Supplementary material

Functional genetic elements for controlling gene expression in Cupriavidus necator H16

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The authors wish it to be known that the first three authors, Swathi Alagesan, Erik K. R. Hanko and Naglis Malys, should be regarded as joint First Authors

SUPPLEMENTARY MATERIAL AND METHODS

Construction of modular vector pMTL71101

Vector pMTL71101 was built on the basis of plasmid pMTL83141, which is formed of four modules flanked by 8 bp recognition sites of the type II restriction endonucleases AscI, FseI, PmeI and SbfI (1). Three modifications using plasmid pMTL83141 were carried out as following. First, the 892 bp PmeI-SbfI-flanked DNA fragment encoding the E. coli Gram negative replicon ColEI RNA II was replaced with a fragment containing broad-host-range origin of replication (pBBR1) and mobilisation gene mob. This 4045 bp DNA fragment was amplified by PCR using primers P015_71101_f and P016_71101_r from plasmid pME6000 (2), followed by restriction digestion with PmeI and SbfI. Second, the 2418 bp AscI-PmeIflanked DNA fragment including the Gram positive replicon pCB102 and chloramphenicol resistance marker (*catP*) was replaced with a 805 bp DNA fragment containing solely *catP*, which was amplified by PCR using primers P017_CatP_f and P018_CatP_r from pMTL83141, followed by restriction digestion with AscI and PmeI. Third, the NotI restriction site in the *mob* gene locus was removed by conservative change using the QuikChange Site-Directed Mutagenesis Kit (Agilent Technologies) and primers P019_Qmob_f and P020_Qmob_r by following manufacturer's instructions. The resulting plasmid pMTL71101 contains the pBBR1 origin of replication and mobilisation gene mob (PmeI-SbfI module), the chloramphenicol resistance marker *catP* (FseI-PmeI module) and a multiple cloning site (SbfI-AscI module) (Supplementary Figure S1).

Codon-optimised *P. alba ispS* gene encoding for isoprene synthase, 49 amino acids truncated at N-terminus

ATGGAAGCCCGCCGCTCGGCCAACTACGAGCCGAACTCGTGGGACTACGACTACCTGCTGTC GTCGGACACCGACGAGTCGATCGAGGTGTACAAGGACAAGGCCAAGAAGCTGGAAGCCGAGG TCCGCCGCGAGATCAACAACGAGAAGGCCGAGTTCCTGACGCTGCTGGAACTGATCGACAAC GTGCAGCGCCTGGGCCTGGGCTACCGCTTCGAAAGCGACATCCGCGGCGCCCTGGACCGCTT CGTGAGCAGCGGCGGCTTCGACGCCGTGACCAAGACCTCGCTGCATGGCACCGCGCTGTCGT TCCGCCTGCTGCGCCAGCACGGCTTCGAGGTGTCGCAGGAAGCCTTCTCGGGCTTCAAGGAC CAGAACGGCAACTTCCTGGAAAACCTGAAGGAAGATATCAAGGCCATCCTGTCGCTGTACGA GGCCAGCTTCCTGGCGCTGGAAGGCGAGAACATCCTGGACGAGGCCAAGGTGTTCGCCATCT CGCATCTGAAGGAACTGTCGGAGGAAAAGATCGGCAAGGAACTGGCCGAACAGGTGAACCAT CTACCGCAAGAAGGAAGACGCCAACCAGGTCCTGCTGGAACTGGCCATCCTGGACTACAACA TGATCCAGTCGGTGTACCAGCGCGACCTGCGCGAAACCAGCCGCTGGTGGCGCCGCGTCGGC CTGGCCACCAAGCTGCACTTCGCCCGCGACCGCCTGATCGAGTCGTTCTACTGGGCCGTGGG CGTCGCCTTCGAGCCGCAGTATTCGGACTGCCGCAACTCGGTGGCCAAGATGTTCAGCTTCG TGACCATCATCGACGACATCTACGACGTGTACGGCACCCTGGACGAACTGGAACTGTTCACC GACGCGGTGGAACGCTGGGACGTGAACGCCATCAACGACCTGCCGGACTATATGAAGCTGTG CTTCCTGGCGCTGTACAACACCATCAACGAGATCGCCTACGACAATCTGAAGGACAAGGGCG AAAATATCCTGCCGTACCTGACCAAGGCCTGGGCCGACCTGTGCAACGCCTTCCTGCAGGAA GCGAAGTGGCTGTATAACAAGTCGACCCCGACCTTCGACGACTACTTCGGCAACGCGTGGAA GTCGTCGTCGGGCCCGCTGCAGCTGGTGTTCGCCGTACTTCGCCGTGGTGCAGAACATCAAGA AGGAAGAGATCGAGAACCTGCAGAAGTACCACGACACCATCTCGCGCCCGTCGCACATCTTC CGCCTGTGCAATGACCTGGCCTCGGCCGGGGAAATCGCCCGCGGCGAAACCGCCAACAG CGTGTCGTGCTACATGCGCACCAAGGGCATCTCGGAAGAACTGGCGACCGAGTCGGTGATGA ACCTGATCGACGAAAACCTGGAAGAAGATGAACAAGGAAAAGCTGGGCGGCTCGCTGTTCGCC AAGCCGTTCGTGGAAACCGCGATCAATCTGGCCCGCCAGTCGCACTGCACCTACCACAACGG CGACGCGCACACCTCGCCGGATGAGCTGACCCGCAAGCGCGTGCTGAGCGTGATCACCGAGC CGATCCTGCCGTTCGAGCGCTGA

SUPPLEMENTARY TABLES

Supplementary Table S1. Oligonucleotide primers used in this study.

