

# Supporting Information

## Synthesis of randomly substituted anionic cyclodextrins in a ball mill

László Jicsinszky<sup>1,\*</sup>, Marina Caporaso<sup>1</sup>, Emanuela Calcio Gaudino<sup>1</sup>, Cristina Giovannoli<sup>2</sup>, and Giancarlo Cravotto<sup>1,\*</sup>

<sup>1</sup> University of Turin, Department of Drug Science and Technology, Via P. Giuria 7, 10125 Turin, Italy; [laszlo.jicsinszky@unito.it](mailto:laszlo.jicsinszky@unito.it) (LJ); [marina.caporaso@unito.it](mailto:marina.caporaso@unito.it) (MC); [Giancarlo.cravotto@unito.it](mailto:Giancarlo.cravotto@unito.it) (GC).

<sup>2</sup> University of Turin, Department of Chemistry, Via P. Giuria 7 - 10125 Turin, Italy; [cristina.giovannoli@unito.it](mailto:cristina.giovannoli@unito.it) (CG)

\* Correspondence: [ljicsinszky@gmail.com](mailto:ljicsinszky@gmail.com) (LJ), [Giancarlo.cravotto@unito.it](mailto:Giancarlo.cravotto@unito.it) (GC). Tel.: +39-011-670-7663, +39-011-670-7684

## Content

Figure S 1: Proton NMR spectrum of carboxymethylated $\beta$ CD ( <b>2</b> ) prepared in solution (item 1 in Table 1), DS $\approx$ 4.3-4.5	S4
Figure S 2: HSQC-DEPT spectrum of carboxymethylated $\beta$ CD ( <b>2</b> ) prepared in solution (item 1 in Table 1), DS $\approx$ 4.3-4.5	S4
Figure S 3: <sup>13</sup> C-NMR spectrum of carboxymethylated $\beta$ CD ( <b>2</b> ) prepared in solution (item 1 in Table 1), DS $\approx$ 4.3-4.5	S5
Figure S 4: ESI+ mass spectrum of carboxymethylated $\beta$ CD ( <b>2</b> ) prepared in solution (item 1 in Table 1), DS $\approx$ 4.3-4.5	S5
Figure S 5: Peak identification in ESI+ mass spectrum of carboxymethylated $\beta$ CD ( <b>2</b> ) prepared in solution (item 1 in Table 1), DS $\approx$ 4.3-4.5 [1,2]	S6
Figure S 6: Proton NMR spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 2 in Table 1), DS $\approx$ 4.3-4.6	S7
Figure S 7: HSQC-DEPT spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 2 in Table 1), DS $\approx$ 4.3-4.6	S7
Figure S 8: Proton NMR spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 3 in Table 1), DS $\approx$ 4.4-4.6	S8
Figure S 9: HSQC-DEPT spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 3 in Table 1), DS $\approx$ 4.4-4.6	S8
Figure S 10: Proton NMR spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 4 in Table 1), DS $\approx$ 4.4-4.6	S9
Figure S 11: HSQC-DEPT spectrum of carboxymethylated $\beta$ CD ( <b>2'</b> ) prepared in ball mill (item 4 in Table 1), DS $\approx$ 4.4-4.6	S9
Figure S 12: Capillary electropherogram of carboxymethylated $\beta$ CDs	S10
Figure S 13: Proton NMR spectrum of carboxyethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4	S11
Figure S 14: HSQC-DEPT spectrum of carboxyethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4	S11
Figure S 15: <sup>13</sup> C-NMR spectrum of carboxyethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4	S12
Figure S 16: ESI+ mass spectrum of carboxyethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4	S12
Figure S 17: Peak identification in ESI+ mass spectrum of carboxyethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4	S13
Figure S 18: Proton NMR spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 5 in Table 1), DS $\approx$ 3.0-3.3	S14

Figure S 19: HSQC-DEPT spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 5 in Table 1), DS $\approx$ 3.0-3.3	<b>S14</b>
Figure S 20: Proton NMR spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 6 in Table 1), DS $\approx$ 2.8-3.2	<b>S15</b>
Figure S 21: HSQC-DEPT spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 6 in Table 1), DS $\approx$ 2.8-3.2	<b>S15</b>
Figure S 22: Proton NMR spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 7 in Table 1), DS $\approx$ 2.8-3.1	<b>S16</b>
Figure S 23: HSQC-DEPT spectrum of carboxyethylated $\beta$ CD ( <b>3'</b> ) prepared in ball mill (item 7 in Table 1), DS $\approx$ 2.8-3.1	<b>S16</b>
Figure S 24: IR spectrum of carboxylethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1) as acid, DS $\approx$ 3.0-3.4	<b>S17</b>
Figure S 25: IR spectrum of carboxylethylated $\beta$ CD ( <b>3</b> ) prepared in solution (item 8 in Table 1) as sodium salt, DS $\approx$ 3.0-3.4	<b>S17</b>
Figure S 26: Overlay of IR spectrum of carboxylethylated $\beta$ CD acid & salt form ( <b>3</b> ) prepared in solution (item 8 in Table 1), DS $\approx$ 3.0-3.4 and carbamoylethylated $\beta$ CD prepared in ball mill (item 10 in table 1)	<b>S18</b>
Figure S 27: Proton NMR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 9 in Table 1), DS $\approx$ 3.8-4.3	<b>S22</b>
Figure S 29: $^{13}\text{C}$ -NMR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 9 in Table 1), DS $\approx$ 3.8-4.3	<b>S20</b>
Figure S 30: ESI+ mass spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 9 in Table 1), DS $\approx$ 3.8-4.3	<b>S20</b>
Figure S 31: Peak identification in ESI+ mass spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 9 in Table 1), DS $\approx$ 3.8-4.3	<b>S21</b>
Figure S 32: Proton NMR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S22</b>
Figure S 33: HSQC-DEPT spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S22</b>
Figure S 34: $^{13}\text{C}$ -NMR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S31</b>
Figure S 35: ESI+ mass spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S23</b>
Figure S 36: Peak identification in ESI+ mass spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.8-4.3	<b>S23</b>
Figure S 37: IR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S25</b>
Figure S 38: IR spectrum of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 10 in Table 1), DS $\approx$ 3.3-3.5	<b>S25</b>
Figure S 39: Overlay of IR spectra of carbamoylethylated $\beta$ CD ( <b>5</b> ) prepared in ball mill (item 9&10 in Table 1), DS $\approx$ 3.3-3.5 carboxyethylated $\beta$ CD prepared in solution (item 8 in Table 1)	<b>S26</b>
Figure S 40: Capillary electropherogram of carboxyethylated $\beta$ CDs	<b>S26</b>
Figure S 41: Proton NMR spectrum of sulfobutylated $\beta$ CD ( <b>4</b> ) prepared in solution (item 11 in Table 1), DS $\approx$ 6.2-6.5	<b>S28</b>
Figure S 42: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD ( <b>4</b> ) prepared in solution (item 11 in Table 1), DS $\approx$ 6.2-6.5	<b>S28</b>
Figure S 43: ESI+ mass spectrum of sulfobutylated $\beta$ CD ( <b>4</b> ) prepared in solution (item 11 in Table 1), DS $\approx$ 6.2-6.5	<b>S29</b>
Figure S 44: Peak identification in ESI+ mass spectrum of sulfobutylated $\beta$ CD ( <b>4</b> ) prepared in solution (item 11 in Table 1), DS $\approx$ 6.2-6.5	<b>S30</b>

Figure S 45: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 12 in Table 1), DS $\approx$ 3.1-3.3	<b>S31</b>
Figure S 46: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 12 in Table 1), DS $\approx$ 3.1-3.3	<b>S31</b>
Figure S 47: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 13 in Table 1), DS $\approx$ 5.2-5.8	<b>S32</b>
Figure S 48: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 13 in Table 1), DS $\approx$ 5.2-5.8	<b>S32</b>
Figure S 49: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 14 in Table 1), DS $\approx$ 6.2-6.6	<b>S33</b>
Figure S 50: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 14 in Table 1), DS $\approx$ 6.2-6.6	<b>S33</b>
Figure S 51: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 15 in Table 1), DS $\approx$ 7.5-7.8	<b>S34</b>
Figure S 52: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 15 in Table 1), DS $\approx$ 7.5-7.8	<b>S34</b>
Figure S 53: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 16 in Table 1), DS $\approx$ 4.4-4.9	<b>S35</b>
Figure S 54: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 16 in Table 1), DS $\approx$ 4.4-4.9	<b>S35</b>
Figure S 55: Proton NMR spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 17 in Table 1), DS $\approx$ 6.9-7.5	<b>S36</b>
Figure S 56: HSQC-DEPT spectrum of sulfobutylated $\beta$ CD (4') prepared in ball mill (item 17 in Table 1), DS $\approx$ 6.9-7.5	<b>S36</b>
Figure S 57: Proton NMR spectrum of 4-hydroxy-1-butanefulfonic acid sodium salt (HOBSANa)	<b>S37</b>
Figure S 58: HSQC-DEPT spectrum of HOBSANa	<b>S37</b>
Figure S 59: Proton NMR spectrum of di(1,1'-sulfonatobutyl)ether disodium salt (disodium 4,4'-oxydibutane-1,1'-disulfonate, BIBSANA)	<b>S38</b>
Figure S 60: HSQC-DEPT spectrum of BIBSANA	<b>S38</b>
Figure S 61: Capillary electropherogram of sulfobutylated $\beta$ CDs	<b>S39</b>
 Scheme S 1: Carbon atom numbering for the NMR assignment (OH: unsubstituted; OR: substituted)	 <b>S40</b>
 Table S 1: Proton assignment of cyclodextrin derivatives prepared in solution (based on $^1\text{H}$ - and HSQC-DEPT experiments)	 <b>S40</b>
Table S 2: Carbon assignment of cyclodextrin derivatives prepared in solution (based on $^{13}\text{C}$ - and HSQC-DEPT experiments)	<b>S41</b>
 Table S 3: Characteristic C=O IR bands of the prepared (carbonyl)ethyl group containing CD derivatives	 <b>S41</b>
 Table S 4: $R_F$ -values of the prepared compounds after twice run of the same plate in the same solvent mixture at 7 cm distance in 100 $\mu\text{g}$	 <b>S41</b>
 References	 <b>S42</b>

Acquisition Time (sec)	3.6438	Comment	CMB CD LJ00SOL_1H_D2O_131016_2056_rg=90	Date	13 Oct 2016 14:05:20
Date Stamp	13 Oct 2016 14:05:20	File Name	D:\Docs\1\1\Notebooks\1M_Reactions\NMR\CMB CD LJ00SOL\1fid	Nucleus	1H
Frequency (MHz)	300.13	Number of Transients	64	Origin	spect
Owner	root	Pulse Sequence	zg	Receiver Gain	90.50
Solvent	CDCl3	Spectrum Offset (Hz)	1093.5470	Sweep Width (Hz)	4496.37
		Spectrum Type	STANDARD	Original Points Count	16384
				SW(cyclical) (Hz)	4496.40
				Temperature (degree C)	20.360

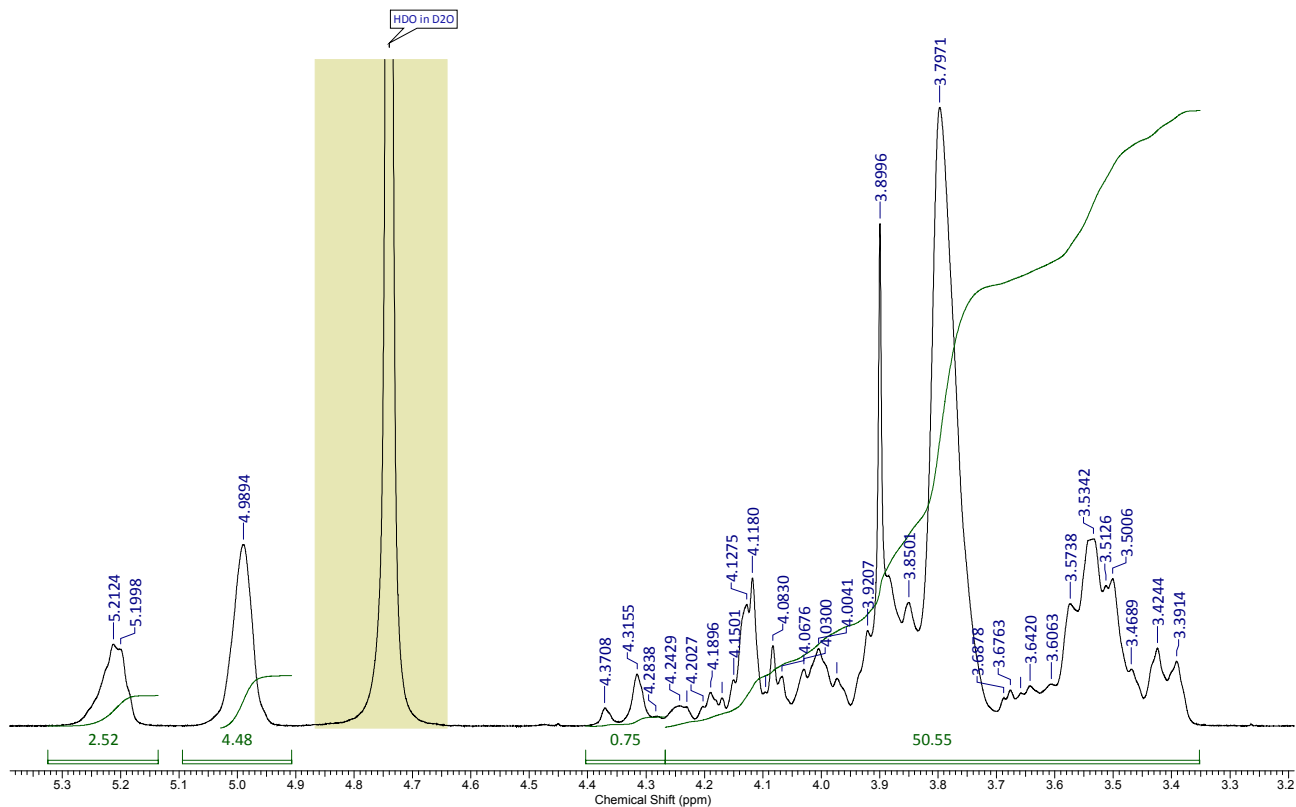


Figure S 1: Proton NMR spectrum of carboxymethylated  $\beta$ CD (2) prepared in solution (item 1 in Table 1),  $DS \approx 4.3-4.5$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	13 Oct 2016 13:59:34
File Name	D:\Docs\1\1\Notebooks\1M_Reactions\NMR\CMB CD LJ00SOL\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(4096, 256)	Pulse Sequence	hsqcdeptg	Solvent	DMSO
Sweep Width (Hz)	(2996.87, 12451.17)	Temperature (degree C)	20.460	Title	CMB CD LJ00SOL HSQC_D2O_131016_2056_rg=90

