

**A novel β -glucuronidase from *Talaromyces pinophilus* Li-93 precisely
hydrolyzes glycyrrhizin into glycyrrhetic acid
3-O-mono- β -D-glucuronide**

Yinghua Xu^{#a}, Xudong Feng^{#a}, Jintong Jia^a, Xinyi Chen^a, Tian Jiang^a, Aamir Rasool^a,
Bo Lv^a, Liangti Qu^b, Chun Li^{*a}

^a Institute for Synthetic Biosystem/Department of Biochemical Engineering, School
of Chemistry and Chemical Engineering, Beijing Institute of Technology, Beijing
100081, P. R. China

^b Beijing Key Laboratory of Photoelectronic/Electrophotonic Conversion Materials,
Key Laboratory of Cluster Science, Ministry of Education, School of Chemistry and
Chemical Engineering, Beijing Institute of Technology, Beijing 100081, P. R. China.

[#]These authors contributed equally to this paper.

^{*}Corresponding author: Chun Li, Email: lichun@bit.edu.cn

TABLE S1 The specific activity and catalytic efficiency of the published GUSs on GL

| Enzyme | specific activity ($\mu\text{mol}^{-1} \text{min}^{-1} \text{mg}^{-1}$) | k_{cat}/K_m ($\text{mM}^{-1}\text{s}^{-1}$) | Reference |
|---------------------------|--|--|------------|
| TpGUS79A | 11.97 | 11.14 | This study |
| β -glucuronidase I | 0.32 | NA* | (1) |
| β -glucuronidase II | 0.19 | NA | (1) |
| PGUS-P | 9.79E-3 | 6.40E-4 | (2) |
| PGUS-E | 8.62E-3 | 3.86E-3 | (2) |
| AtGUS-E | NA | 2.24 | (3) |

* Not available

TABLE S2 The specific activity and catalytic efficiency of the published GUSs on baicalin

| Enzyme | Specific activity ($\mu\text{mol}^{-1}\text{min}^{-1}\text{mg}^{-1}$) | k_{cat}/K_m ($\text{mM}^{-1}\text{s}^{-1}$) | Reference |
|----------|--|--|------------|
| TpGUS79A | 0.56 | 0.29 | This study |
| LcGUS30 | 7.57 | 14.0 | (4) |
| SvGUS | 9.37 | NA* | (5) |

* Not available

References

1. Hye-Young P, Kim NY, Han MJ, Bae EA, Kim DH. 2005. Purification and characterization of two novel beta-D-glucuronidases converting glycyrrhizin to 18 beta-glycyrrhetic acid-3-O-beta-D-glucuronide from *Streptococcus* LJ-22. J Microbiol Biotechnol 15:792-799.
2. Zou S, Guo S, Kaleem I, Li C. 2013. Purification, characterization and comparison of *Penicillium purpurogenum* β -glucuronidases expressed in *Escherichia coli* and *Pichia pastoris*. J Chem Technol Biotechnol 88:1913-1919.
3. Liu Y, Huangfu J, Qi F, Kaleem I, E W, Li C. 2012. Effects of a non-conservative sequence on the properties of beta-glucuronidase from *Aspergillus terreus* Li-20. PLoS One 7:e30998.
4. Sakurama H, Kishino S, Uchibori Y, Yonejima Y, Ashida H, Kita K, Takahashi S, Ogawa J. 2014. beta-Glucuronidase from *Lactobacillus brevis* useful for baicalin hydrolysis belongs to glycoside hydrolase family 30. Appl Microbiol Biotechnol 98:4021-4032.
5. Zhang CZ, Zhang YF, Chen JP, Liang XM. 2005. Purification and characterization of baicalin-beta-D-glucuronidase hydrolyzing baicalin to baicalein from fresh roots of *Scutellaria viscidula* Bge. Process Biochem 40:1911-1915.

