Applied Microbiology and Biotechnology

Supplementary Information

Microbial Community Composition and Diversity in Rice Straw Co-digestion Bioreactors with Dairy Manure

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Additional Methods in Next Generation Sequencing:

The PCRs included 1 to 10 ng of DNA extract (total volume 1µl), 15 pmol of each forward primer and reverse primer (in 20 µL volume of 1 x MyTaq buffer containing 1.5 units MyTaq DNA polymerase (Bioline) and 2 µl of BioStabII PCR Enhancer (Sigma). For each sample, the forward and reverse primers had the same 10-nt barcode sequence. PCRs were carried out for 30 cycles using the following parameters: 2 min 96°C pre-denaturation; 96°C for 15 s, 50°C for 30 s, 70°C for 90 s. DNA concentration of amplicons of interest was determined by gel electrophoresis. About 20 ng of amplicon DNA from each sample were pooled for up to 48 samples carrying different barcodes. If needed, PCRs with low yields were further amplified for 5 cycles.

The amplicon pools were purified with one volume AMPure XP beads (Agencourt) to remove primer dimer and other small mispriming products, followed by an additional purification on MinElute columns (Qiagen). About 100 ng of each purified amplicon pool DNA was used to construct Illumina libraries using the Ovation Rapid DR Multiplex System 1-96 (NuGEN). Illumina libraries were pooled and size selected by preparative gel electrophoresis. Sequencing was done on an Illumina MiSeq using V3 Chemistry (Illumina).

A total of 419,747 quality filtered samples were obtained from 15 samples with the number of 16S rDNA sequences ranging from 10,864 to 48,127 (mean, 27,983). After removing the chimera sequences the operational taxonomic units (OTUs) were picked at more than 97 % similarity representing 9,720 OTUs. The alpha and beta diversity of the microbial community were then assessed.

List of Figures:

- Figure S1: Time-course data of digester performance including acclimation period (experiment 'Time 0' shown by dashed line) for pH, %Volatile Solids Removal (VSR), g VS/L, specific methane yields and VFA levels for: a) RS100, b) RS90, c) RS70, d) RS30.
- Figure S2: Shared predominant OTU table to the genus level (only ≥ 0.5 % abundance) in bioreactor and raw dairy manure based on sample appearances. "H1", "H2", and "H3" refer to each HRT cycle (25 days) the reactors were operated.
- Figure S3: Phylogenetic tree of predominant OTUs (only ≥ 0.5 % abundance) shared across all samples. Branch closeness and small white, grey, and black circles on branch junctions indicate percentage of similarity (50 - 69 %, 70 - 89 %, and 90+ %).
- **Figure S4**: Extended error bar plot of significant differences between predominant OTUs in the RS100 reactor versus raw dairy manure (DM).
- **Figure S5**: Extended error bar plot of significant differences between predominant OTUs in the RS90 reactor versus raw dairy manure (DM).
- **Figure S6**: Extended error bar plot of significant differences between predominant OTUs in the RS70 reactor versus raw dairy manure (DM).
- **Figure S7**: Extended error bar plot of significant differences between predominant OTUs in the RS30 reactor versus raw dairy manure (DM).







Figure S2