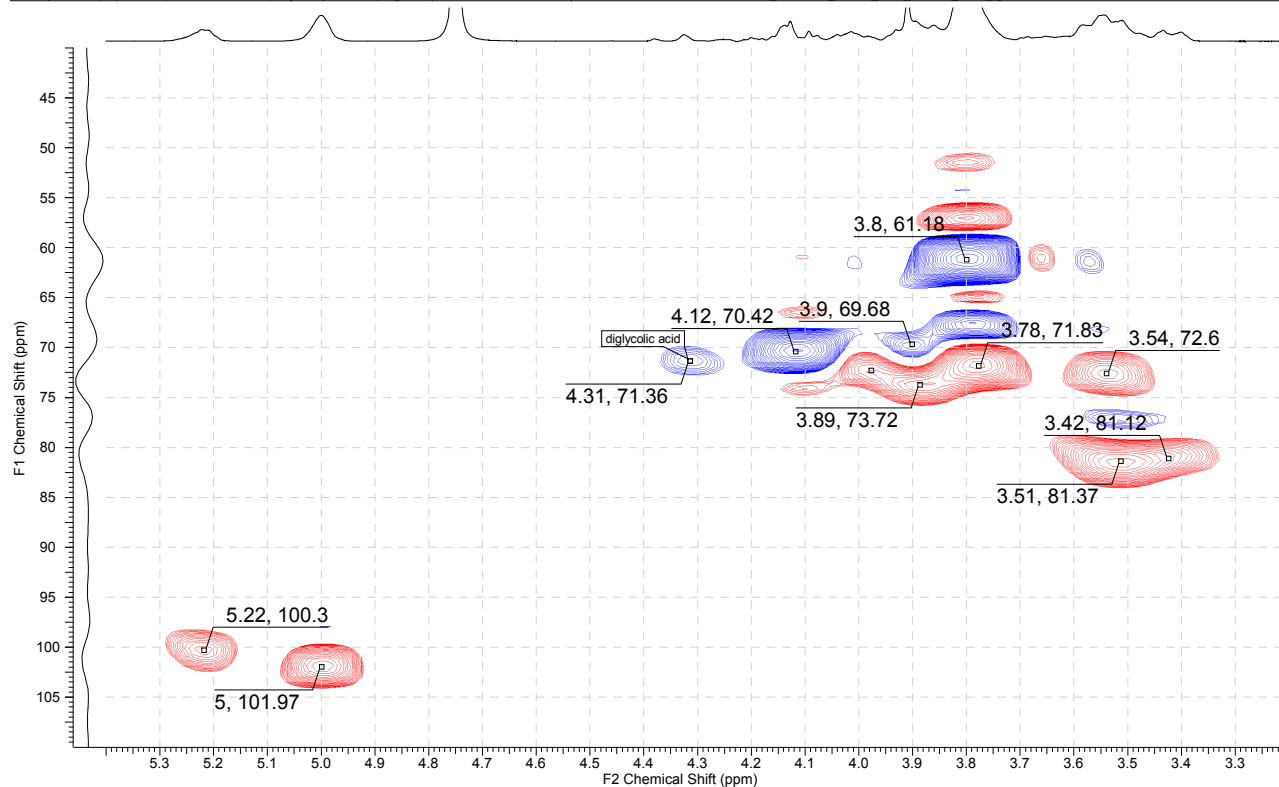


Figure S 2: HSQC-DEPT spectrum of carboxymethylated  $\beta$ CD (2) prepared in solution (item 1 in Table 1),  $DS \approx 4.3-4.5$

Acquisition Time (sec)	1.8088	Comment	CMBCD_LJ02_13C_D2O_230117_2134_RG=100	Date	23 Jan 2017 08:10:56
Date Stamp	23 Jan 2017 08:10:56	File Name	E:\Documents\1\Molecules\Anal\CarbonNMR\CMBCD_LJ02\2fid	Origin	spect
Frequency (MHz)	75.47	Nucleus	13C	Number of Transients	627
Owner	root	Points Count	262144	Pulse Sequence	zgpq.save.txt
Solvent	D2O	Spectrum Offset (Hz)	8301.4463	Receiver Gain	13004.00
		Spectrum Type	STANDARD	SW(cyclical) (Hz)	18115.87
				Sweep Width (Hz)	18115.87
				Temperature (degree C)	20.460

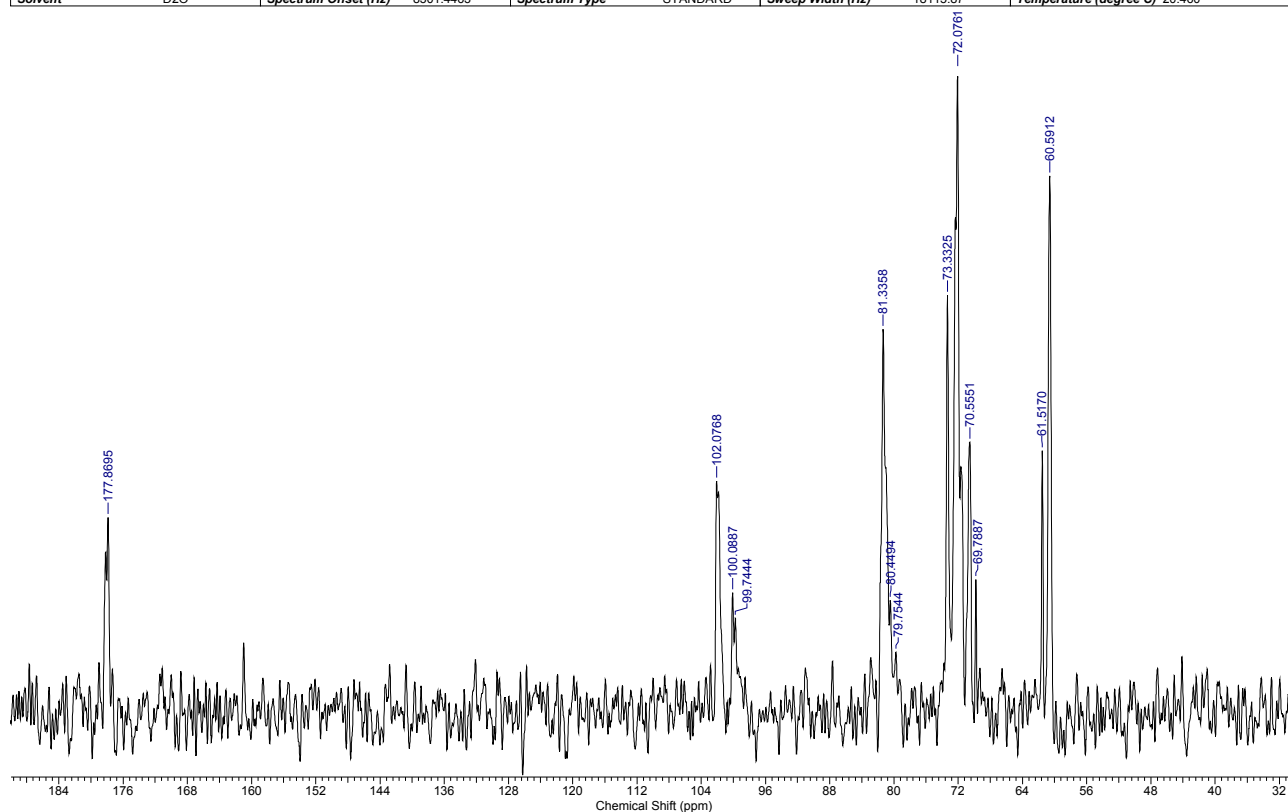


Figure S 3:  $^{13}\text{C}$ -NMR spectrum of carboxymethylated  $\beta\text{CD}$  (2) prepared in solution (item 1 in Table 1), DS  $\approx 4.3$ -4.5

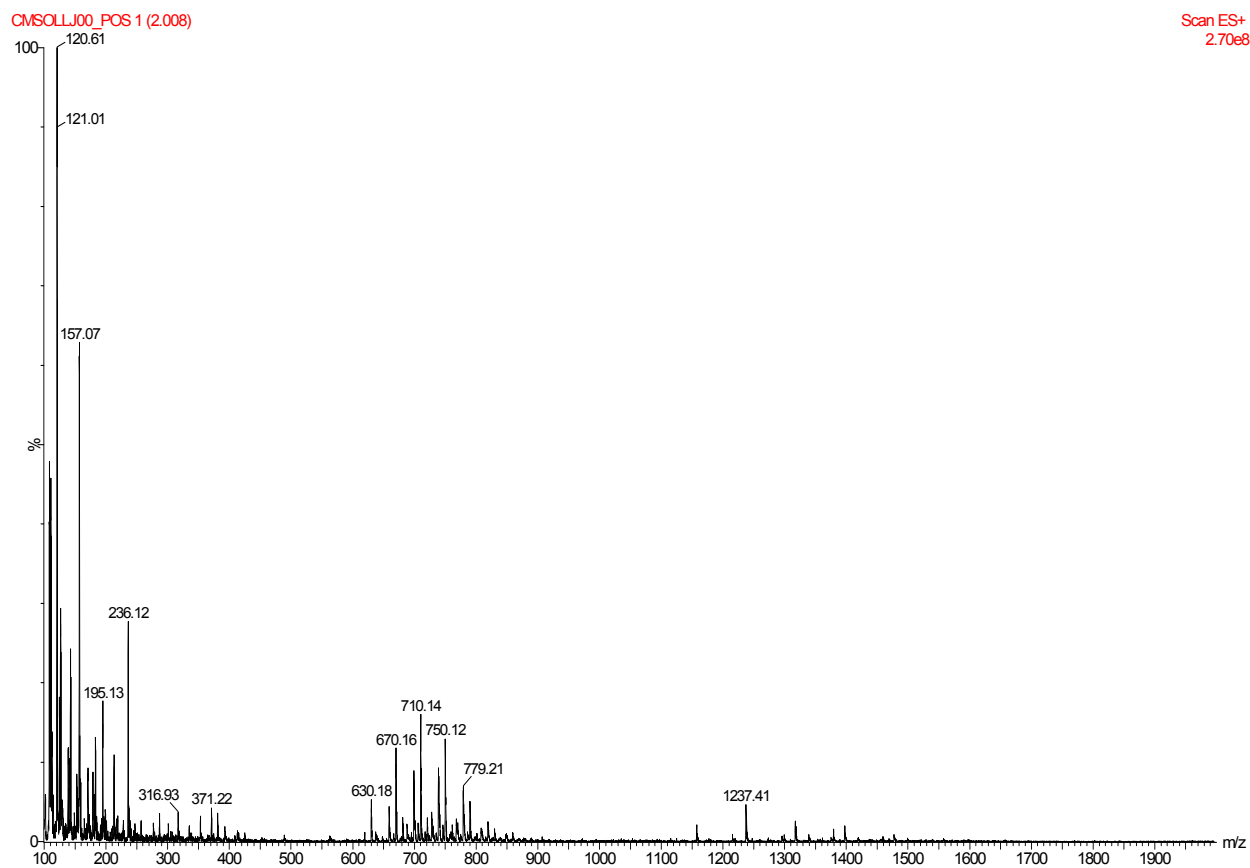
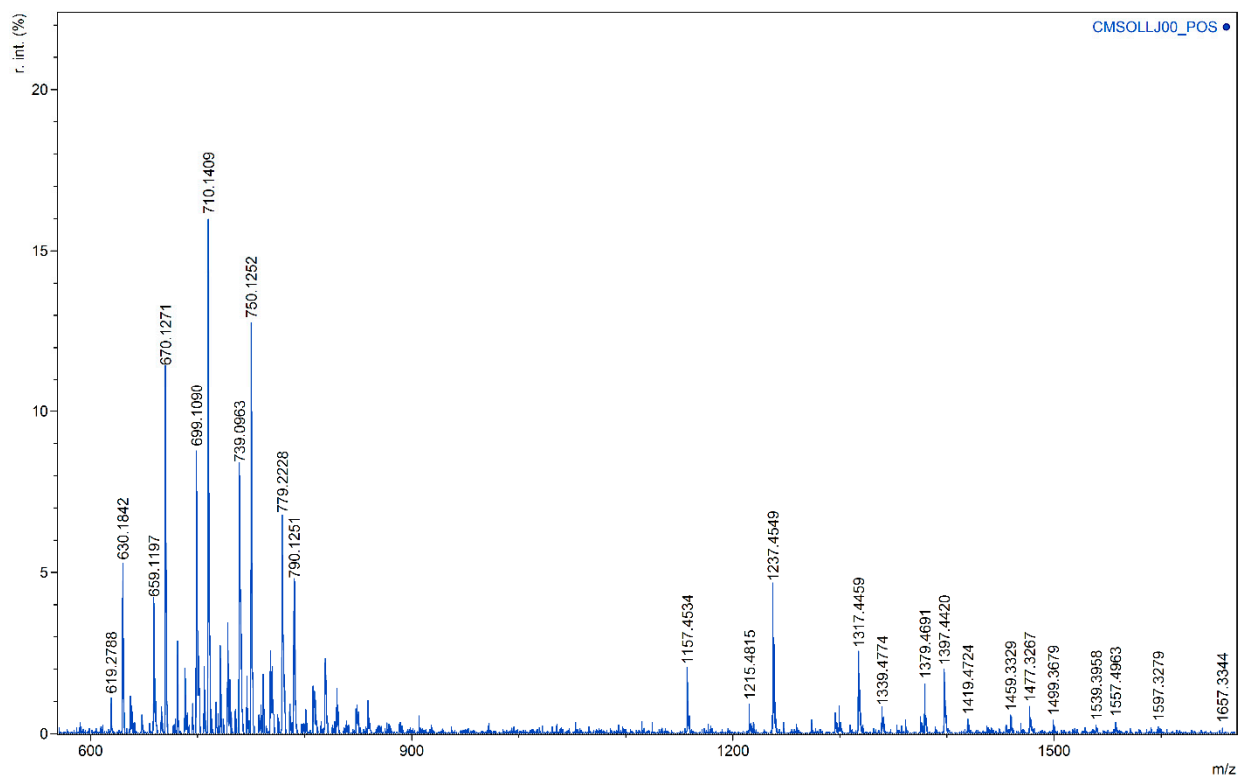


