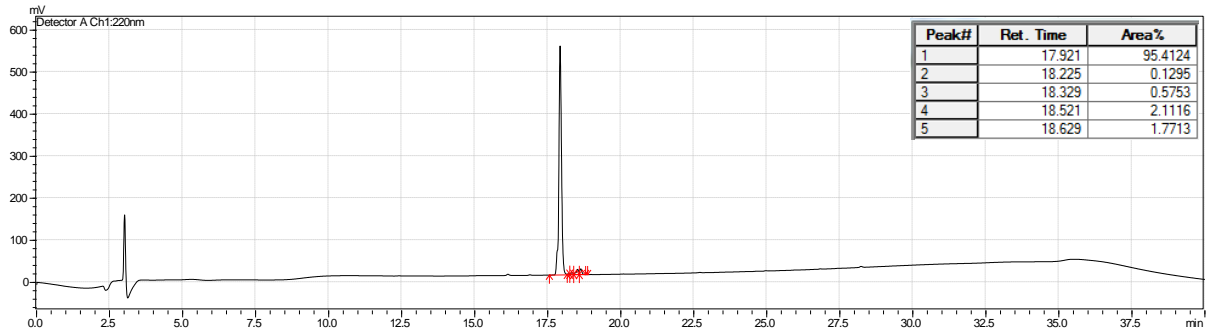
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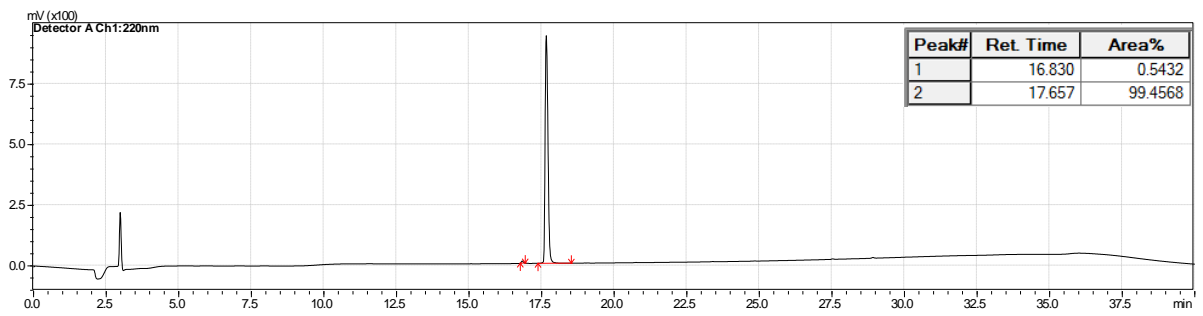
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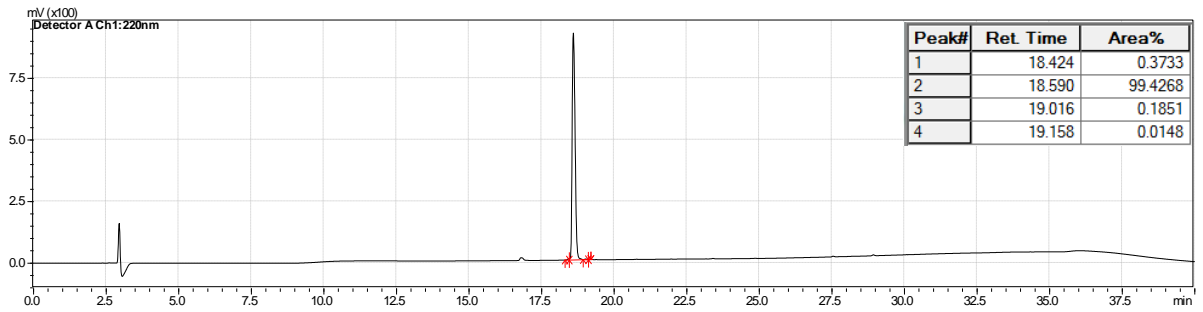
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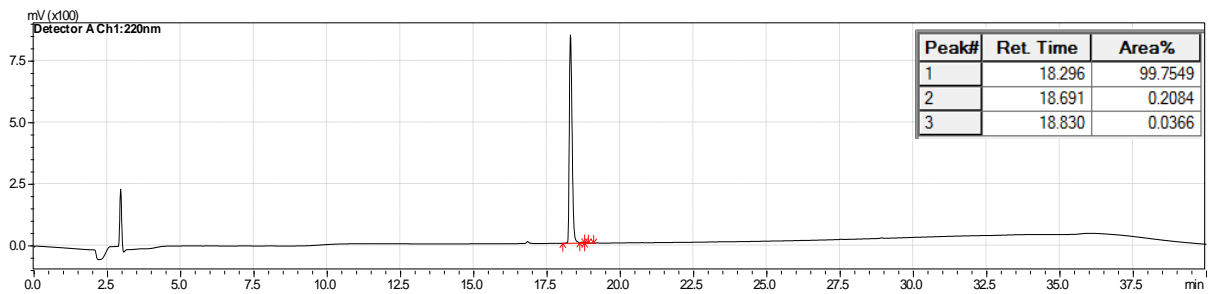
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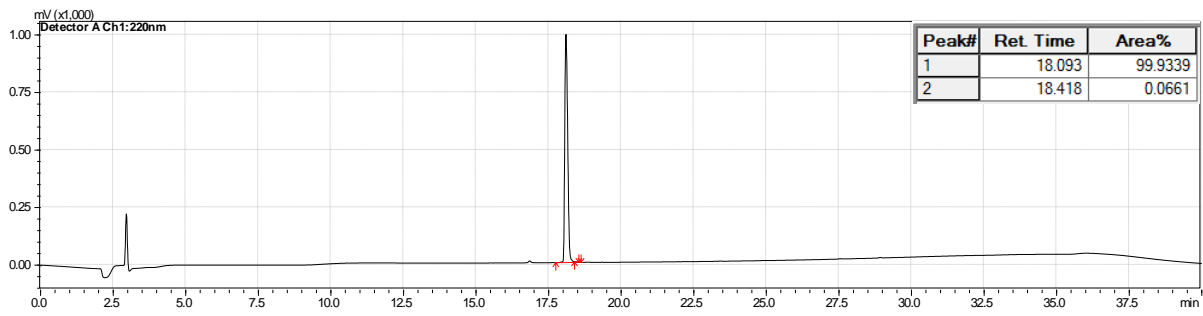
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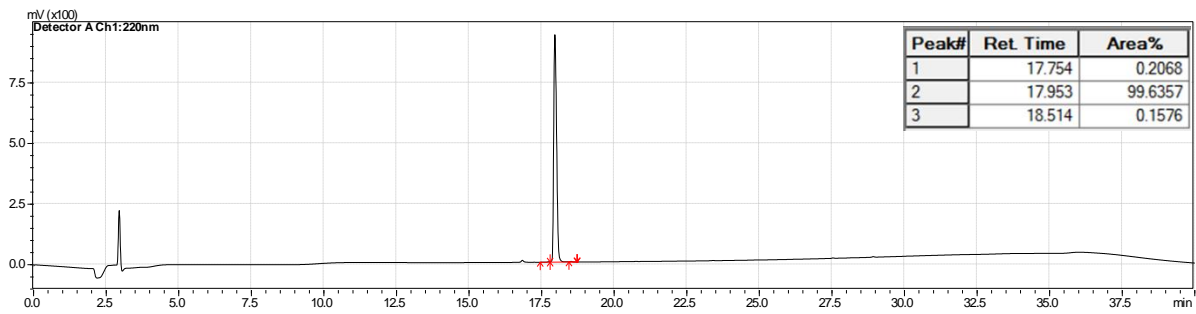
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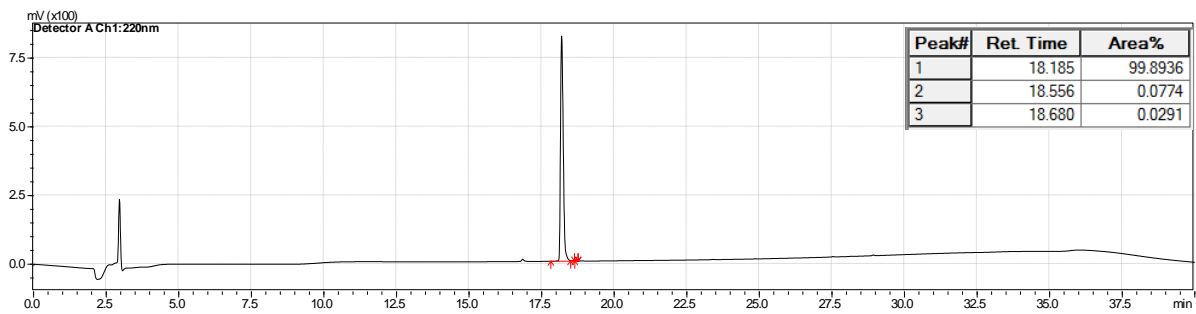
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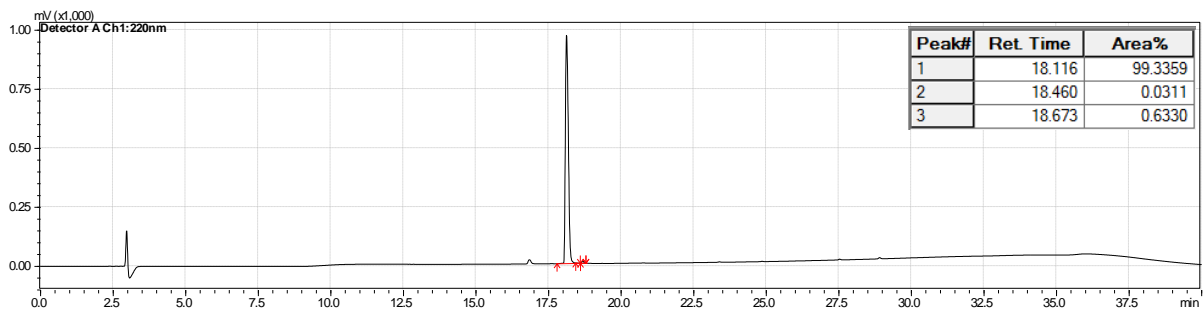
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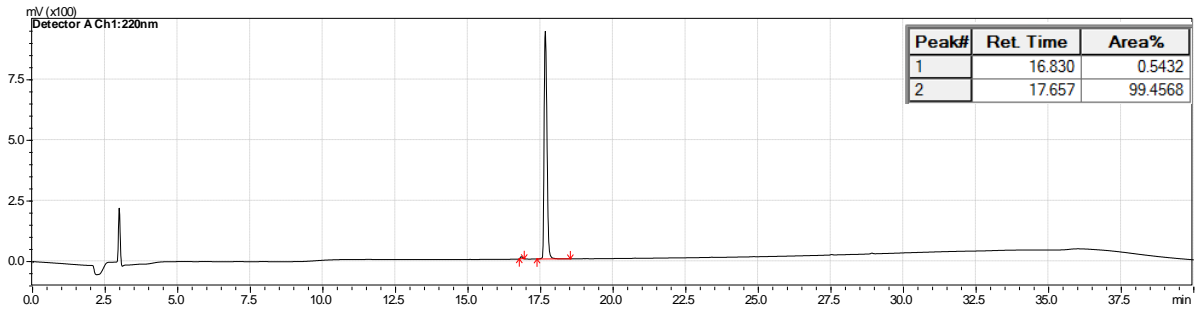
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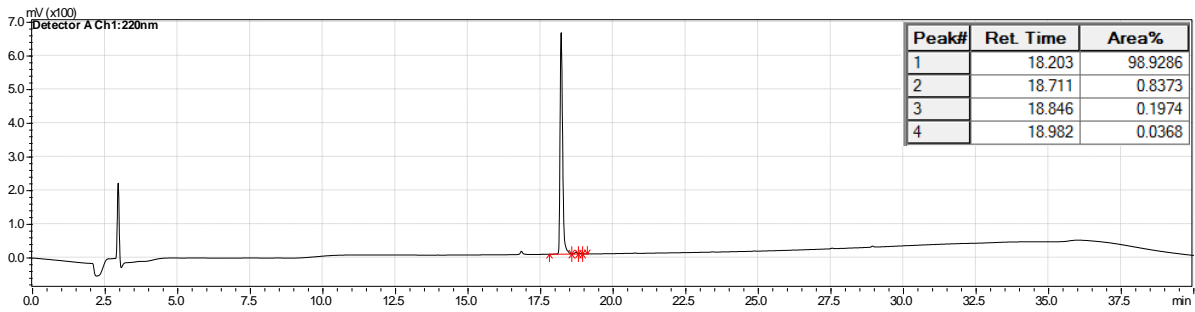
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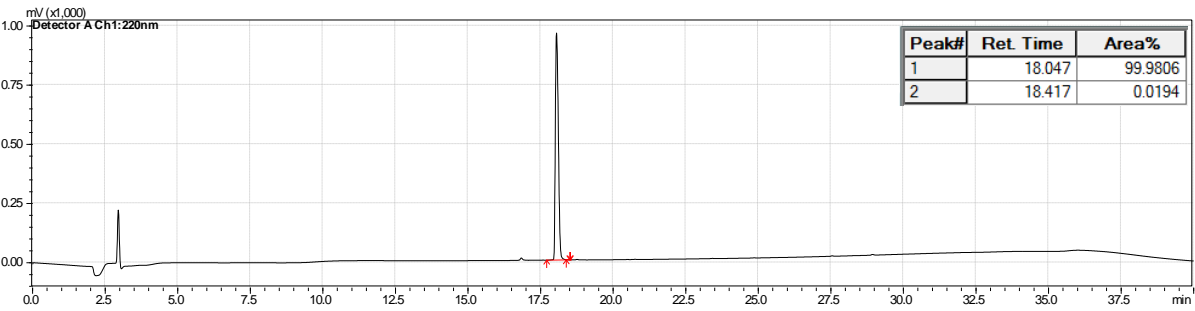
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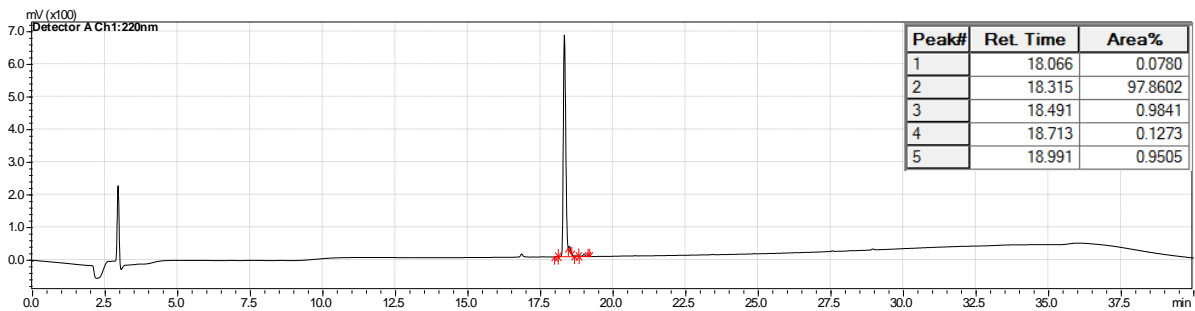
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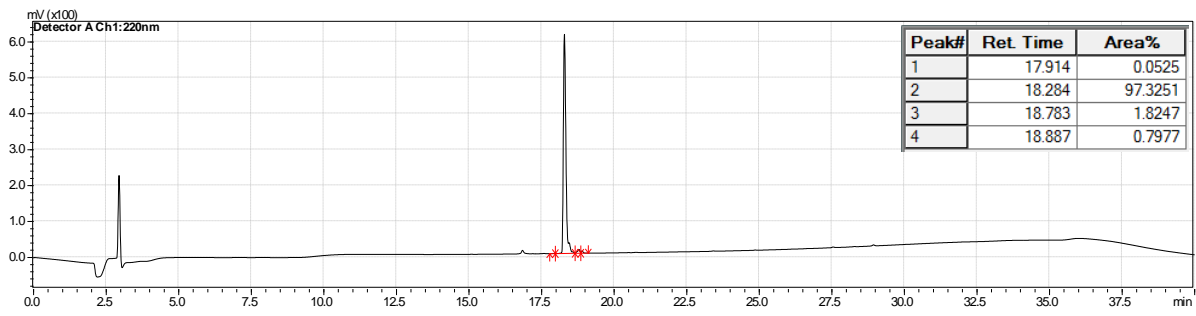
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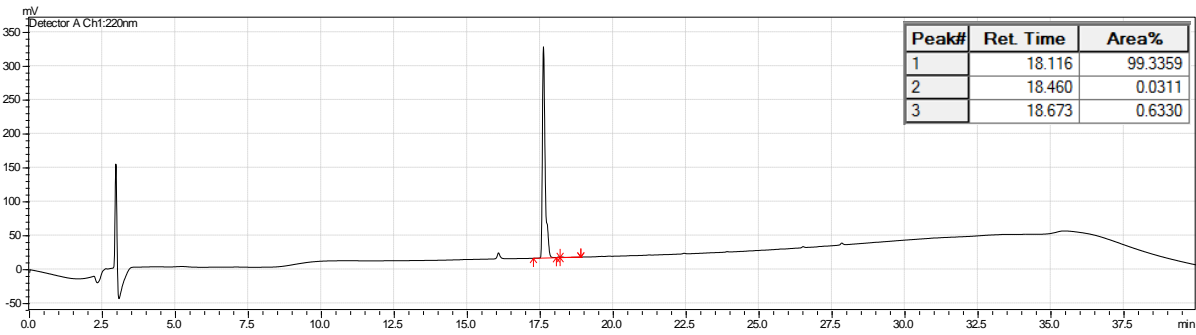
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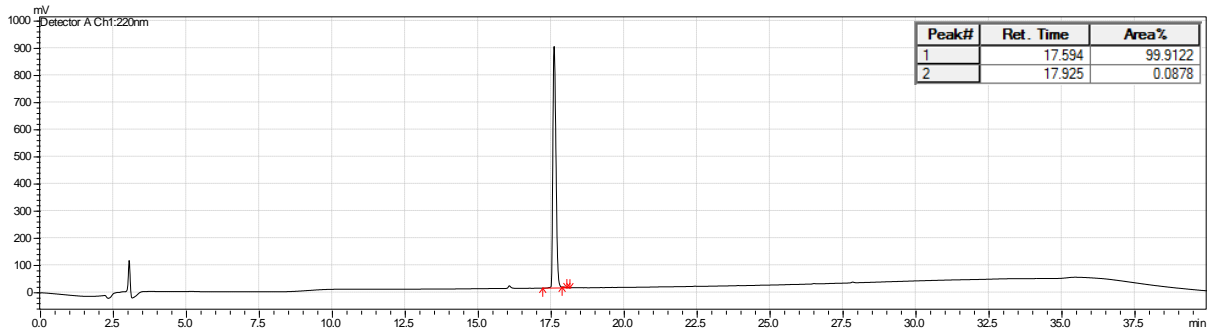
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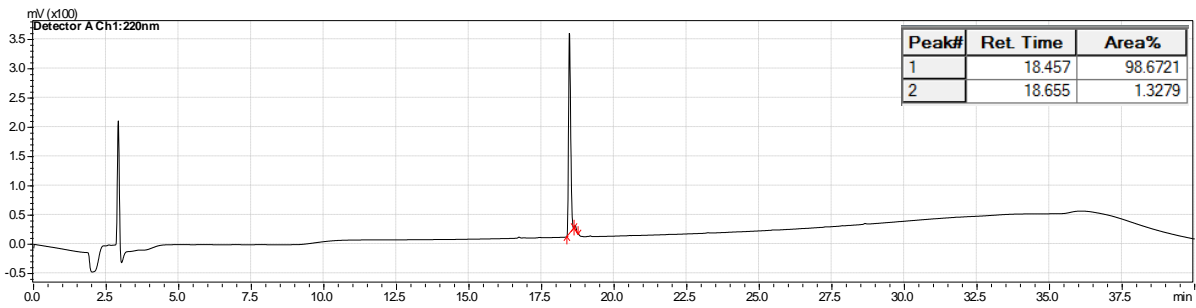
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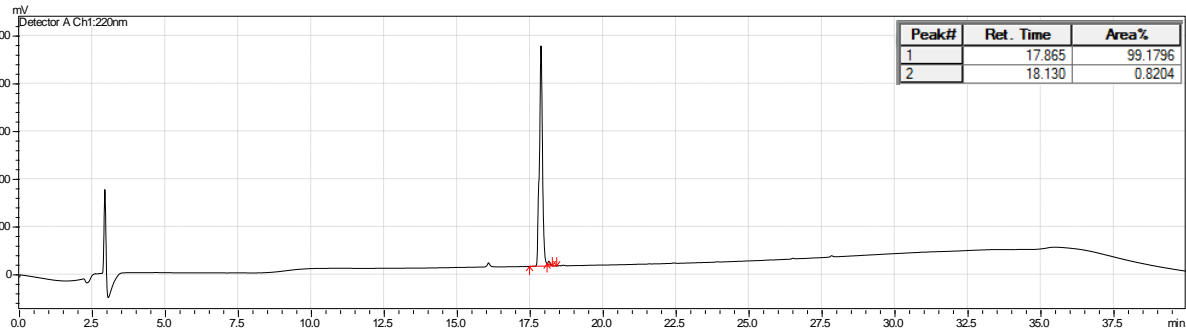
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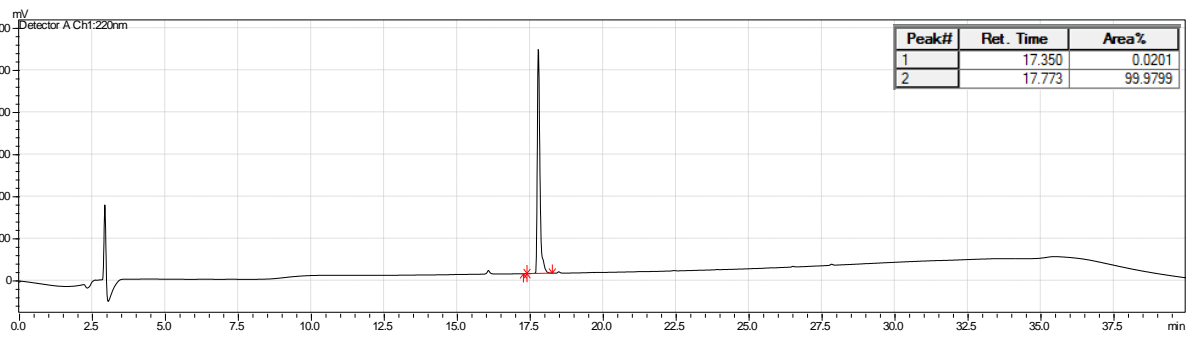
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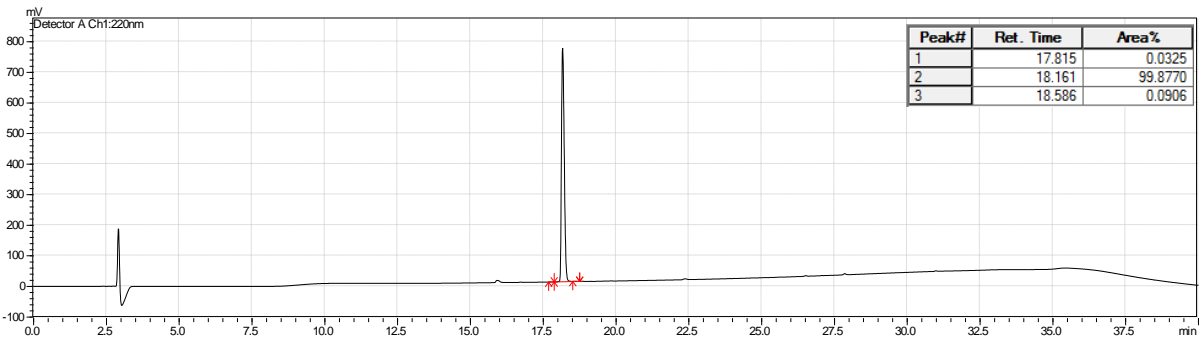
18-CSP-q17