Primer	Sequence 5' to 3'
P001	${\sf atatggtaccaaattaagcagaaggccatcctgacggatggcctttttgcgtttcttaaggctgaggaaagtctagacgattcattaatgcagctggc$
P002	aattaccggtaaacaacagataaaacgaaaggcccgaaggcctttcgttttatttgatgctagctgaggagacatctagaaaggaagg
P003	gggaaaguacccaaaaattcatccttctcg
P004	ggagacauggatccaagcttacttgtacagctcgtccatgc
P005_p0_1_r	attgtctcuctgccgtcactattcg
P005_p0_1-sd_r	attgagagugagccgtcactattcgaaccg
P005_p0_1+SD_r	agtgaattcugacatttgagtccattgttgcc
P005_p0_2_r	attgtctcuctgccgacatttgagtccattgttgcc
P005_p1_r	attataattgutatccgctcacaaagcaaataaattttttatcgattaatccgcctcggcactgc
P005_p2_r	aaagtcaguctagggtagtcactatcatcgtgcaagttcgtttttaatccgcctcggcactgc
P005_p3_r	attcattauacgagccgatgattaattgtcaacagctcatcgattaatccgcctcggcactgc
P005_p4_r	agtatattcuaaatttccacctgtgtcaataacggttttatcgattaatccgcctcggcactgc
P005_p5_r	acgagaauttgaagcgtttagcaaatgaattttttaatcgattaatccgcctcggcactgc
P005_p6_r	attctattauacagaaaaattttcctgaaagcaaataaattttttatcgattaatccgcctcggcactgc
P005_p7_r	${\sf aggtctataauaccacaatagtatcagtatgtaaacaactttctgaaattttttatcgattaatccgcctcggcactgc$
P005_p8_r	${\sf atgaatctautatatcgccgcaagggataaaaagcaaataaattttttatcgattaatccgcctcggcactgc$
P005_p9_r	${\sf atgaatctautataggtacaaaaagatgcgaagtcaatactctttttatcgattaatccgcctcggcactgc$
P005_p10_r	atcttcaauattgagtggcacggttaagctccaaaatgacgatcgattaatccgcctcggcactgc
P005_p11_r	${\sf acgttaa} {\sf auctaggtaca} {\sf aaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaaa$
P005_p12_r	agcacaauacctaggactgagctagccgtcaaatcgattaatccgcctcggcactgc
P005_p13_r	acacattauacgagccggatgattaaaagggaaatcgattaatccgcctcggcactgc
P005_p14_r	attcgtcccuccacacatcctaggagccggatgattaattgtcaaatcgattaatccgcctcggcactgc
P005_p15_r	attcaggtugctagcacagtacctaggactgagctagctgtcaaatcgattaatccgcctcggcactgc
P005_p16_r	attaagcuagcactgtacctaggactgagctagccgtcaaatcgattaatccgcctcggcactgc

P005_p17_r	atgaaataautatgggtacaaaaagatgcgatgtcaaatcgattaatccgcctcggcactgc
P005_p18_r	atgaaataautatgcctataggttagactttatgtcaaatcgattaatccgcctcggcactgc
P005_p19_r	acgttaaauctatcgccgcaagggataaatgtcaaatcgattaatccgcctcggcactgc
P005_p20_r	${\sf atg}$ aaataautatgggtacaaaagatgcgaaagcaaataaattttttatcgattaatccgcctcggcactgc
P005_p21_r	actccacauactctacgagccggatgattaattgtcaaatcgattaatccgcctcggcactgc
P005_p22_r	attcttcgugctagcattatacctaggactgagctagctgtcaaatcgattaatccgcctcggcactgc
P005_p23_r	${\sf actat}{\sf gaauctattat}{\sf acctat}{\sf aggttag}{\sf actttat}{\sf gtcaaatcgattaatccgcctcggcactgc}$
P005_p24_r	${\tt atccacacautatacagaaaaattttcctgaagtcaatactctttttatcgattaatccgcctcggcactgc$
P005_p25_r	${\tt atctcacaautccacacattatacgagccggatgattaattgtcaaatcgattaatccgcctcggcactgc$
P006_p0_f	gggaaagugacggcagagagacaatc
P006_p0_1_f	agagacaaucaacatatggtgagc
P006_p0_1-sd_f	actctcaaucaacatatggtgagcaagg
P006_p0_1+SD_f	agaattcacuagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p0_2_f	agagacaaucaacatatggtgagc
P006_p1_f	${\sf a}$ caattataauagaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p2_f	actgacttucaaatcgatagcaagcaggtctctcgaggaagccaatcatggtgagcaagggcgag
P006_p3_f	ataatgaautcactagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p4_f	${\sf a}$ gaatatacutgaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p5_f	${\tt attctcguataatagaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p6_f	${\sf a}$ taatagaautcactagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p7_f	${\tt attatagaccuat} {\tt gaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p8_f	${\tt atagattcaucttagaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p9_f	${\tt atagattcautgctagaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p10_f	${\sf att}{\sf g}{\sf a}{\sf g}{\sf a}{\sf u}{\sf a}{\sf g}{\sf a}{\sf a}{\sf t}{\sf c}{\sf a}{\sf c}{\sf t}{\sf a}{\sf g}{\sf a}{\sf g}{\sf g}{\sf a}{\sf g}{\sf a}{\sf d}{\sf a}{\sf t}{\sf a}{\sf c}{\sf a}{\sf t}{\sf a}{\sf g}{\sf d}{\sf g}{\sf d}{\sf g}{\sf d}{\sf g}{\sf d}{\sf g}{\sf g}{\sf g}{\sf g}{\sf c}{\sf a}{\sf g}{\sf g}{\sf g}{\sf g}{\sf c}{\sf d}{\sf d}{\sf g}{\sf d}{\sf d}{\sf g}{\sf d}{\sf g}{\sf d}{\sf d}{\sf g}{\sf d}{\sf d}{\sf g}{\sf d}{\sf d}{\sf d}{\sf d}{\sf d}{\sf d}{\sf d}{\sf d$
P006_p11_f	${\tt atttaacguatcccgagaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p12_f	${\tt attgtgcuagccgtcggaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p13_f	${\tt ataatgtguggagacttgaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
P006_p14_f	${\sf agggacgaautcactagtttaactttaagaaggagatatacatatggtgagcaagggc}$