Figure S 4: ESI+ mass spectrum of carboxymethylated  $\beta\text{CD}$  (2) prepared in solution (item 1 in Table 1), DS  $\approx 4.3$ -4.5



Meas. m/z	Calc. m/z	$\delta$ (Da)	z	Annotation	Formula
630.1842	630.1678	0.0164	2	(+1 CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>1</sub>
659.1197	659.1706	-0.0509	2	(+1 CH <sub>2</sub> COOH+1 CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>1</sub>
699.1090	699.1760	-0.0670	2	(+1 CH <sub>2</sub> COOH +2 HCOOH +1 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>2</sub> (HCOONa) <sub>1</sub> (H <sub>2</sub> O) <sub>0</sub>
710.1409	710.1670	-0.0261	2	(+1 CH <sub>2</sub> COOH +1 HCOOH +2 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>1</sub> (HCOONa) <sub>2</sub> (H <sub>2</sub> O) <sub>0</sub>
739.0963	739.1697	-0.0734	2	(+2 CH <sub>2</sub> COOH +1 HCOOH +2 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>2</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>1</sub> (HCOONa) <sub>2</sub> (H <sub>2</sub> O) <sub>0</sub>
750.1252	750.1607	-0.0355	2	(+2 CHCOOH +3 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>2</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>3</sub> (H <sub>2</sub> O) <sub>0</sub>
779.2228	779.1635	0.0593	2	(+3 CH <sub>2</sub> COOH +3 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>3</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>3</sub> (H <sub>2</sub> O) <sub>0</sub>
790.1251	790.1544	-0.0293	2	(+2 CH <sub>2</sub> COOH +1 CH <sub>2</sub> COONa +3 HCOONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>2</sub> (CH <sub>1</sub> COONa) <sub>1</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>3</sub> (H <sub>2</sub> O) <sub>0</sub>
1157.4534	1157.3590	0.0944	1	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> (βCD)	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na
1215.4815	1215.3645	0.1171	1	+1 CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na)CH <sub>1</sub> COOH
1237.4549	1237.3464	0.1085	1	+1 CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na)CH <sub>1</sub> COONa
1317.4459	1317.3338	0.1120	1	+2 CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>0</sub> (CH <sub>1</sub> COONa) <sub>2</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>0</sub>
1379.4691	1379.4177	0.0515	1	+1 CH <sub>2</sub> COOH +2 HCOOH +4 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>2</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>4</sub>
1397.4420	1397.3213	0.1207	1	+3 CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>0</sub> (CH <sub>1</sub> COONa) <sub>3</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>0</sub>
1419.4724	1419.3267	0.1457	1	+1 CH <sub>2</sub> COOH +3 HCOONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>3</sub> (H <sub>2</sub> O) <sub>0</sub>
1459.3329	1459.4051	-0.0722	1	+1 CH <sub>2</sub> COOH +1 CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>1</sub> (CH <sub>1</sub> COONa) <sub>1</sub> (HCOOH) <sub>2</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>4</sub>
1477.3267	1477.4181	-0.0913	1	+3 CH <sub>2</sub> COOH +2 HCOOH +3 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>3</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>2</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>3</sub>
1499.3679	1499.4000	-0.0322	1	+3 CH <sub>2</sub> COOH +1 HCOOH +1 HCOONa +3 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>3</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>1</sub> (HCOONa) <sub>1</sub> (H <sub>2</sub> O) <sub>3</sub>
1539.3958	1539.3925	0.0032	1	+3 CH <sub>2</sub> COOH +2 HCOONa +4 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>3</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>2</sub> (H <sub>2</sub> O) <sub>4</sub>
1557.4963	1557.3196	0.1767	1	+2 CH <sub>2</sub> COOH +1 CH <sub>2</sub> COONa +3 HCOONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>2</sub> (CH <sub>1</sub> COONa) <sub>1</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>3</sub> (H <sub>2</sub> O) <sub>0</sub>
1597.3279	1597.3980	-0.0701	1	+4 CH <sub>2</sub> COOH +2 HCOONa +4 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>4</sub> (CH <sub>1</sub> COONa) <sub>0</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>2</sub> (H <sub>2</sub> O) <sub>4</sub>
1657.3344	1657.4215	-0.0872	1	+6 CH <sub>2</sub> COOH +1 CH <sub>2</sub> COONa +4 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> COOH) <sub>6</sub> (CH <sub>1</sub> COONa) <sub>1</sub> (HCOOH) <sub>0</sub> (HCOONa) <sub>0</sub> (H <sub>2</sub> O) <sub>4</sub>

Figure S 5: Peak identification in ESI+ mass spectrum of carboxymethylated βCD (2) prepared in solution (item 1 in Table 1), DS ≈4.3-4.5 [1,2]

Acquisition Time (sec)	3.6438	Comment	CMBcD_LJ01 (20mg in 0.6ml D2O)		Date	28 Sep 2016 11:33:52			
Date Stamp	28 Sep 2016 11:33:52	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\CMBcD_LJ01\1\fid						
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Owner	root	Points Count	131072	Pulse Sequence	zg	Receiver Gain	50.80	SW(cyclical) (Hz)	4496.40
Solvent	CDCI3	Spectrum Offset (Hz)	1096.0339	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37	Temperature (degree C)	22.260

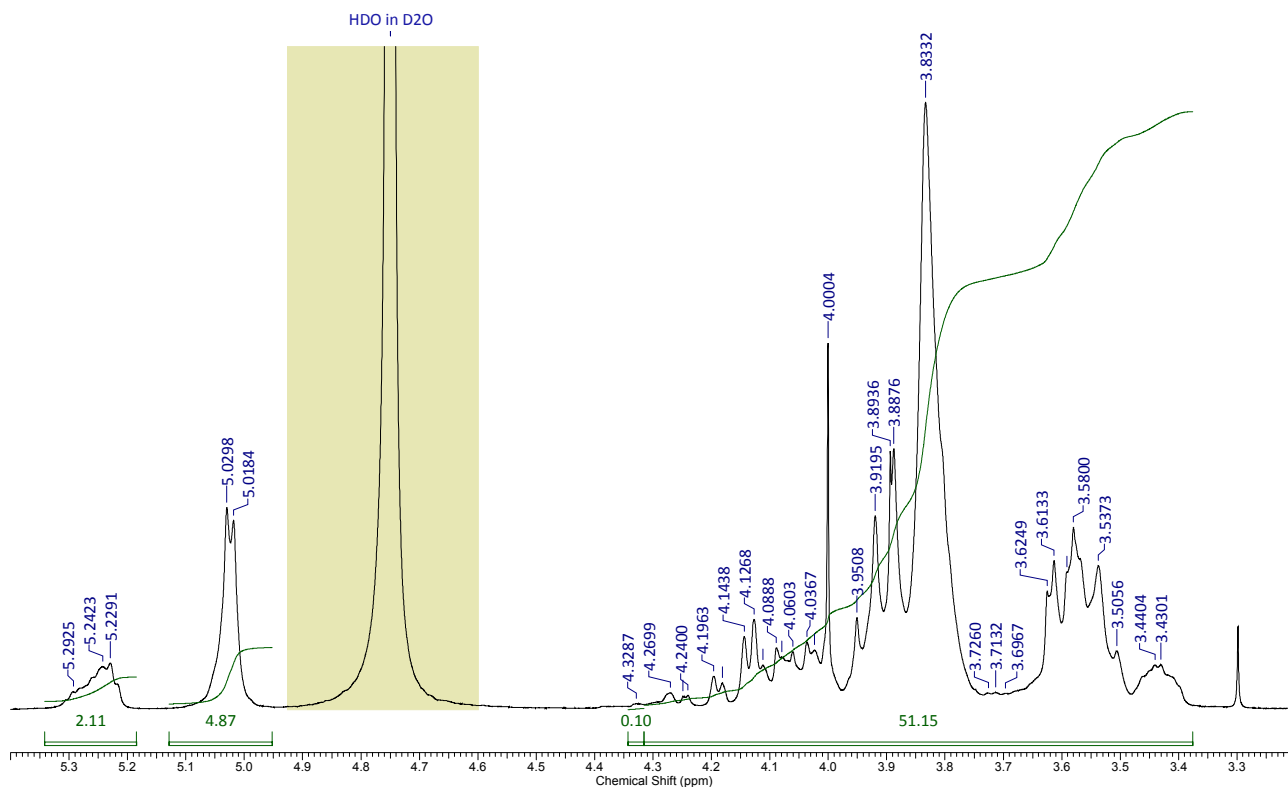


Figure S 6: Proton NMR spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 2 in Table 1),  $DS \approx 4.3-4.6$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059		Date	28 Sep 2016 12:27:24	
File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\CMBcD_LJ01\2\ser	Frequency (MHz)	(300.13, 75.47)		Nucleus	(1H, 13C)	
Number of Transients	32	Origin	spect		Original Points Count	(128, 64)	
Points Count	(512, 256)	Pulse Sequence	hsqcetdtp		Solvent	DMSO	
Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	22.260		Spectrum Type	HSQC-DEPT	

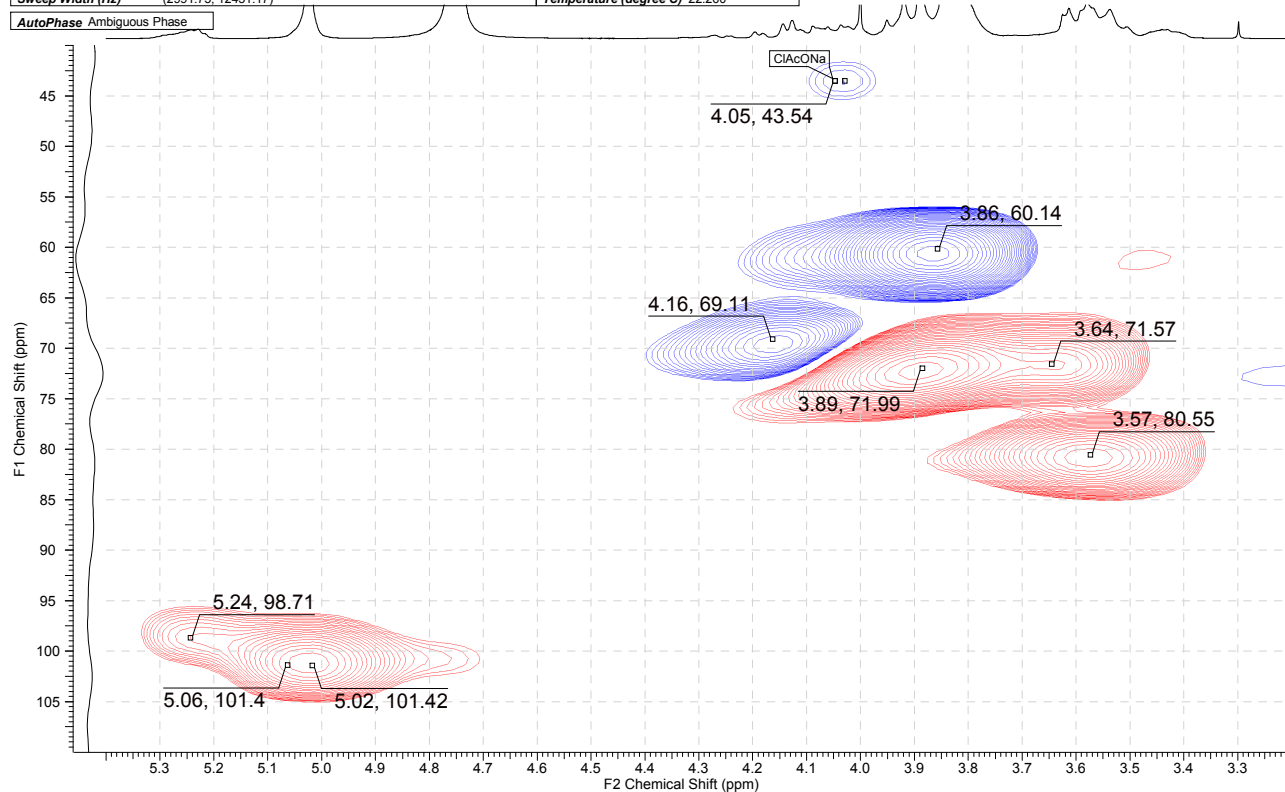


Figure S 7: HSQC-DEPT spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 2 in Table 1),  $DS \approx 4.3-4.6$