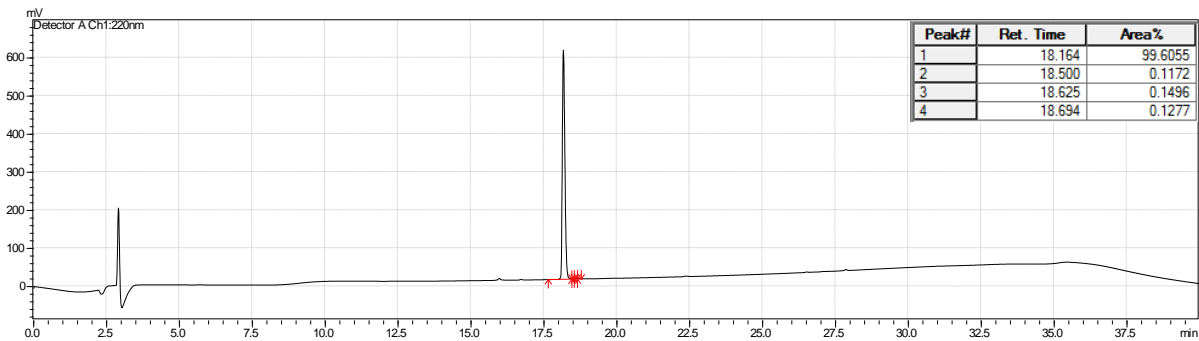
18-CSP-a18



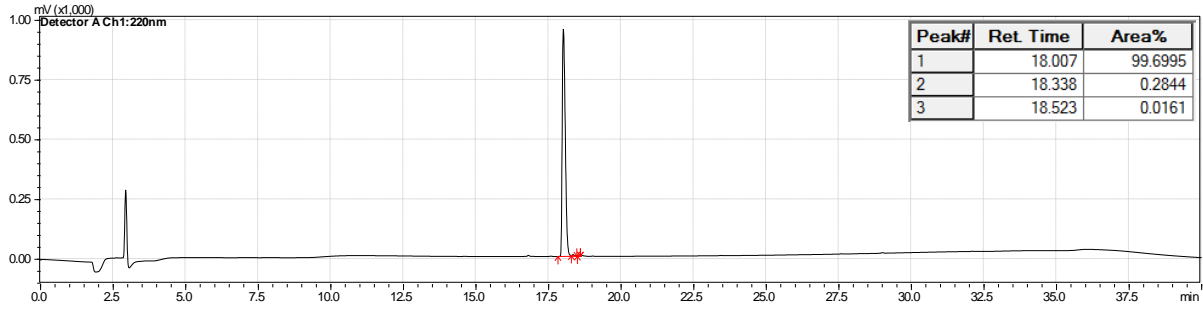
21-CSP-des-S1



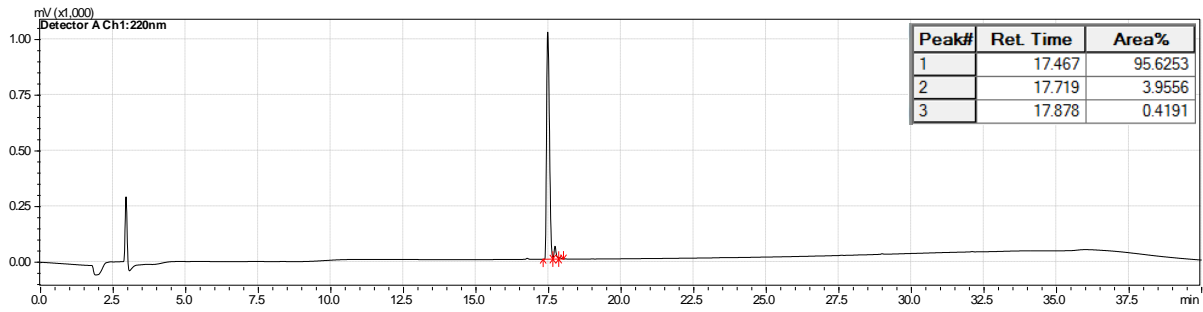
21-CSP-des-S1G2



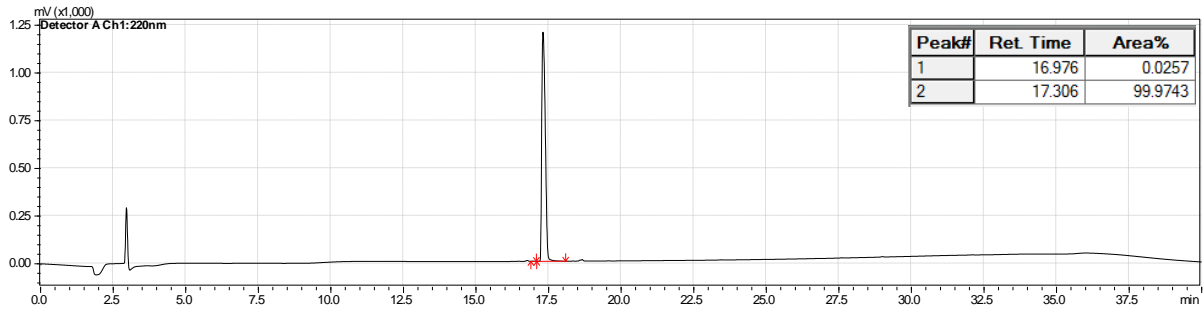
21-CSP-des-S1G2S3



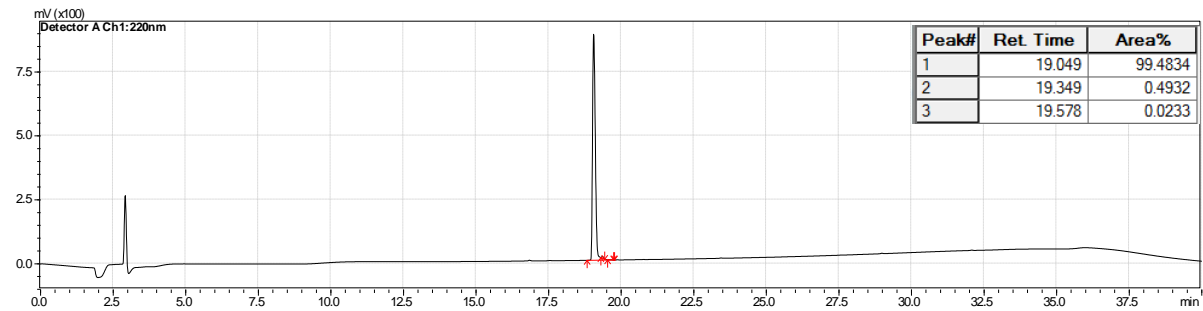
21-CSP-des-S1G2S3L4



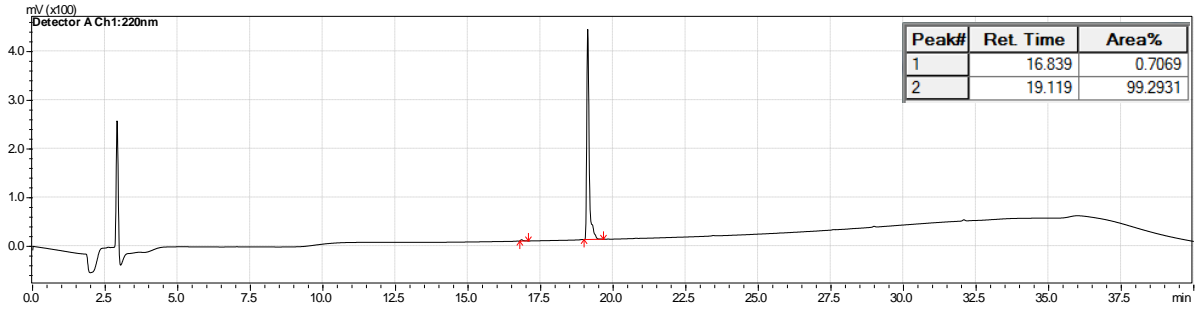
21-CSP-des-S1G2S3L4S5



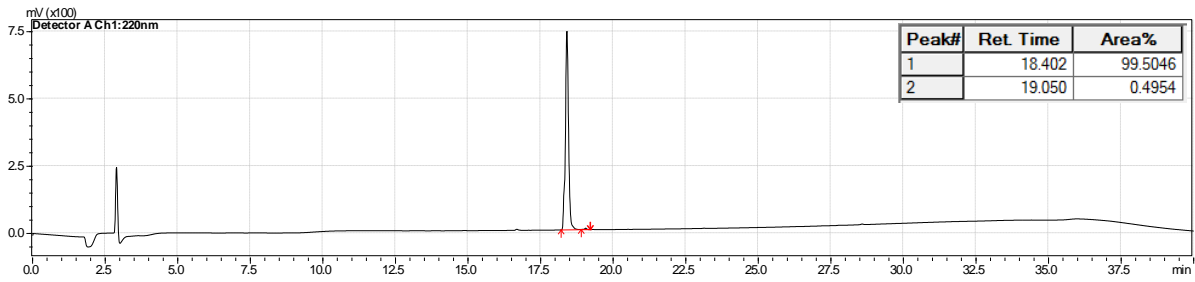
21-CSP-des-K21



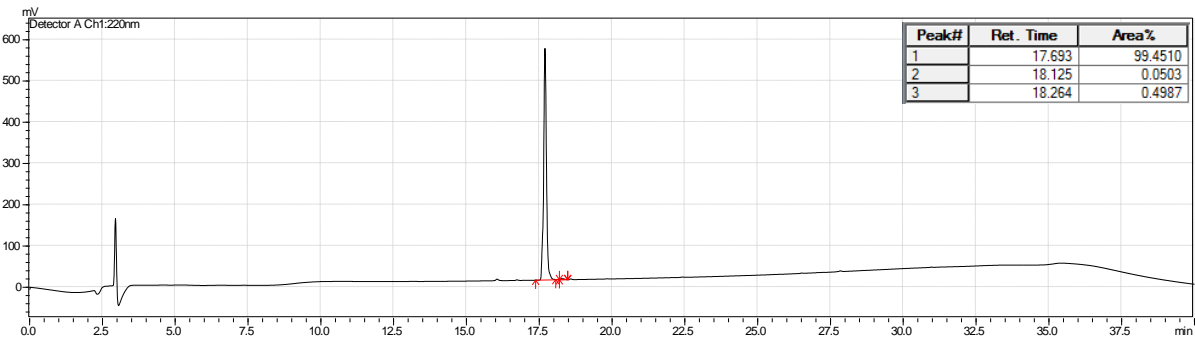
21-CSP-des-G20K21



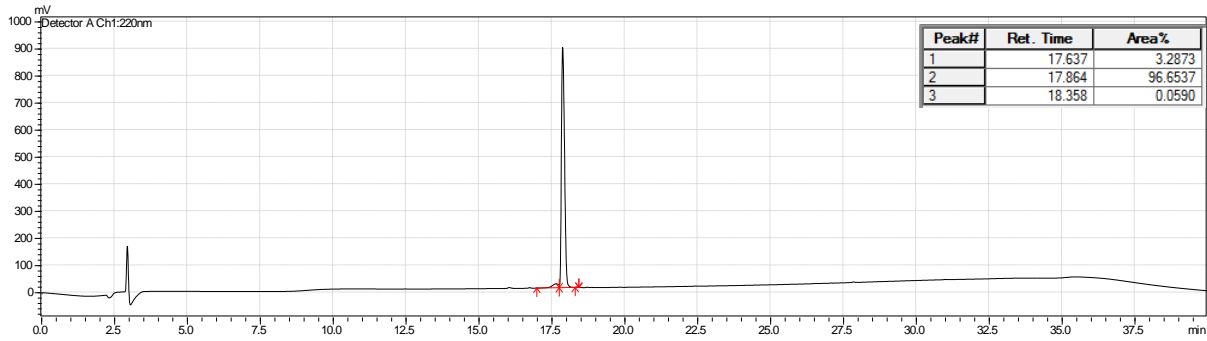
21-CSP-des-L19G20K21 (18-CSP)



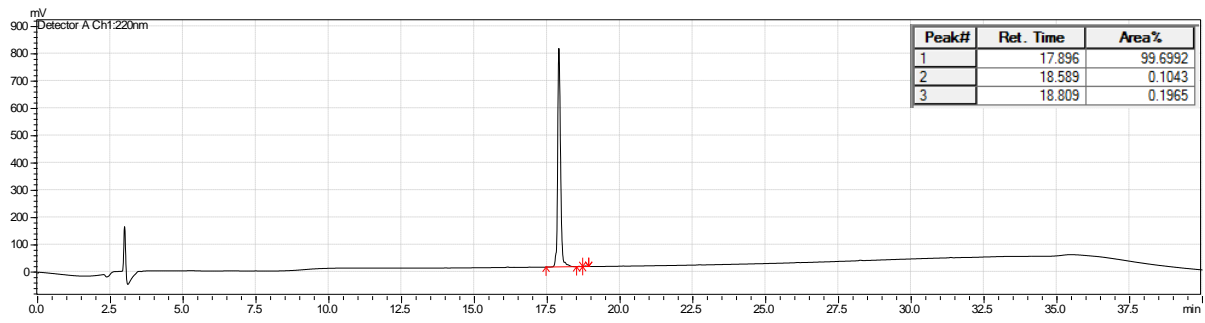
18-CSP-des-A18



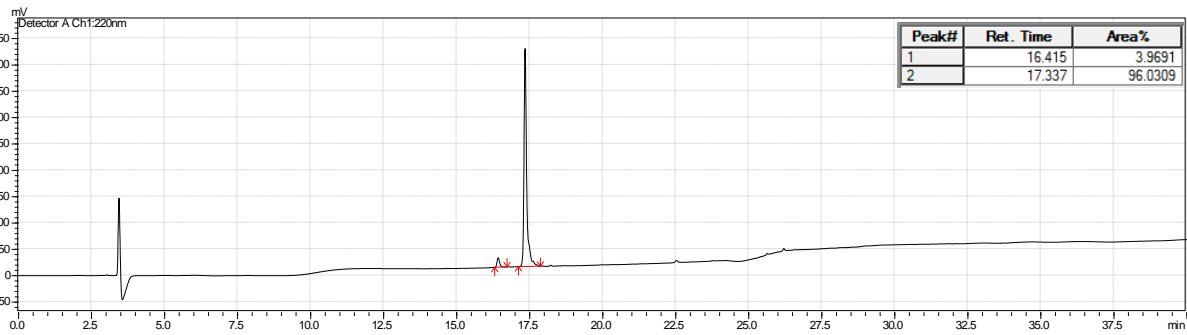
18-CSP-des-Q17A18



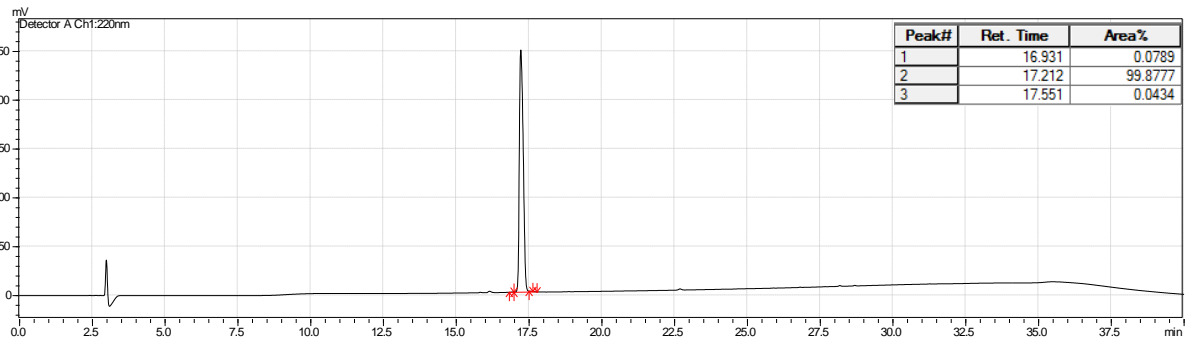
18-CSP-des-S1



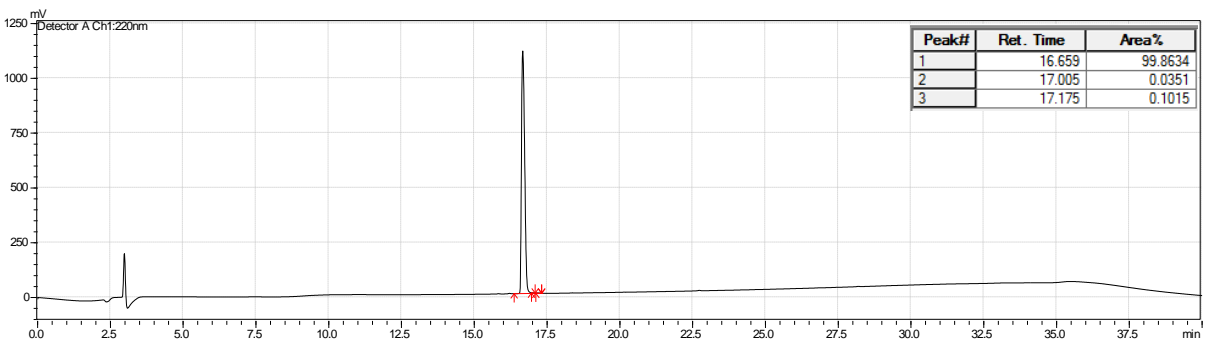
18-CSP-des-S1G2



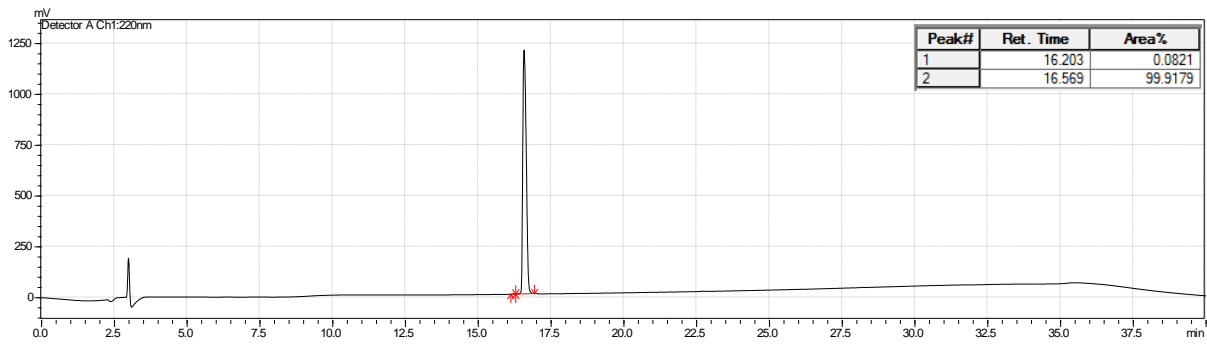
18-CSP-des-S1G2S3



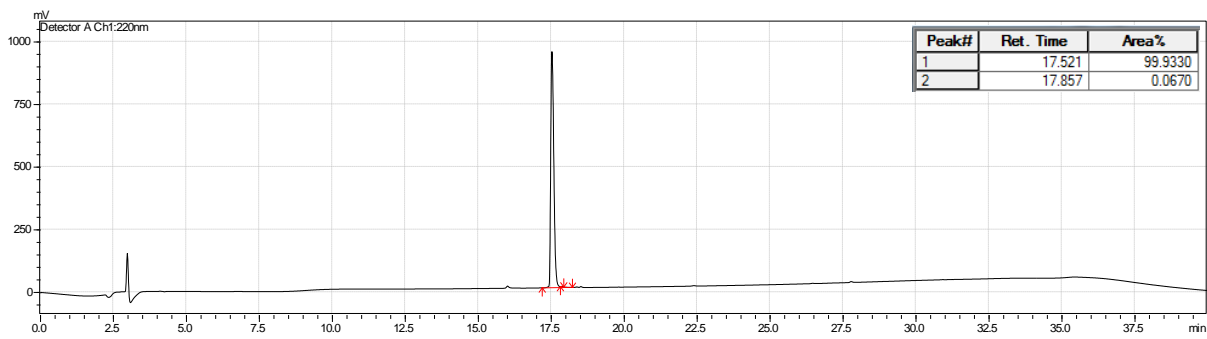
18-CSP-des-S1G2S3L4



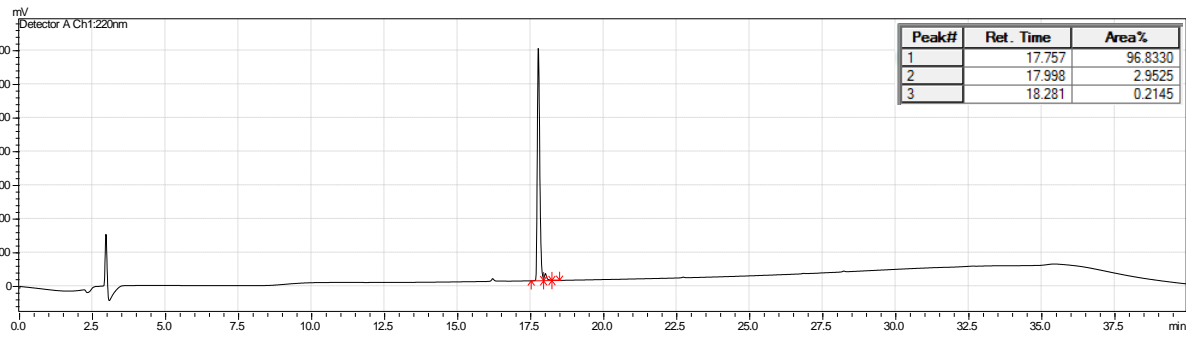
18-CSP-des-S1G2S3L4S5



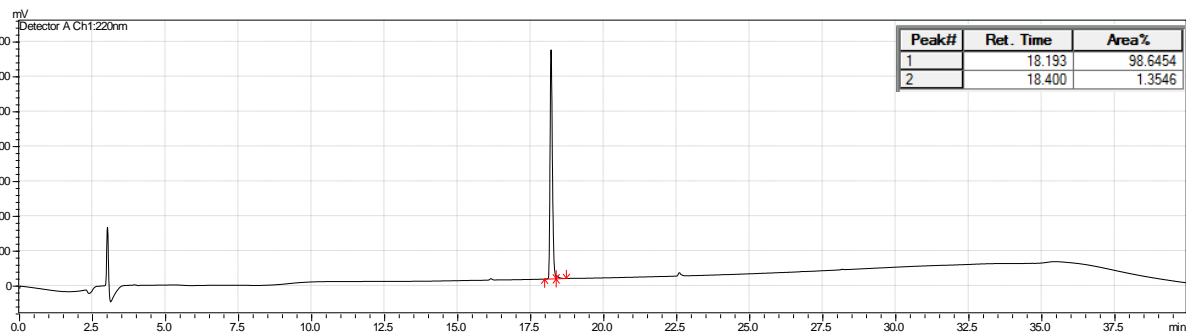
18-CSP-s1s314



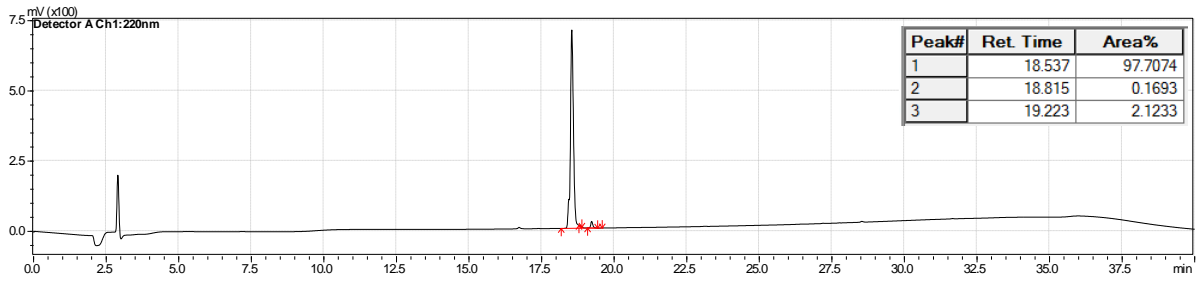
18-CSP-s1s314L10AR13AS14A



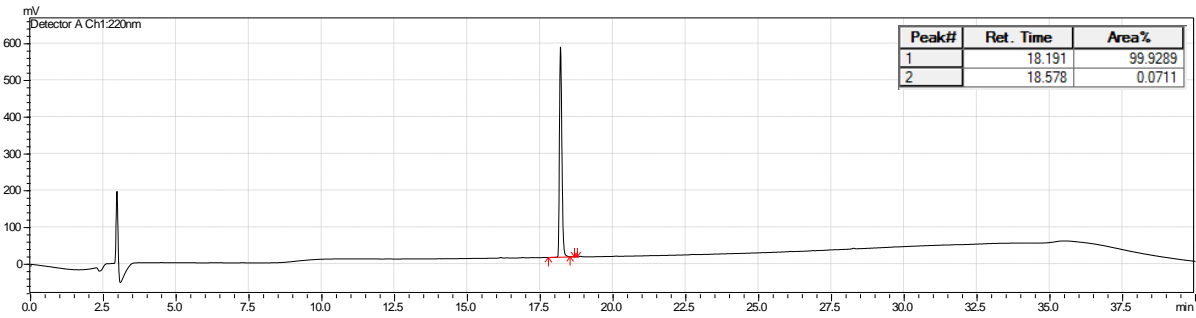
18-CSP-S1AL10AR13AS14A



18-CSP-S3T



21-CSP-S3T



MS and HPLC data for CSP analogs

Table S-1. MS and HPLC data for 21-CSP alanine & D-amino acid scanning analogs.

Compound Name	Calc. EM MH ₂ ²⁺	Obs. EM MH ₂ ²⁺	Purity (%)	Compound Name	Calc. EM MH ₂ ²⁺	Obs. EM MH ₂ ²⁺	Purity (%)
21-CSP	1183.1280	1183.1260	>99				
21-CSP-S1A	1175.6291	1175.6294	>99	21-CSP-s1	1183.1280	1183.1249	>97
21-CSP-G2A	1190.1358	1190.1329	>99				
21-CSP-S3A	1175.6291	1175.6287	>99	21-CSP-s3	1183.1280	1183.1296	>99
21-CSP-L4A	1162.1045	1162.1029	>99	21-CSP-l4	1183.1280	1183.1330	>97
21-CSP-S5A	1175.6291	1175.6292	>98	21-CSP-s5	1183.1280	1183.1324	>99
21-CSP-T6A	1168.1230	1168.1203	>99	21-CSP-t6	1183.1280	1183.1332	>99
21-CSP-F7A	1145.1124	1145.1091	>96	21-CSP-f7	1183.1280	1183.1329	>99
21-CSP-F8A	1145.1124	1145.1103	>99	21-CSP-f8	1183.1280	1183.1300	>99
21-CSP-R9A	1140.5960	1140.5929	>95	21-CSP-r9	1183.1280	1183.1296	>99
21-CSP-L10A	1162.1045	1162.1024	>99	21-CSP-l10	1183.1280	1183.1325	>96
21-CSP-F11A	1145.1124	1145.1099	>99	21-CSP-f11	1183.1280	1183.1322	>99
21-CSP-N12A	1161.6251	1161.6229	>98	21-CSP-n12	1183.1280	1183.1294	>99
21-CSP-R13A	1140.5960	1140.5932	>98	21-CSP-r13	1183.1280	1183.1282	>99
21-CSP-S14A	1175.6291	1175.6281	>98	21-CSP-s14	1183.1280	1183.1287	>99
21-CSP-F15A	1145.1124	1145.1095	>98	21-CSP-f15	1183.1280	1183.1284	>99
21-CSP-T16A	1168.1230	1168.1195	>98	21-CSP-t16	1183.1280	1183.1288	>98
21-CSP-Q17A	1154.6175	1154.6138	>95	21-CSP-q17	1183.1280	1183.1257	>99
				21-CSP-a18	1183.1280	1183.1301	>99
21-CSP-L19A	1162.1045	1162.1008	>99	21-CSP-l19	1183.1280	1183.1300	>99
21-CSP-G20A	1190.1358	1190.1330	>99				
21-CSP-K21A	1154.5991	1154.5965	>97	21-CSP-k21	1183.1280	1183.1258	>97

EM = Exact Mass. See main text for methods.

Table S-2. MS and HPLC data for 21-CSP and 18-CSP truncated analogs.

Compound Name	Calc. EM MH ₂ ²⁺	Obs. EM MH ₂ ²⁺	Purity (%)
21-CSP-des-S1	1139.6120	1139.6116	>99
21-CSP-des-S1G2	1111.1013	1111.1006	>99
21-CSP-des-S1G2S3	1067.5853	1067.5871	>99
21-CSP-des-S1G2S3L4	1011.0432	1011.0430	>95
21-CSP-des-S1G2S3L4S5	967.5272	967.5232	>99
21-CSP-des-K21	1119.0805	1119.0827	>99
21-CSP-des-G20K21	1090.5698	1090.5706	>99
18-CSP-des-S1	990.5118	990.5131	>99
18-CSP-des-S1G2	962.0010	961.9970	>96
18-CSP-des-S1G2S3	918.4850	918.4848	>99
18-CSP-des-S1G2S3L4	861.4415	861.4402	>99
18-CSP-des-S1G2S3L4S5	817.9255	817.9263	>99
18-CSP-des-A18	998.5092	998.5065	>99
18-CSP-des-Q17A18	934.4799	934.4802	>96

EM = Exact Mass. See main text for methods.

Table S-3. MS and HPLC data for 18-CSP alanine & D-amino acid scanning analogs.

Compound Name	Calc. EM MH₂²⁺	Obs. EM MH₂²⁺	Purity (%)	Compound Name	Calc. EM MH₂²⁺	Obs. EM MH₂²⁺	Purity (%)
18-CSP	1034.0278	1034.0261	>99				
18-CSP-S1A	1026.0303	1026.0315	>99	18-CSP-s1	1034.0278	1034.0305	>99
18-CSP-G2A	1041.0356	1041.0354	>99				
18-CSP-S3A	1026.0303	1026.0330	>99	18-CSP-s3	1034.0278	1034.0323	>99
18-CSP-L4A	1013.0043	1013.0085	>99	18-CSP-l4	1034.0278	1034.0284	>99
18-CSP-S5A	1026.0303	1026.0289	>98	18-CSP-s5	1034.0278	1034.0299	>99
18-CSP-T6A	1019.0225	1019.0255	>96	18-CSP-t6	1034.0278	1034.0287	>99
18-CSP-F7A	996.0121	996.0167	>96	18-CSP-f7	1034.0278	1034.0304	>99
18-CSP-F8A	996.0121	996.0125	>97	18-CSP-f8	1034.0278	1034.0313	>96
18-CSP-R9A	991.4958	991.4975	>99	18-CSP-r9	1034.0278	1034.0317	>99
18-CSP-L10A	1013.0043	1013.0057	>99	18-CSP-l10	1034.0278	1034.0283	>98
18-CSP-F11A	996.0121	996.0125	>99	18-CSP-f11	1034.0278	1034.0295	>99
18-CSP-N12A	1012.5249	1012.5214	>95	18-CSP-n12	1034.0278	1034.0302	>97
18-CSP-R13A	991.4958	991.4958	>98	18-CSP-r13	1034.0278	1034.0299	>97
18-CSP-S14A	1026.0303	1026.0318	>95	18-CSP-s14	1034.0278	1034.0296	>99
18-CSP-F15A	996.0121	996.0110	>97	18-CSP-f15	1034.0278	1034.0298	>99
18-CSP-T16A	1019.0225	1019.0230	>98	18-CSP-t16	1034.0278	1034.0300	>98
18-CSP-Q17A	1005.5170	1005.5187	>95	18-CSP-q17	1034.0278	1034.0303	>99
				18-CSP-a18	1034.0278	1034.0307	>99

EM = Exact Mass. See main text for methods.

Table S-4. MS and HPLC data for 21-CSP and 18-CSP multiple mutation analogs.

Compound Name	Calc. EM MH₂²⁺	Obs. EM MH₂²⁺	Purity (%)
18-CSP-S3T	1041.0356	1041.0364	>97
21-CSP-S3T	1190.1358	1190.1328	>99
18-CSP-s1s3l4	1034.0278	1034.0262	>99
18-CSP-s1s3l4L10AR13AS14A	962.4746	962.4786	>96
18-CSP-S1AL10AR13AS14A	954.4774	954.4801	>98

EM = Exact Mass. See main text for methods.

Primary reporter gene assay data

S. mutans SAB249 (pCipB::lacZ)

Agonism assays were performed at 10 μ M concentration. 21-CSP was used as the positive control (100%) with DMSO as the negative control (0%). Percent (%) ComD activation was measured by normalizing the miller units obtained for each peptide to that of 21-CSP. All peptides were screened in triplicates over three separate trials. Error bars indicate standard error of the mean of nine values.