P006_p15_f	aacctgaautcactagtttaactttaagaaggagatatacatatggtgagcaagggc
P006_p16_f	${\sf agcttaaugaattcactagtttaactttaagaaggagatatacatatggtgagcaagggc$
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P007_(A/U) ₁₂ /rbs2_r	aagtagtccuccttaaaagatcttttgccgtcactattcgaaccg
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P008_rbs7_f	aggataagguatggtgagcaagggcgag
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P008_rbs13_f	aaggaggtucatatggtgagcaagggcgag
P008_rbs14_f	aaggaggtuaacatatggtgagcaagggcgag
P008_rbs20_f	agagagacaaucaacatatggtgagcaagggcgag
P008_rbs21_f	aaaaggaauggtgagcaagggcgag
P008_rbs22_f	aaatagaauggtgagcaagggcgag
P008_rbs23_f	aggagaccauatggtgagcaagggcgag
P008_rbs24_f	aggaggaaguatggtgagcaagggcgag
P008_rbs25_f	aggaggtgguatggtgagcaagggcgag
P008_rbs26_f	aggaggacguatggtgagcaagggcgag
P008_rbs27_f	aaggagauttagatggtgagcaagggcgag
P008_rbs28_f	aggagatauagatggtgagcaagggcgag
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P008_SL/rbs8_f	agaaataautttgaaaggaggacaaggatggtgagcaagggcgag
P009_ispS_f	gacggcgaaggagatatacatatgg
P010_ispS_r	tatacttaagtctagattcagcgctcgaacgg
P011_16S_f	agagtttgatcctggctcag
P012_16S_r	ttaccgcggctgctggcac
P013_yfp_f	agaagaacggcatcaaggtg
P014_yfp_r	gaactccagcaggaccatgt

P015_71101_f	tatagtttaaacgtgctacgcctgaataagtg
P016_71101_r	tatacctgcaggctgaggtctgcctcgtgaag
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EH007_f	atttctagagggaaaccgttgtggtctccctacgaccagtctaaaaagcgcct
EH008_r	atcgacgtcttaatctttctgcgaattgagatgacgc
EH011_f	aatccaagcgtttaaacggaggcagacaaggtatagggc
EH012_r	tctgcctccgtttaaacgcttggattctcaccaataaaaaacgc
EH016_r	cgaaaggctcagtcgaaagactgggcctttcgttttatgacgtcttatgacaacttgacggctacatcattc
EH025_f	tcagaaggccatcctgacggatggccttttggcgcgccaggccggcc
EH026_r	catacgagccggaagcataaagtgtaaagccgatcgttgttgacactctatcattgatagagttattttaccacgggagaccacaacggtttcc
EH027_f	${\sf tacactttatgcttccggctcgtatgttgtgtggaattgtgagcggataacaatttcacacaggaaacagctatgacaaagttgcagccgaat$
EH028_r	ggccatccgtcaggatggccttctgacgtctcaatcgtcaccctttctcgg
EH048_f	gacgtcagaaggccatcct
EH055_r	atccgtcaggatggccttctgacgtcttattcctgtgtccgggtcacg
EH056_f	${\sf tacactttatgcttccggctcgtatgttgtgtggaattgtgagcggataacaatttcacacaggaaacagctatgcctctgacagaca$
EH057_r	tacgagccggaagcataaagtgtaaagccgatcggcttcacaaccgcacttgatttaatagaccataccgtctattatttctgggggagaccacaacggtttcc
EH109_r	tttaagaaggagatatacatatggcgagtagc
EH111_r	atccgtcaggatggccttctgacgtcctagcgcttgaatttcgcgtac
EH112_f	${\sf ccggctcgtatgttgtgtggaattgtgagcggataacaatttcacacaggaaacagctatggtgatcatgagtccaaagagaa$
EH113_r	${\sf a}$ ca attcc a ca catacg agc cgg a agc at a a agt gt a a agc cg at cg a a at cata a a a a att tatt t
EH114_f	atgtatatctccttcttaaagttaaacaaaattatttctagtaac
EH310_f	gctactcgccatatgtatatctccttcttaaaagatcttttgaattcccaaaaaaacgggtatggagaaac

Supplementary Table S2. Plasmids used and generated in this study.