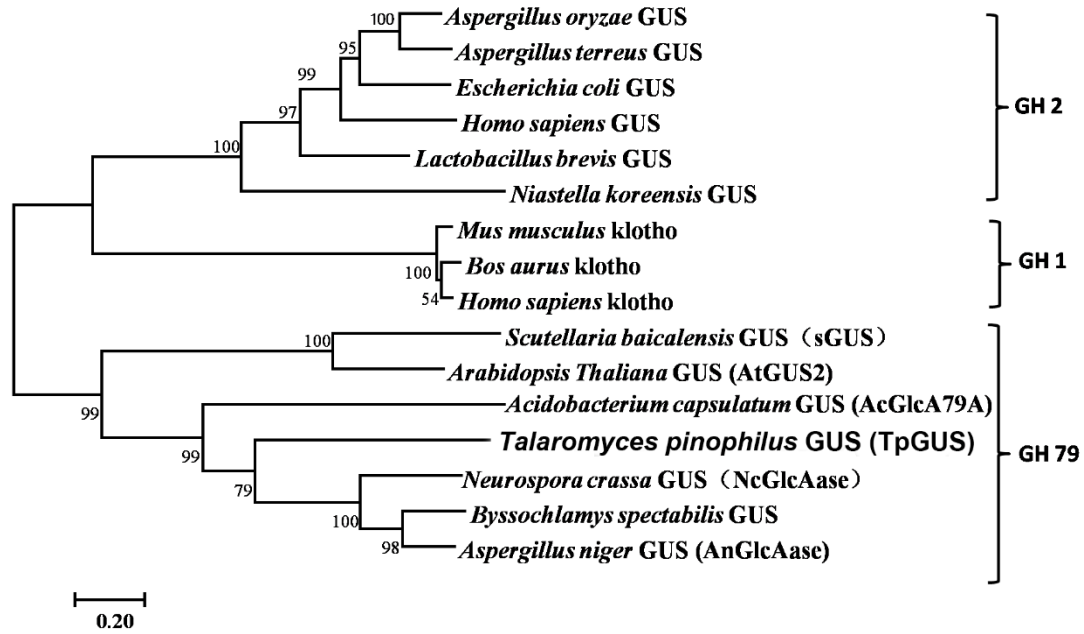


FIG S1 The phylogenetic analysis of TpG79A with other GUSs from GH1, GH2 and GH79 based on their amino acid sequences. The phylogenetic tree was constructed using MEGA 7 with a neighbor-joining method. On the phylogenetic tree, the GH79 GUSs include enzymes from *S. baicalensis* (10/29(34%) identity, AB040072), *A. Thaliana* (26/111(23%) identity, NP_196400), *A. capsulatum* (114/459 (25%) identity, PDB: 3VNY_A), *N. crassa* (144/432(33%) identity, XP_964763), *B. spectabilis* (160/462 (35%) GAD97470), and *A. niger* (128/377(34%) identity, GAQ47476); The GH2 GUSs include enzymes from *A. oryzae* (ABU68712), *A. terreus* (AEP39213), *E. coli* (PDB: 3K46_A), *Homo sapiens* (PDB: 3HN3_A), *L. brevis* (ACU21612) and *N. koreensis* (AEW00660); The GH1 GUSs include enzymes from *M. musculus* (BAA23381), *B. aurus* (NP_001178124), *Homo sapiens* (BAA23382).