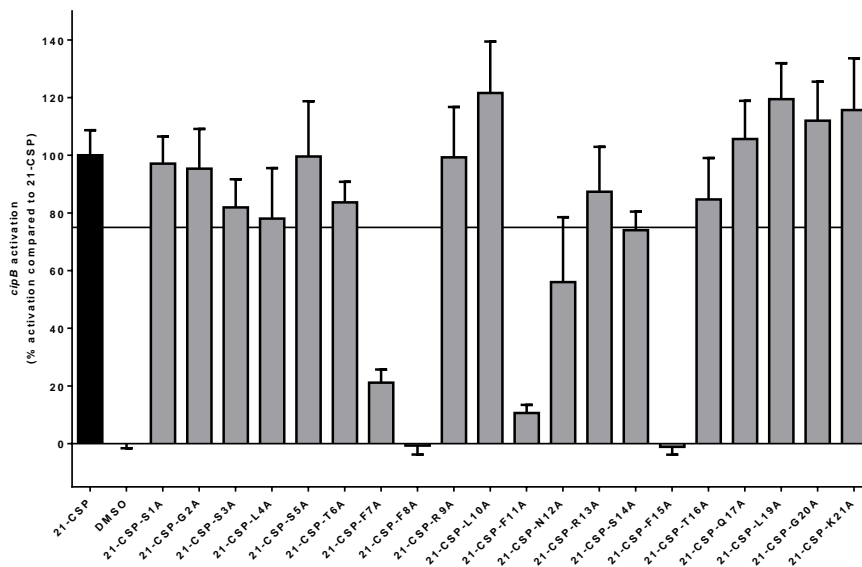


Figure S-1. Primary agonism screening assay data for the 21-CSP alanine scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

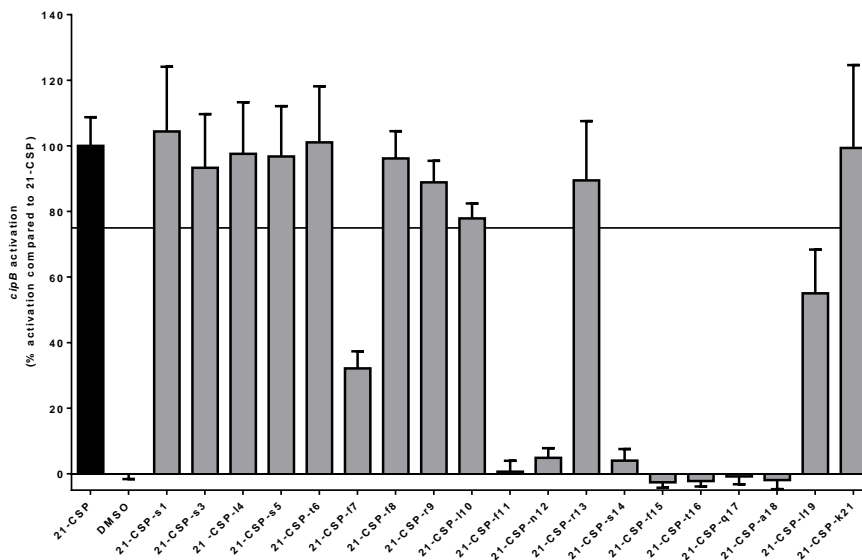


Figure S-2. Primary agonism screening assay data for the 21-CSP D-amino acid scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC_{50} while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

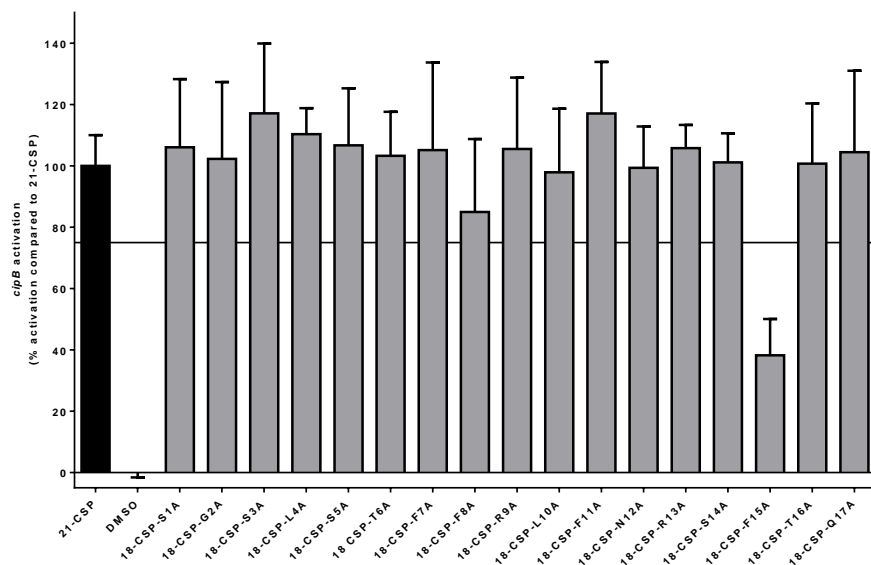


Figure S-3. Primary agonism screening assay data for the 18-CSP alanine scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

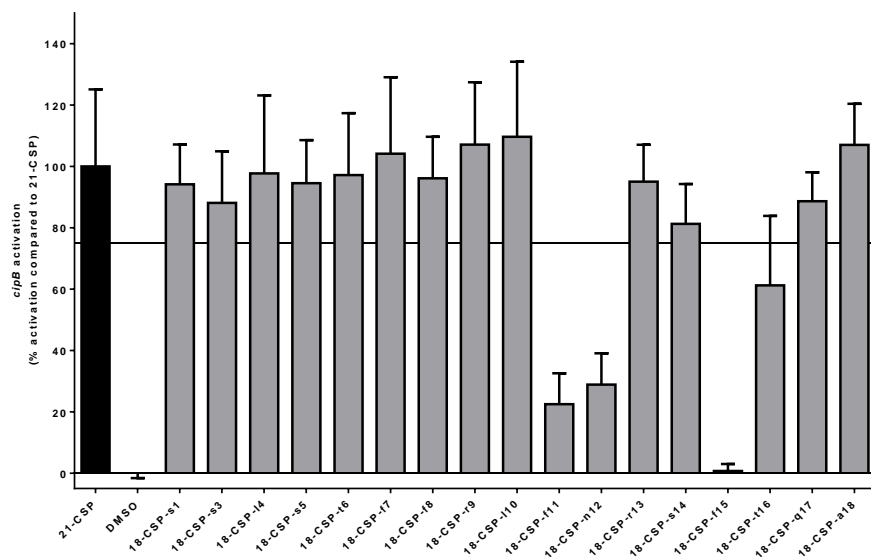


Figure S-4. Primary agonism screening assay data for the 18-CSP D-amino acid scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

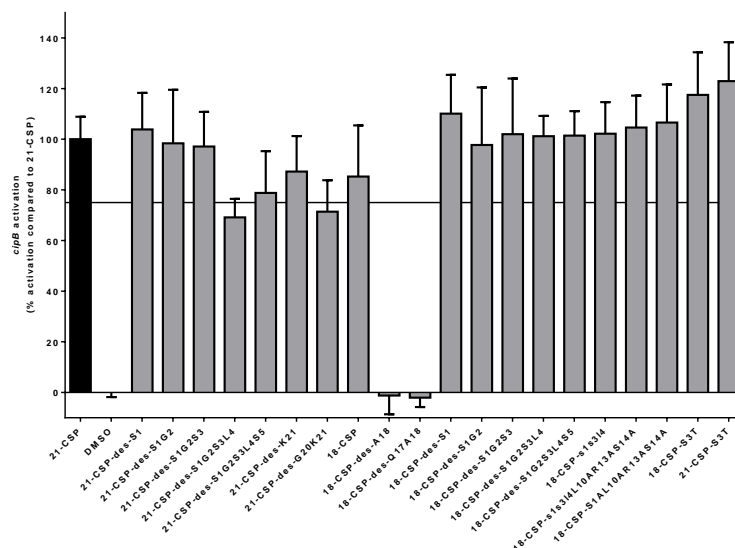


Figure S-5. Primary agonism screening assay data for the 21-CSP & 18-CSP truncated peptides and second-generation peptide analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

Antagonism assays were performed at 10 μM concentration of peptides against 1000 nM concentration of 21-CSP. 21-CSP (1000 nM) was used as the positive control (100%) with DMSO as the negative control (0%). Percent (%) ComD activation was measured by normalizing the miller units obtained for each peptide to that of 21-CSP. All peptides were screened in triplicates over three separate trials. Error bars indicate standard error of the mean of nine values.

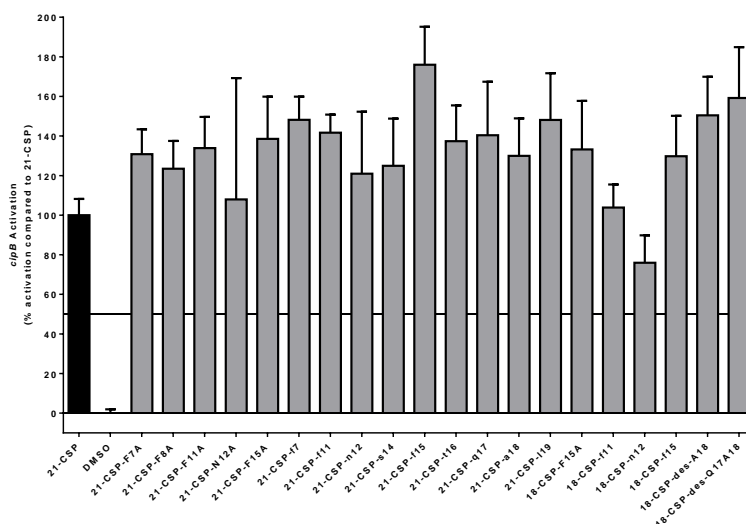


Figure S-6. Primary antagonism screening assay data for the 21-CSP and 18-CSP alanine, D-amino acid scan libraries and truncated peptides.

***S. mutans* SMCOM2 ($\Delta comC$, $pcomX::lacZ$)**

Agonism assays were performed at 10 μ M concentration. 21-CSP was used as the positive control (100%) with DMSO as the negative control (0%). Percent (%) ComD activation was measured by normalizing the miller units obtained for each peptide to that of 21-CSP. All peptides were screened in triplicates over three separate trials. Error bars indicate standard error of the mean of nine values.

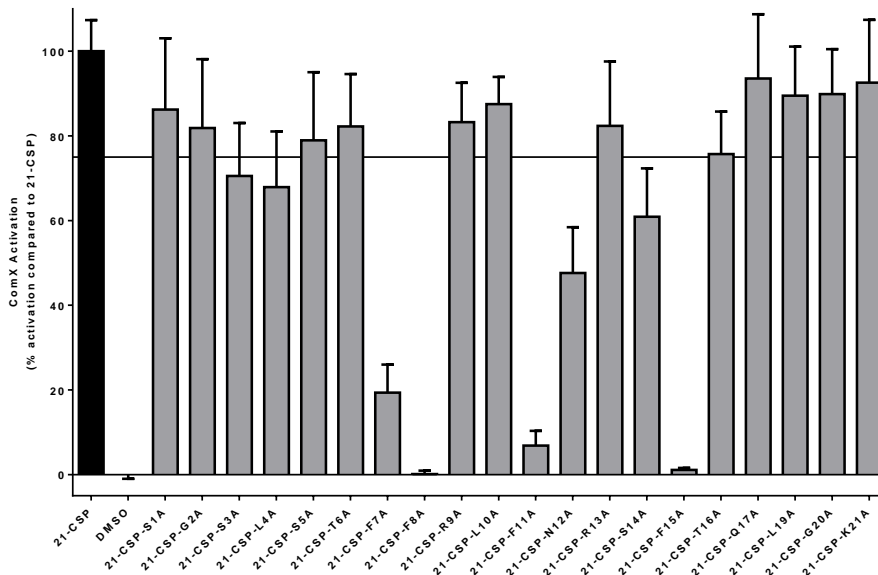


Figure S-7. Primary agonism screening assay data for the 21-CSP alanine scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

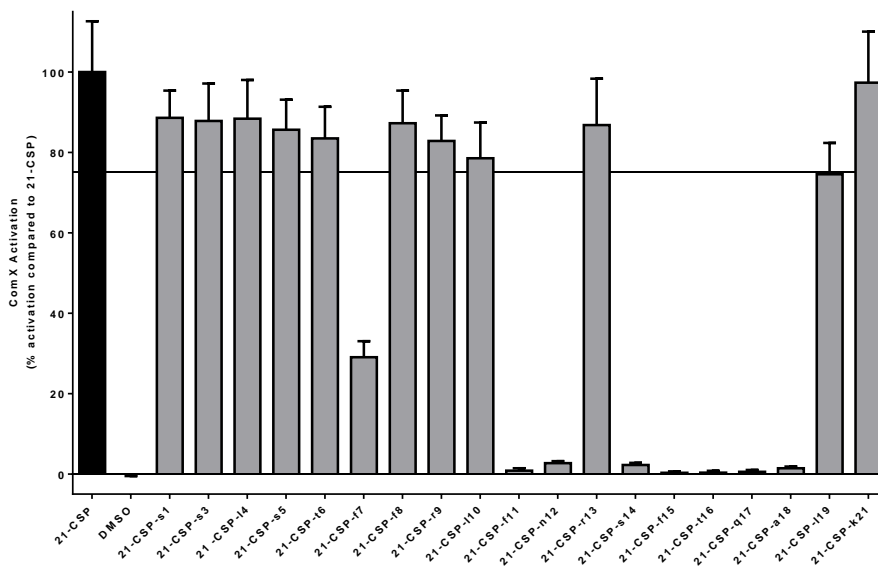


Figure S-8. Primary agonism screening assay data for the 21-CSP D-amino acid scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

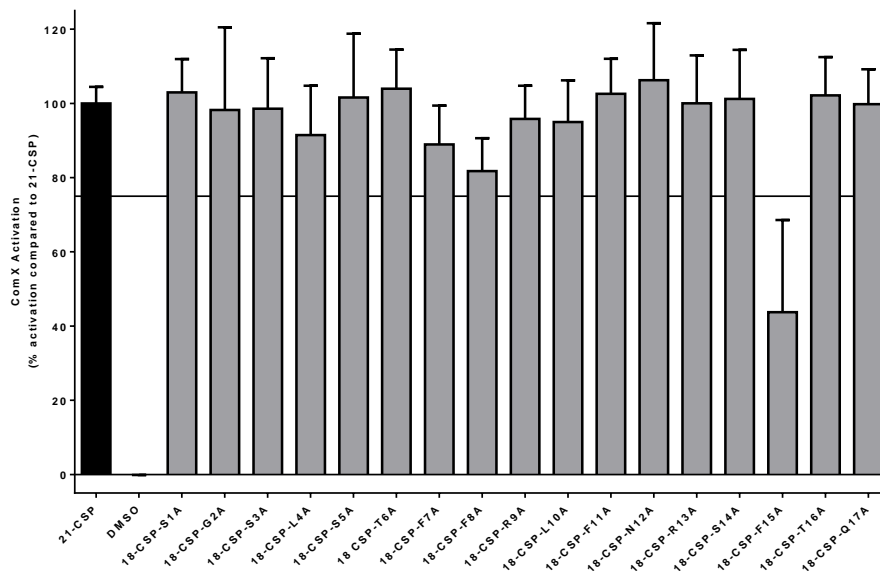


Figure S-9. Primary agonism screening assay data for the 18-CSP alanine scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

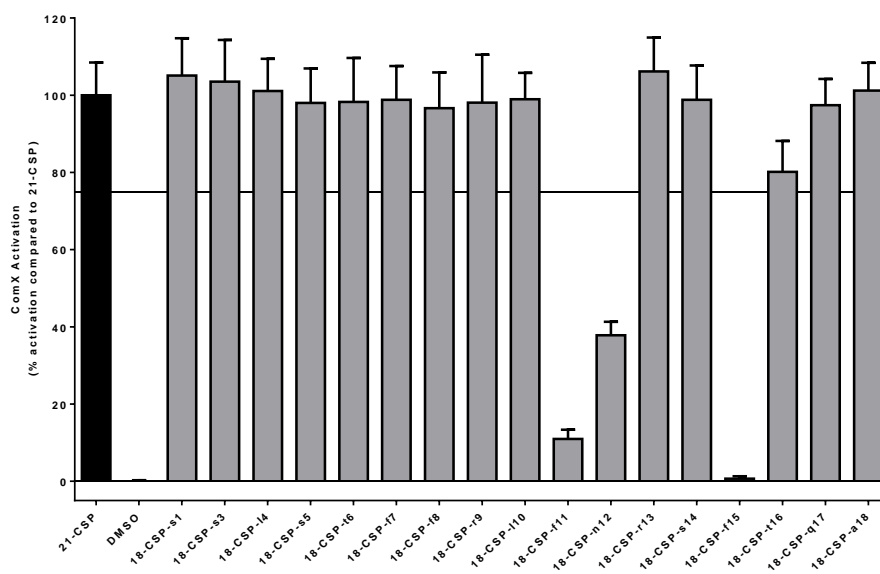


Figure S-10. Primary agonism screening assay data for the 18-CSP D-amino acid scan library. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

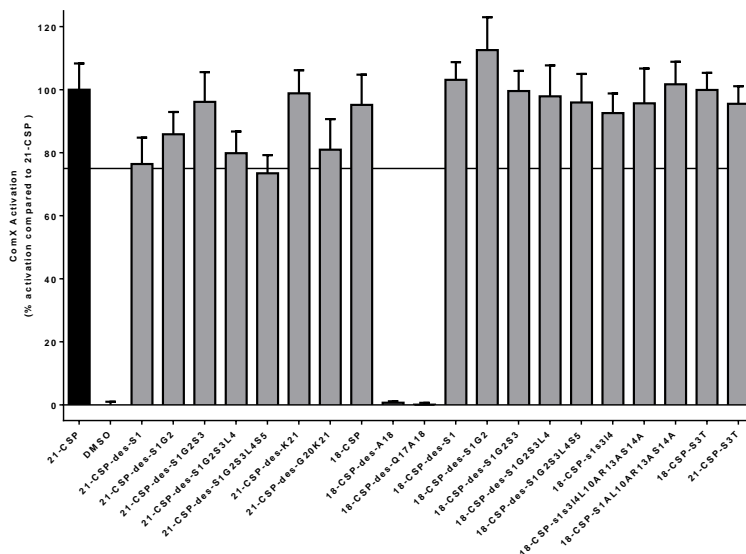


Figure S-11. Primary agonism screening assay data for the 21-CSP & 18-CSP truncated peptides and second-generation peptide analogs. Peptides that exhibited over 75% activation were further evaluated to determine their EC₅₀ while peptides that exhibited less than 50% activation were evaluated as potential competitive inhibitors.

Antagonism assays were performed at 10 μM concentration of peptides against 350 nM concentration of 21-CSP. 21-CSP (350 nM) was used as the positive control (100%) with DMSO as the negative control (0%). Percent (%) ComD activation was measured by normalizing the miller units obtained for each peptide to that of 21-CSP. All peptides were screened in triplicates over three separate trials. Error bars indicate standard error of the mean of nine values.

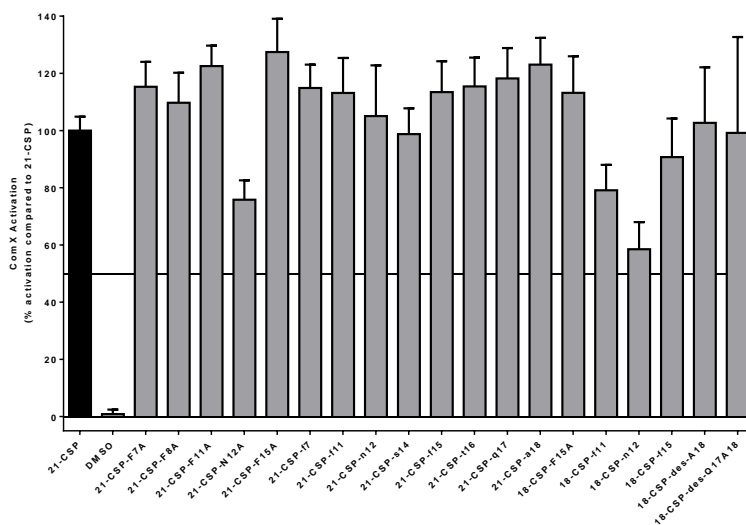


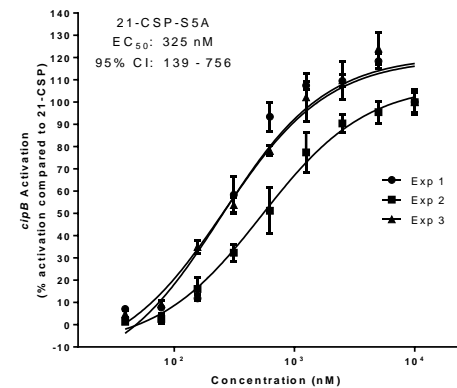
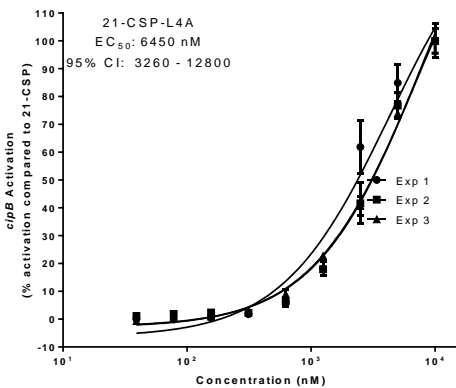
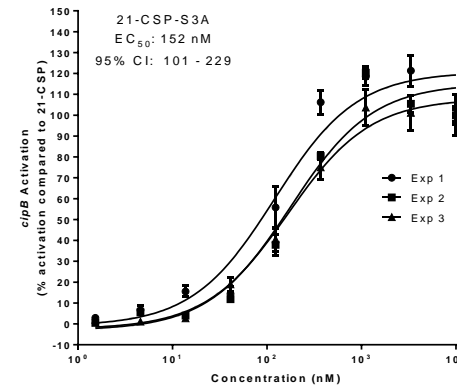
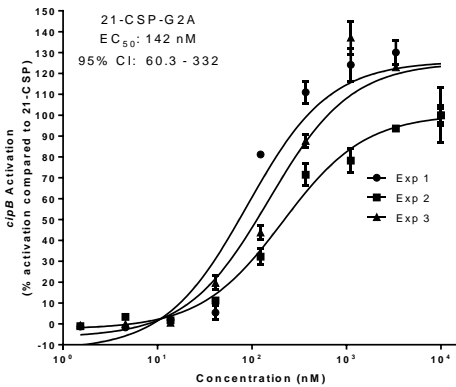
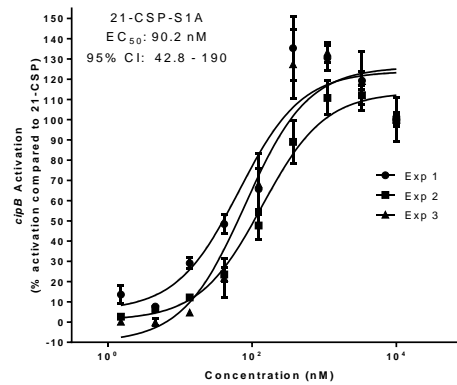
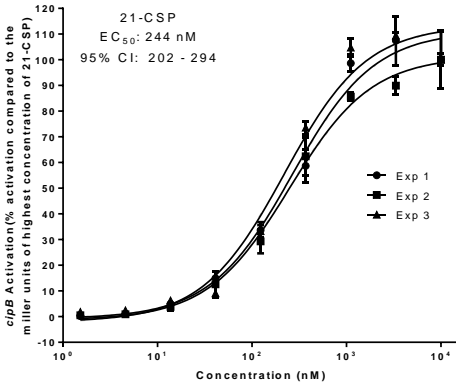
Figure S-12. Primary antagonism screening assay data for the 21-CSP and 18-CSP alanine, D-amino acid scan libraries and truncated peptides.

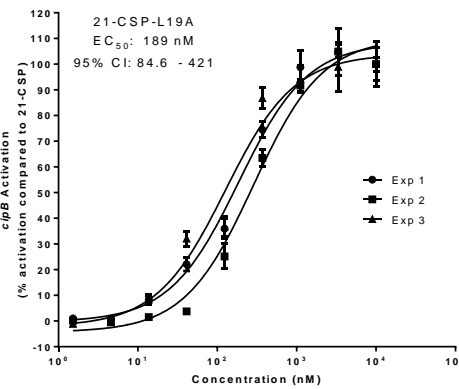
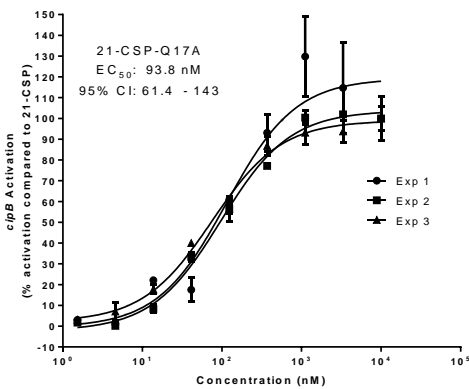
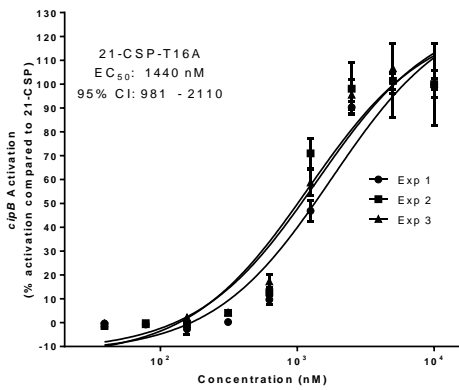
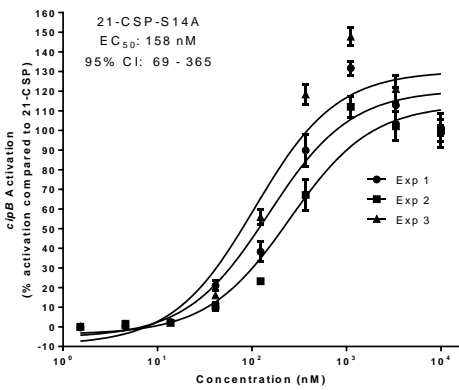
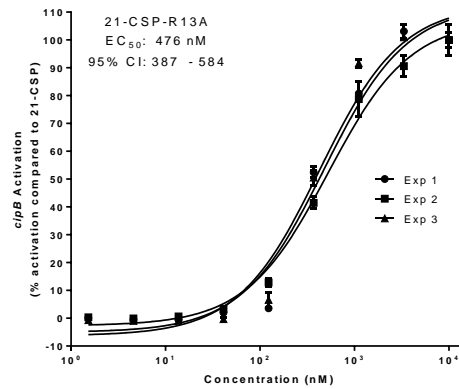
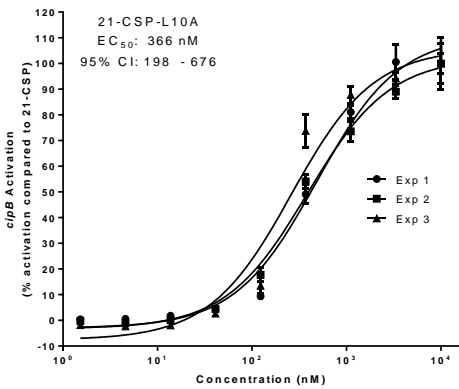
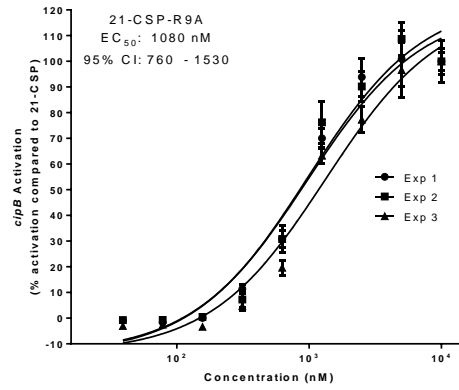
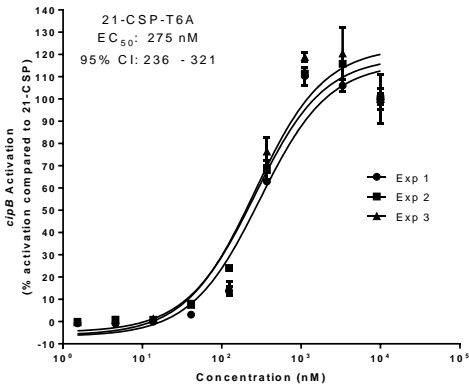
Agonism dose response curves

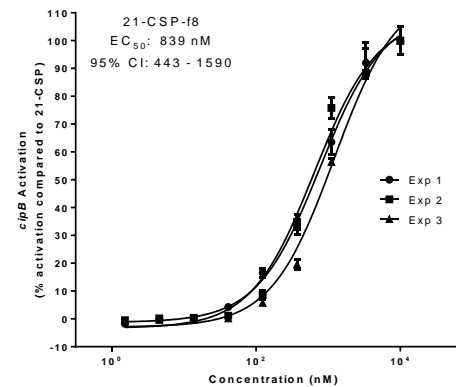
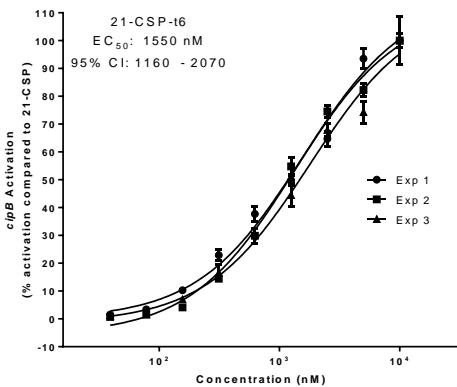
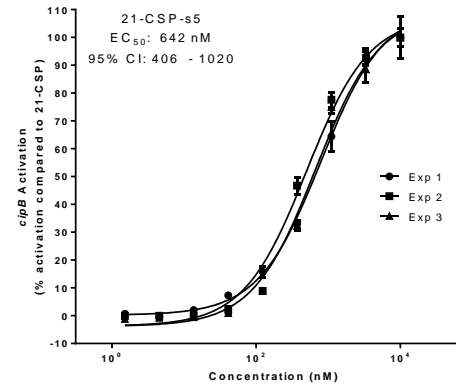
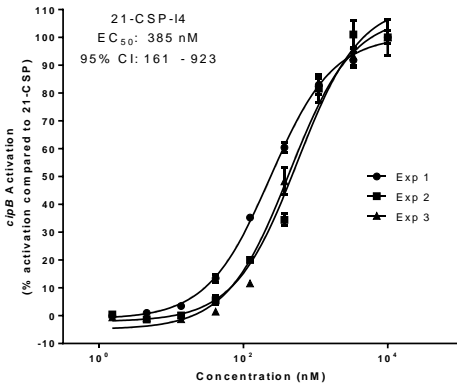
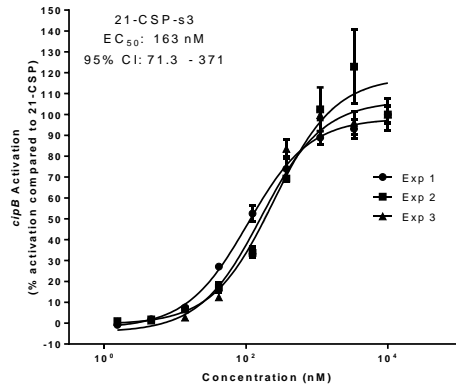
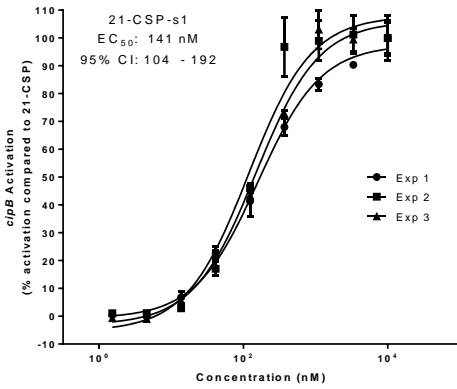
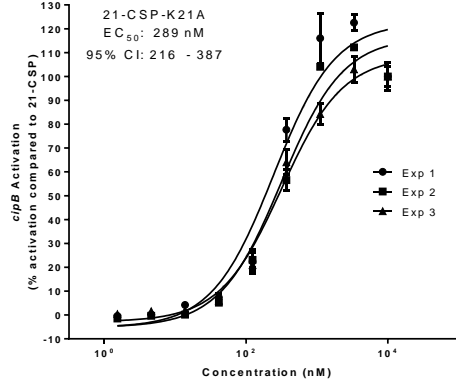
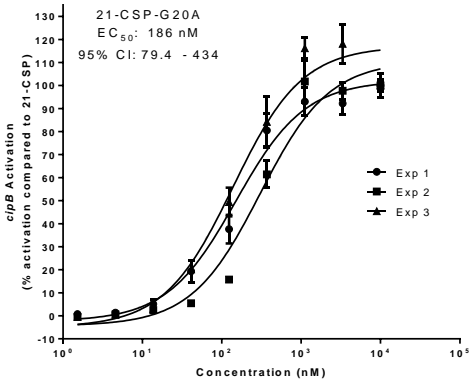
Both the 21-CSP and 18-CSP analogs were screened over varying concentrations in the two indicated *S. mutans* beta-galactosidase reporter strains (SAB249 and SMCOM2). Each dose response experiment was performed in triplicate on three separate occasions (i.e., experiments (Exp.) #1-3; shown for each peptide below). Error bars indicate standard error of the mean of triplicate values. In each plot the peptide as well as its EC₅₀ value and 95% confidence interval values (95% CI) are indicated at top left.