Plasmid	Characteristic	Reference or source
pMTL83141	Cm ^r , ColE1 RNA II, CatP, <i>lacZ, pCB102</i>	(1)
pME6000 pBBR1MCS-2-P <i>phaC-eyfp</i> -	Tet ^r , pBBR1 ori, <i>lacZ, mob</i> ⁺	(2)
c1	Kan ^r ; broad host range vector used as a backbone and to amplify the eyfp gene	(3)
pGEM-T	Amp ^r ; <i>E. coli</i> vector used to amplify the <i>lacZ</i> gene for blue/white colony screening	Promega
pEH006	Cm ^r ; ParaC-araC-TrrnB1 and ParaBAD-T7sI-EcRBS-rfp-Tdbl	(4)
pJOE7784.1	Kan ^r ; vector used to amplify P _{rhask} -rhaSR and P _{rhaBAD}	(5)
pJOE7801.1	Kan ^r ; vector used to amplify <i>tetR</i>	(5)
pMTL71101	Cm ^r ; broad host range modular vector used as a backbone	This study
pBBR1MCS-2-USER	Kan ^r ; broad host range vector with USER cassette	This study
pMTL71107	Cm ^r ; broad host range modular vector with USER cassette	This study
pBBR1MCS-2-P ₀	Kan ^r ; P ₀ - <i>eyfp</i> , assembled from pBBR1MCS-2-USER1 and PCR fragments generated by PCR using primer pair P006_P0_f-P004 from pBBR1MCS-2-PphaC-eyfp-c1	This study
pBBR1MCS-2-P _{0_1}	Kan ^r ; P _{0_1} -eyfp, assembled from pBBR1MCS-2-USER1 and PCR fragments generated by PCR using primer pairs P003-P005_P0_1_r and P006_P0_1_f-P004 from pBBR1MCS-2-PphaC-eyfp- c1	This study
pBBR1MCS-2-P _{0_1-sd}	Kan ^r ; P _{0_1-sd} -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P _{0_1+SD}	Kan ^r ; P _{0_1+SD} -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-P _{0_2}	Kan ^r ; P _{0_2} - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁	Kan ^r ; P ₁ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₂	Kan ^r ; P ₂ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P₃	Kan ^r ; P ₃ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₄	Kan ^r ; P ₄ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P ₅	Kan ^r ; P ₅ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₆	Kan ^r ; P ₆ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P7	Kan ^r ; P ₇ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P ₈	Kan ^r ; P ₈ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P9	Kan ^r ; P ₉ - <i>eyfp,</i> assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P ₁₀	Kan ^r ; P ₁₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₁	Kan ^r ; P ₁₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-P ₁₂	Kan ^r ; P ₁₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₃	Kan ^r ; P ₁₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₄	Kan ^r ; P ₁₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₅	Kan ^r ; P ₁₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₆	Kan ^r ; P ₁₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₇	Kan ^r ; P ₁₇ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₈	Kan ^r ; P ₁₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₁₉	Kan ^r ; P ₁₉ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₂₀	Kan ^r ; P ₂₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> Methods	This study
pBBR1MCS-2-P ₂₁	Kan ^r ; P ₂₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₂₂	Kan ^r ; P ₂₂ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pBBR1MCS-2-P ₂₃	Kan ^r ; P ₂₃ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-P ₂₄	Kan ^r ; P ₂₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-P ₂₅	Kan ^r ; P ₂₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₀	Cm ^r ; P ₀ - <i>eyfp</i> , assembled from pMTL71107 and PCR fragments generated by PCR using primer pair P006_P0_f-P004 from pBBR1MCS-2-PphaC-eyfp-c1	This study
pMTL71101-P _{0_1}	Cm ^r ; P _{0_1} -eyfp, assembled from pMTL71107 and PCR fragments generated by PCR using primer pairs P003-P005_P0_1_r and P006_P0_1_f-P004 from pBBR1MCS-2-PphaC-eyfp-c1	This study
pMTL71101-P _{0_1-sd}	Cm ^r ; P _{0_1-sd} -eyfp, assembled as above using corresponding primers as described in <i>Material</i> and Methods	This study
pMTL71101-P _{0_1+SD}	Cm ^r ; P _{0_1+SD} -eyfp, assembled as above using corresponding primers as described in <i>Material</i> and Methods	This study
pMTL71101-P _{0_2}	Cm ^r ; P _{0_2} -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P₁	Cm ^r ; P ₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> Methods	This study
pMTL71101-P2	Cm ^r ; P ₂ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P₃	Cm ^r ; P ₃ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pMTL71101-P4	Cm ^r ; P ₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pMTL71101-P ₅	Cm ^r ; P ₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pMTL71101-P ₆	Cm ^r ; P ₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P7	Cm ^r ; P ₇ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pMTL71101-P ₈	Cm ^r ; P ₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pMTL71101-P9	Cm ^r ; P ₉ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> <i>Methods</i>	This study
pMTL71101-P ₁₀	Cm ^r ; P ₁₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₁	Cm ^r ; P ₁₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₂	Cm ^r ; P ₁₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₃	Cm ^r ; P ₁₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₄	Cm ^r ; P ₁₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₅	Cm ^r ; P ₁₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₆	Cm ^r ; P ₁₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₁₇	Cm ^r ; P ₁₇ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pMTL71101-P ₁₈	Cm ^r ; P ₁₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and</i> Methods	This study
pMTL71101-P ₁₉	Cm ^r ; P ₁₉ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₀	Cm ^r ; P ₂₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₁	Cm ^r ; P ₂₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₂	Cm ^r ; P ₂₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₃	Cm ^r ; P ₂₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₄	Cm ^r ; P ₂₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-P ₂₅	Cm ^r ; P ₂₅ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₀	Kan ^r ; P _{phaC} -RBS ₀ - <i>eyfp</i> , assembled from pBBR1MCS-2-USER1 and PCR fragments generated by PCR using primer pairs P003-P007_rbs0_r and P008_rbs0_f-P004 from pBBR1MCS-2-PphaC-eyfp-c1	This study
pBBR1MCS-2-RBS1	Kan ^r ; P _{phaC} -RBS ₁ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₂	Kan ^r ; P _{phaC} -RBS ₂ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study