Acquisition Time (sec)	3.6438	Comment	CBCD_LJ02 (20mg in 0.6ml D2O)		Date	30 Sep 2016 15:20:00			
Date Stamp	30 Sep 2016 15:20:00	File Name	D:\Docs\1\1\Notebooks\BM_Reactions\NMR\CMB\CD_LJ02\1.tif						
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Owner	root	Points Count	131072	Pulse Sequence	zg	Receiver Gain	101.60	SW(cyclical) (Hz)	4496.40
Solvent	DEUTERIUM OXIDE	Spectrum Offset (Hz)	1096.2397	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37		
Temperature (degree C)	22.160								

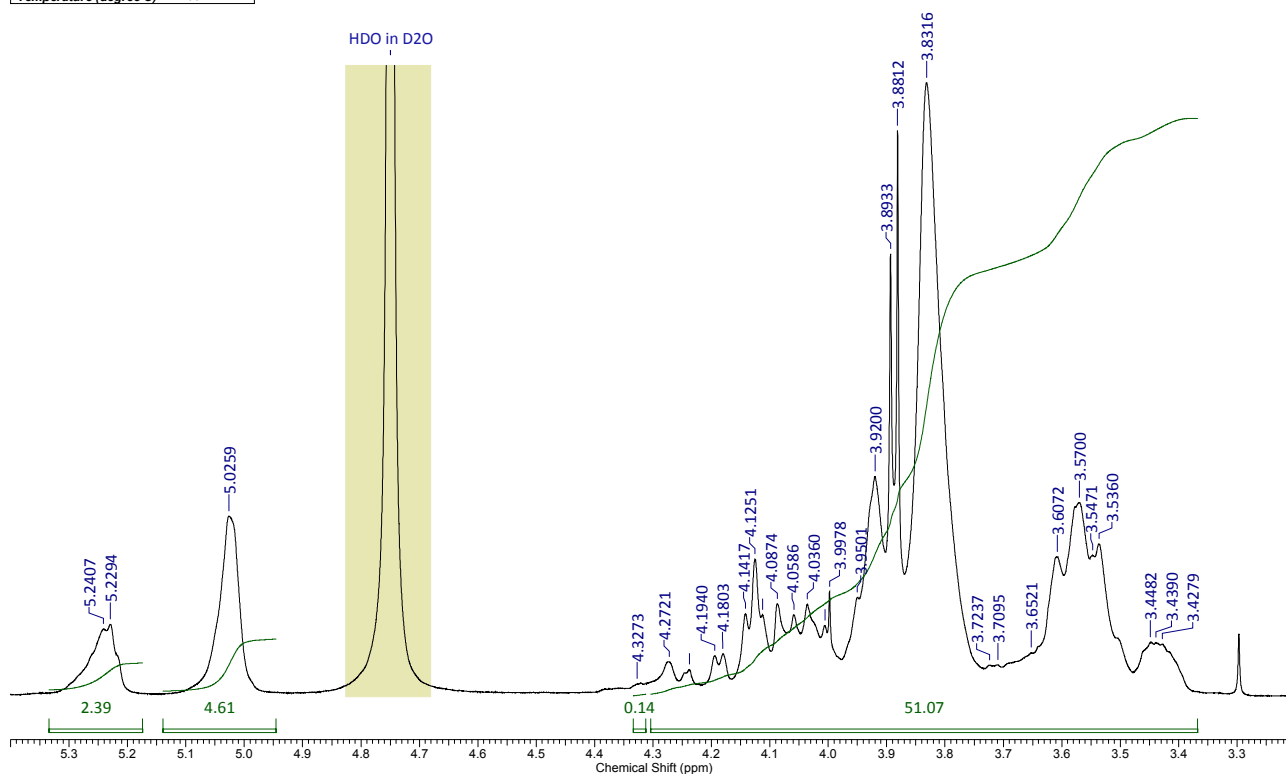


Figure S 8: Proton NMR spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 3 in Table 1),  $DS \approx 4.4-4.6$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059		Date	03 Oct 2016 09:51:00			
File Name	D:\Docs\1\1\Notebooks\BM_Reactions\NMR\CMB\CD_LJ02\2\ser	Frequency (MHz)	(300.13, 75.47)		Nucleus	(1H, 13C)			
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)	Owner	psm		
Points Count	(512, 256)	Pulse Sequence	hsqcetdgp	Solvent	DMSO	Spectrum Type	HSQC-DEPT		
Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	21.760	Title	CBCD_LJ02 (20mg in 0.6ml D2O)_HSQC_03102016_2032				

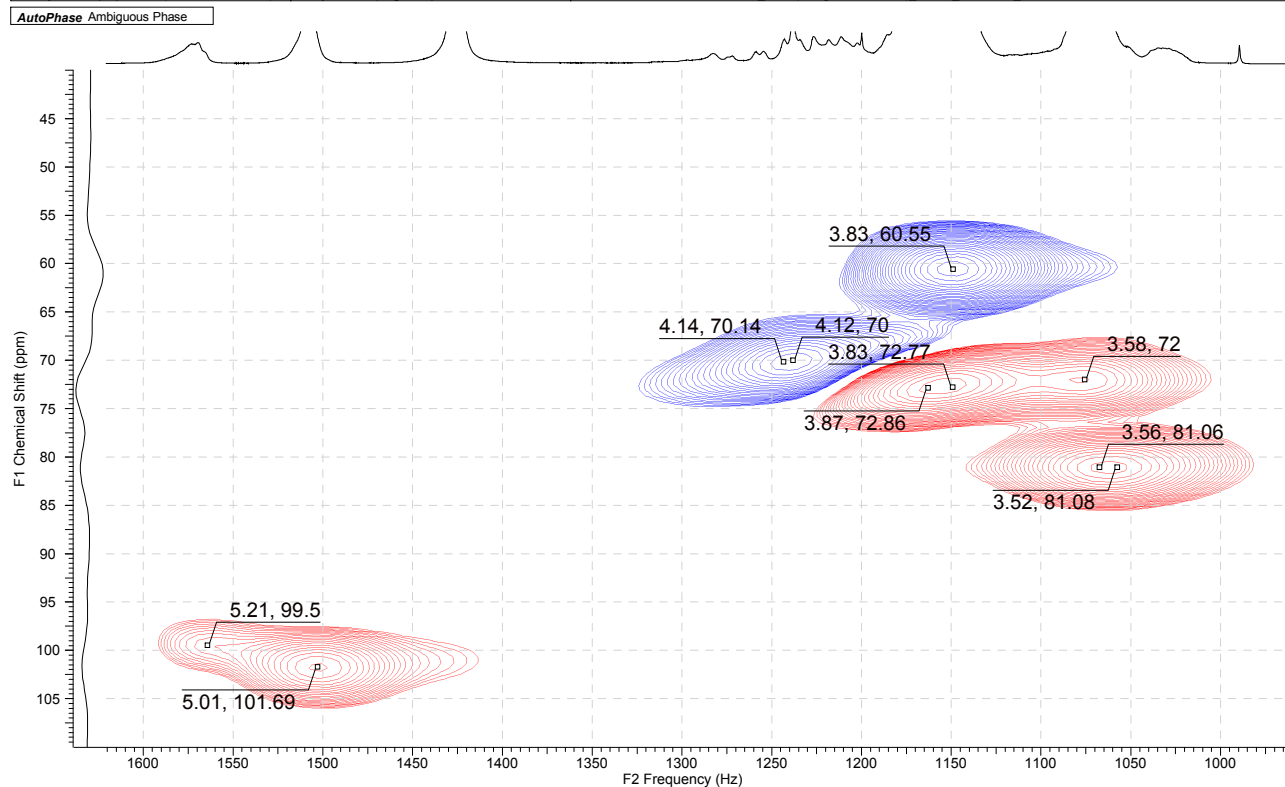


Figure S 9: HSQC-DEPT spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 3 in Table 1),  $DS \approx 4.4-4.6$



Acquisition Time (sec)	3.6438	Comment	CMBCD-LJ03 1H D2O 041016 2036 RG=50	Date	04 Oct 2016 15:34:56
Date Stamp	04 Oct 2016 15:34:56	File Name	D:\Docs\1\1\Notebooks\BM_Reactions\NMR\CMBCD_LJ03\11fid	Nucleus	1H
Frequency (MHz)	300.13	Points Count	131072	Number of Transients	64
Owner	root	Pulse Sequence	zg	Origin	spect
Solvent	CDCI3	Spectrum Offset (Hz)	1095.7936	Receiver Gain	50.80
		Spectrum Type	STANDARD	SW(cyclical) (Hz)	4496.40
				Sweep Width (Hz)	4496.37
				Temperature (degree C)	22.660

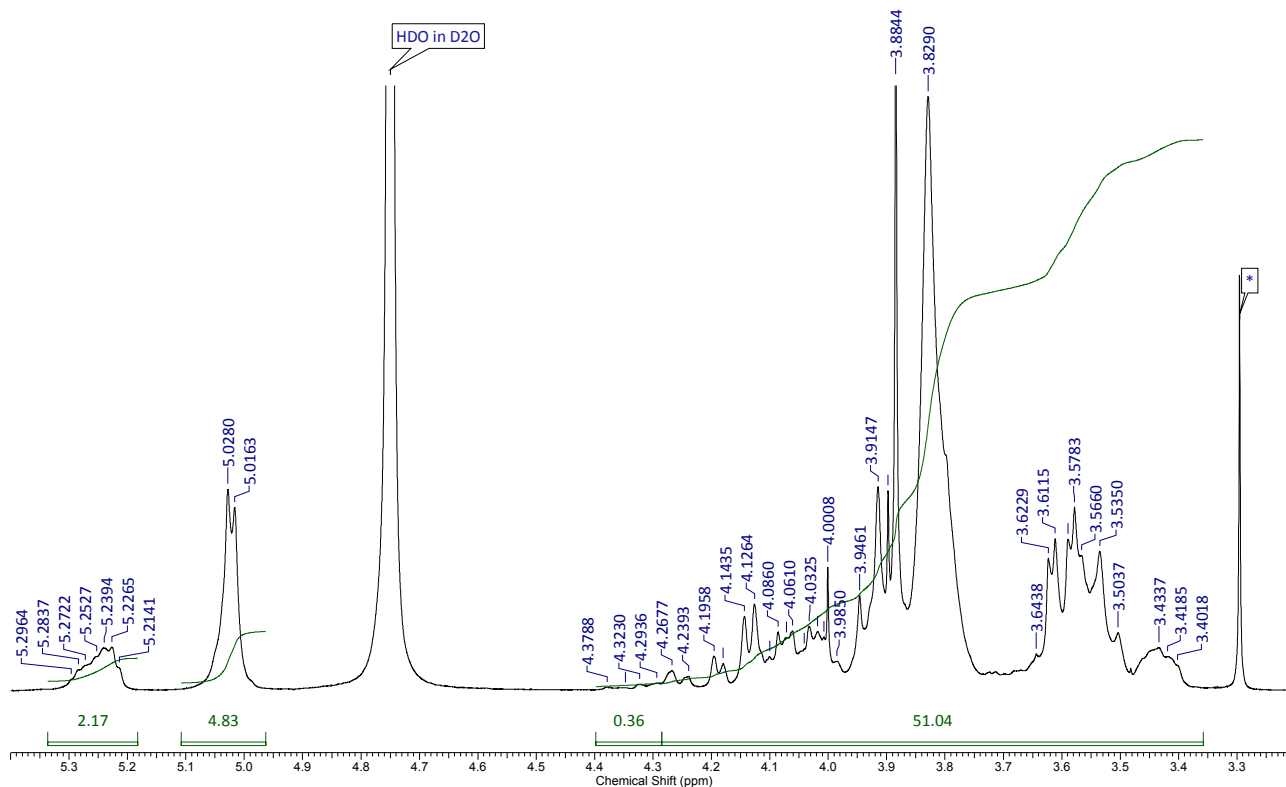


Figure S 10: Proton NMR spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 4 in Table 1),  $DS \approx 4.4-4.6$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	04 Oct 2016 16:28:22
File Name	D:\Docs\1\1\Notebooks\BM_Reactions\NMR\CMBCD_LJ03\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(512, 256)	Pulse Sequence	hsqcetdtp	Solvent	DMSO
Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	22.760	Title	CMBCD-LJ03 HSQC D2O 041016 2036 RG=50