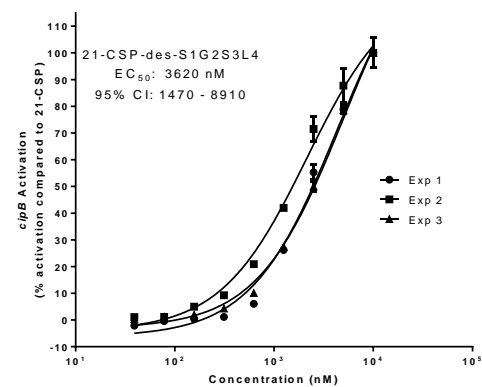
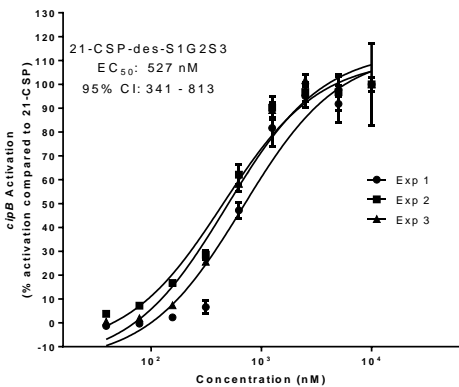
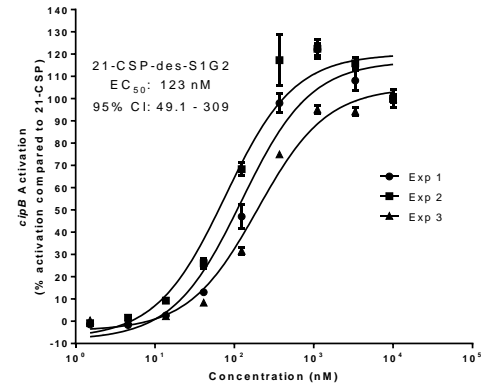
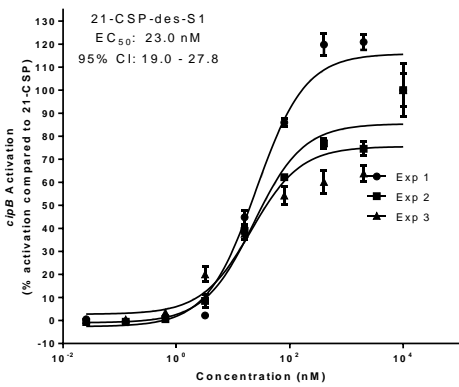
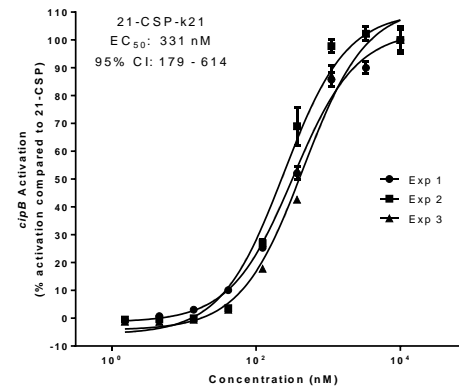
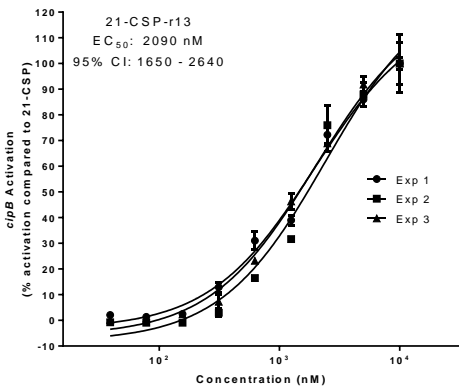
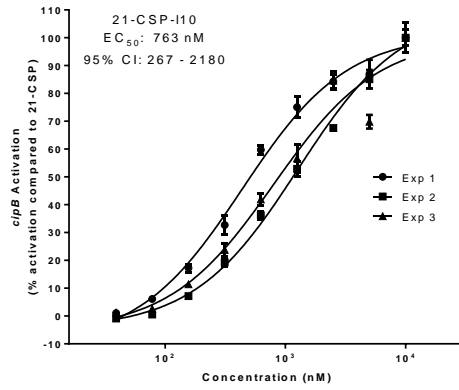
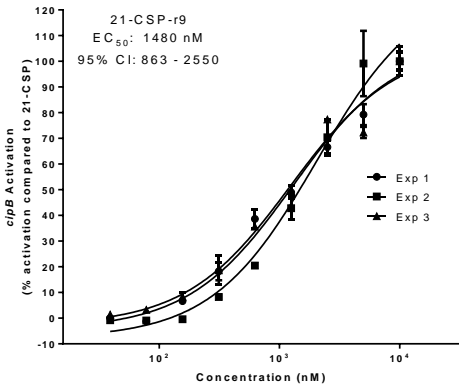
S. mutans SAB249 (pCipB::lacZ)

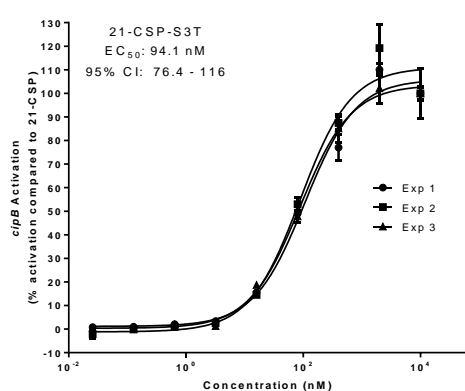
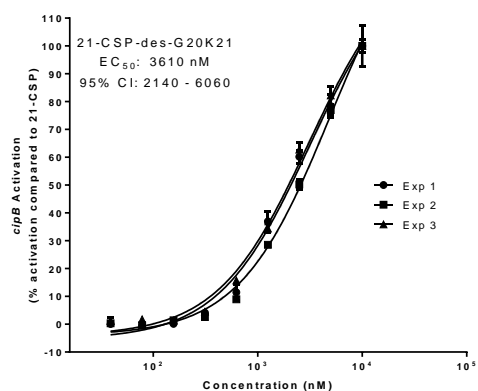
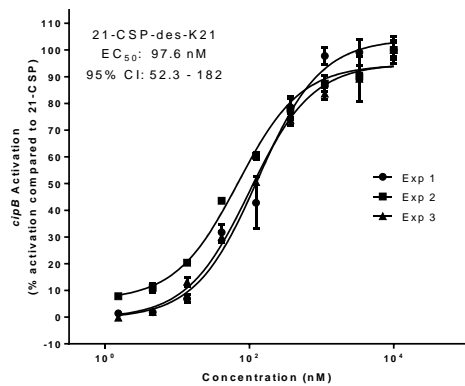
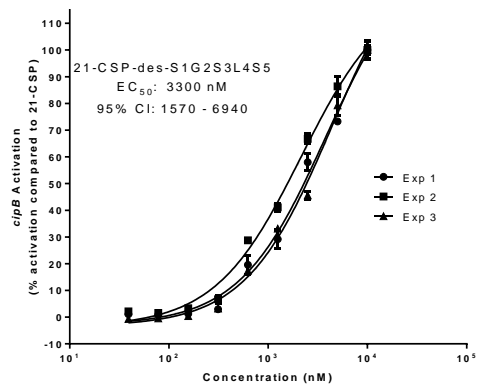
Activation dose response curves (21-CSP)



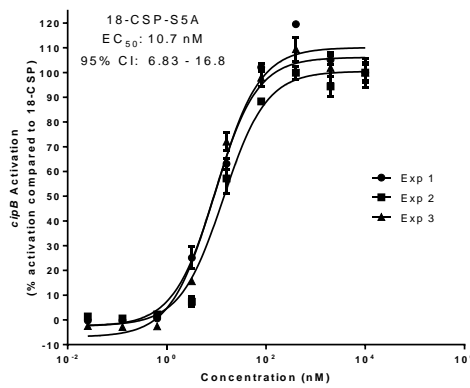
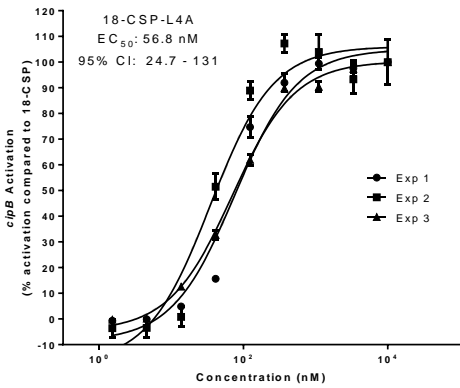
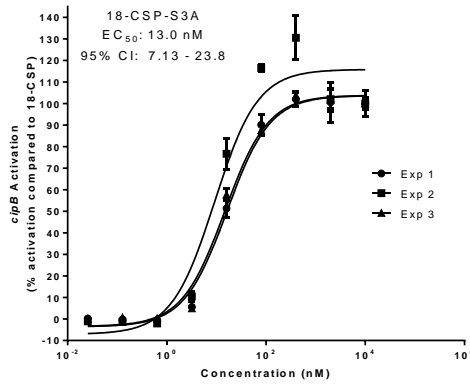
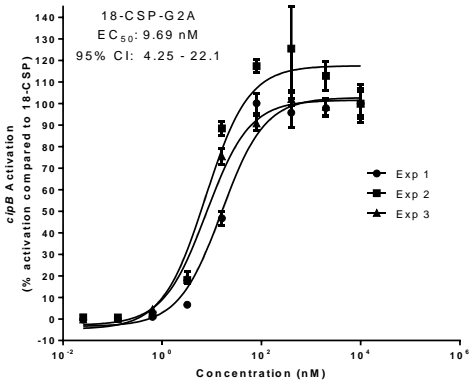
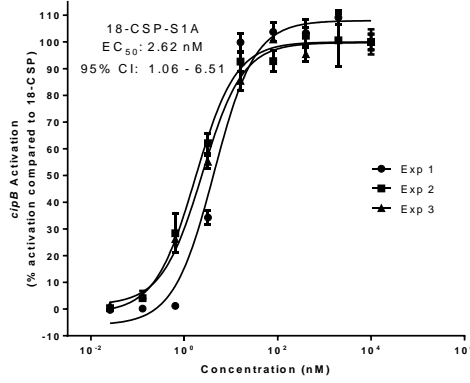
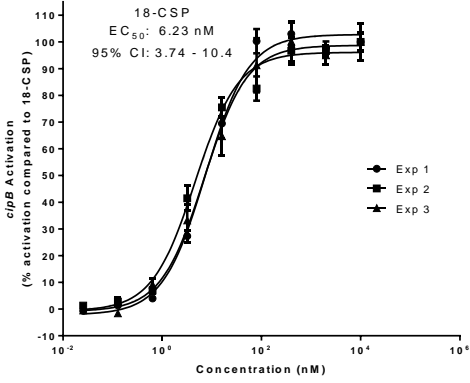


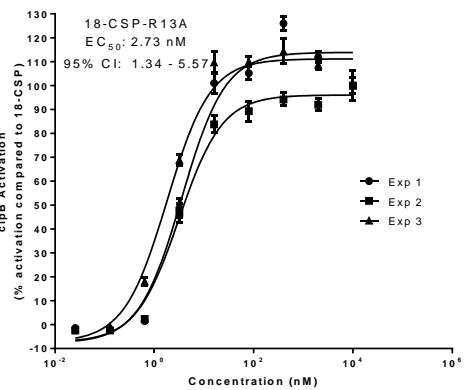
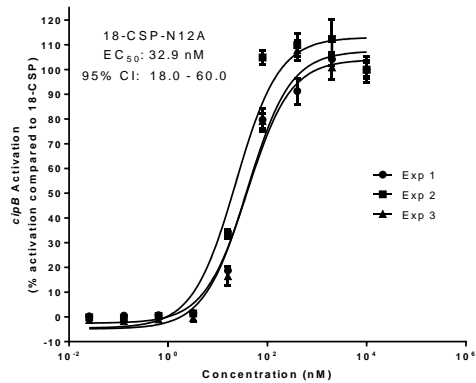
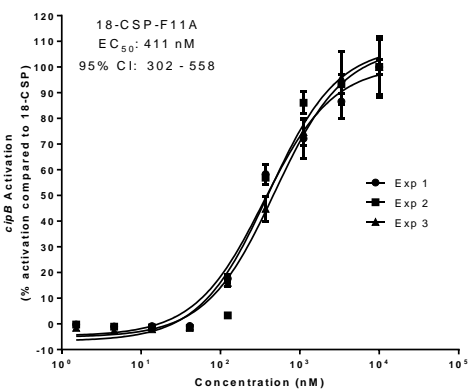
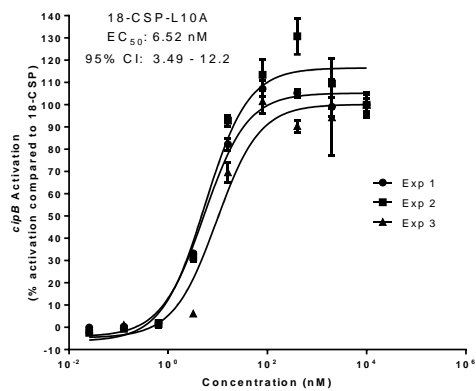
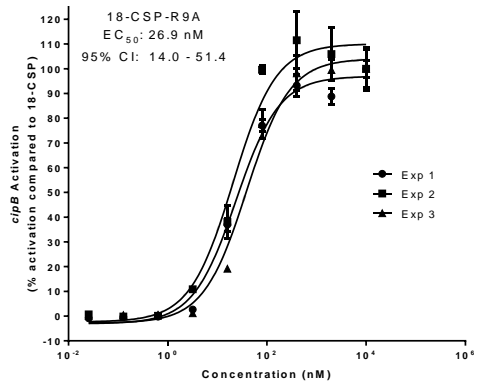
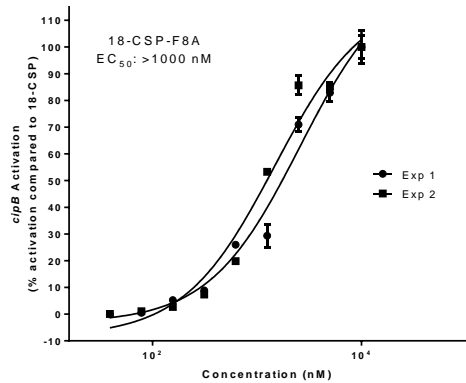
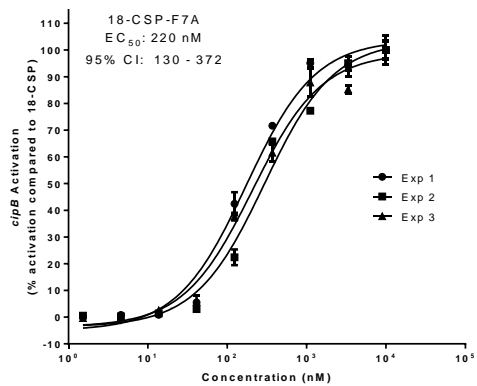
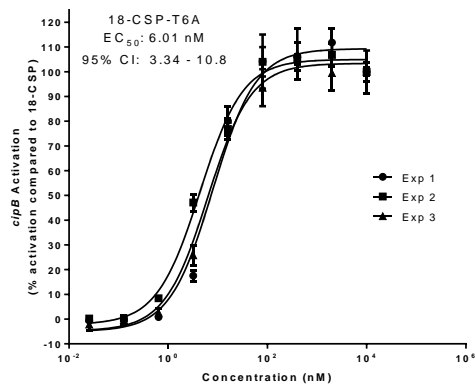


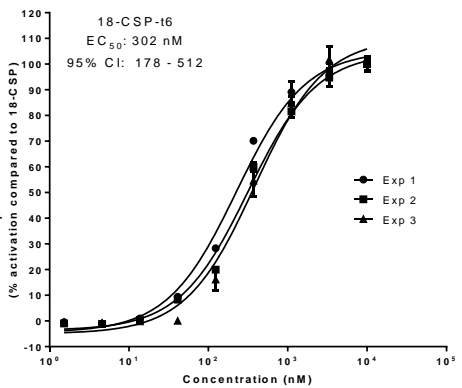
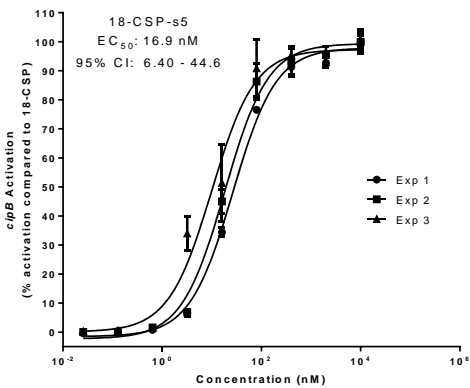
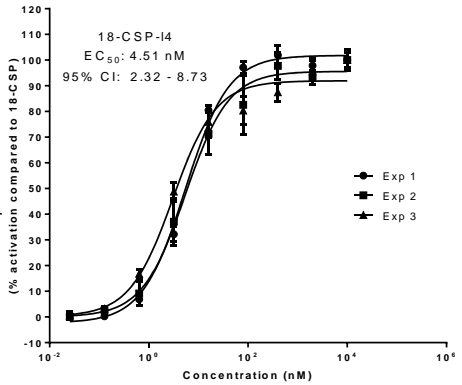
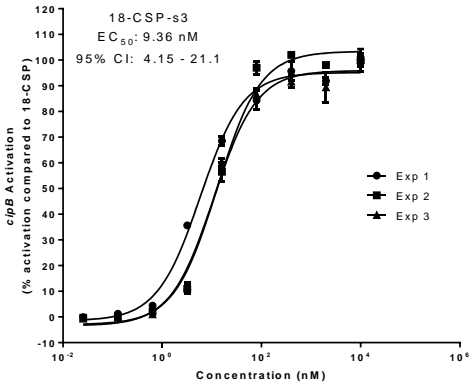
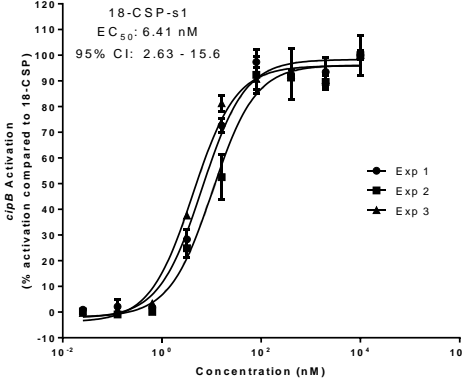
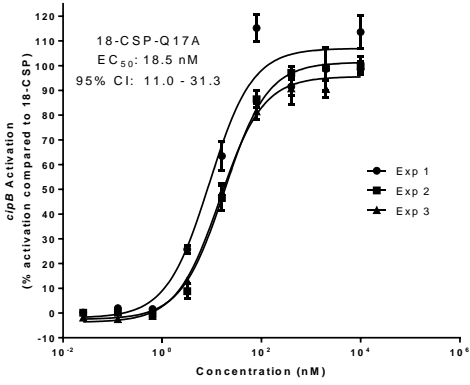
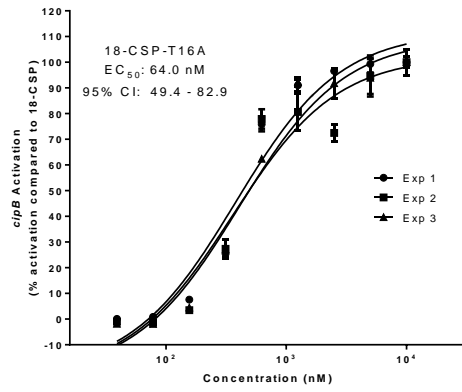
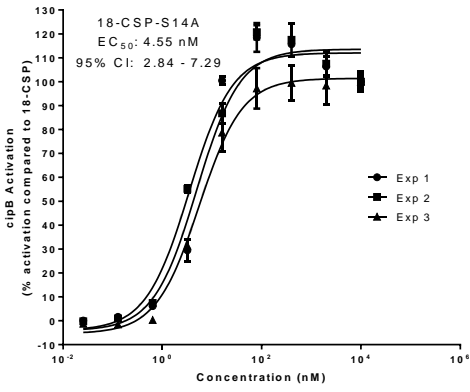


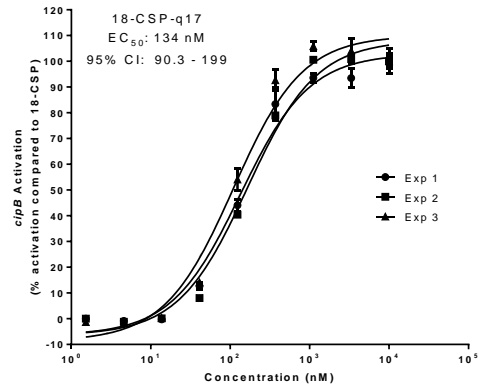
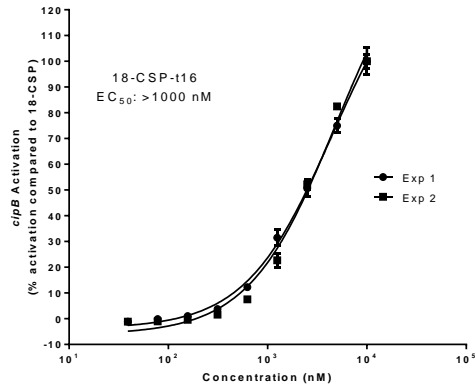
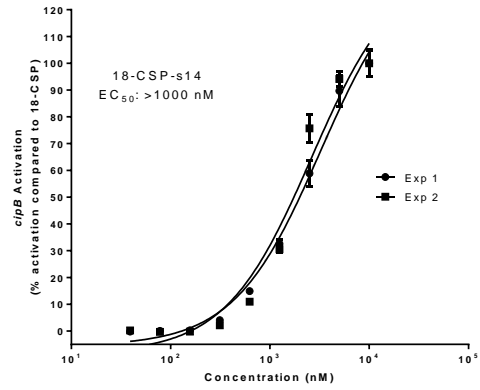
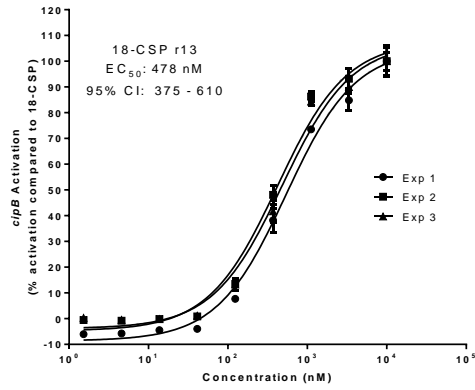
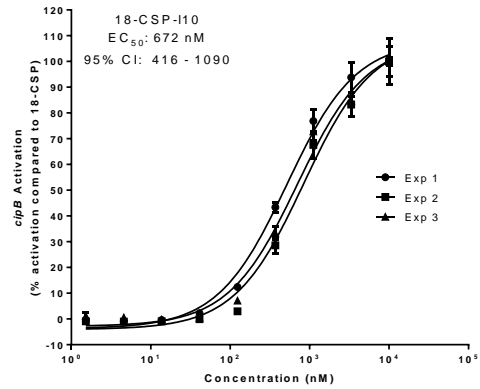
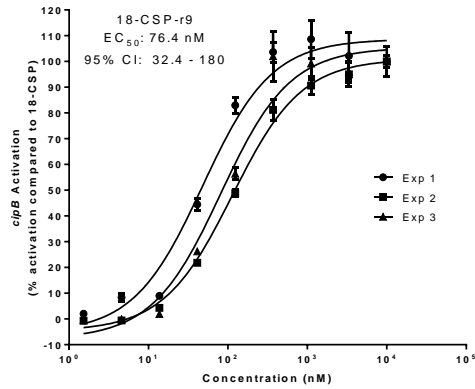
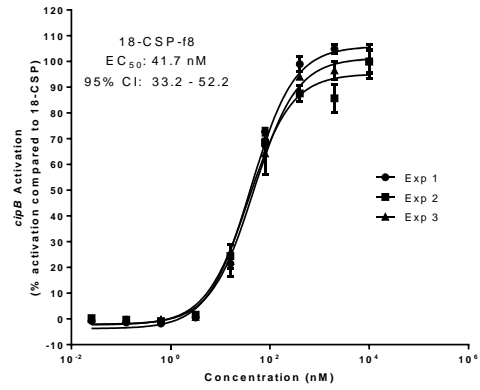
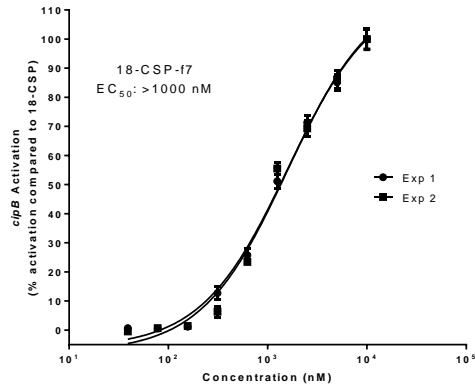