pBBR1MCS-2-RBS₃	Kan ^r ; P _{phaC} -RBS ₃ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS₄	Kan ^r ; P _{phaC} -RBS ₄ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₅	Kan ^r ; P _{phaC} -RBS ₅ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₆	Kan ^r ; P _{phaC} -RBS ₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS7	Kan ^r ; P _{phaC} -RBS <i>7-eyfp</i> , assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₈	Kan ^r ; P _{phaC} -RBS ₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS9	Kan ^r ; P _{phaC} -RBS ₉ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS10	Kan ^r ; P _{phaC} -RBS ₁₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS11	Kan ^r ; P _{phaC} -RBS ₁₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₁₂	Kan ^r ; P _{phaC} -RBS1 ₂ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₁₃	Kan ^r ; P _{phaC} -RBS ₁₃ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₁₄	Kan ^r ; P _{phaC} -RBS ₁₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-RBS ₂₀	Kan ^r ; P _{phaC} -RBS ₂₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₁	Kan ^r ; P _{phaC} -RBS ₂₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₂	Kan ^r ; P _{phaC} -RBS ₂₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₃	Kan ^r ; P _{phaC} -RBS ₂₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₄	Kan ^r ; P _{phaC} -RBS ₂₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₅	Kan ^r ; P _{phaC} -RBS ₂₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₆	Kan ^r ; P _{phaC} -RBS ₂₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS27	Kan ^r ; P _{phaC} -RBS ₂₇ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₂₈	Kan ^r ; P _{phaC} -RBS ₂₈ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS29	Kan ^r ; P _{phaC} -RBS ₂₉ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₃₀	Kan ^r ; P _{phaC} -RBS ₃₀ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-RBS ₃₁	Kan ^r ; P _{phaC} -RBS ₃₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-RBS ₃₂	Kan ^r ; P _{phaC} -RBS ₃₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-SL/RBS ₁	Kan ^r ; P _{phaC} -T7g10 mRNA stemloop-RBS ₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-SL/RBS ₂	Kan ^r ; P _{phaC} -T7g10 mRNA stemloop-RBS ₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-SL/RBS8	Kan ^r ; P _{phaC} -T7g10 mRNA stemloop-RBS ₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-(A/U)₄/RBS1	Kan ^r ; P _{phaC} -(A/U) ₄ -RBS ₁ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-(A/U)₀/RBS1	Kan ^r ; P _{phaC} -(A/U) ₆ -RBS ₁ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-(A/U)9/RBS1	Kan ^r ; P _{phaC} -(A/U) ₉ -RBS ₁ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pBBR1MCS-2-(A/U) ₁₂ /RBS ₁	Kan ^r ; P _{phaC} -(A/U) ₁₂ -RBS ₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-(A/U) ₁₂ /RBS ₂	Kan ^r ; P _{phaC} -(A/U) ₁₂ -RBS ₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-(A/U) ₁₂ /RBS ₈	Kan ^r ; P _{phaC} -(A/U) ₁₂ -RBS ₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS₁-ispS	Kan ^r ; P _{phaC} -RBS ₁ - <i>ispS</i> , constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₁ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂ -ispS	Kan ^r ; P _{phaC} -RBS ₂ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₂ as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-RBS₅-ispS	Kan ^r ; P _{phaC} -RBS ₅ - <i>ispS</i> , constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₅ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₆ -ispS	Kan ^r ; P _{phaC} -RBS ₆ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₆ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₈ -ispS	Kan ^r ; P _{phaC} -RBS ₈ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₈ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS9-ispS	Kan ^r ; P _{phaC} -RBS ₉ - <i>ispS</i> , constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₉ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₁₃ -ispS	Kan ^r ; P _{phaC} -RBS ₁₃ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₁₃ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS14-ispS	Kan ^r ; P _{phaC} -RBS ₁₄ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₁₄ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₀ -ispS	Kan ^r ; P _{phaC} -RBS ₂₀ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₂₀ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₄ -ispS	Kan ^r ; P _{phaC} -RBS ₂₄ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₂₄ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₅ -ispS	Kan ^r ; P _{phaC} -RBS ₂₅ - <i>ispS</i> , constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₂₅ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₂₈ -ispS	Kan ^r ; P _{phaC} -RBS ₂₈ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₂₈ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₃₁ -ispS	Kan ^r ; P _{phaC} -RBS ₃₁ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₃₁ as described in <i>Material and Methods</i>	This study
pBBR1MCS-2-RBS ₃₂ -ispS	Kan ^r ; P _{phaC} -RBS ₃₂ -ispS, constructed by replacing <i>eyfp</i> with <i>ispS</i> in pBBR1MCS-2-RBS ₃₂ as described in <i>Material and Methods</i>	This study