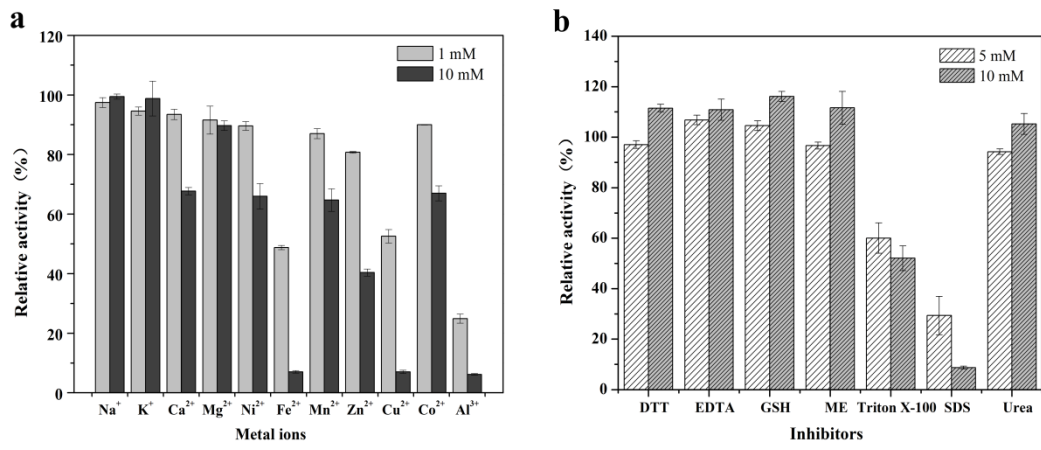


FIG S2 Effect of metal ions (a) and inhibitors (b) on TpGUS79A activity. The experiment was performed in triplicate measurements, and the errors stand for one standard deviation.