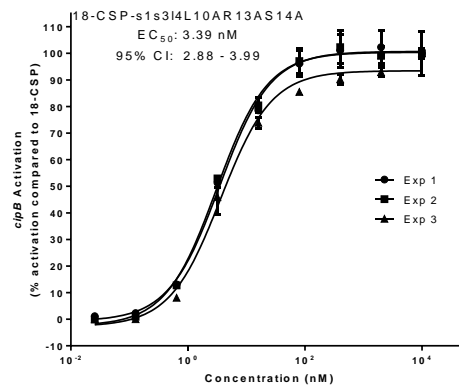
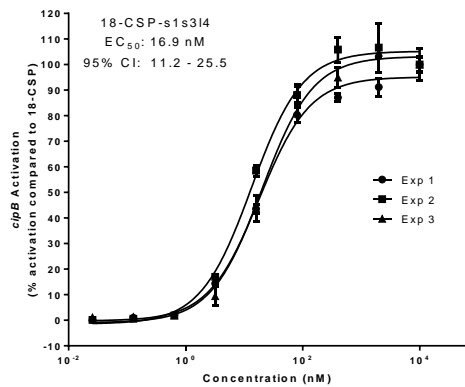
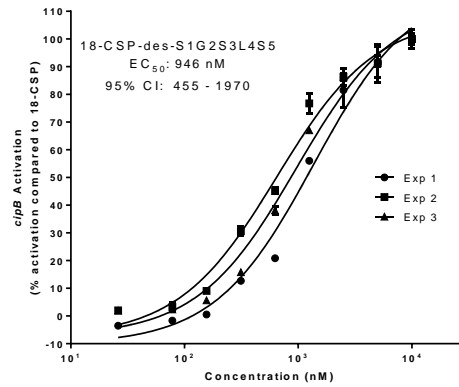
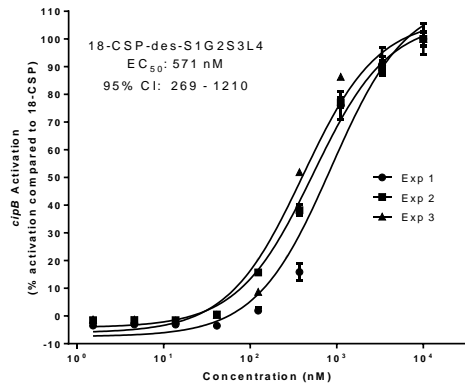
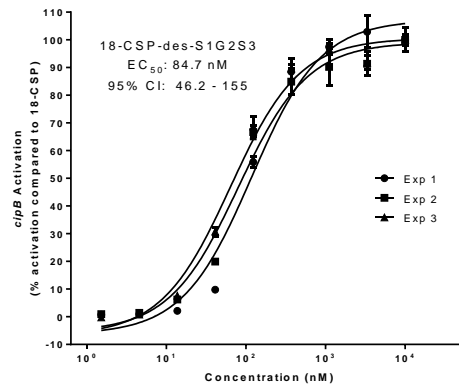
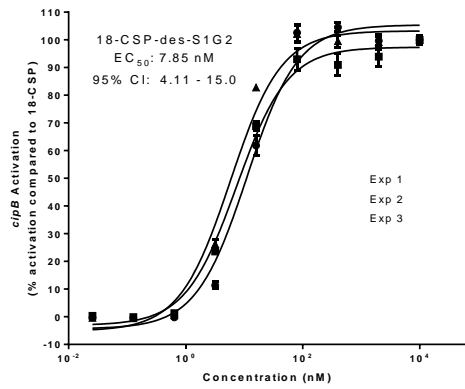
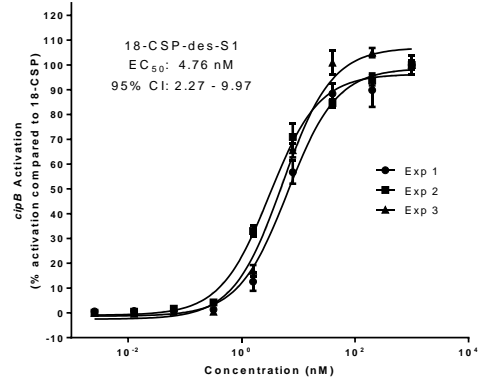
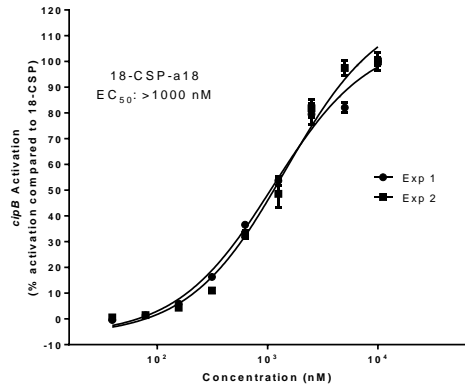
***S. mutans* SAB249 (pCipB::lacZ)**
 Activation dose response curves (18-CSP)

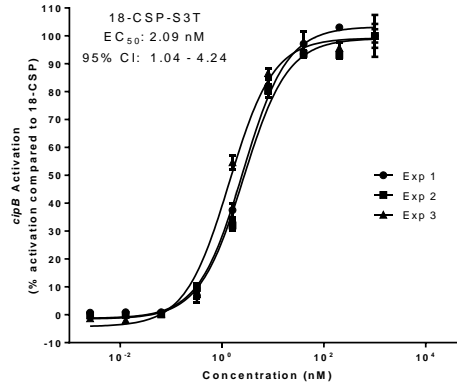
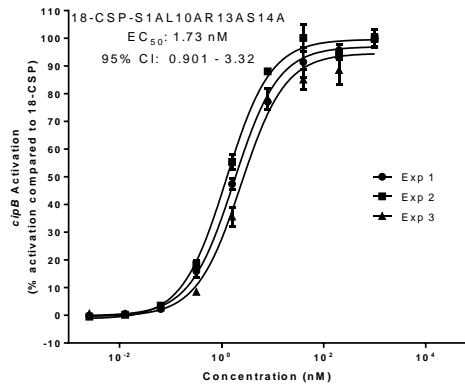




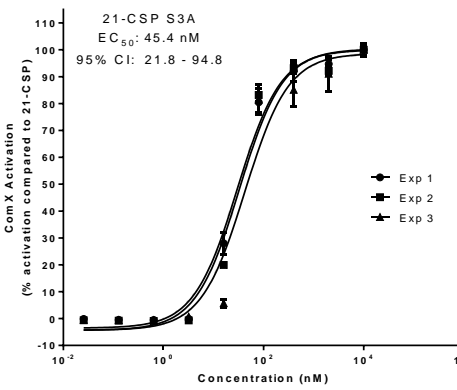
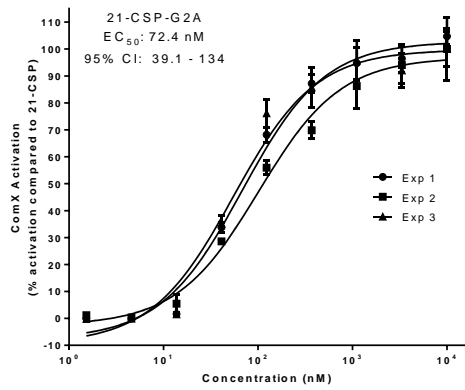
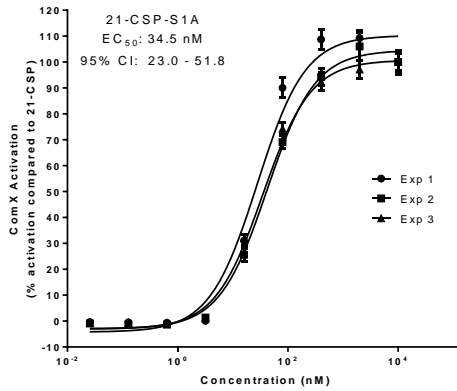
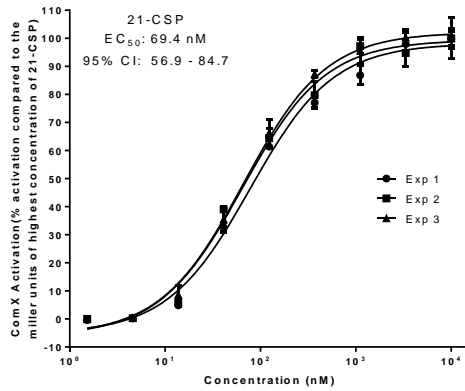


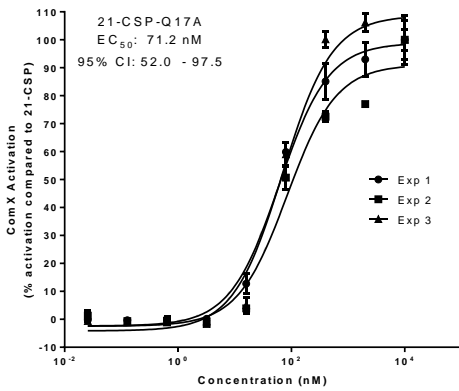
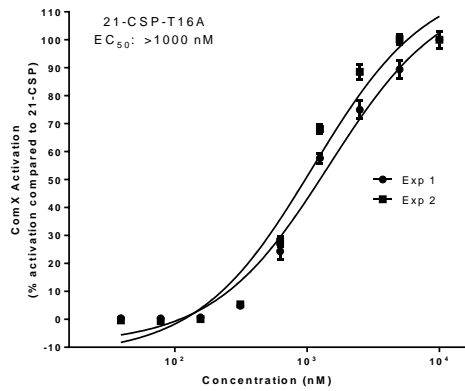
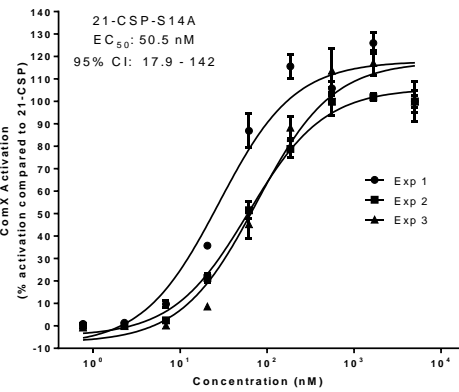
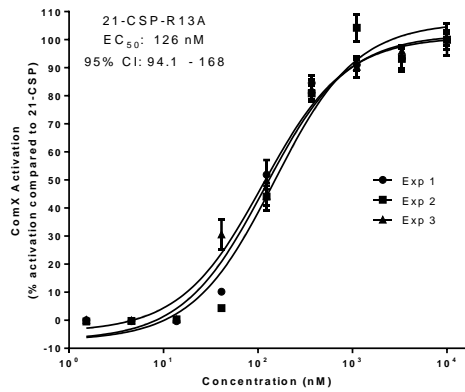
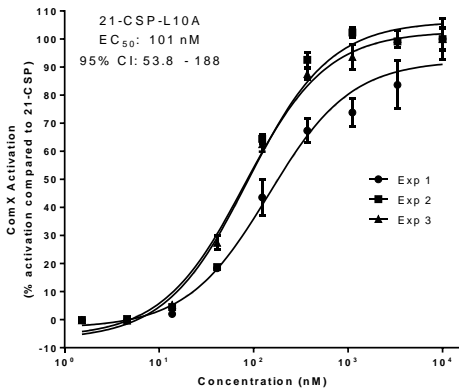
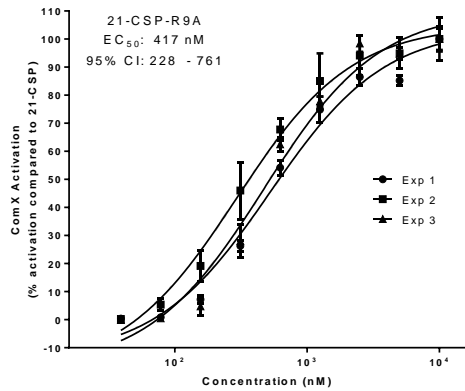
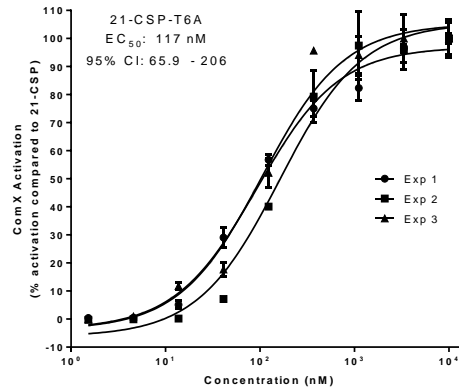
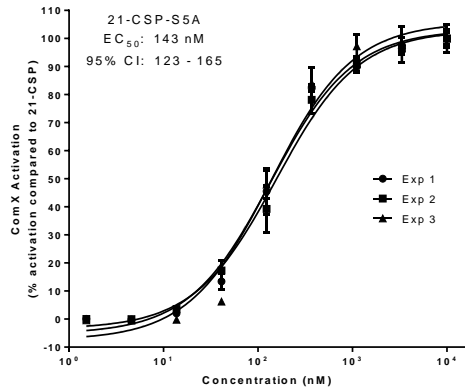


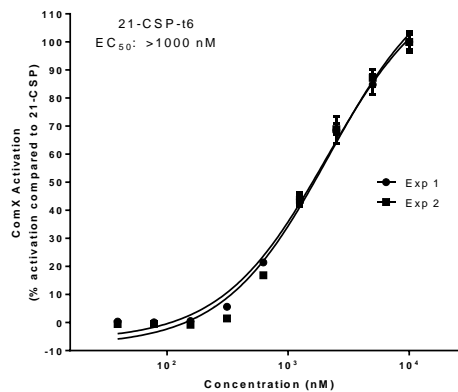
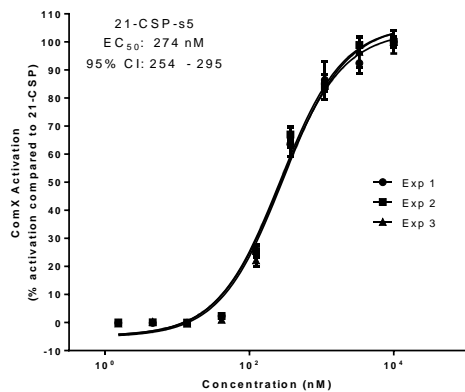
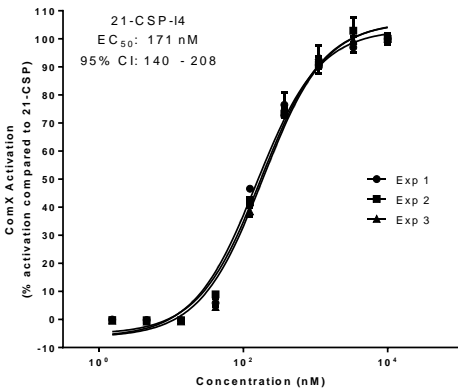
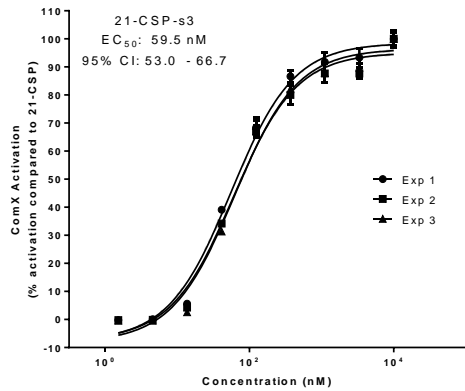
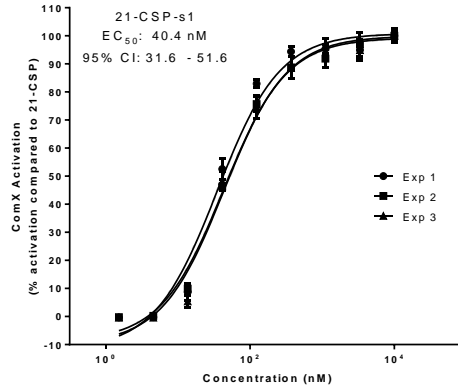
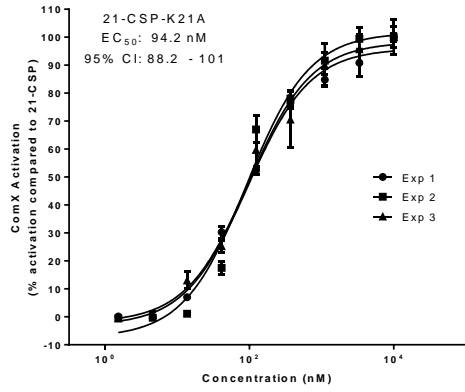
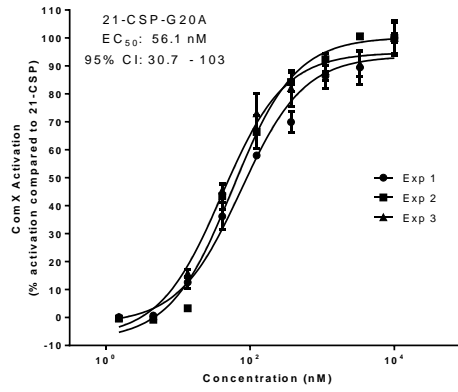
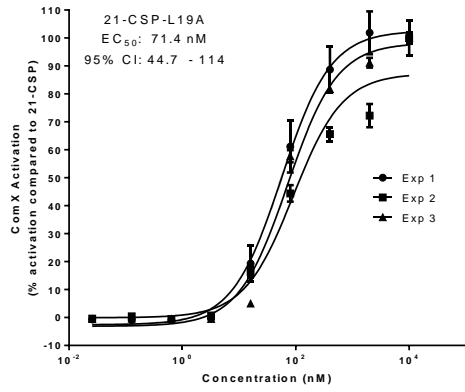


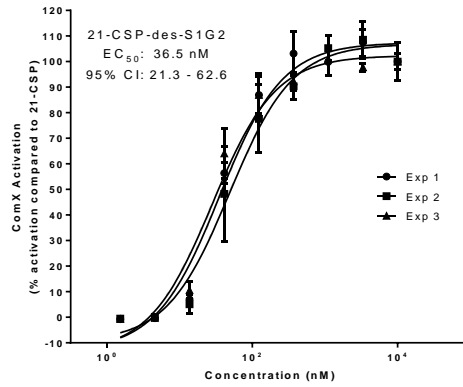
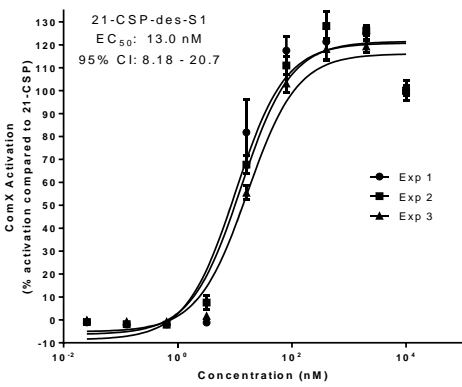
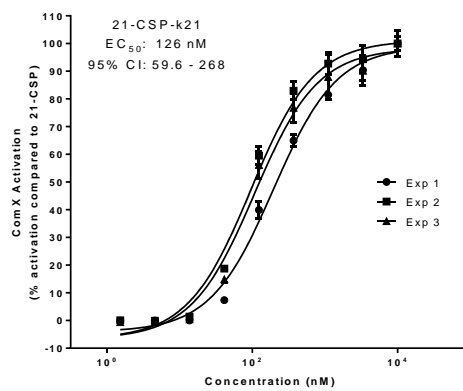
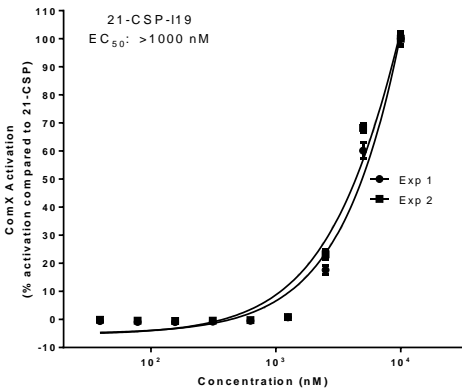
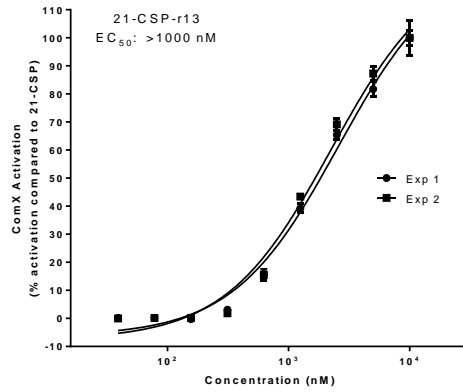
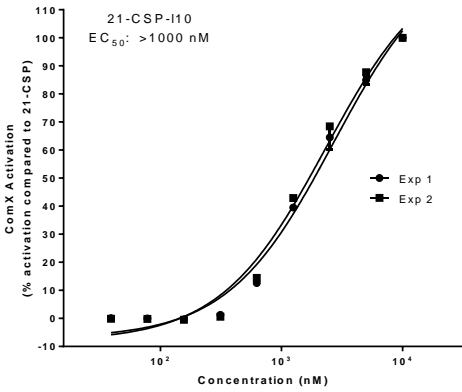
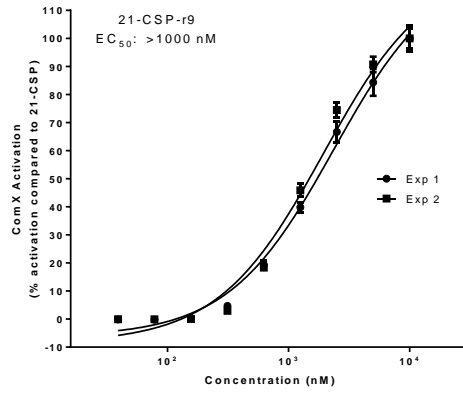
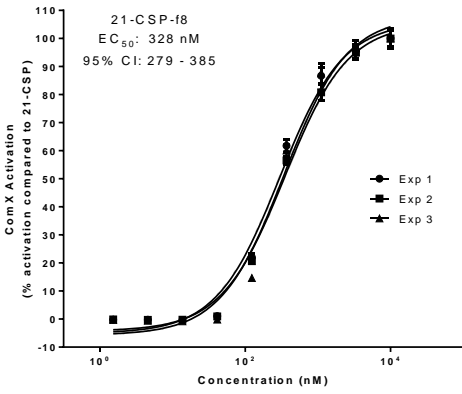


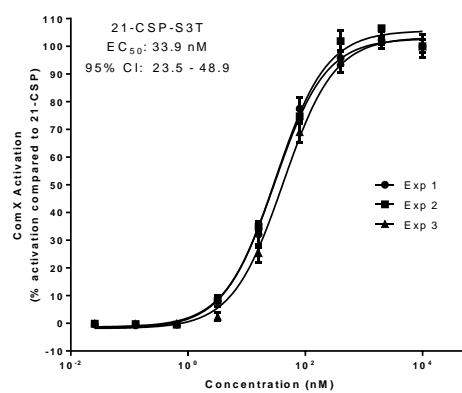
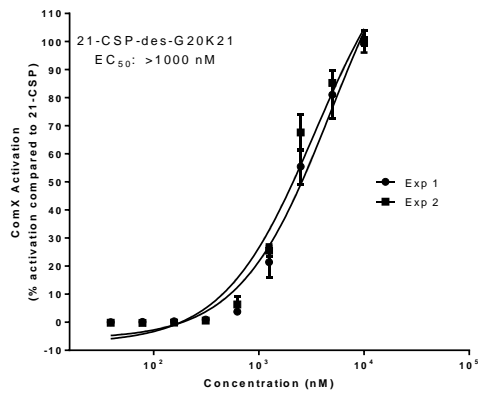
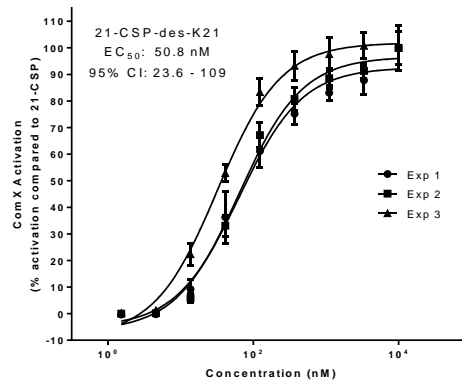
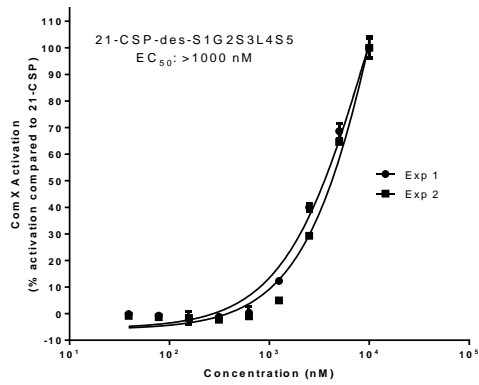
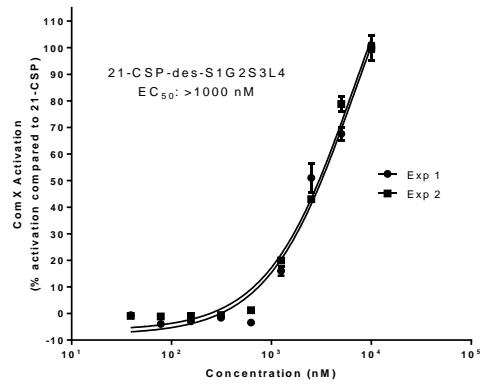
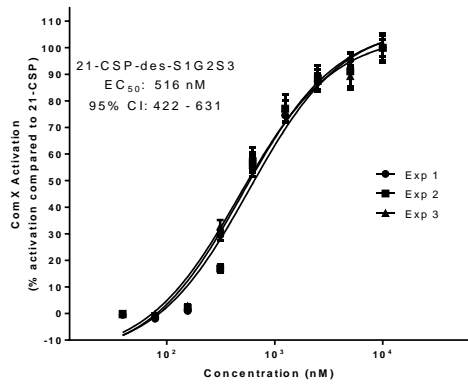
***S. mutans* SMCOM2 ($\Delta comC$, $pcomX::lacZ$)**
 Activation dose response curves (21-CSP)



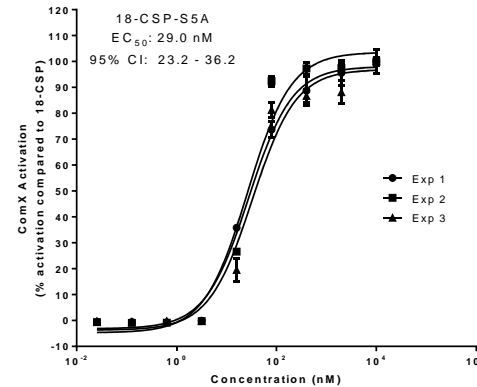
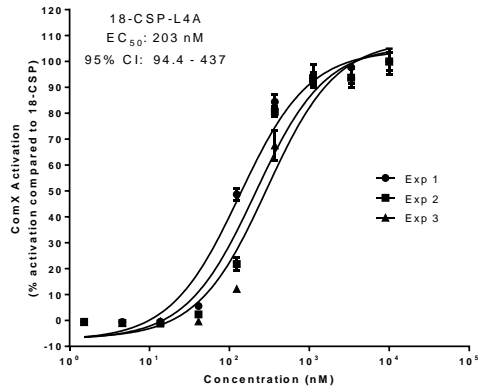
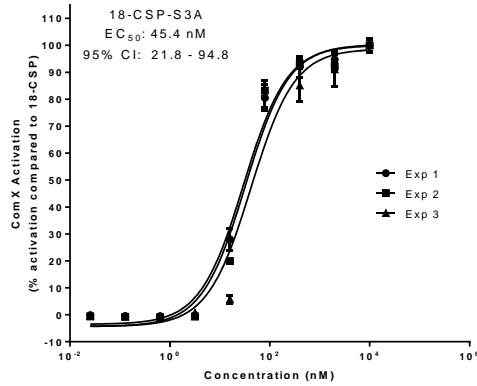
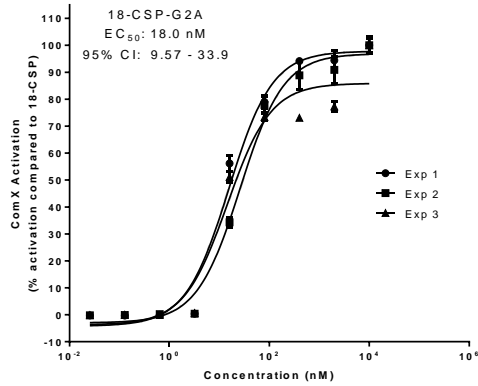
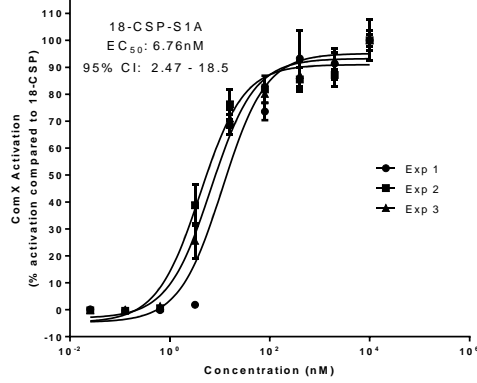
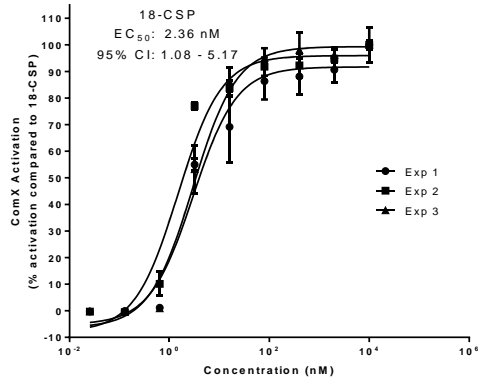