pBBR1MCS-2-P ₁ -RBS ₁ Kan ^r ; P ₁ -RBS ₁ - <i>eyfp</i> , constructed by replacing P_{phac} with P ₁ in pBBR1MCS-2-RBS ₁			
pBBR1MCS-2-P ₁ -RBS ₂	Kan ^r ; P ₁ -RBS ₂ - <i>eyfp,</i> constructed by replacing P _{phaC} with P ₁ in pBBR1MCS-2-RBS ₂	This study	
pBBR1MCS-2-P ₁ -RBS ₈	Kan ^r ; P ₁ -RBS ₈ -eyfp, constructed by replacing P _{phaC} with P ₁ in pBBR1MCS-2-RBS ₈	This study	
pBBR1MCS-2-P _{rhaBAD} -RBS ₁	Kan ^r ; P _{rhaBAD} -RBS ₁ - <i>eyfp,</i> constructed by replacing P _{phaC} with P _{rhaBAD} in pBBR1MCS-2-RBS ₁	This study	
pBBR1MCS-2-P _{rhaBAD} -RBS ₂	Kan ^r ; P _{rhaBAD} -RBS ₂ -eyfp, constructed by replacing P _{phaC} with P _{rhaBAD} in pBBR1MCS-2-RBS ₂	This study	
pBBR1MCS-2-P _{rhaBAD} -RBS ₈	Kan ^r ; P _{rhaBAD} -RBS ₈ -eyfp, constructed by replacing P _{phaC} with P _{rhaBAD} in pBBR1MCS-2-RBS ₈	This study	
pBBR1MCS-2-RBS1-rfp	Kan ^r ; P _{phaC} -RBS ₁ - <i>rfp,</i> constructed by replacing <i>eyfp</i> with <i>rfp</i> in pBBR1MCS-2-RBS ₁	This study	
pBBR1MCS-2-RBS ₂ -rfp	Kan ^r ; P _{phaC} -RBS ₂ - <i>rfp</i> , constructed by replacing <i>eyfp</i> with <i>rfp</i> in pBBR1MCS-2-RBS ₂	This study	
pBBR1MCS-2-RBS ₈ -rfp	Kan ^r ; P _{phaC} -RBS ₈ -rfp, constructed by replacing eyfp with rfp in pBBR1MCS-2-RBS ₈	This study	
pMTL71101-RBS ₀	Cm ^r ; P _{phaC} -RBS ₀ - <i>eyfp</i> , assembled from pMTL71107 and PCR fragments generated by PCR using primer pairs P003-P007_rbs0_r and P008_rbs0_f-P004 from pBBR1MCS-2-PphaC-eyfp-c1	This study	
pMTL71101-RBS₁	Cm ^r ; P _{phaC} -RBS ₁ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	
pMTL71101-RBS ₂	Cm ^r ; P _{phaC} -RBS ₂ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	
pMTL71101-RBS₃	Cm ^r ; P _{phaC} -RBS ₃ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	
pMTL71101-RBS ₄	Cm ^r ; P _{phaC} -RBS ₄ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	
pMTL71101-RBS₅	Cm ^r ; P _{phaC} -RBS ₅ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	
pMTL71101-RBS ₆	Cm ^r ; P _{phaC} -RBS ₆ - <i>eyfp</i> , assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study	

pMTL71101-RBS ₇	Cm ^r ; P _{phaC} -RBS ₇ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₈	Cm ^r ; P _{phaC} -RBS ₈ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₉	Cm ^r ; P _{phaC} -RBS ₉ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₁₀	Cm ^r ; P _{phaC} -RBS ₁₀ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS11	Cm ^r ; P _{phaC} -RBS ₁₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₁₂	Cm ^r ; P _{phaC} -RBS ₁₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS13	Cm; P _{phaC} -RBS ₁₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS14	Cm ^r ; P _{phaC} -RBS ₁₄ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₂₀	Cm ^r ; P _{phaC} -RBS ₂₀ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₂₁	Cm ^r ; P _{phaC} -RBS ₂₁ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₂₂	Cm ^r ; P _{phaC} -RBS ₂₂ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₂₃	Cm ^r ; P _{phaC} -RBS ₂₃ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study

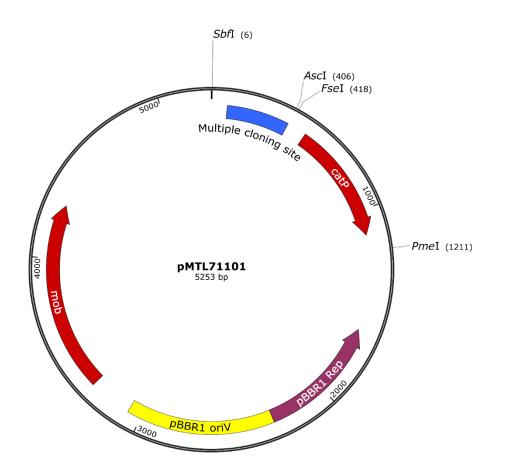
pMTL71101-RBS ₂₄	Cm ^r ; P _{phaC} -RBS ₂₄ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₂₅	Cm ^r ; P _{phaC} -RBS ₂₅ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₂₆	Cm ^r ; P _{phaC} -RBS ₂₆ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₂₇	Cm ^r ; P _{phaC} -RBS ₂₇ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₂₈	Cm ^r ; P _{phaC} -RBS ₂₈ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₂₉	Cm; P _{phaC} -RBS ₂₉ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₃₀	Cm ^r ; P _{phaC} -RBS ₃₀ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pMTL71101-RBS ₃₁	Cm ^r ; P _{phaC} -RBS ₃₁ -eyfp, assembled as above using corresponding primers as described in <i>Material and Methods</i>	This study
pMTL71101-RBS ₃₂	Cm ^r ; P _{phaC} -RBS ₃₂ -eyfp, assembled as above using corresponding primers as described in Material and Methods	This study
pEH002	Cm ^r ; P _{rhasR} -rhaSR-T _{rrnB1} and P _{rhaBAD} -T7sI- <i>Ec</i> RBS-rfp-T _{dbl} from pJOE7784.1 cloned into pEH006 by AatII and XbaI sites	This study
pEH005	Cm ^r ; P _{lac} -tetR-T _{rrnB2} and P _{tetA} -T7sI-EcRBS-rfp-T _{dbl} from pJOE7801.1 assembled by NEBuilder	This study
pEH020	Cm ^r ; P _{lac} -acuR-T _{rrnB2} and P _{acuR} -T7sl-EcRBS- <i>rfp</i> -T _{dbl} from <i>R. sphaeroides</i> 2.4.1 genomic DNA assembled by NEBuilder	This study