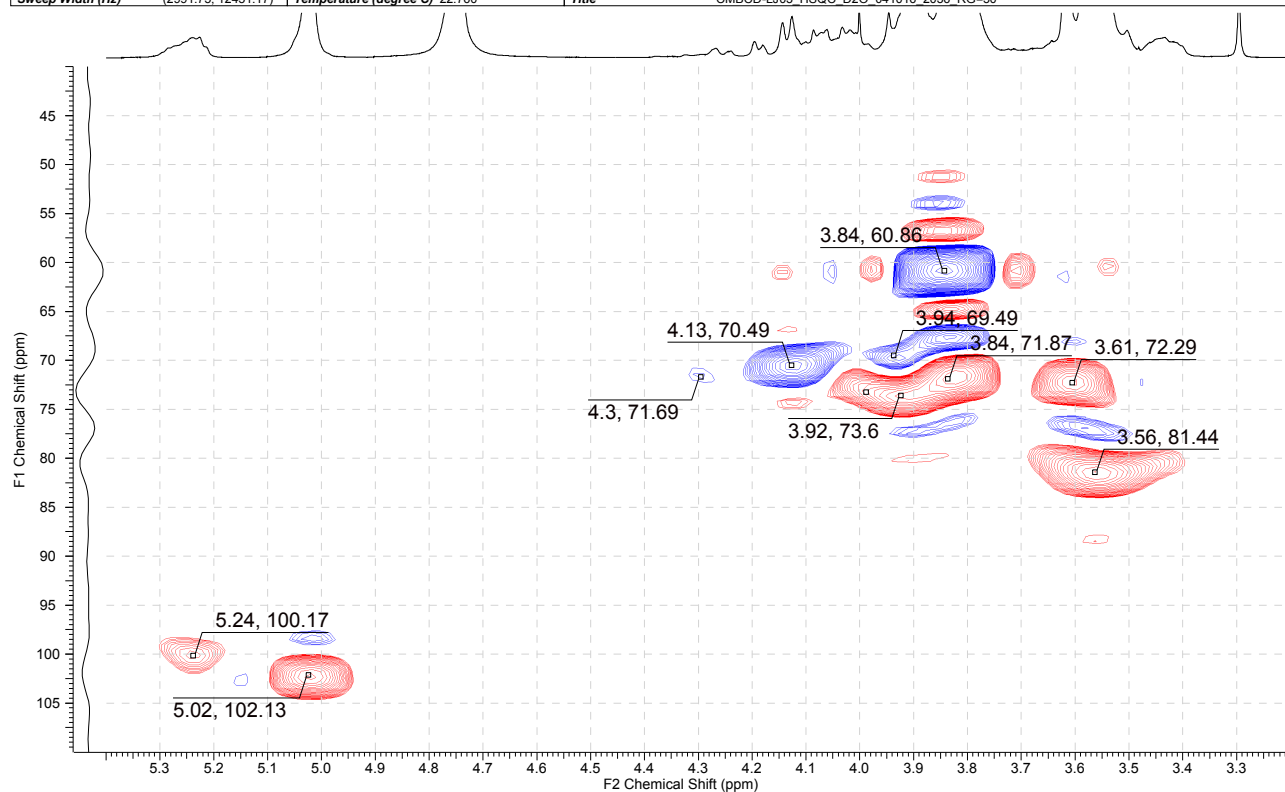
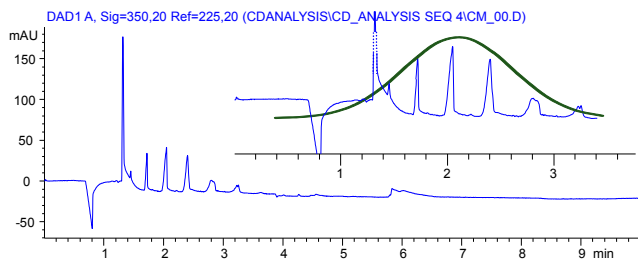
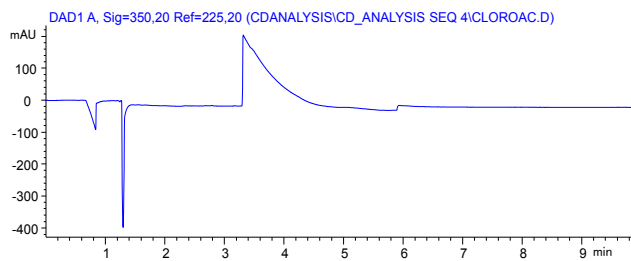


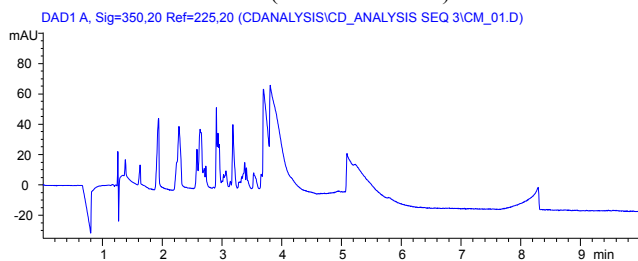
Figure S 11: HSQC-DEPT spectrum of carboxymethylated  $\beta$ CD ( $2'$ ) prepared in ball mill (item 4 in Table 1),  $DS \approx 4.4-4.6$



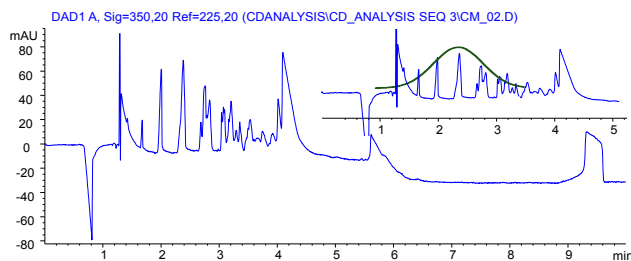
Carboxymethylated  $\beta$ CD (2) DS 4.3-4.5, synthesized in solution (item 1 in Table 1)



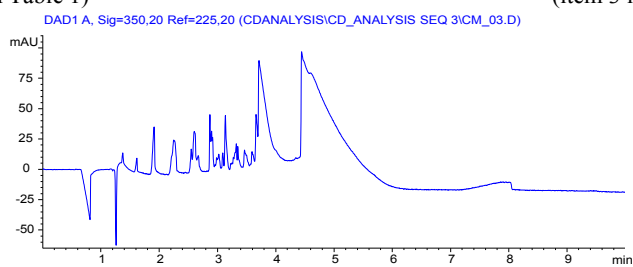
Chloroacetic acid sodium salt



Carboxymethylated  $\beta$ CD (2') DS 4.3-4.6, synthesis in ball mill (item 2 in Table 1)



Carboxymethylated  $\beta$ CD (2') DS 4.4-4.6, synthesis in ball mill (item 3 in Table 1)



Carboxymethylated  $\beta$ CD (2') DS 4.4-4.6, synthesis in ball mill (item 4 in Table 1)

Figure S 12: Capillary electropherogram of carboxymethylated  $\beta$ CDs

Acquisition Time (sec)	3.6438	Comment	CE_LJ00SOL_1H_D2O_131016_2057_rg=90	Date	13 Oct 2016 15:17:52
Date Stamp	13 Oct 2016 15:17:52	File Name	E:\docs\1\molecules\Anah\NMR\CE_LJ00SOL\1\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	262144	Pulse Sequence	zg	Original Points Count	16384
Spectrum Offset (Hz)	1096.8828	Spectrum Type	STANDARD	Receiver Gain	90.50
				SW(cyclical) (Hz)	4496.40
				Solvent	DEUTERIUM OXIDE
				Sweep Width (Hz)	4496.39
				Temperature (degree C)	20.160

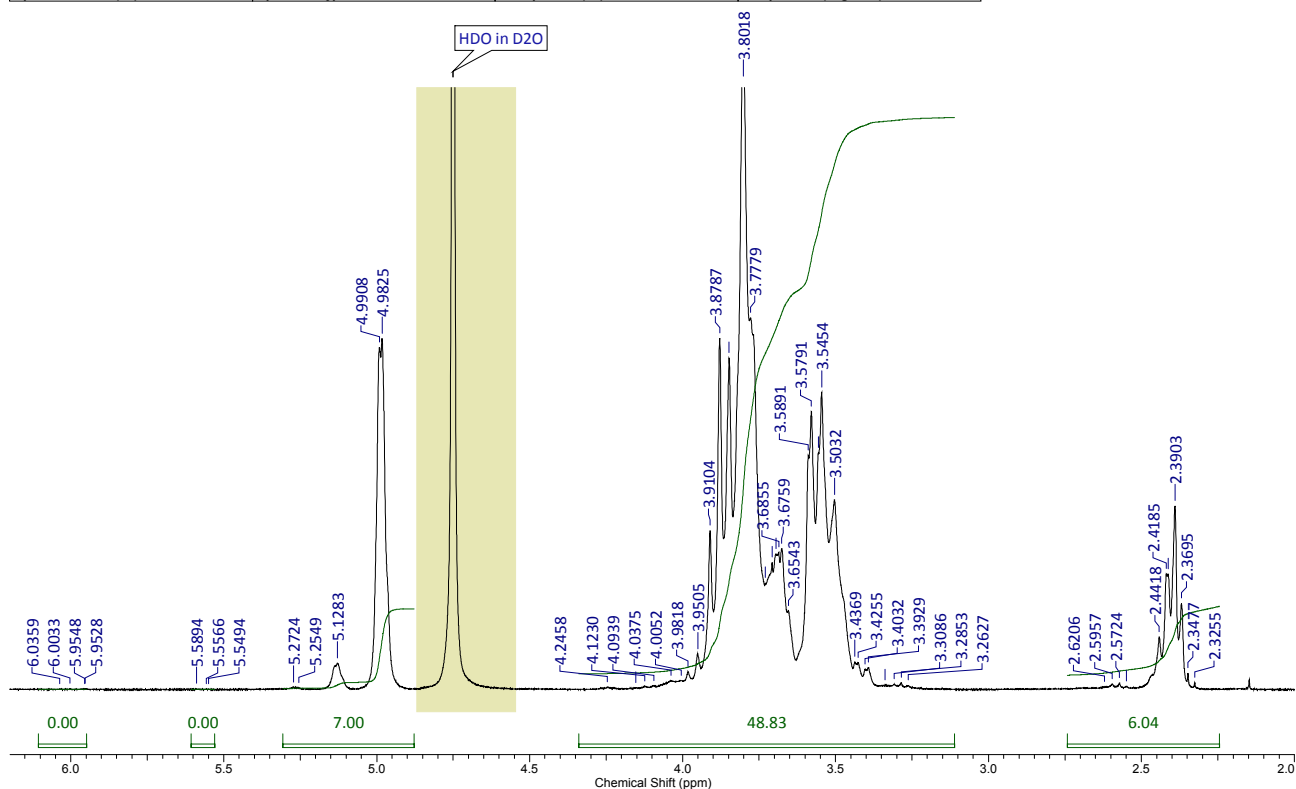


Figure S 13: Proton NMR spectrum of carboxyethylated  $\beta$ CD (3) prepared in solution (item 8 in Table 1),  $DS \approx 3.0-3.4$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	13 Oct 2016 16:11:42
File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\CE_LJ00SOL\2\iser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(1024, 512)	Pulse Sequence	hsqcetgtp	Solvent	DMSO
Sweep Width (Hz)	(2994.67, 12475.59)	Temperature (degree C)	20.260	Title	CE_LJ00SOL_HSQC_D2O_131016_2057_rg=90