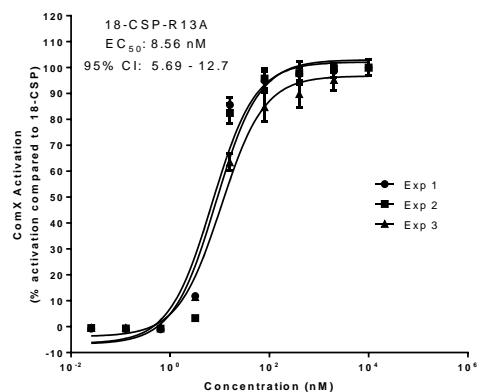
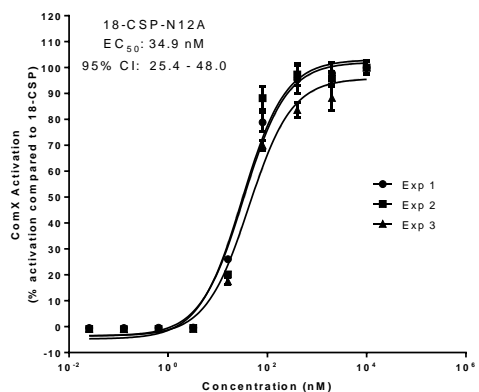
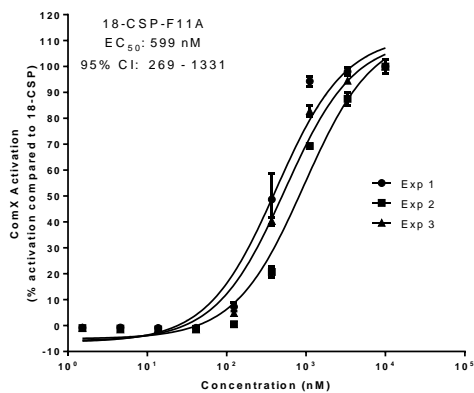
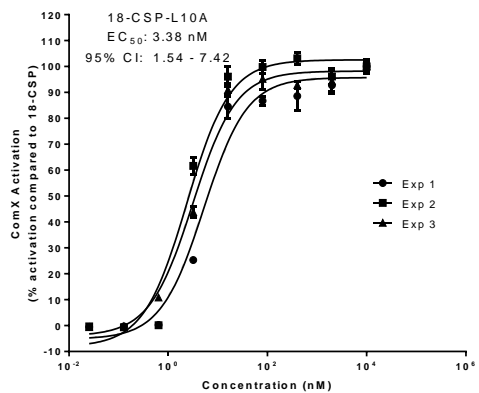
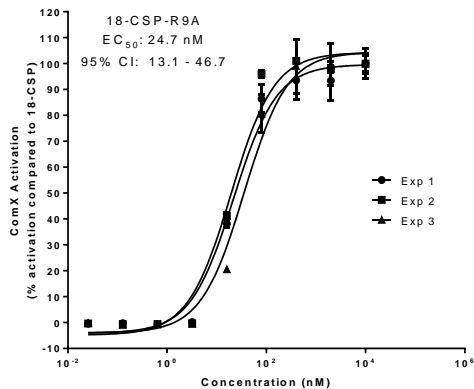
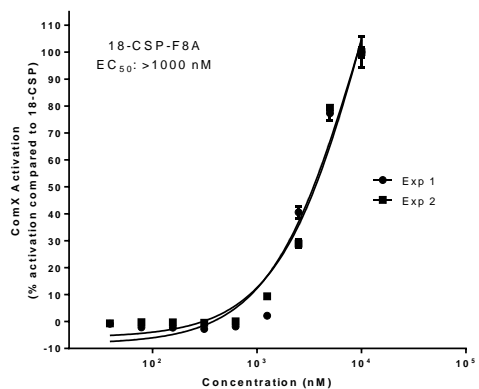
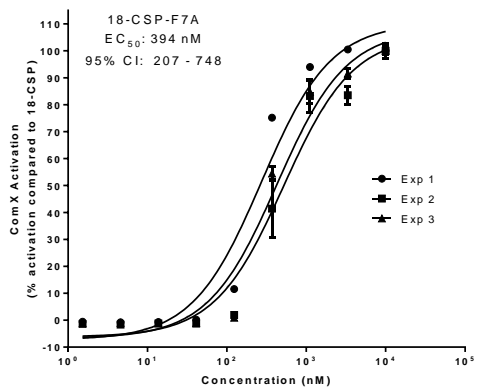
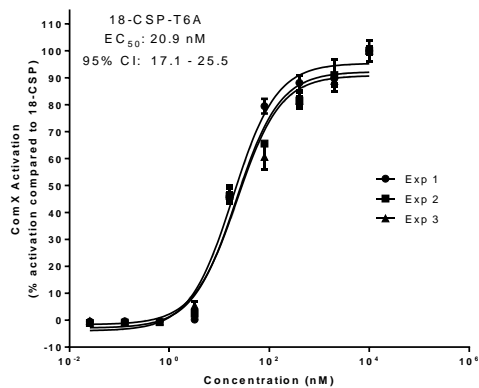


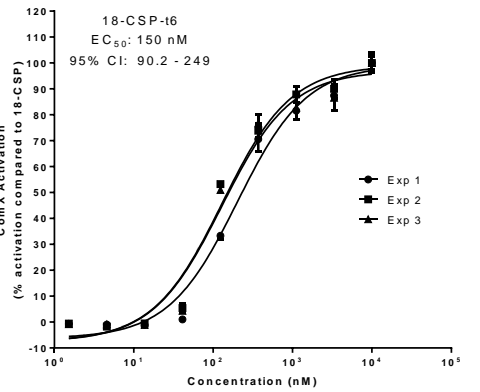
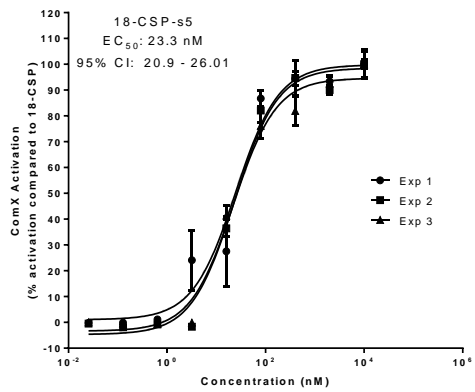
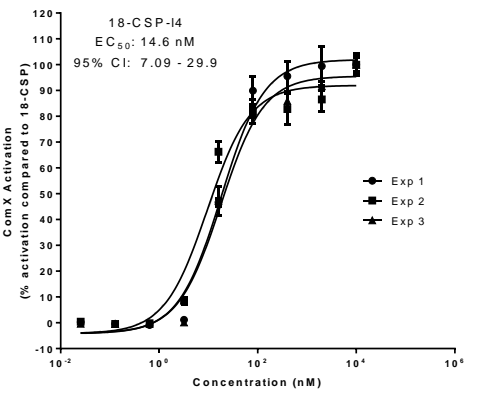
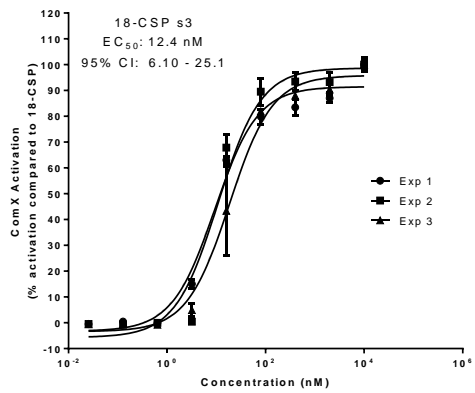
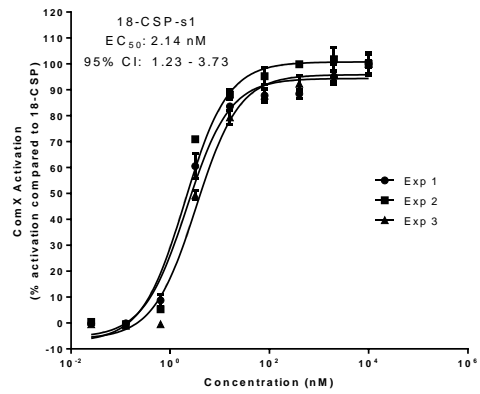
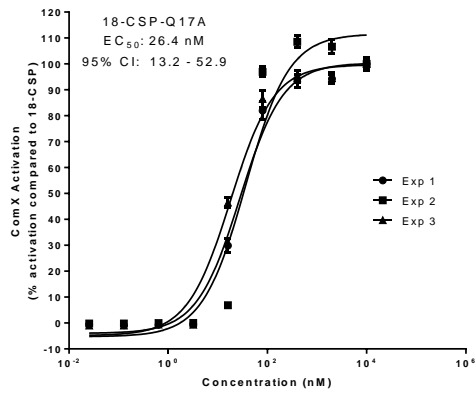
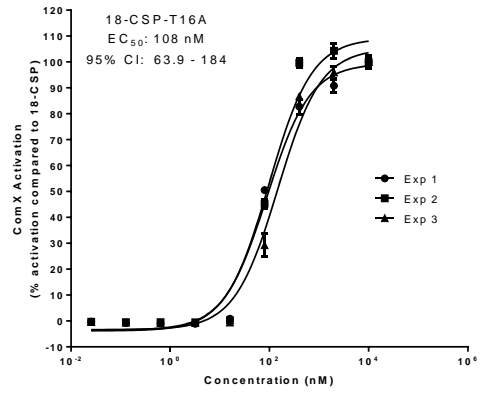
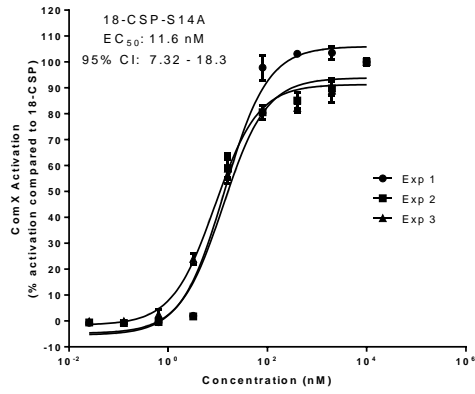


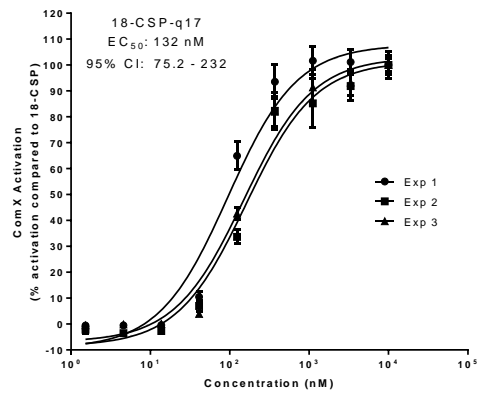
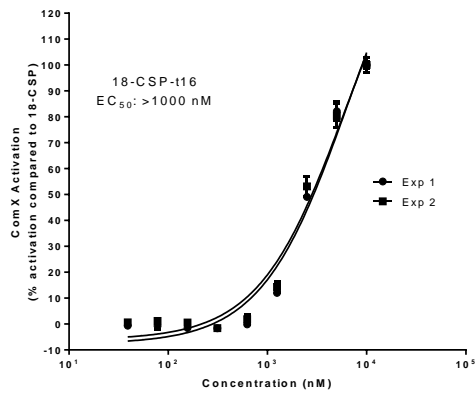
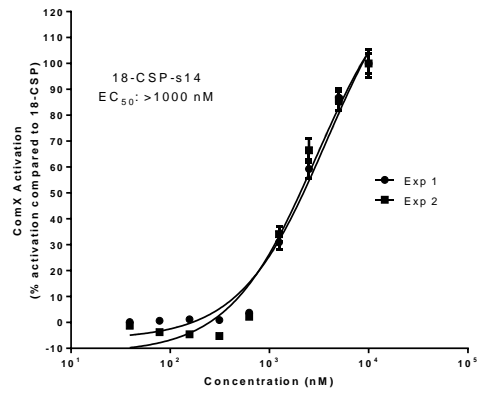
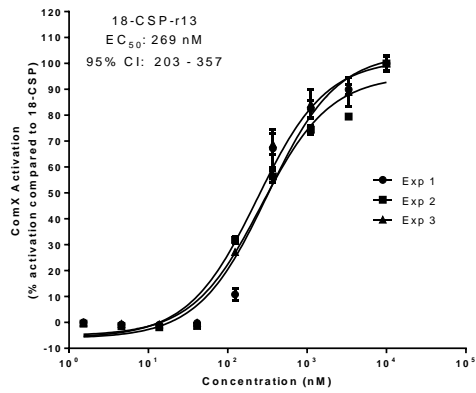
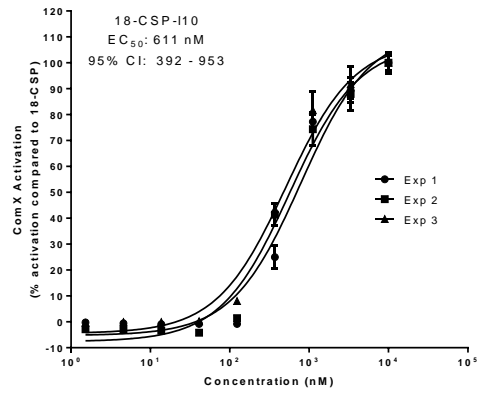
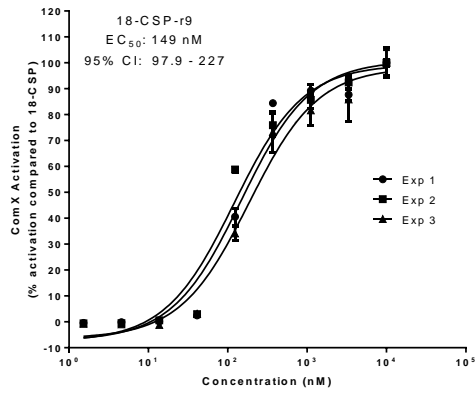
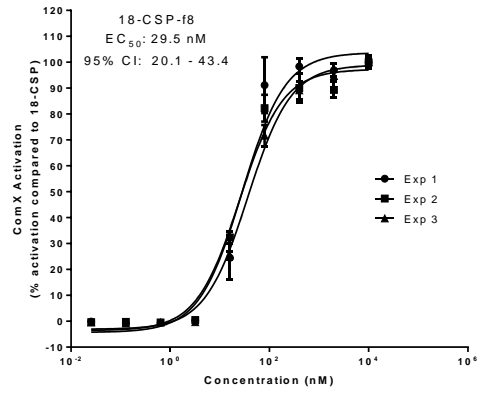
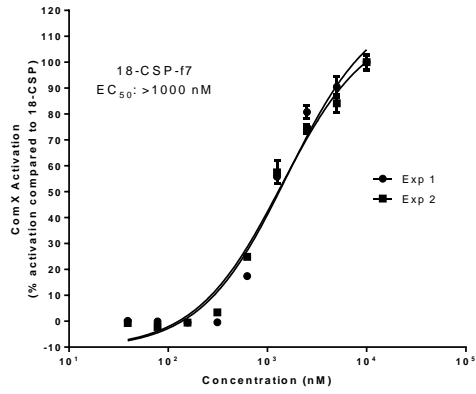


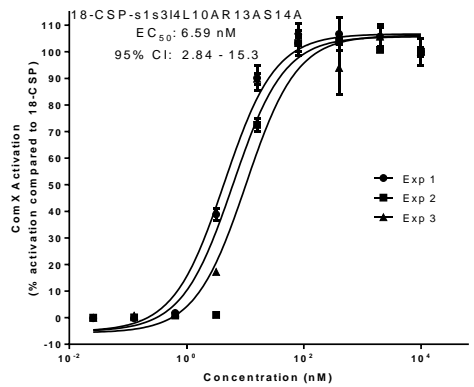
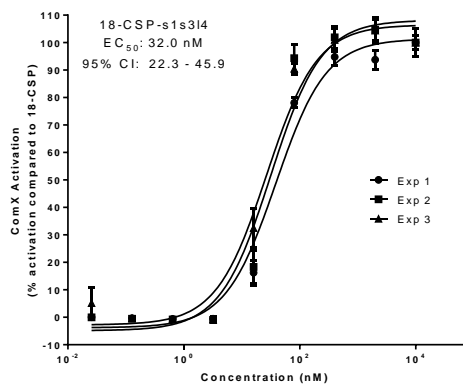
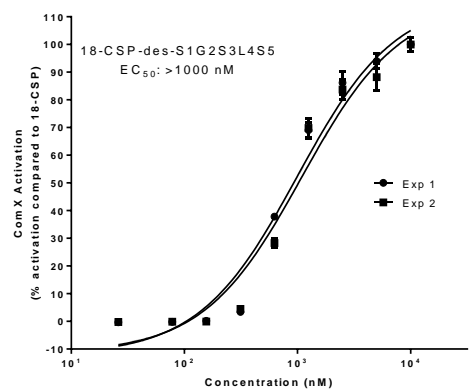
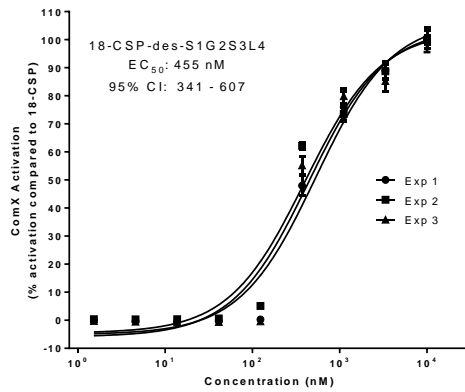
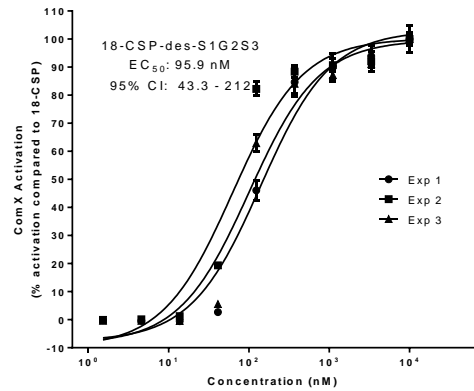
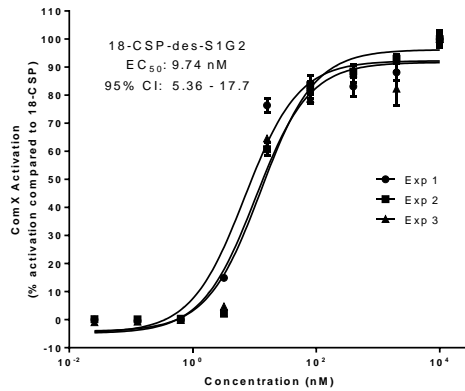
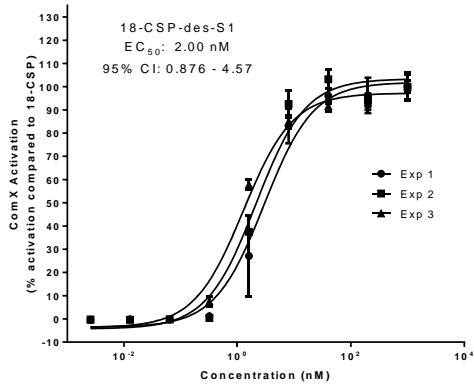
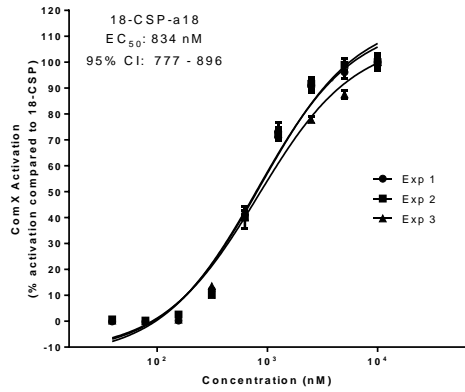
S. mutans SMCOM2 ($\Delta comC$, $pcomX::lacZ$)
 Activation dose response curves (18-CSP)

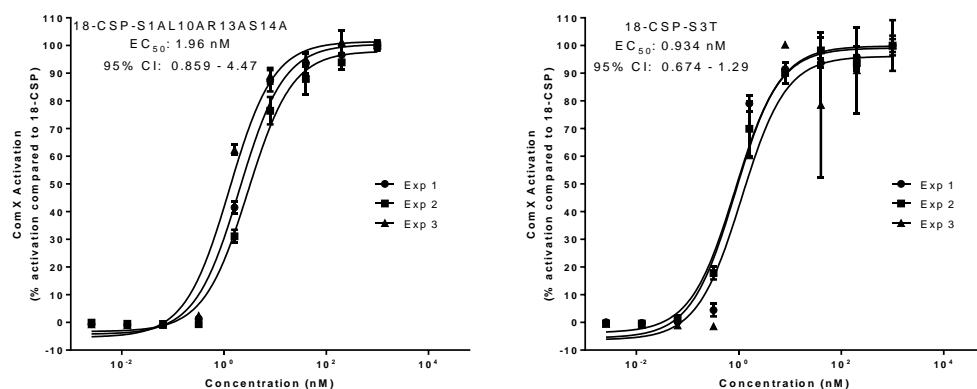












Complete tables of EC₅₀ values for CSP analogs (SMCOM2)

Table S-5. EC₅₀ values of the alanine scan of 21-CSP and 18-CSP against the *S. mutans* SMCOM2^a

Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d	Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d
21-CSP	69	57 – 85		18-CSP	2.4	1.1 – 5.2	
21-CSP-S1A	35	23 – 52	0.51	18-CSP-S1A	6.8	2.5 – 19	2.8
21-CSP-G2A	72	39 – 130	1.0	18-CSP-G2A	18	9.6 – 34	7.5
21-CSP-S3A	58	31 – 110	0.84	18-CSP-S3A	45	22 – 95	19
21-CSP-L4A	-- ^e	--	--	18-CSP-L4A	200	94 – 440	83
21-CSP-S5A	140	120 – 170	2.0	18-CSP-S5A	29	23 – 36	12
21-CSP-T6A	120	66 – 210	1.7	18-CSP-T6A	21	17 – 25.5	8.8
21-CSP-F7A	-- ^e	--	--	18-CSP-F7A	390	210 – 750	163
21-CSP-F8A	-- ^e	--	--	18-CSP-F8A	>1,000	--	--
21-CSP-R9A	420	230 – 760	6.1	18-CSP-R9A	25	13 – 47	10
21-CSP-L10A	100	54 – 190	1.5	18-CSP-L10A	3.4	1.5 – 7.4	1.4
21-CSP-F11A	-- ^e	--	--	18-CSP-F11A	600	270 – 1300	250
21-CSP-N12A	-- ^e	--	--	18-CSP-N12A	35	25 – 48	15
21-CSP-R13A	130	94 – 170	1.9	18-CSP-R13A	8.6	5.7 – 13	3.6
21-CSP-S14A	51	18 – 140	0.74	18-CSP-S14A	12	7.3 – 18	5.0
21-CSP-F15A	-- ^e	--	--	18-CSP-F15A	-- ^e	--	--
21-CSP-T16A	>1,000	--	--	18-CSP-T16A	110	64 – 180	46
21-CSP-Q17A	71	52 – 98	1.0	18-CSP-Q17A	26	13 – 53	11
21-CSP-L19A	71	45 – 110	1.0				
21-CSP-G20A	56	31 – 100	0.81				
21-CSP-K21A	94	88 – 100	1.4				

^a See experimental section for details on reporter strain and methods. See supporting information for primary screening assay results and plots of agonism dose response curves. All assays were performed in triplicate. ^b EC₅₀ values determined by testing peptides over a range of concentrations. ^c 95% confidence interval. ^d Ratio where each analog's EC₅₀ is divided by native CSP EC₅₀; a value <1 indicates a better activator than the parent CSP. ^e EC₅₀ not determined due to the analog's low induction in primary agonism screening assay.

Table S-6. EC₅₀ values of the D-amino acid scan of 21-CSP and 18-CSP against the *S. mutans* SMCOM2^a

Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d	Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d
21-CSP	69	57 – 85		18-CSP	2.4	1.1 – 5.2	
21-CSP-s1	40	32 – 52	0.58	18-CSP-s1	2.1	1.2 – 3.7	0.88
21-CSP-s3	60	53 – 67	0.87	18-CSP-s3	12	6.1 – 25	5.0
21-CSP-l4	170	140 – 210	2.5	18-CSP-l4	15	7.1 – 30	6.3
21-CSP-s5	270	250 – 300	3.9	18-CSP-s5	23	21 – 26	9.6
21-CSP-t6	>1,000	--	--	18-CSP-t6	150	90 – 250	63
21-CSP-f7	-- ^e	--	--	18-CSP-f7	>1,000	--	--
21-CSP-f8	330	280 – 390	4.8	18-CSP-f8	30	20 – 43	13
21-CSP-r9	>1,000	--	--	18-CSP-r9	150	98 – 230	63
21-CSP-l10	>1,000	--	--	18-CSP-l10	610	390 – 950	254
21-CSP-f11	-- ^e	--	--	18-CSP-f11	-- ^e	--	--
21-CSP-n12	-- ^e	--	--	18-CSP-n12	-- ^e	--	--
21-CSP-r13	>1,000	--	--	18-CSP-r13	270	200 – 360	113
21-CSP-s14	-- ^e	--	--	18-CSP-s14	>1,000	--	--
21-CSP-f15	-- ^e	--	--	18-CSP-f15	-- ^e	--	--
21-CSP-t16	-- ^e	--	--	18-CSP-t16	>1,000	--	--
21-CSP-q17	-- ^e	--	--	18-CSP-q17	130	75 – 230	54
21-CSP-a18	-- ^e	--	--	18-CSP-a18	830	780 – 900	346
21-CSP-l19	>1,000	--	--				
21-CSP-k21	130	60 – 270	1.9				

^a See experimental section for details on the reporter strain and methods. See supporting information for primary screening assay results and plots of agonism dose response curves. All assays performed in triplicate. ^b EC₅₀ values determined by testing peptides over a range of concentrations. ^c 95% confidence interval. ^d Ratio where each analog's EC₅₀ is divided by native CSP EC₅₀; a value <1 indicates a better activator than the parent CSP. ^e EC₅₀ not determined due to the analog's low induction in primary agonism screening assay.

Table S-7. EC₅₀ Values of the 2nd generation Analogs of 21-CSP and 18-CSP against the *S. mutans* SMCOM2^a

Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d	Name	EC ₅₀ (nM) ^b	95% CI ^c	Fold change ^d
21-CSP	69	57 – 85		18-CSP	2.4	1.1 – 5.2	
21-CSP-des-S1	13	8.2 – 21	0.19	18-CSP-des-S1	2.0	0.88 – 4.6	0.83
21-CSP-des-S1G2	37	21 – 63	0.53	18-CSP-des-S1G2	9.7	5.4 – 18	4.0
21-CSP-des-S1G2S3	520	420 – 630	7.5	18-CSP-des-S1G2S3	96	43 – 210	40
21-CSP-des-S1G2S3L4	>1,000	--	--	18-CSP-des-S1G2S3L4	460	340 – 610	192
21-CSP-des-S1G2S3L4S5	>1,000	--	--	18-CSP-des-S1G2S3L4S5	>1,000	--	--
21-CSP-des-K21	51	24 – 110	0.74	18-CSP-des-A18	-- ^e	--	--
21-CSP-des-G20K21	>1,000	--	--	18-CSP-des-Q17A18	-- ^e	--	--
21-CSP-S3T	34	24 – 49	0.49	18-CSP-S3T	0.93	0.67 – 1.3	0.39
				18-CSP-s1s3l4	32	22 – 46	13
				18-CSP-s1s3l4L10AR13AS14A	6.6	2.8 – 15	2.8
				18-CSP-S1AL10AR13AS14A	2.0	0.86 – 4.5	0.83

^a See experimental section for details on the reporter strain and methods. See supporting information for primary screening assay results and plots of agonism dose response curves. All assays performed in triplicate. ^b EC₅₀ values determined by testing peptides over a range of concentrations. ^c 95% confidence interval. ^d Ratio where each analog's EC₅₀ is divided by native CSP EC₅₀; a value <1 indicates a better activator than the parent CSP. ^e EC₅₀ not determined due to the analog's low induction in primary agonism screening assay.