pEH040	040 Cm^r ; P_{lac} -cymR-T _{rrnB2} and P_{cmt} -EcRBS-rfp-T _{dbl} from pNEW assembled by NEBuilder	
pEH002-ispS	Cm ^r ; P _{rhasR} -rhaSR-T _{rrnB1} and P _{rhaBAD} -T7sl- <i>Ec</i> RBS- <i>ispS</i> -T _{dbl} , rfp replaced with <i>ispS</i> in pEH002 through NdeI and BamHI restriction sites	This study
pEH005-ispS	Cm ^r ; P _{lac} -tetR-T _{rrnB2} and P _{tetA} -T7sl-EcRBS-rfp-T _{dbl} , rfp replaced with ispS in pEH002 through NdeI and BamHI restriction sites	This study
pEH020-ispS	Cm ^r ; P _{lac} -acuR-T _{rrnB2} and P _{acuR} -T7sl-EcRBS-rfp-T _{dbl} , rfp replaced with ispS in pEH002 through NdeI and BamHI restriction sites	This study
pEH040-ispS	Cm ^r ; P _{lac-} cymR-T _{rrnB2} and P _{cmt} -EcRBS-rfp-T _{dbl} , rfp replaced with <i>ispS</i> in pEH002 through NdeI and AfIII restriction sites	This study
pEH176	Cm ^r ; ParaC-araC-T _{rrnB1} and ParaBAD-EcRBS-rfp-T _{dbl} from <i>E. coli</i> MG1655 genomic DNA cloned into pEH006 by AatII and NdeI sites	This study

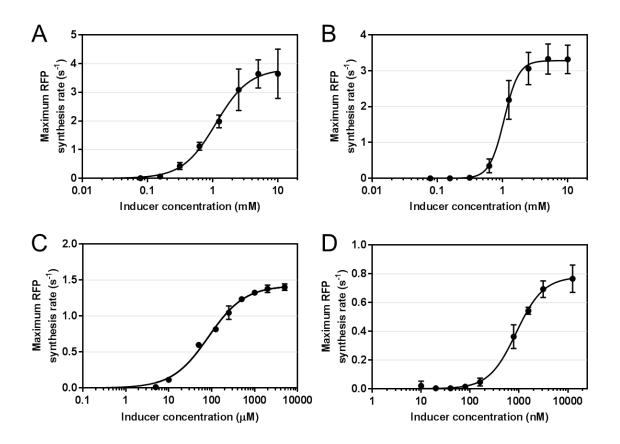
Supplementary Table S3. Hill functio	n fitting parameters.
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Plasmid identifier	Inducer	Minimum RFP synthesis rate, v _{min} (s ⁻¹)	Maximum RFP synthesis rate, v _{max} (s ⁻¹)	Hill coefficient, h	Half-maximal RFP synthesis, <i>K_m</i> (μM)
pEH002	L-rhamnose	0.0058 ± 0.0022	3.292 ± 0.032	3.88 ± 0.23	1057 ± 19
pEH006	L-arabinose	0.0122 ± 0.0023	3.836 ± 0.085	1.70 ± 0.11	1116 ± 56
pEH020	acrylate	0.0474 ± 0.0056	1.471 ± 0.035	1.03 ± 0.09	84.32 ± 8.08
pEH040	cumate	0.0472 ± 0.0076	0.828 ± 0.011	1.56 ± 0.08	0.877 ± 0.029

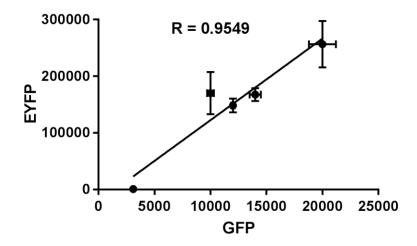
SUPPLEMENTARY FIGURES



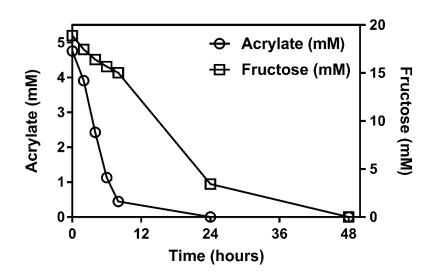
Supplementary Figure S1. Plasmid map of modular vector pMTL71101. Restriction endonucleases SbfI, AscI, FseI, and PmeI, flanking modules of the vector, are shown with their restriction site positions in the brackets. The pBBR1 origin of replication and mobilisation gene *mob* (PmeI-SbfI module), the chloramphenicol resistance marker *catP* (FseI-PmeI module) and a multiple cloning site (SbfI-AscI module) are highlighted.