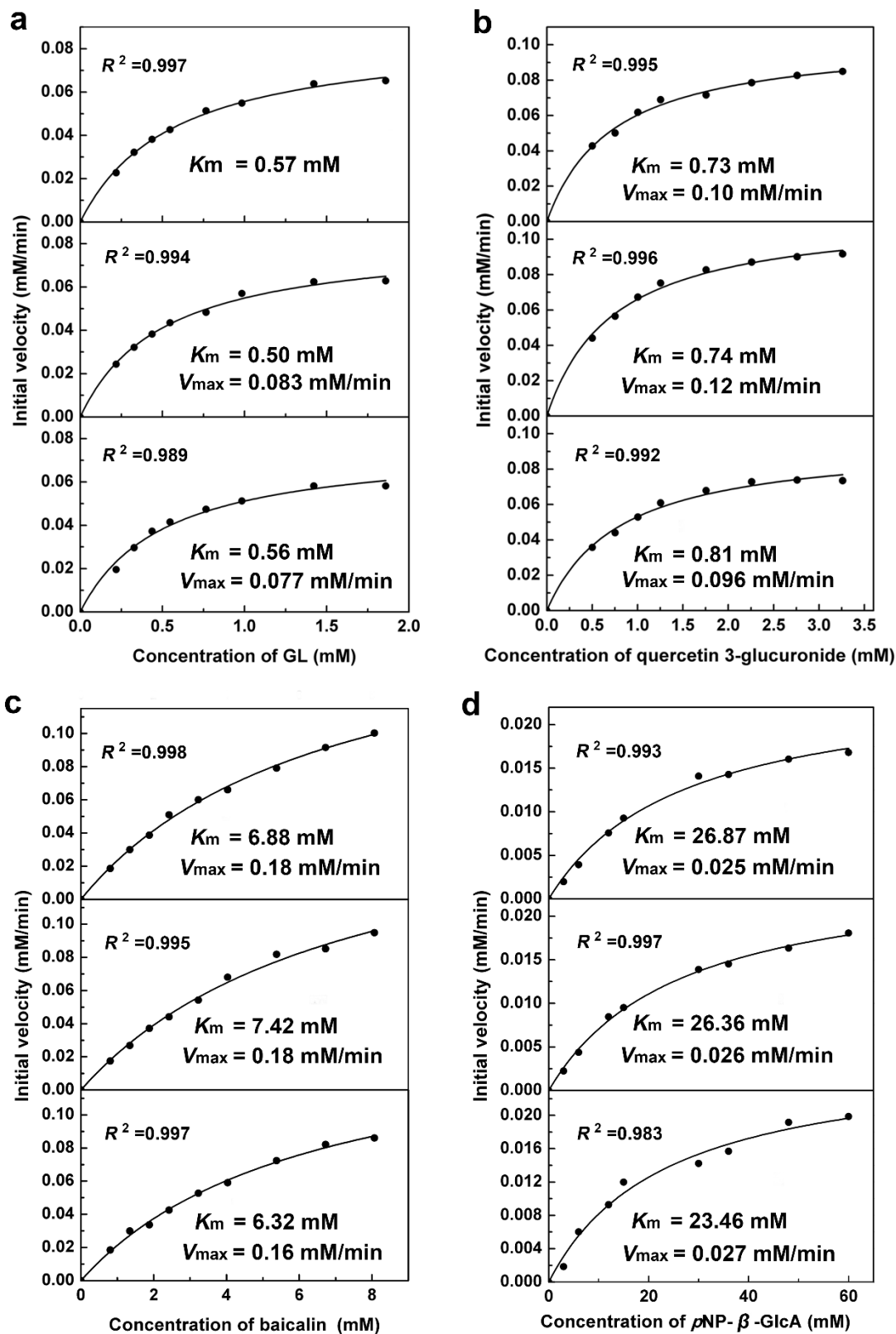


FIG S3 Michaelis-Menten plots of TpGUS79A hydrolyzing the different substrates. a: glycyrrhizin (GL), b: quercetin 3-glucuronide, c: baicalin, d: pNP- β -GlcA.

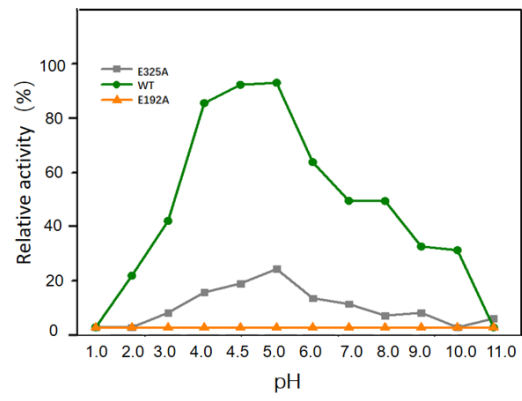


FIG S4 Activity rescue test of TpGUS79A mutants E192A and E325A.

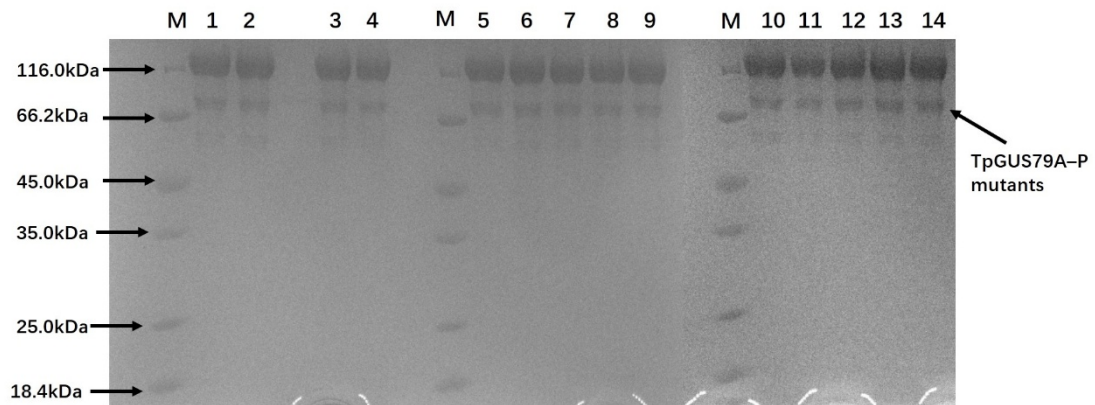


FIG S5 The SDS-PAGE of the purified TpGUS79A-P mutants. Lane M: marker, Lane 1: S330A, Lane 2: H365D, Lane 3: E192A, Lane 4: N191A, Lane 5: C331A, Lane 6: Y372K, Lane 7: C331D, Lane 8: E325A, Lane 9: Y372D, Lane 10: C331K, Lane 11: Y276A, Lane 12: S330R, Lane 13: S95A, Lane 14: Y372A.