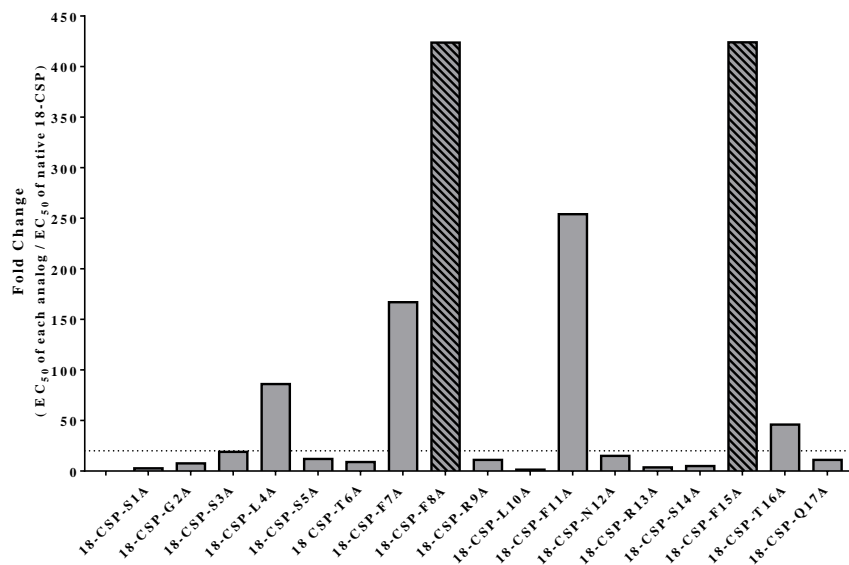


Figure S-13. Plot of the ratio of 18-CSP alanine analogs EC₅₀ values divided by the EC₅₀ value of 18-CSP in the SMCOM2 strain. The plot reveals a periodic sharp reduction in activity every 3-4 residues along the peptide sequence, consistent with an α -helix conformation. Striped bars represent analogs with EC₅₀ values >1,000 nM, thus the EC₅₀ ratio is >425. >20-fold decrease in potency was considered significant (dashed line).

Comparison of results found from both reporter strains SAB249 and SMCOM2

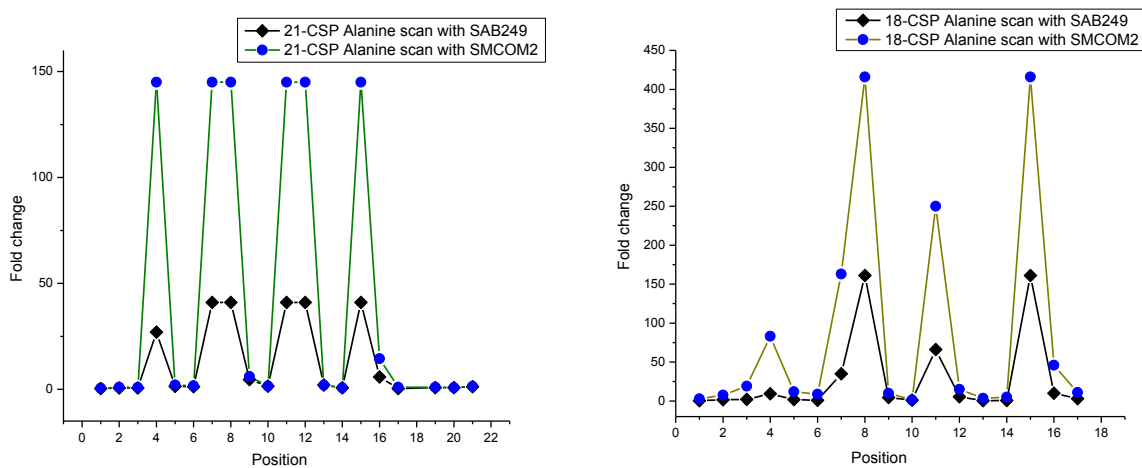


Figure S-14. Comparison of alanine scan results of 21-CSP and 18-CSP with both reporter strains SAB249 and SMCOM2. These results showed a similar trend in changing activity with both reporter systems.

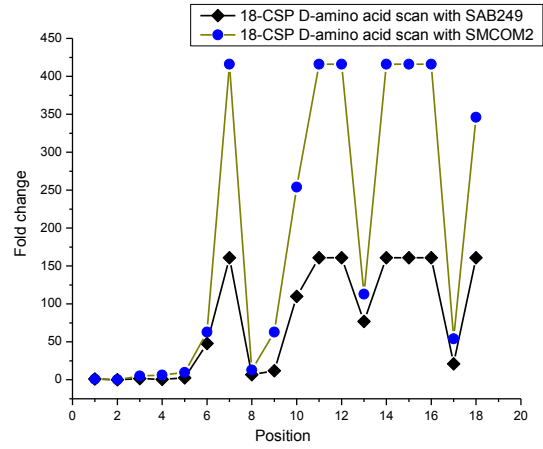
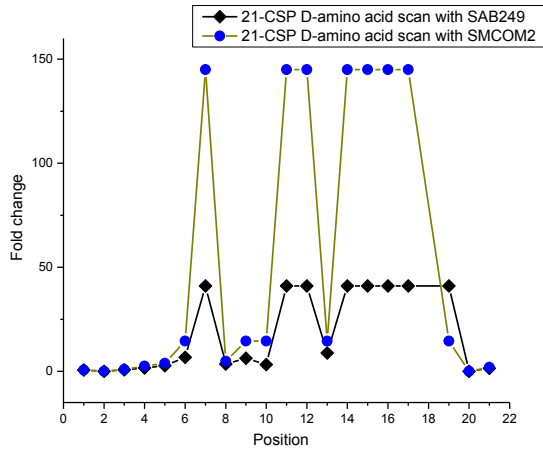


Figure S-15. Comparison of D-amino acid scan results of 21-CSP and 18-CSP with both reporter strains SAB249 and SMCOM2. These results showed a similar trend in changing activity with both reporter systems.

Circular dichroism (CD) spectra

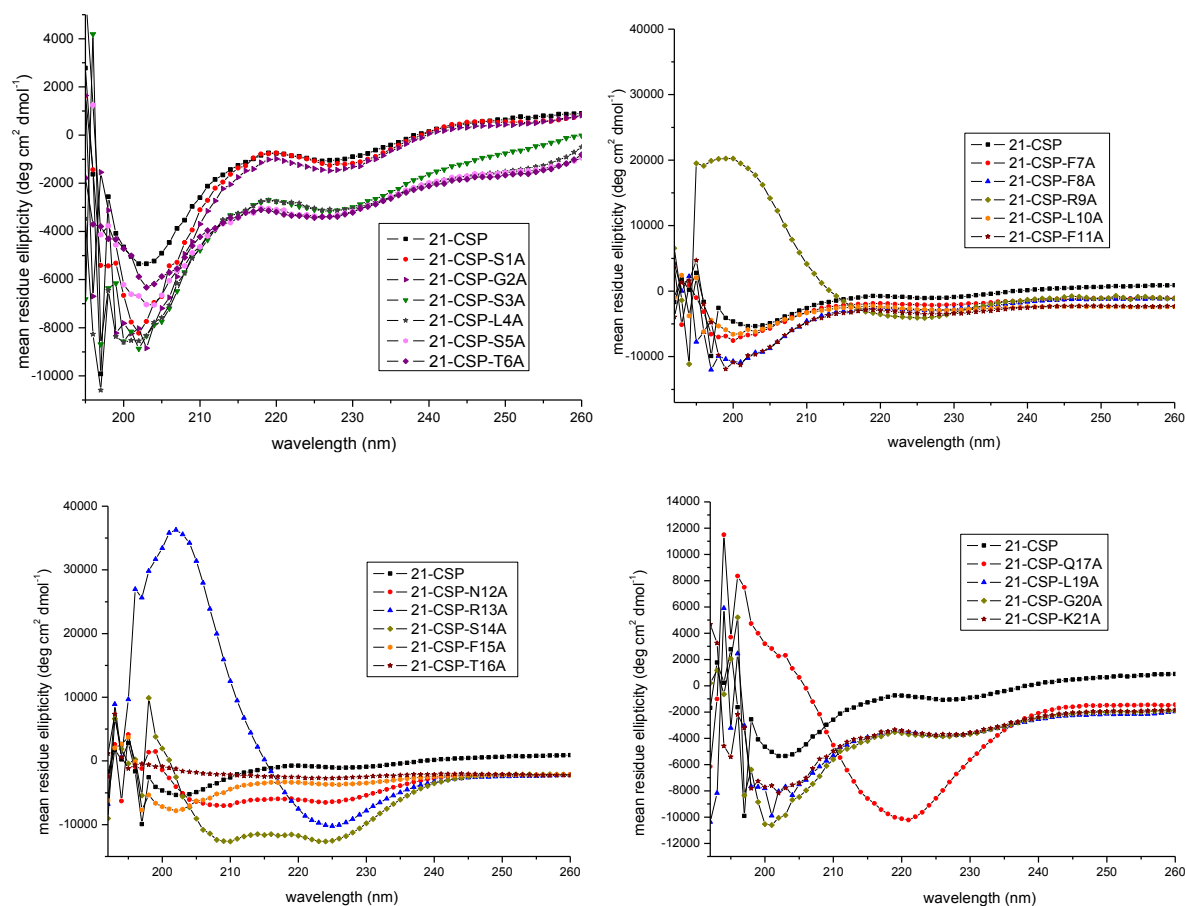


Figure S-16. CD spectra of the 21-CSP alanine scan library in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most peptides exhibit a random coil pattern, while 21-CSP-R9A and 21-CSP-R13A exhibit a β -turn pattern, 21-CSP-Q17A exhibits a β -sheet pattern, and 21-CSP-S14A exhibits an α -helix pattern.

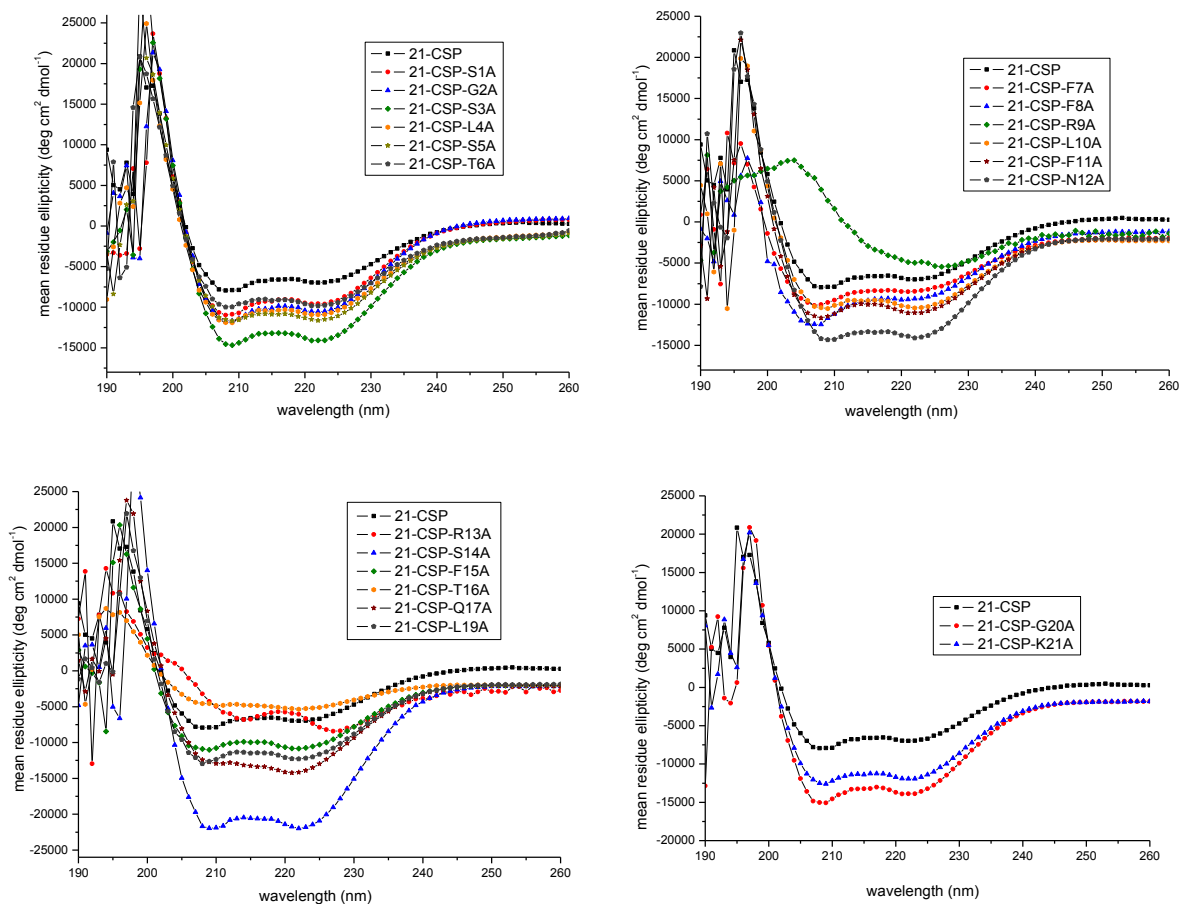


Figure S-17. CD spectra of the 21-CSP alanine scan library in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μM . 21-CSP was added as a control. Peptides exhibit varying degrees of an α -helix pattern. Measurements of 21-CSP-R9A and 21-CSP-R13A were performed with a peptide concentration of 100 μM . 21-CSP-R9A exhibits a distorted β -sheet pattern and 21-CSP-R13A exhibits a distorted α -helix (mixed with β -sheets and β -turns) pattern.¹

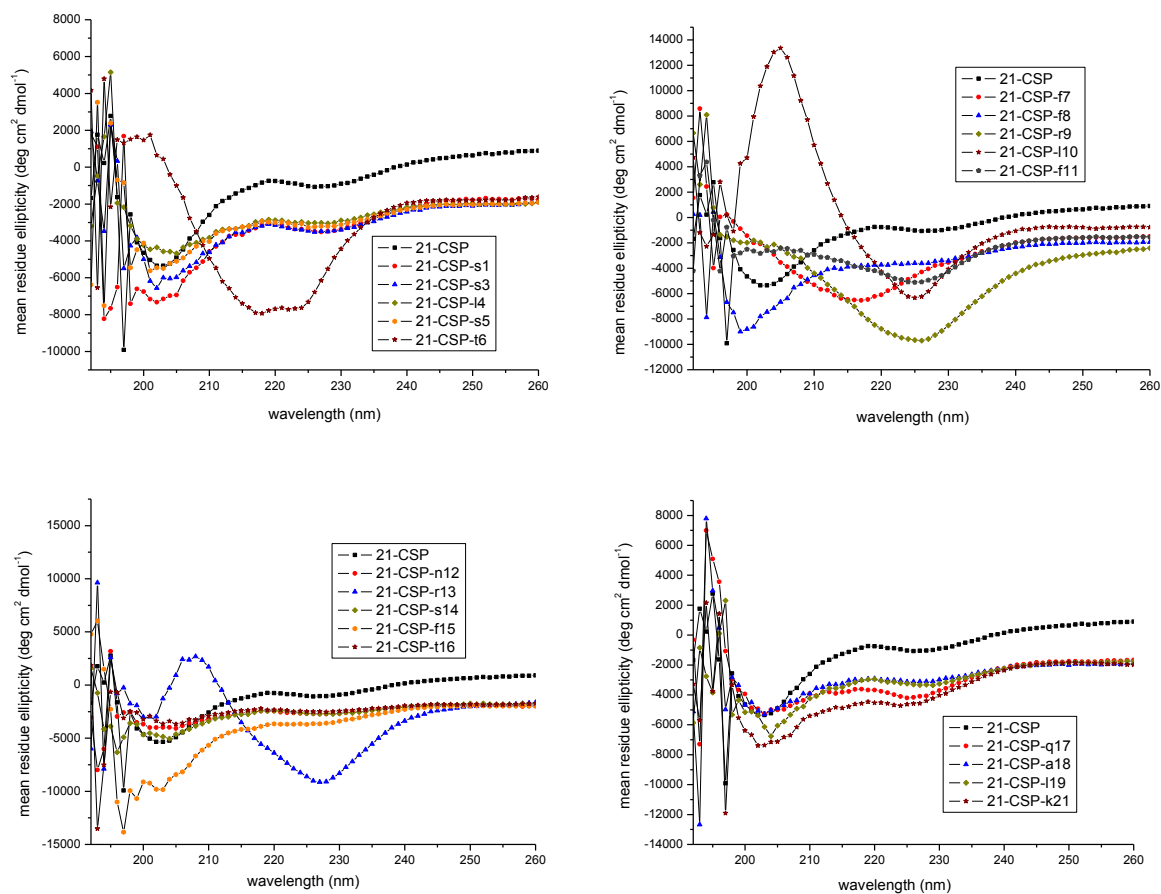


Figure S-18. CD spectra of the 21-CSP D-amino acid scan library in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most peptides exhibit a random coil pattern, while 21-CSP-t6 exhibits an α -helix pattern, 21-CSP-r9 and 21-CSP-f11 exhibit a β -sheet pattern, and 21-CSP-110 and 21-CSP-r13 exhibit a β -turn pattern.

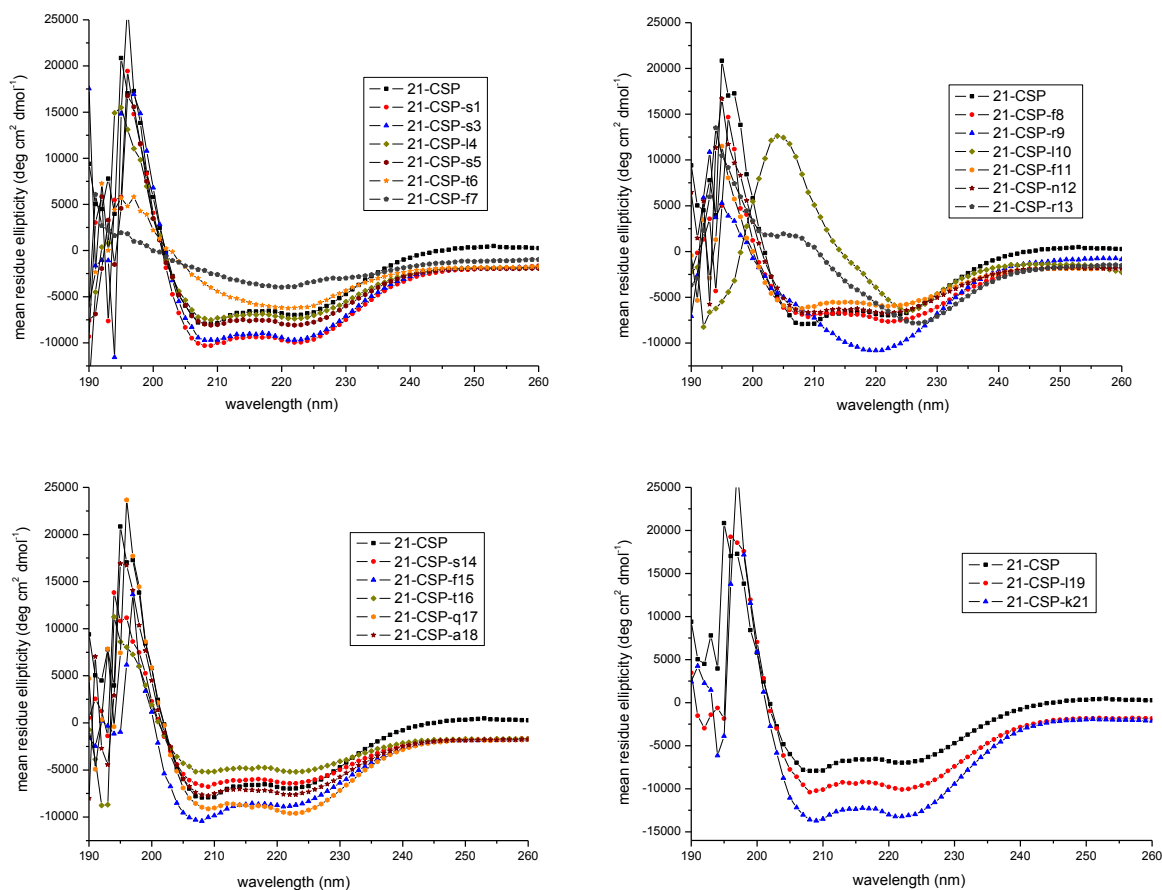


Figure S-19. CD spectra of the 21-CSP D-amino acid scan library in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most Peptides exhibit varying degrees of an α -helix pattern, while 21-CSP-t6 and 21-CSP-r9 exhibit a β -sheet pattern, 21-CSP-f7 exhibits a distorted β -sheet (mixed with α -helix) pattern, and 21-CSP-t10 and 21-CSP-r13 exhibit a β -turn pattern.

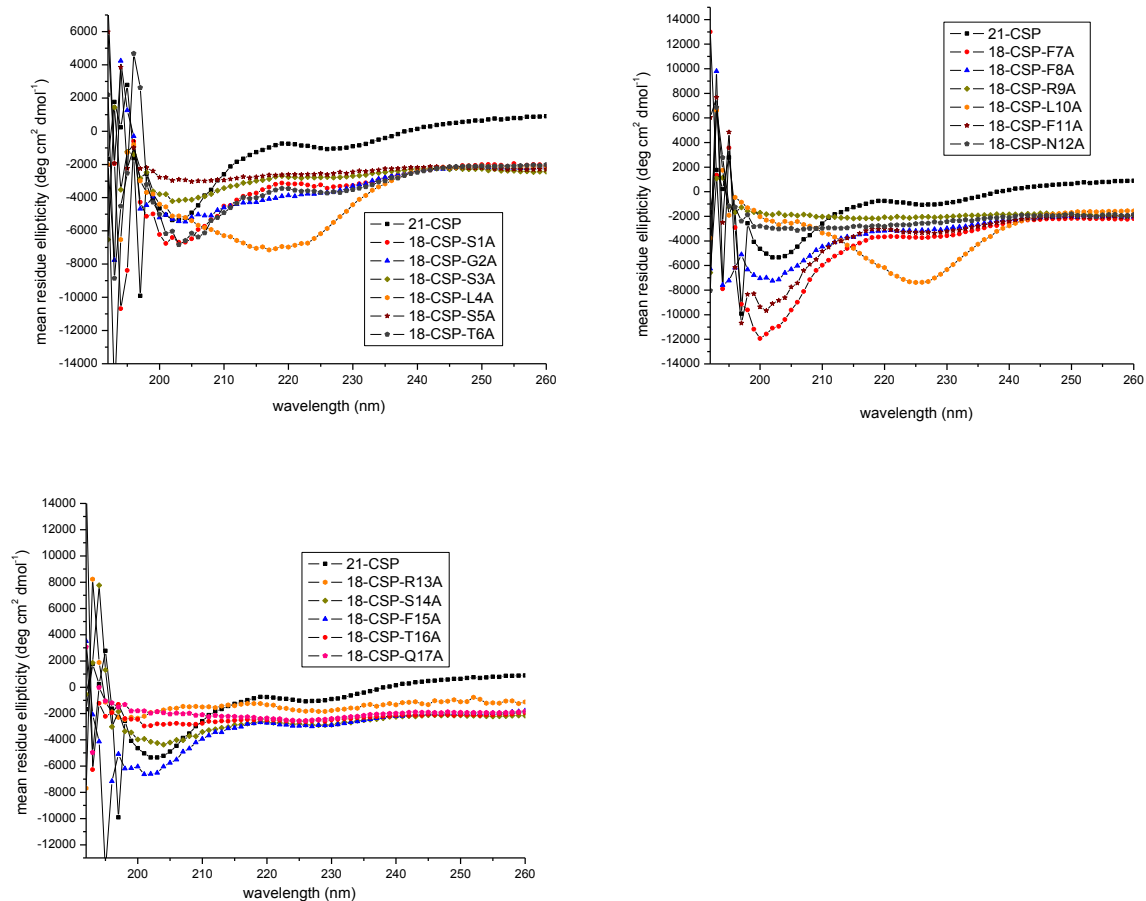


Figure S-20. CD spectra of the 18-CSP alanine scan library in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. Measurements of 18-CSP-R9A and 18-CSP-R13A were performed with a peptide concentration of 100 μ M as these peptides showed low solubility in PBS. 21-CSP was added as a control. Most peptides exhibit a random coil pattern, while few peptides exhibit a β -sheet pattern.

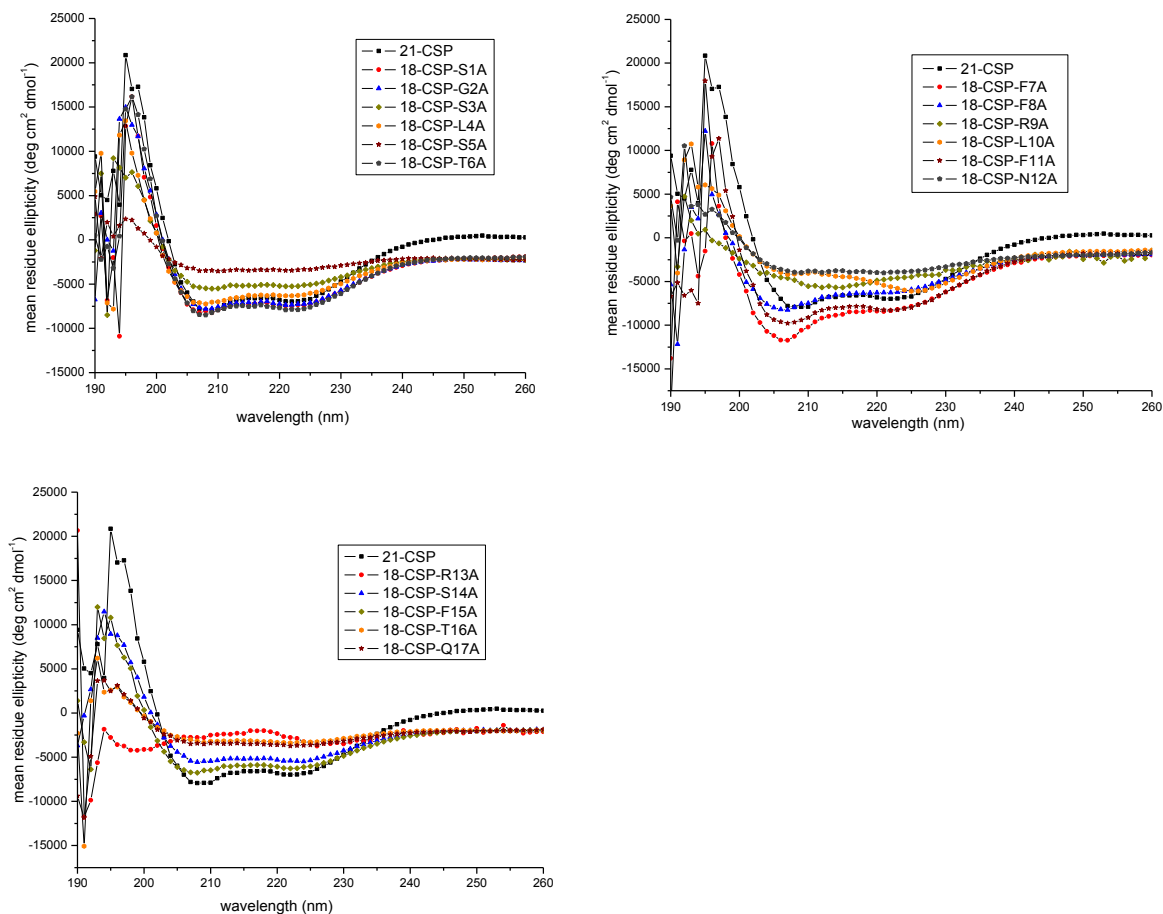


Figure S-21. CD spectra of the 18-CSP alanine scan library in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. Measurements of 18-CSP-R9A and 18-CSP-R13A were performed with a peptide concentration of 100 μ M as these peptides showed low solubility in PBS. 21-CSP was added as a control. Peptides exhibit varying degrees of an α -helix pattern, while 18-CSP-L10A exhibits a distorted α -helix (mixed with β -sheet) pattern, 18-CSP-R9A exhibits a distorted α -helix (mixed with β -sheets) pattern, and 18-CSP-R13A exhibits a distorted α -helix (mixed with β -turns) pattern.