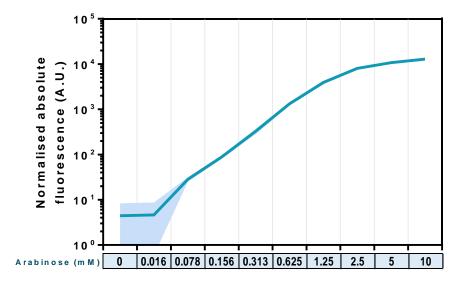
Supplementary Figure S2. Promotor activity of inducible systems. The normalised maximum rate of RFP synthesis was fit to the corresponding inducer concentration using a Hill function for the (A) L-arabinose-, (B) L-rhamnose-, (C) acrylate-, and (D) cumate-inducible systems. Error bars represent standard deviations of three biological replicates.



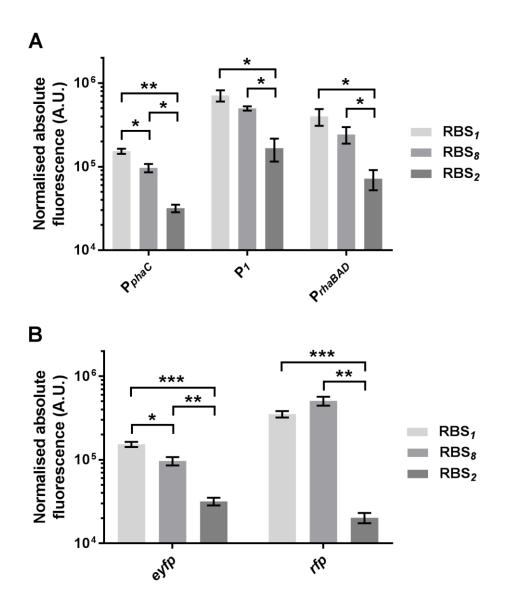
Supplementary Figure S3. Correlation between promoter activities, measured as normalised fluorescence arbitrary units (A.U.) using either GFP as reporter in previous study (6) and EYFP in this study. Core promoters derived from bacteriophage T5 and *C. necator* H16 (promoters P_{T5} , P_{j5} , P_{h207} , P_{h25} and P_{H16_B1772} (6)) were used in analysis. In this study they comprise core part of promoters P_1 , P_4 , P_5 , P_6 , and P_2 , respectively.



Supplementary Figure S4. Consumption of acrylate in *C. necator*. Acrylate was added at time zero. Consumption of acrylate and fructose was monitored using HPLC-UV in supernatant over the time course of 48 hours. The means of three biological replicates are presented. Error bars are too small to be visible.



Supplementary Figure S5. Induction dynamics for the L-arabinose-inducible system lacking the stem-loop structure. Normalised absolute fluorescence of *C. necator* cells in exponential growth phase six hours after L-arabinose addition. The standard deviation of three biological replicates is illustrated as band.



Supplementary Figure S6. Effect of sequence content on the strength of RBS. Normalised absolute fluorescence determined using plasmid constructs with promoters P_{phaC} , $(P_{0_{-}I})$, P_{I} , and P_{rhaBAD} (A), and reporter genes *eyfp* and *rfp* (B) located upstream and downstream, respectively, of RBS₁, RBS₈ and RBS₂. Error bars represent standard deviation of three biological replicas. Asterisks indicate statistically significant difference between RBS strengths: *p < 0.01; **p < 0.001; and ***p < 0.001 (unpaired t-test).

REFERENCES

- 1. **Heap JT, Pennington OJ, Cartman ST, Minton NP.** 2009. A modular system for *Clostridium* shuttle plasmids. Journal of Microbiological Methods **78**:79-85.
- Maurhofer M, Reimmann C, Schmidli-Sacherer P, Heeb S, Haas D, Defago G. 1998. Salicylic acid biosynthetic genes expressed in *Pseudomonas fluorescens* strain P3 improve the induction of systemic resistance in tobacco against tobacco necrosis virus. Phytopathology 88:678-684.
- 3. **Pfeiffer D, Jendrossek D.** 2011. Interaction between poly(3-hydroxybutyrate) granule-associated proteins as revealed by two-hybrid analysis and identification of a new phasin in *Ralstonia eutropha* H16. Microbiology **157:**2795-2807.
- 4. **Hanko E, Minton N, Malys N.** 2017. Characterisation of a 3-hydroxypropionic acid-inducible system from *Pseudomonas putida* for orthogonal gene expression control in *Escherichia coli* and *Cupriavidus necator*. Scientific Reports **7:**1724.
- 5. **Hoffmann J, Altenbuchner J.** 2015. Functional characterization of the mannitol promoter of *Pseudomonas fluorescens* DSM 50106 and its application for a mannitol-inducible expression system for *Pseudomonas putida* KT2440. PLoS ONE **10**.
- 6. **Gruber S, Hagen J, Schwab H, Koefinger P.** 2014. Versatile and stable vectors for efficient gene expression in *Ralstonia eutropha* H16. Journal of Biotechnology **186:**74-82.