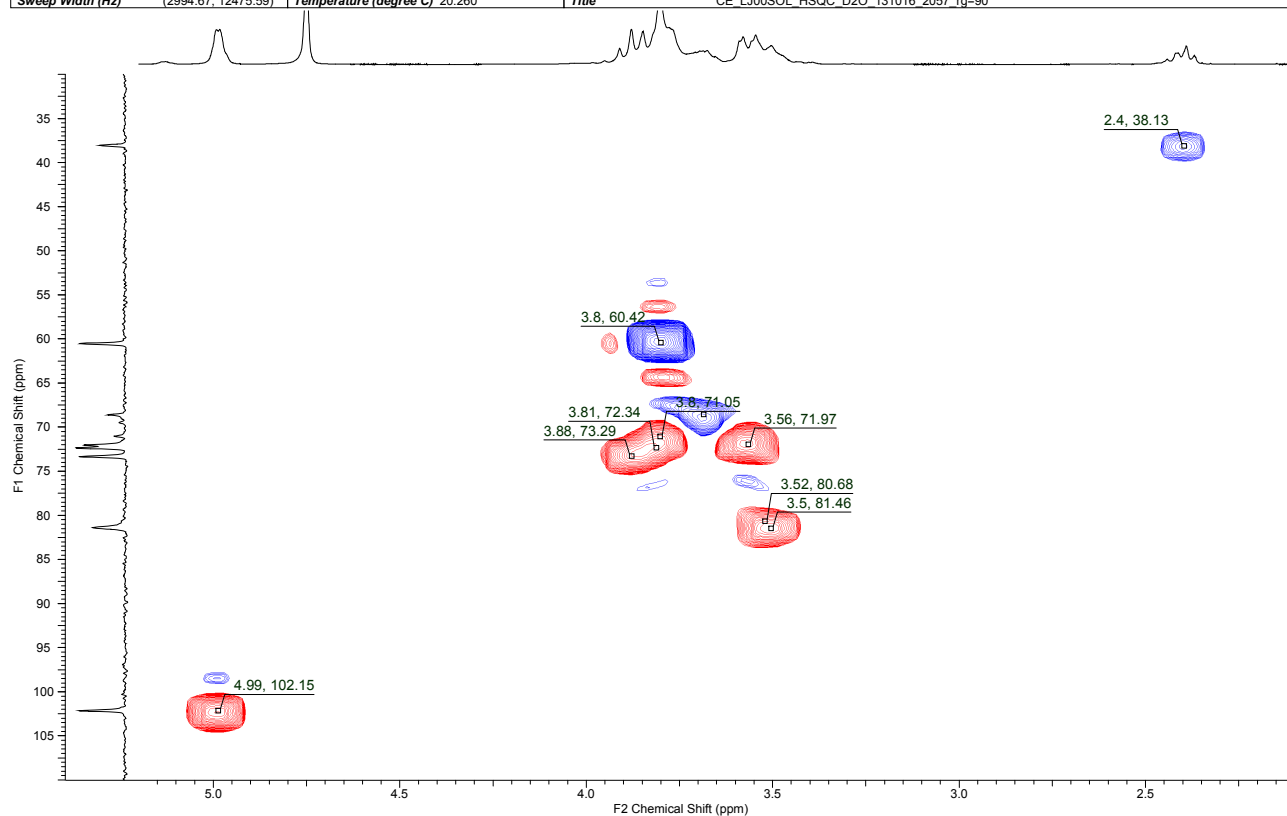


Figure S 14: HSQC-DEPT spectrum of carboxyethylated  $\beta$ CD (3) prepared in solution (item 8 in Table 1),  $DS \approx 3.0-3.4$

Acquisition Time (sec)	1.8088	Comment	CEbCD_LJ00sol (700 ul D2O)	Date	20 Jan 2017 16:10:56
Date Stamp	20 Jan 2017 16:10:56	File Name	E:\Documents\11\Molecules\Anal\CarbonNMR\CEbCD_LJ00SOL\1.tif		
Frequency (MHz)	75.47	Nucleus	13C	Number of Transients	480
Owner	root	Points Count	262144	Pulse Sequence	zgpg.save.txt
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	8301.4463	Receiver Gain	13004.00
Temperature (degree C)	20.960	Spectrum Type	STANDARD	SW(cyclical) (Hz)	18115.87
				Sweep Width (Hz)	18115.87

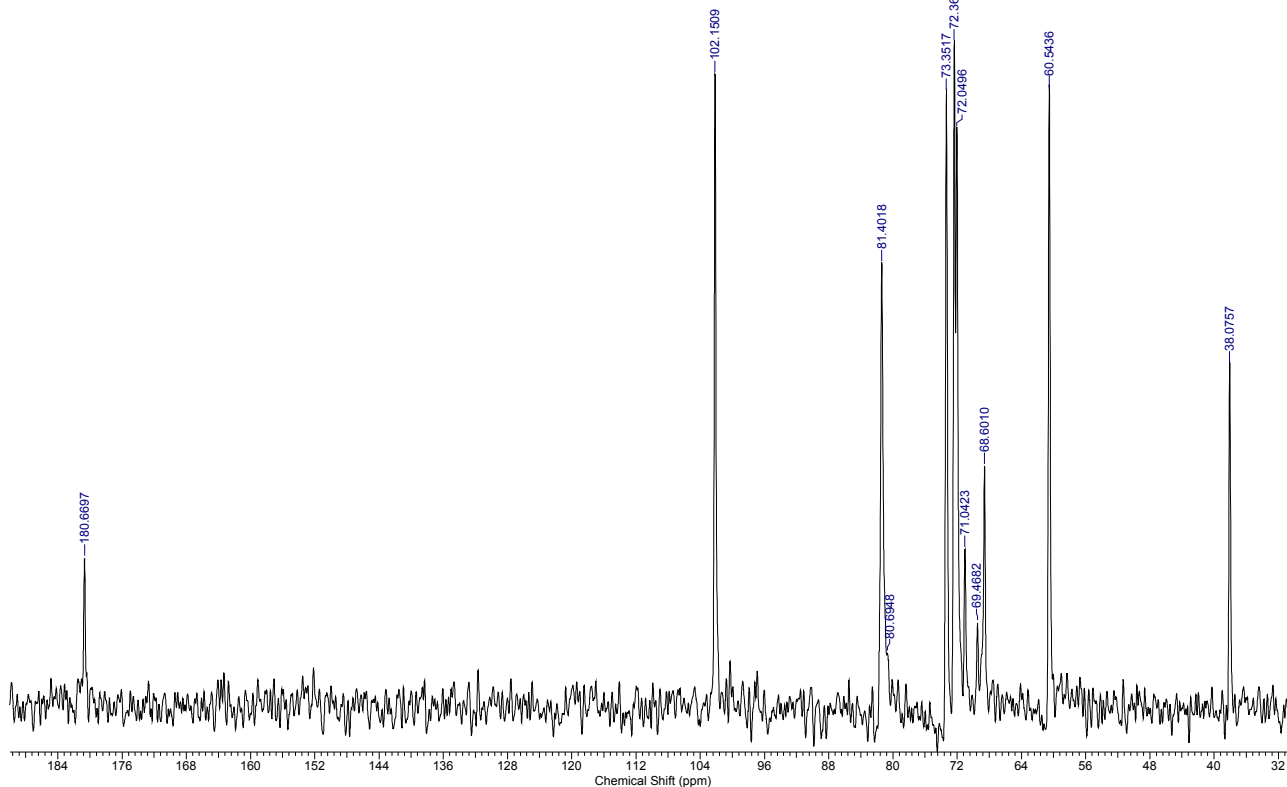


Figure S 15:  $^{13}\text{C}$ -NMR spectrum of carboxyethylated  $\beta\text{CD}$  (**3**) prepared in solution (item 8 in Table 1), DS  $\approx 3.0$ -3.4

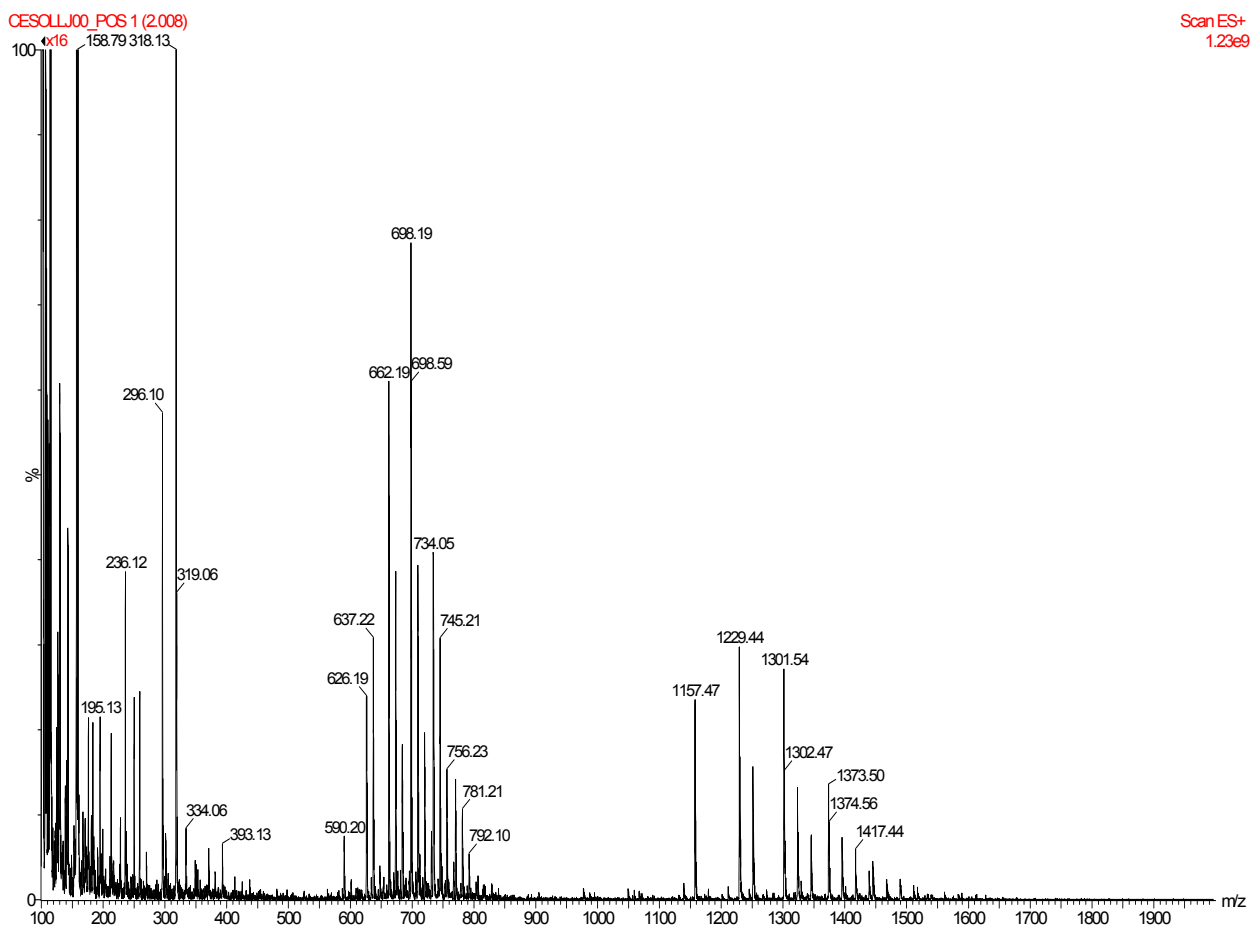
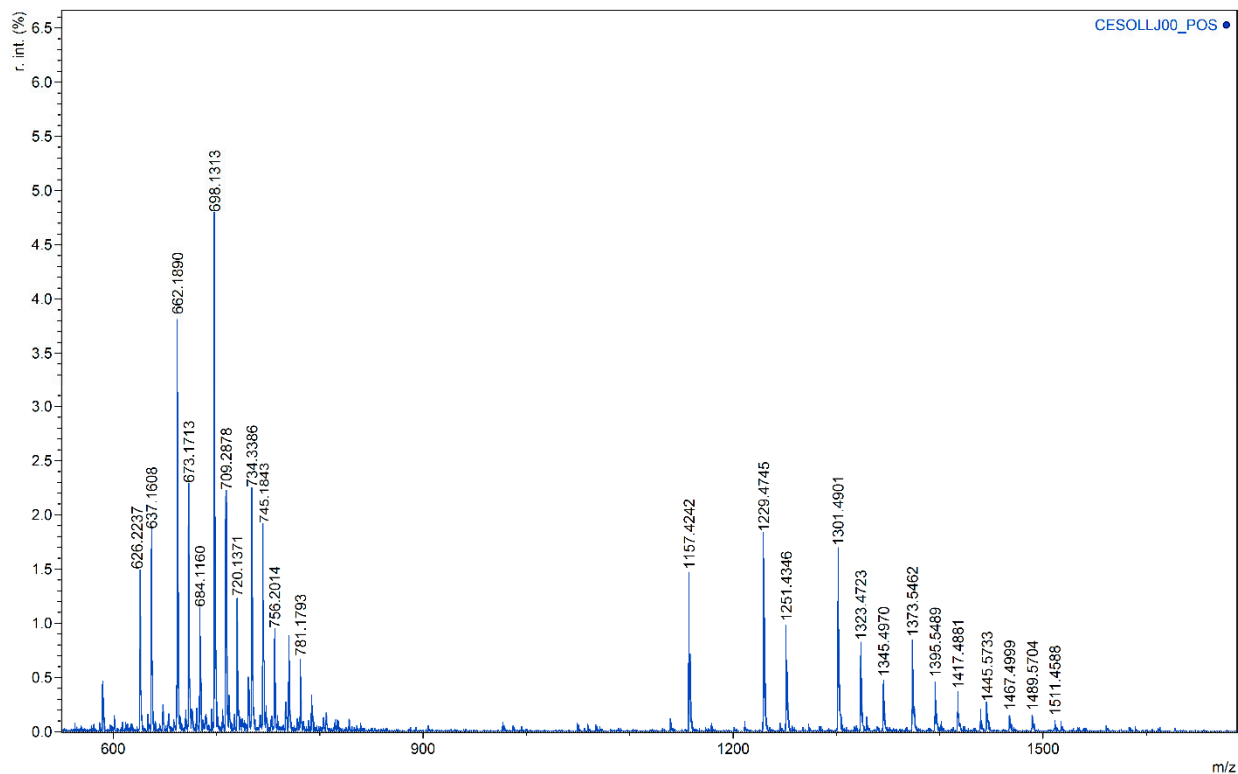


Figure S 16: ESI+ mass spectrum of carboxyethylated  $\beta\text{CD}$  (**3**) prepared in solution (item 8 in Table 1), DS  $\approx 3.0$ -3.4