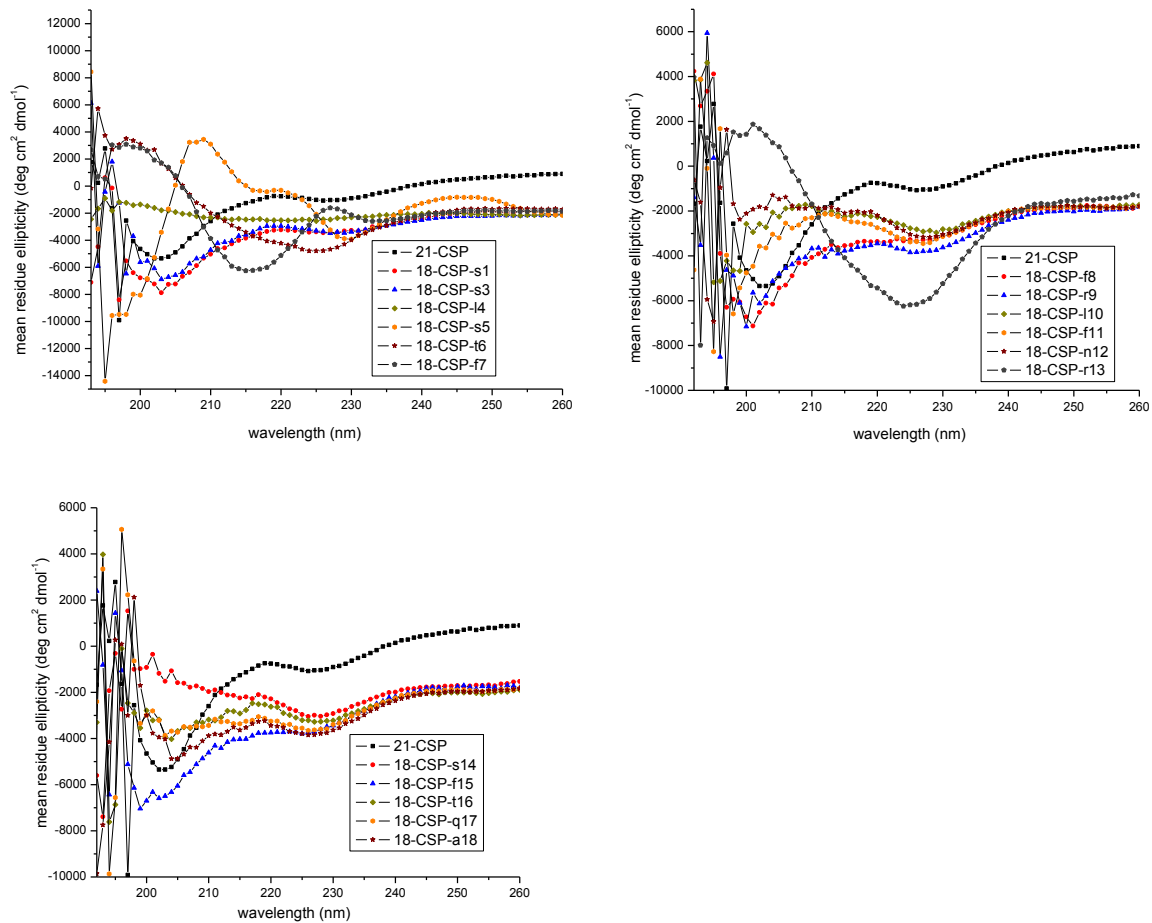


Figure S-22. CD spectra of the 18-CSP D-amino acid scan library in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μM . 21-CSP was added as a control. Most peptides exhibit a random coil pattern, while few peptides exhibit a β -sheet pattern.

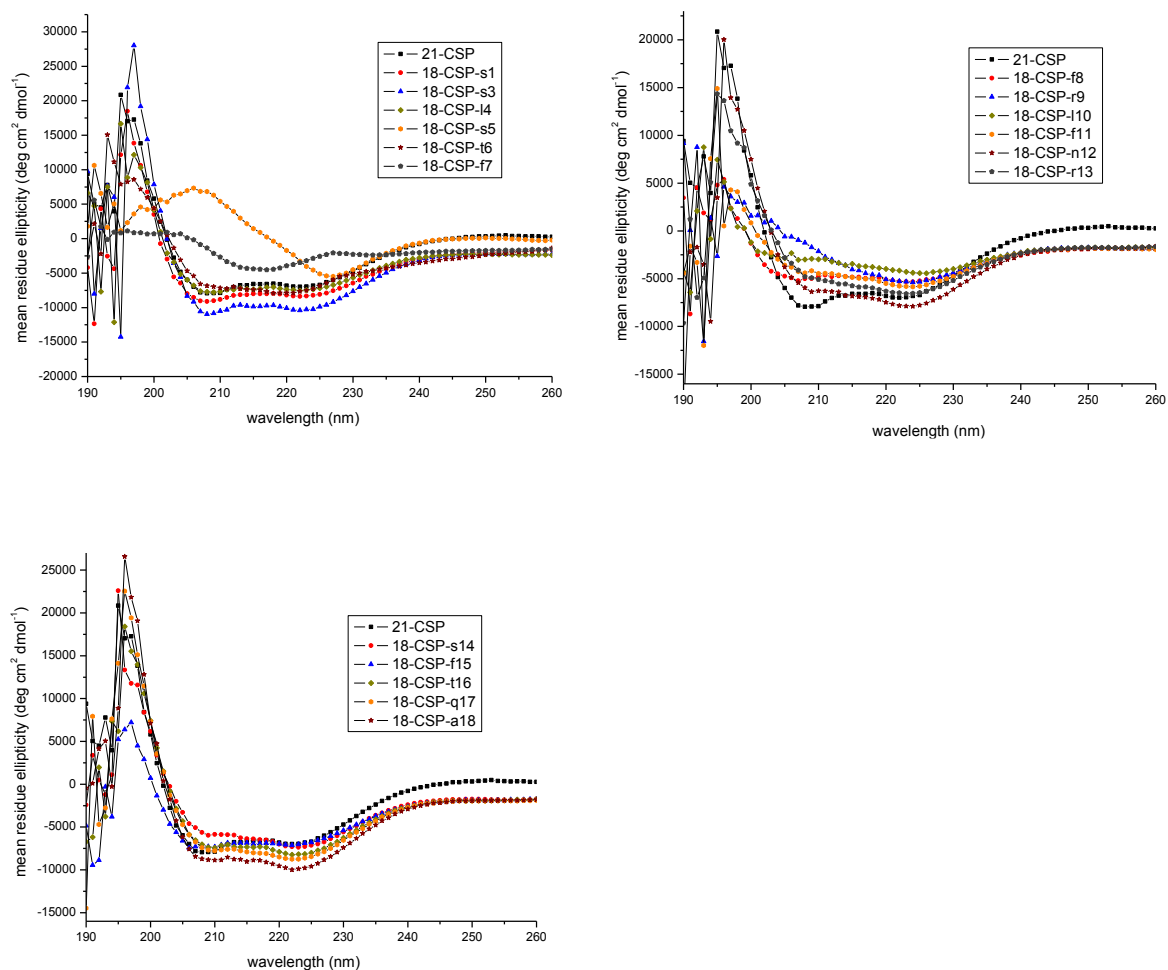


Figure S-23. CD spectra of the 18-CSP D-amino acid scan library in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most peptides exhibit varying degrees of an α -helix pattern, while 18-CSP-s5 exhibits a β -turn pattern, and 18-CSP-f7 and 18-CSP-r9 exhibit a β -sheet pattern.

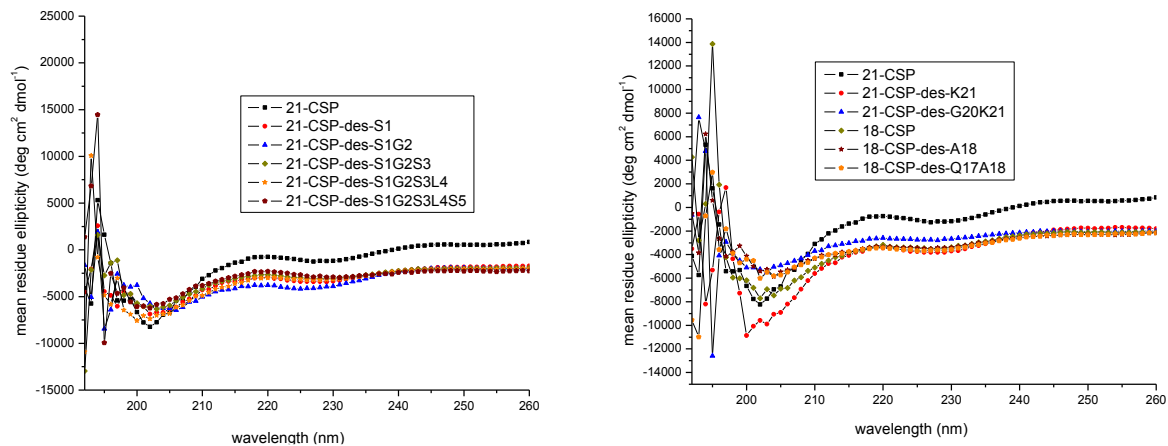


Figure S-24. CD spectra of the 21-CSP and 18-CSP truncated analogs in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μM . 21-CSP was added as a control. All peptides exhibit a random coil pattern.

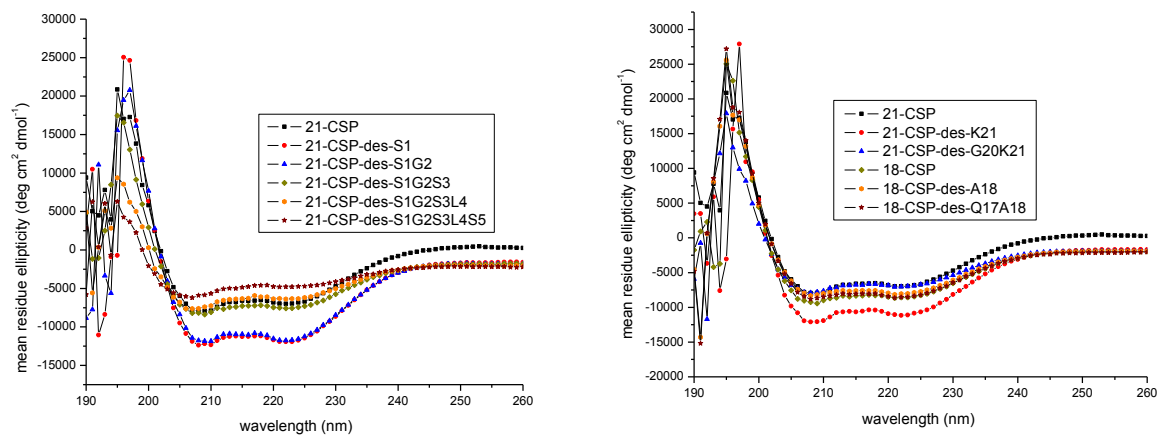


Figure S-25. CD spectra of the 21-CSP and 18-CSP truncated analogs in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μM . 21-CSP was added as a control. Peptides exhibit varying degrees of an α -helix pattern.

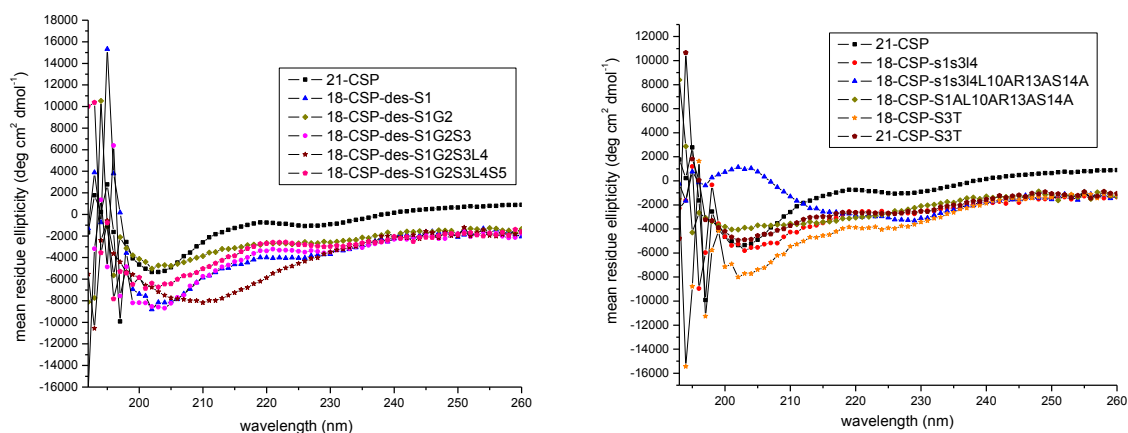


Figure S-26. CD spectra of 18-CSP truncated and 2nd generation CSP analogs in aqueous solution (PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most peptides exhibit a random coil pattern, while 18-CSP-S1G2S3L4 exhibits a β -sheet pattern and 18-CSP-s1s3l4L10AR13AS14A exhibits a distorted β -sheet pattern.

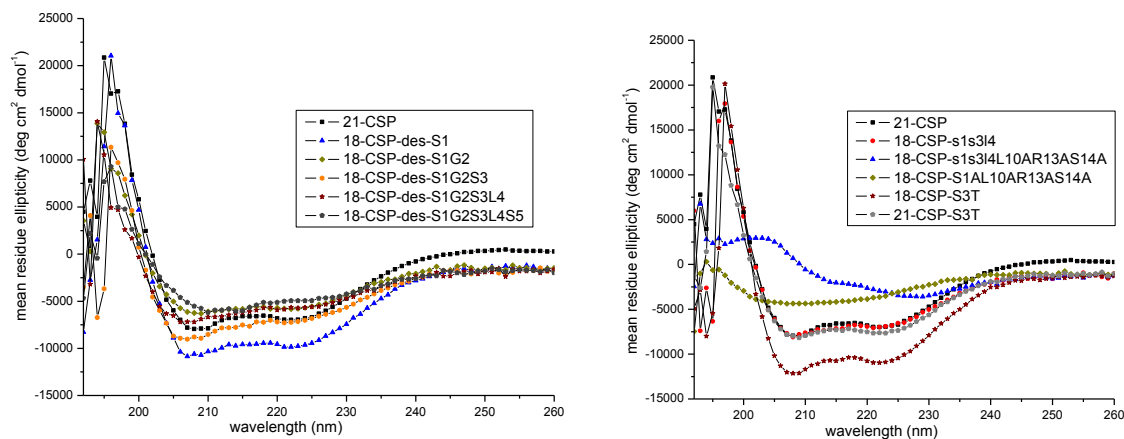


Figure S-27. CD spectra of 18-CSP truncated and 2nd generation CSP analogs in membrane mimicking conditions (20% TFE: 80% PBS, pH 7.4). All the measurements were performed with a peptide concentration of 200 μ M. 21-CSP was added as a control. Most peptides exhibit varying degrees of an α -helix pattern, while 18-CSP-s1s3l4L10AR13AS14A exhibits a distorted β -sheet (mixed with β -turn) pattern, and 18-CSP-S1AL10AR13AS14A exhibits a distorted α -helix (mixed with β -sheet) pattern.

Table S-8. EC₅₀ values (SAB249) and percent helicity (in 20% TFE) data for 21-CSP alanine & D-amino acid scanning analogs.

Compound Name	EC ₅₀ (nM)	Helicity (%)	Compound Name	EC ₅₀ (nM)	Helicity (%)
21-CSP	240	18.0%			
21-CSP-S1A	90	24.7%	21-CSP-s1	140	25.7%
21-CSP-G2A	140	27.2%	--		
21-CSP-S3A	150	36.3%	21-CSP-s3	160	25.0%
21-CSP-L4A	6500	28.2%	21-CSP-l4	390	19.0%
21-CSP-S5A	330	30.0%	21-CSP-s5	640	20.9%
21-CSP-T6A	280	25.4%	21-CSP-t6	1600	β -sheet
21-CSP-F7A	--	21.8%	21-CSP-f7	--	Distorted β -sheet
21-CSP-F8A	--	24.1%	21-CSP-f8	840	19.8%
21-CSP-R9A	1100	Distorted β -sheet	21-CSP-r9	1500	β -sheet
21-CSP-L10A	370	27.0%	21-CSP-l10	760	β -turn
21-CSP-F11A	--	28.5%	21-CSP-f11	--	15.4%
21-CSP-N12A	--	36.4%	21-CSP-n12	--	17.5%
21-CSP-R13A	480	Distorted α -helix	21-CSP-r13	2100	β -turn
21-CSP-S14A	160	56.7%	21-CSP-s14	--	16.6%
21-CSP-F15A	--	28.0%	21-CSP-f15	--	22.8%
21-CSP-T16A	1400	13.8%	21-CSP-t16	--	13.4%
21-CSP-Q17A	94	36.5%	21-CSP-q17	--	24.8%
--			21-CSP-a18	--	19.7%
21-CSP-L19A	190	31.7%	21-CSP-l19	--	26.0%
21-CSP-G20A	190	35.9%	--		
21-CSP-K21A	290	30.8%	21-CSP-k21	330	34.1%

Table S-9. EC₅₀ values (SAB249) and percent helicity (in 20% TFE) data for 18-CSP alanine & D-amino acid scanning analogs.

Compound Name	EC ₅₀ (nM)	Helicity (%)	Compound Name	EC ₅₀ (nM)	Helicity (%)
18-CSP	6.2	22.6%			
18-CSP-S1A	2.6	19.8%	18-CSP-s1	6.4	22.1%
18-CSP-G2A	9.7	19.4%	--		
18-CSP-S3A	13	13.9%	18-CSP-s3	9.4	27.5%
18-CSP-L4A	57	16.7%	18-CSP-l4	4.5	19.9%
18-CSP-S5A	11	9.13%	18-CSP-s5	17	β -turn
18-CSP-T6A	6.0	20.7%	18-CSP-t6	300	20.3%
18-CSP-F7A	220	22.0%	18-CSP-f7	>1,000	β -sheet
18-CSP-F8A	>1,000	16.6%	18-CSP-f8	42	13.4%
18-CSP-R9A	27	Distorted α -helix	18-CSP-r9	76	β -sheet
18-CSP-L10A	6.5	Distorted α -helix	18-CSP-l10	670	10.9%
18-CSP-F11A	410	21.9%	18-CSP-f11	--	15.2%
18-CSP-N12A	33	10.4%	18-CSP-n12	--	20.7%
18-CSP-R13A	2.7	Distorted α -helix	18-CSP-r13	480	17.1%
18-CSP-S14A	4.5	14.3%	18-CSP-s14	>1,000	19.3%
18-CSP-F15A	--	16.6%	18-CSP-f15	--	18.8%
18-CSP-T16A	64	8.90%	18-CSP-t16	>1,000	21.8%
18-CSP-Q17A	19	9.90%	18-CSP-q17	130	23.1%
--			18-CSP-a18	>1,000	26.4%

Table S-10. EC₅₀ values (SAB249) and percent helicity (in 20% TFE) data for 21-CSP and 18-CSP truncated & 2nd generation analogs.

Compound Name	EC ₅₀ (nM)	Helicity (%)
21-CSP-des-S1	23	31.0%
21-CSP-des-S1G2	120	30.8%
21-CSP-des-S1G2S3	530	20.0%
21-CSP-des-S1G2S3L4	3600	16.9%
21-CSP-des-S1G2S3L4S5	3300	12.9%
21-CSP-des-K21	98	28.9%
21-CSP-des-G20K21	3600	18.4%
18-CSP-des-S1	4.8	26.3%
18-CSP-des-S1G2	7.9	15.5%
18-CSP-des-S1G2S3	85	19.7%
18-CSP-des-S1G2S3L4	570	15.7%
18-CSP-des-S1G2S3L4S5	950	13.4%
18-CSP-des-A18	--	21.4%
18-CSP-des-Q17A18	--	23.0%
18-CSP-s1s3l4	17	18.4%
18-CSP-s1s3l4L10AR13AS14A	3.4	Distorted β -sheet
18-CSP-S1AL10AR13AS14A	1.7	Distorted α -helix
18-CSP-S3T	2.1	28.9%
21-CSP-S3T	94	19.7%

Table S-11. EC₅₀ values (SAB249) and estimated secondary structure contents (in 20% TFE) data for 21-CSP alanine scan analogs.

Compound Name	EC ₅₀ (nM)	Estimated secondary structure contents (%) ^a				
		α -helix	Antiparallel β -sheet	Parallel β -sheet	β -turn	Others
21-CSP	240	27.1	16.4	0.0	9.7	46.7
21-CSP-S1A	90	34.2	17.0	0.0	5.8	43.1
21-CSP-G2A	140	37.3	19.1	0.0	7.4	36.2
21-CSP-S3A	150	38.0	9.5	0.0	5.1	47.4
21-CSP-L4A	6500	32.9	5.3	0.0	8.5	53.4
21-CSP-S5A	330	34.3	5.5	0.0	8.1	52.1
21-CSP-T6A	280	30.0	5.9	0.9	10.3	52.9
21-CSP-F7A	--	22.0	10.1	3.1	14.8	50.1
21-CSP-F8A	--	24.9	11.5	1.9	12.0	49.8
21-CSP-R9A	1100	4.1	28.6	19.3	14.6	33.5
21-CSP-L10A	370	32.8	17.6	0.0	10.5	39.1
21-CSP-F11A	--	29.7	11.1	0.0	7.7	51.5
21-CSP-N12A	--	33.4	9.7	0.0	7.6	49.3
21-CSP-R13A	480	13.8	9.1	17.9	19.6	39.6
21-CSP-S14A	160	41.9	25.5	23.2	1.7	7.7
21-CSP-F15A	--	30.1	14.3	0.0	9.5	46.1
21-CSP-T16A	1400	10.2	12.0	15.3	15.2	47.3
21-CSP-Q17A	94	39.3	17.5	0.0	5.9	37.3
--						
21-CSP-L19A	190	38.9	16.8	0.0	7.5	36.8
21-CSP-G20A	190	37.8	12.5	0.0	6.6	43.0
21-CSP-K21A	290	36.1	13.1	0.0	9.6	41.2

^a Secondary structure contents were calculated using the BeStSel¹ (Beta Structure Selection) method.

Table S-12. EC₅₀ values (SAB249) and estimated secondary structure contents (in 20% TFE) data for 21-CSP D-amino acid scan analogs.

Compound Name	EC ₅₀ (nM)	Estimated secondary structure contents (%) ^a				
		α -helix	Antiparallel β -sheet	Parallel β -sheet	β -turn	Others
21-CSP	240	27.1	16.4	0.0	9.7	46.7
21-CSP-s1	140	26.6	9.7	0.7	10.9	52.1
--						
21-CSP-s3	160	30.8	27.0	0.0	9.1	33.2
21-CSP-l4	390	13.1	4.7	14.5	10.8	56.9
21-CSP-s5	640	25.7	8.7	3.9	10.0	51.6
21-CSP-t6	1600	11.0	3.8	16.9	17.2	51.1
21-CSP-f7	--	3.0	20.5	14.4	16.4	45.8
21-CSP-f8	840	23.2	9.3	5.6	14.4	47.7
21-CSP-r9	1500	2.4	1.4	18.1	15.2	62.8
21-CSP-l10	760	0.0	65.7	0.0	24.2	10.2
21-CSP-f11	--	15.3	9.8	10.7	15.2	49.0
21-CSP-n12	--	19.9	13.4	8.3	13.5	45.0
21-CSP-r13	2100	5.7	4.4	14.8	22.2	52.9
21-CSP-s14	--	15.1	7.4	13.5	14.8	49.2
21-CSP-f15	--	26.1	14.3	0.0	12.7	46.8
21-CSP-t16	--	18.4	13.3	7.8	12.7	47.8
21-CSP-q17	--	30.5	10.1	0.0	10.2	49.2
21-CSP-a18	--	25.8	7.1	4.1	10.9	52.1
21-CSP-l19	--	35.0	18.0	0.0	8.5	38.4
--						
21-CSP-k21	330	43.9	19.1	0.0	7.1	29.9

^a Secondary structure contents were calculated using the BeStSel¹ (Beta Structure Selection) method.

Table S-13. EC₅₀ values (SAB249) and estimated secondary structure contents (in 20% TFE) data for 18-CSP alanine scan analogs.

Compound Name	EC ₅₀ (nM)	Estimated secondary structure contents (%) ^a				
		α -helix	Antiparallel β -sheet	Parallel β -sheet	β -turn	Others
18-CSP	6.2	27.1	6.8	0.0	9.1	57.0
18-CSP-S1A	2.6	21.1	16.2	0.0	12.4	50.3
18-CSP-G2A	9.7	15.4	6.1	13.3	13.5	51.7
18-CSP-S3A	13	10.5	6.2	16.1	17.6	49.6
18-CSP-L4A	57	18.1	16.9	4.7	14.7	45.6
18-CSP-S5A	11	8.6	22.2	11.2	15.5	42.5
18-CSP-T6A	6.0	24.2	9.7	0.0	11.2	54.9
18-CSP-F7A	220	19.2	8.3	8.3	12.5	51.7
18-CSP-F8A	>1,000	10.9	4.0	14.6	14.3	56.2
18-CSP-R9A	27	10.2	4.3	13.3	18.0	54.1
18-CSP-L10A	6.5	7.0	9.6	16.0	18.5	48.9
18-CSP-F11A	410	20.7	10.7	0.0	11.1	57.5
18-CSP-N12A	33	6.3	20.1	11.6	16.1	45.8
18-CSP-R13A	2.7	3.3	53.1	0.0	16.4	27.2
18-CSP-S14A	4.5	9.5	2.3	14.8	16.2	57.1
18-CSP-F15A	--	10.9	5.9	15.3	16.7	51.2
18-CSP-T16A	64	11.7	14.0	12.0	15.9	46.5
18-CSP-Q17A	19	13.7	5.7	13.3	15.8	51.4

^a Secondary structure contents were calculated using the BeStSel¹ (Beta Structure Selection) method.

Table S-14. EC₅₀ values (SAB249) and estimated secondary structure contents (in 20% TFE) data for 18-CSP D-amino acid scan analogs.

Compound Name	EC ₅₀ (nM)	Estimated secondary structure contents (%) ^a				
		α -helix	Antiparallel β -sheet	Parallel β -sheet	β -turn	Others
18-CSP	6.2	27.1	6.8	0.0	9.1	57.0
18-CSP-s1	6.4	24.2	9.1	0.0	9.9	56.8
--						
18-CSP-s3	9.4	43.6	17.3	0.0	7.2	31.9
18-CSP-l4	4.5	21.5	21.5	0.0	13.3	43.7
18-CSP-s5	17	0.0	47.5	0.0	25.3	27.2
18-CSP-t6	300	8.1	0.0	22.2	16.5	53.2
18-CSP-f7	>1,000	1.3	28.2	14.7	13.6	42.2
18-CSP-f8	42	10.5	11.7	13.4	18.4	46.1
18-CSP-r9	76	4.2	51.7	1.7	14.1	28.3
18-CSP-l10	670	9.7	9.7	14.0	17.8	48.8
18-CSP-f11	--	13.3	7.0	12.5	17.1	50.1
18-CSP-n12	--	20.5	10.9	5.1	10.6	52.9
18-CSP-r13	480	19.7	7.7	11.4	12.7	48.5
18-CSP-s14	>1,000	23.4	8.4	8.9	12.5	46.8
18-CSP-f15	--	18.7	12.0	5.4	13.3	50.5
18-CSP-t16	>1,000	26.8	7.8	5.5	10.0	49.9
18-CSP-q17	130	27.5	6.9	6.1	9.6	50.0
18-CSP-a18	>1,000	33.4	8.7	0.0	8.1	49.8

^a Secondary structure contents were calculated using the BeStSel¹ (Beta Structure Selection) method.

Table S-15. EC₅₀ values (SAB249) and estimated secondary structure contents (in 20% TFE) data for 21-CSP and 18-CSP truncated & 2nd generation analogs.

Compound Name	EC ₅₀ (nM)	Estimated secondary structure contents (%) ^a				
		α -helix	Antiparallel β -sheet	Parallel β -sheet	β -turn	Others
21-CSP	240	27.1	16.4	0.0	9.7	46.7
21-CSP-des-S1	23	40.3	19.9	0.0	5.9	33.8
21-CSP-des-S1G2	120	32.6	9.8	0.0	7.6	50.1
21-CSP-des-S1G2S3	530	23.7	9.5	3.9	11.5	51.4
21-CSP-des-S1G2S3L4	3600	16.1	13.5	9.1	14.6	46.8
21-CSP-des-S1G2S3L4S5	3300	10.4	5.0	16.3	18.5	49.8
21-CSP-des-K21	98	42.0	16.6	0.0	8.4	33.1
21-CSP-des-G20K21	3600	16.7	5.2	11.3	11.8	55.0
18-CSP-des-S1	4.8	29.6	12.6	0.0	7.5	50.2
18-CSP-des-S1G2	7.9	15.1	5.6	11.3	15.3	52.6
18-CSP-des-S1G2S3	85	21.5	15.0	0.8	13.0	49.7
18-CSP-des-S1G2S3L4	570	9.3	2.5	14.5	17.9	55.8
18-CSP-des-S1G2S3L4S5	950	17.9	14.8	10.1	14.5	42.6
18-CSP-des-A18	--	14.2	7.8	14.7	13.7	49.6
18-CSP-des-Q17A18	--	23.7	4.8	11.8	9.7	50.0
18-CSP-s1s3l4	17	27.2	22.2	0.0	9.9	40.7
18-CSP-s1s3l4L10AR13AS14A	3.4	3.4	36.8	11.7	15.2	32.9
18-CSP-S1AL10AR13AS14A	1.7	13.2	12.5	10.3	15.6	48.4
18-CSP-S3T	2.1	38.8	25.7	0.0	7.0	28.6
21-CSP-S3T	94	30.1	10.2	0.0	11.4	48.3

^a Secondary structure contents were calculated using the BeStSel¹ (Beta Structure Selection) method.

References

- [1] Micsonai, A., Wien, F., Kernya, L., Lee, Y. H., Goto, Y., Réfrégiers, M., and Kardos, J. (2015) Accurate secondary structure prediction and fold recognition for circular dichroism spectroscopy, *Proc. Natl. Acad. Sci. U. S. A.* *112*, E3095-3103 DOI 10.1073/pnas.1500851112.