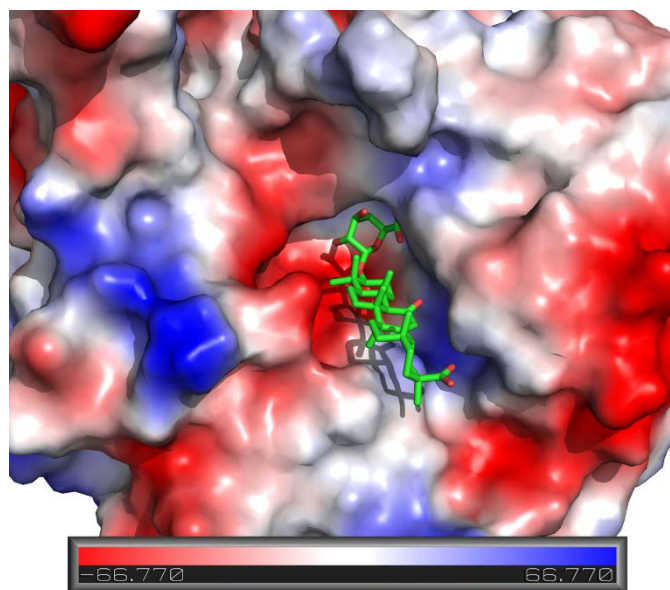


FIG S6 The electrostatic potential distribution of the active pocket of GH2 β -glucuronidase from *Aspergillus oryzae* Li-3. The green stick stands for the substrate glycyrrhetic acid 3-*O*-mono- β -D-glucuronide. The image was reproduced from PDB 5C71.

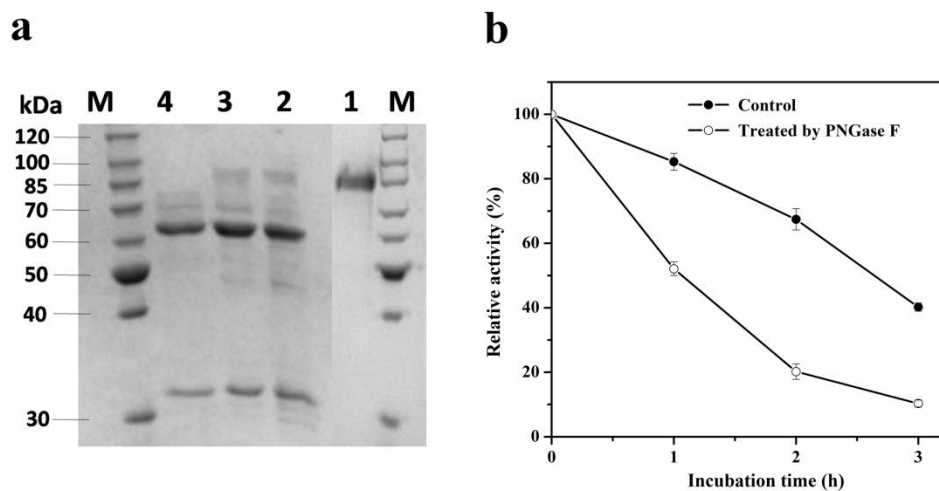


FIG S7 The effect of N-glycosylation on TpGUS79A activity (a) SDS-PAGE analysis of TpGUS79A treated with PNGase F in the non-denaturing reaction condition at different incubation time. Lane 1: 0 h, Lane 2: 1 h, Lane 3: 2 h, Lane 4: 3h. (b) The activity of TpGUS79A natively deglycosylated by PNGase F on GL. The activity for GL was assayed at 50 °C for 5 min in 50 mM NaAc-HAc buffer (pH 4.5). Data represent the mean \pm one standard deviation of results from the experiments in triplicate.