Meas. m/z	Calc. m/z	$\delta$ (Da)	Z	Annotation	Formula
626.2237	626.1847	0.0390	2	(+1 CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
637.1608	637.1756	-0.0148	2	(+1 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>0</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
662.1890	662.1952	-0.0063	2	(+2 CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
673.1713	673.1862	-0.0149	2	(+1 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
684.1160	684.1772	-0.0611	2	(+2 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>0</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
698.1313	698.2058	-0.0745	2	(+3 CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>3</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
709.2878	709.1968	0.0911	2	(+2 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
720.1371	720.1877	-0.0507	2	(+1 CH <sub>2</sub> CH <sub>2</sub> COOH + 2 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
734.3386	734.2164	0.1223	2	(+4 CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>4</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
745.1843	745.2073	-0.0230	2	(+3 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>3</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
756.2014	756.1983	0.0031	2	(+2 CH <sub>2</sub> CH <sub>2</sub> COOH + 2 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
770.2052	770.2269	-0.0217	2	+5 CH <sub>2</sub> CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>5</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
781.1793	781.2179	-0.0386	2	(+4 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>4</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
792.1803	792.2089	-0.0286	2	(+3 CH <sub>2</sub> CH <sub>2</sub> COOH + 2 CH <sub>2</sub> CH <sub>2</sub> COONa) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>3</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
1157.4242	1157.3590	0.0652	1	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> (BCD)	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na
1229.4745	1229.3801	0.0944	1	+1 CH <sub>2</sub> CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
1251.4346	1251.3621	0.0726	1	+1 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>0</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
1301.4901	1301.4012	0.0888	1	+2 CH <sub>2</sub> CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
1323.4723	1323.3832	0.0891	1	+1 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
1345.4970	1345.3651	0.1319	1	+2 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>0</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
1373.5462	1373.4224	0.1239	1	+3 CH <sub>2</sub> CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>3</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
1395.5489	1395.4043	0.1446	1	+2 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
1417.4881	1417.3863	0.1019	1	+1 CH <sub>2</sub> CH <sub>2</sub> COOH + 2 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
1445.5733	1445.4435	0.1298	1	+4 CH <sub>2</sub> CH <sub>2</sub> COOH	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>4</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>0</sub>
1467.4999	1467.4254	0.0744	1	+3 CH <sub>2</sub> CH <sub>2</sub> COOH + 1 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>3</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>1</sub>
1489.5704	1489.4074	0.1630	1	+2 CH <sub>2</sub> CH <sub>2</sub> COOH + 2 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>2</sub>
1511.4588	1511.3893	0.0694	1	+1 CH <sub>2</sub> CH <sub>2</sub> COOH + 3 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COOH) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> COONa) <sub>3</sub>

Figure S 17: Peak identification in ESI+ mass spectrum of carboxyethylated  $\beta$ CD (3) prepared in solution (item 8 in Table 1), DS  $\approx$  3.0-3.4

Acquisition Time (sec)	3.6438	Comment	CE_LJ01A_1H_D2O_281016_2081_rg=80	Date	28 Oct 2016 11:12:32
Date Stamp	28 Oct 2016 11:12:32	File Name	E:\doc\!!molecules\Anal\NMR\CE_LJ01\3\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	524288	Pulse Sequence	zg	Original Points Count	16384
Spectrum Offset (Hz)	1096.4756	Spectrum Type	STANDARD	SW(cyclical) (Hz)	4496.40
				Receiver Gain	80.60
				Sweep Width (Hz)	4496.39
				Temperature (degree C)	20.660

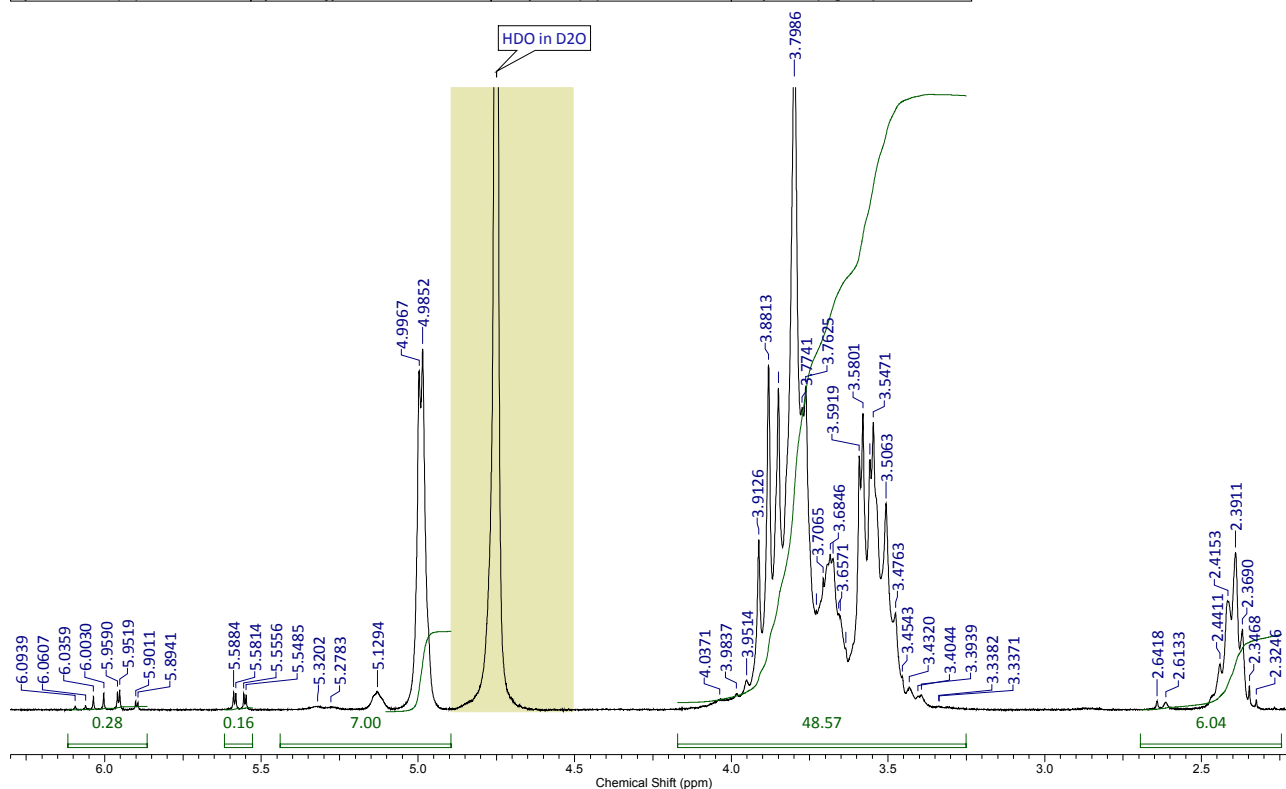


Figure S 18: Proton NMR spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 5 in Table 1),  $DS \approx 3.0-3.3$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	28 Oct 2016 11:06:14
File Name	E:\doc\!!molecules\Anal\NMR\CE_LJ01\4\user	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(4096, 128)	Pulse Sequence	hsqcetgqp	Solvent	DMSO
Sweep Width (Hz)	(2996.87, 12402.34)	Temperature (degree C)	20.660	Spectrum Type	HSQC-DEPT
				Title	CE_LJ01A_HSQC_D2O_281016_2081_rg=80

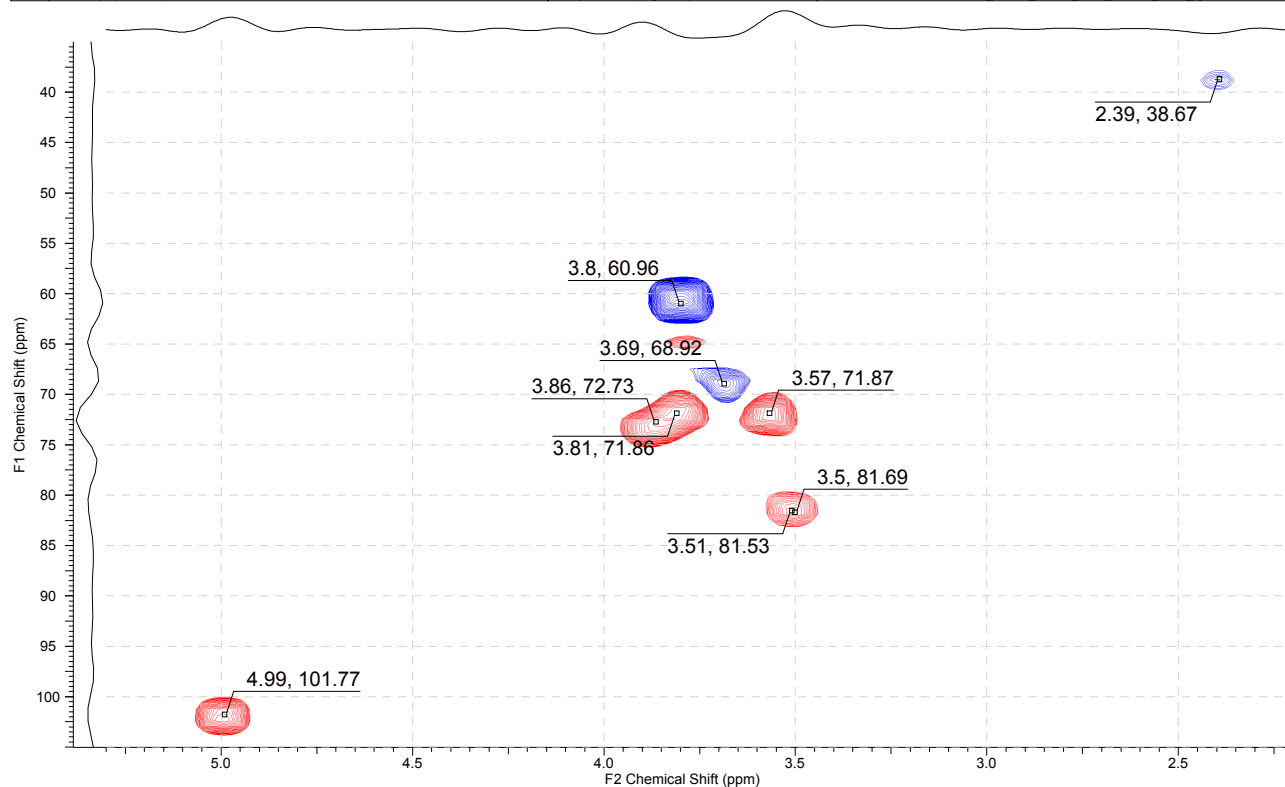


Figure S 19: HSQC-DEPT spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 5 in Table 1),  $DS \approx 3.0-3.3$

Acquisition Time (sec)	3.6438	Comment	CE_LJ02A_1H_D2O_261016_2079_rg=80	Date	26 Oct 2016 11:46:40
Date Stamp	26 Oct 2016 11:46:40	File Name	E:\doc\1\molecules\Anal\NMR\CE_LJ02\3\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	524288	Pulse Sequence	zg	Original Points Count	16384
Spectrum Offset (Hz)	1096.3091	Spectrum Type	STANDARD	SW(cyclical) (Hz)	4496.40
				Receiver Gain	90.50
				Sweep Width (Hz)	4496.39
				Temperature (degree C)	20.460

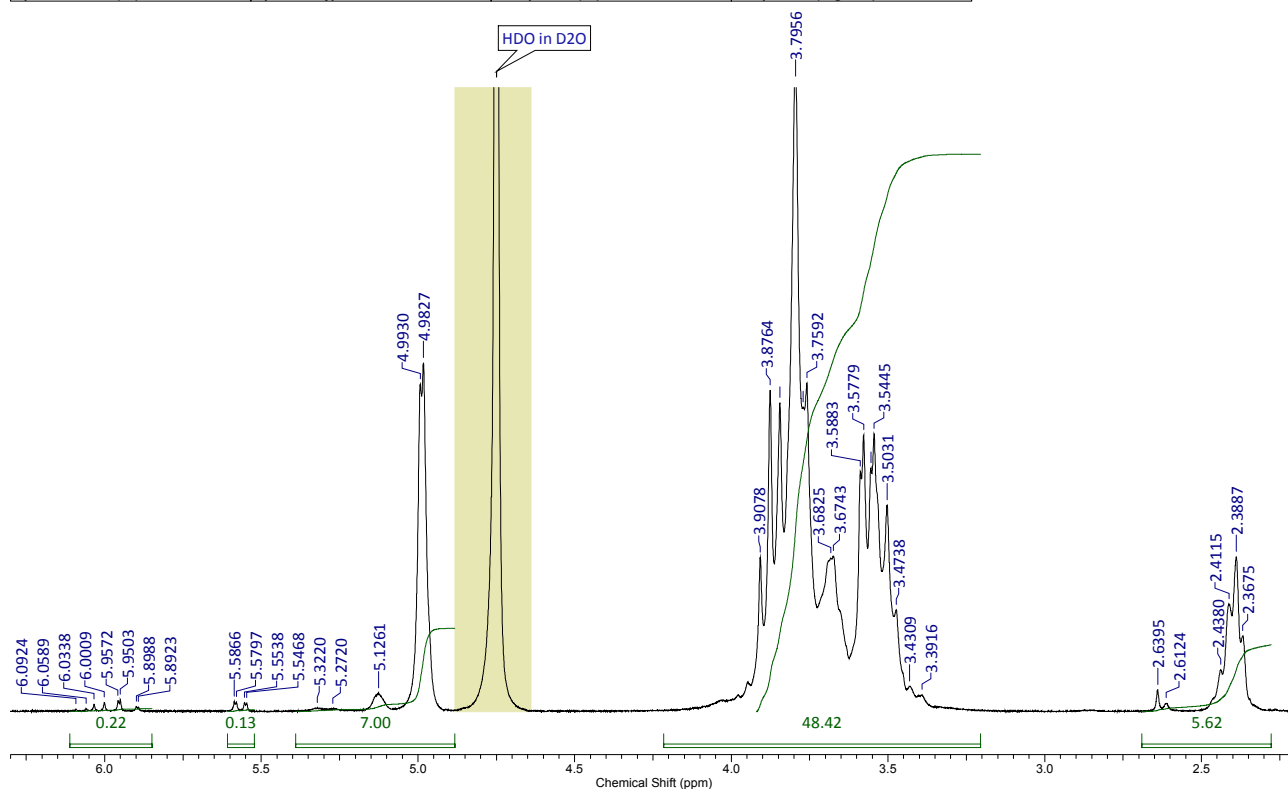


Figure S 20: Proton NMR spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 6 in Table 1),  $DS \approx 2.8-3.2$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	26 Oct 2016 11:40:52
File Name	E:\doc\1\molecules\Anal\NMR\CE_LJ02\4\user	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(256, 512)	Pulse Sequence	hsqcetdgp	Solvent	DMSO
Sweep Width (Hz)	(2885.89, 12475.59)	Temperature (degree C)	20.560	Spectrum Type	HSQC-DEPT
				Title	CE_LJ02A_HSQC_D2O_261016_2079_rg=80

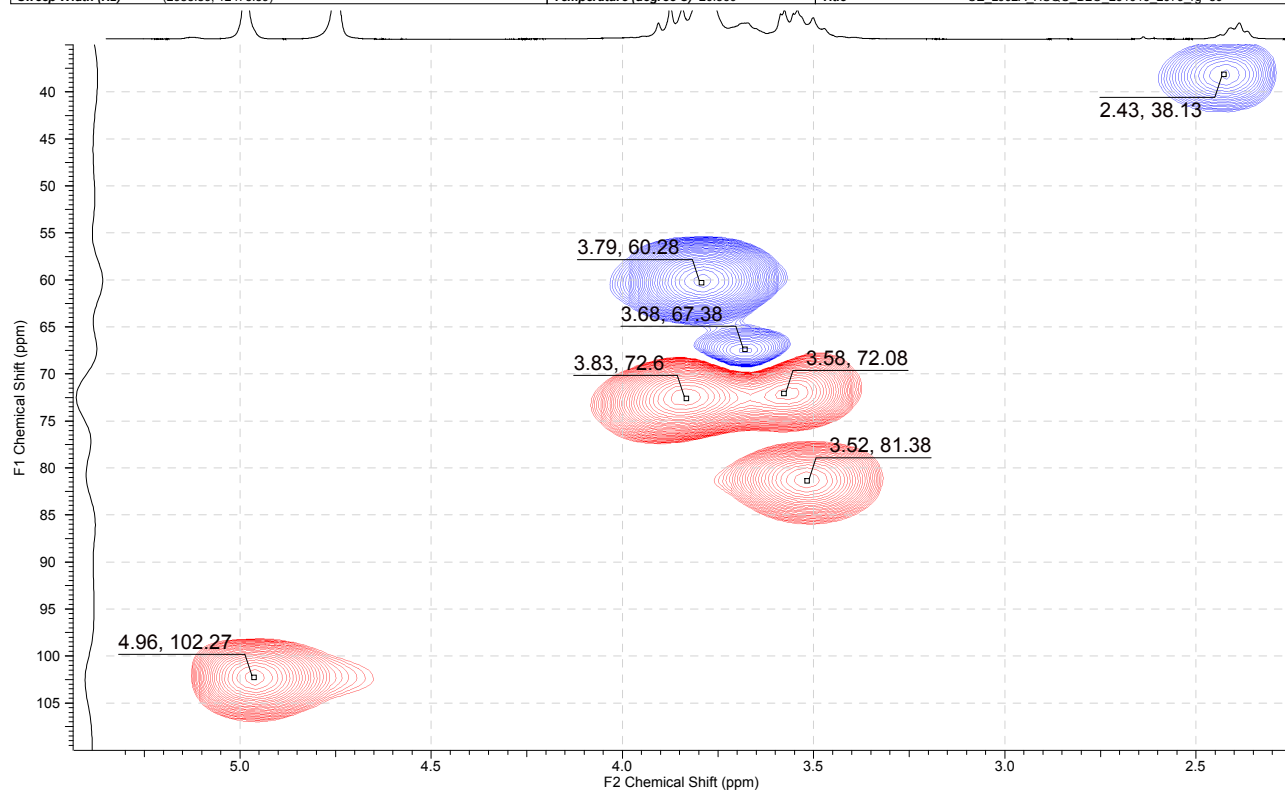


Figure S 21: HSQC-DEPT spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 6 in Table 1),  $DS \approx 2.8-3.2$

Acquisition Time (sec)	3.6438	Comment	CE_LJ03B_1H_D2O_281016_2082_rg=60	Date	28 Oct 2016 13:41:52
Date Stamp	28 Oct 2016 13:41:52	File Name	E:\doc\1\molecules\Anal\NMR\ICE_LJ03\3\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	524288	Pulse Sequence	zg	Original Points Count	16384
Spectrum Offset (Hz)	1096.4291	Spectrum Type	STANDARD	Receiver Gain	64.00
				SW(cyclical) (Hz)	4496.40
				Solvent	CDCl3
				Sweep Width (Hz)	4496.39
				Temperature (degree C)	20.660

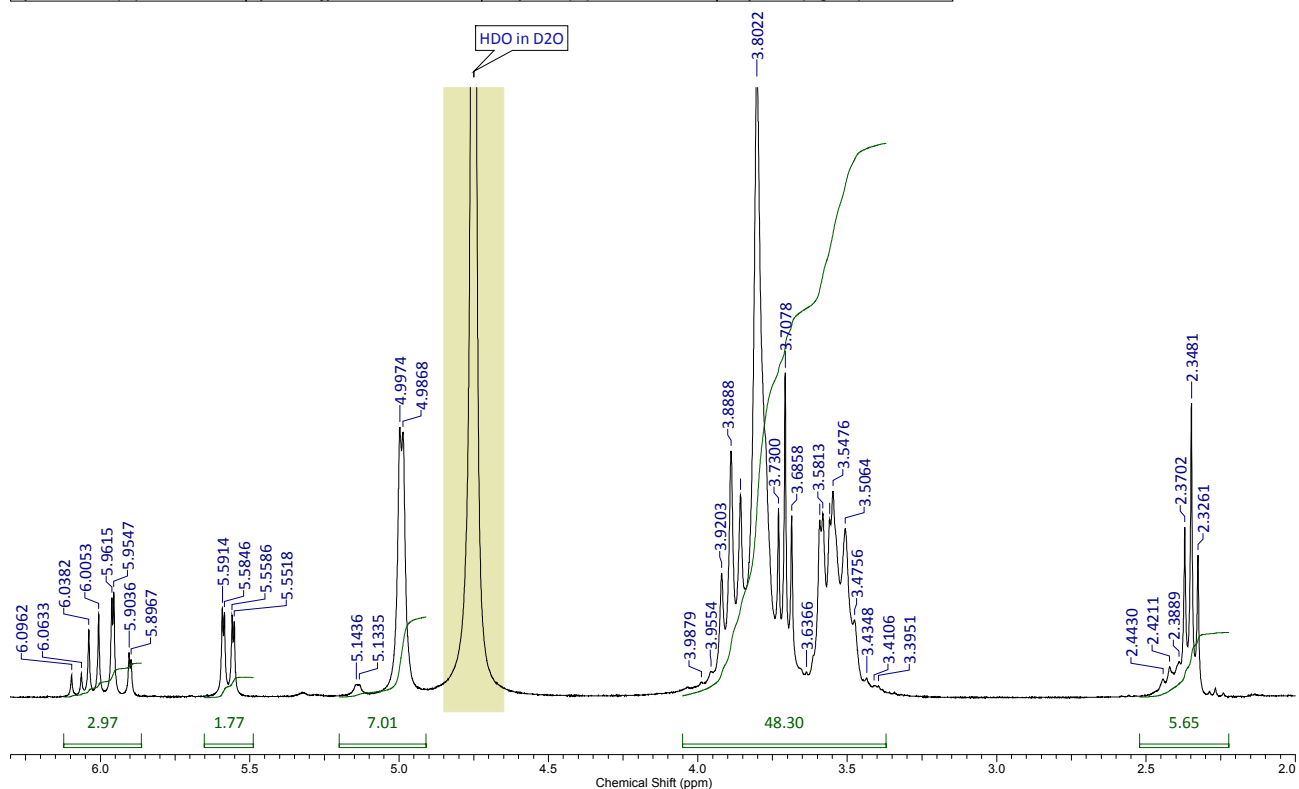


Figure S 22: Proton NMR spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 7 in Table 1),  $DS \approx 2.8-3.1$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	28 Oct 2016 13:36:38
File Name	E:\doc\1\molecules\Anal\NMR\ICE_LJ03\4\iser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(2048, 8192)	Pulse Sequence	hsqcetdgp	Solvent	DMSO
Sweep Width (Hz)	(2996.14, 12498.47)	Temperature (degree C)	20.760	Spectrum Type	HSQC-DEPT
				Title	CE_LJ03B_HSQC_D2O_281016_2082_rg=60

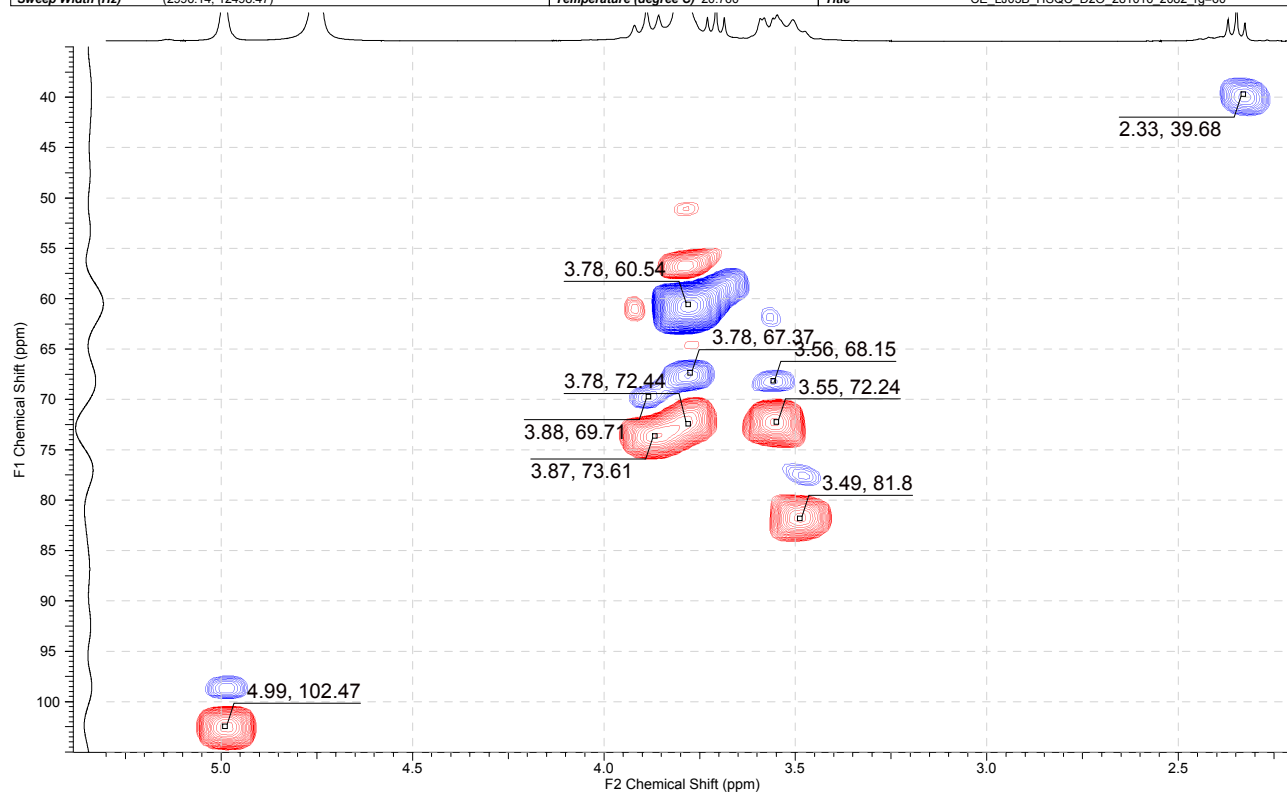


Figure S 23: HSQC-DEPT spectrum of carboxyethylated  $\beta$ CD (3') prepared in ball mill (item 7 in Table 1),  $DS \approx 2.8-3.1$



<b>Title</b>	CEBCD_LJ00sol_H	<b>Origin</b>	Exported PE Spectrum Data File	<b>Owner</b>	Admin
<b>File Name</b>	E:\DOCUMENTS\1\MOLECULES\ANAL\IR2\S\CEBCD_LJ00SOL_H.DX	<b>Date Stamp</b>	17/01/20 13:14:56.00	<b>Date</b>	27 Jan 2017 16:14:32
<b>Technique</b>	Infrared	<b>Instrument</b>	PERKIN-ELMER 1005 IR	<b>Spectral Region</b>	IR
<b>Y Axis</b>	Transmittance	<b>Spectrum Range</b>	400.2000 - 3999.2361	<b>Points Count</b>	3733
		<b>Data Spacing</b>	0.9644	<b>X Axis</b>	Wavenumber (cm-1)

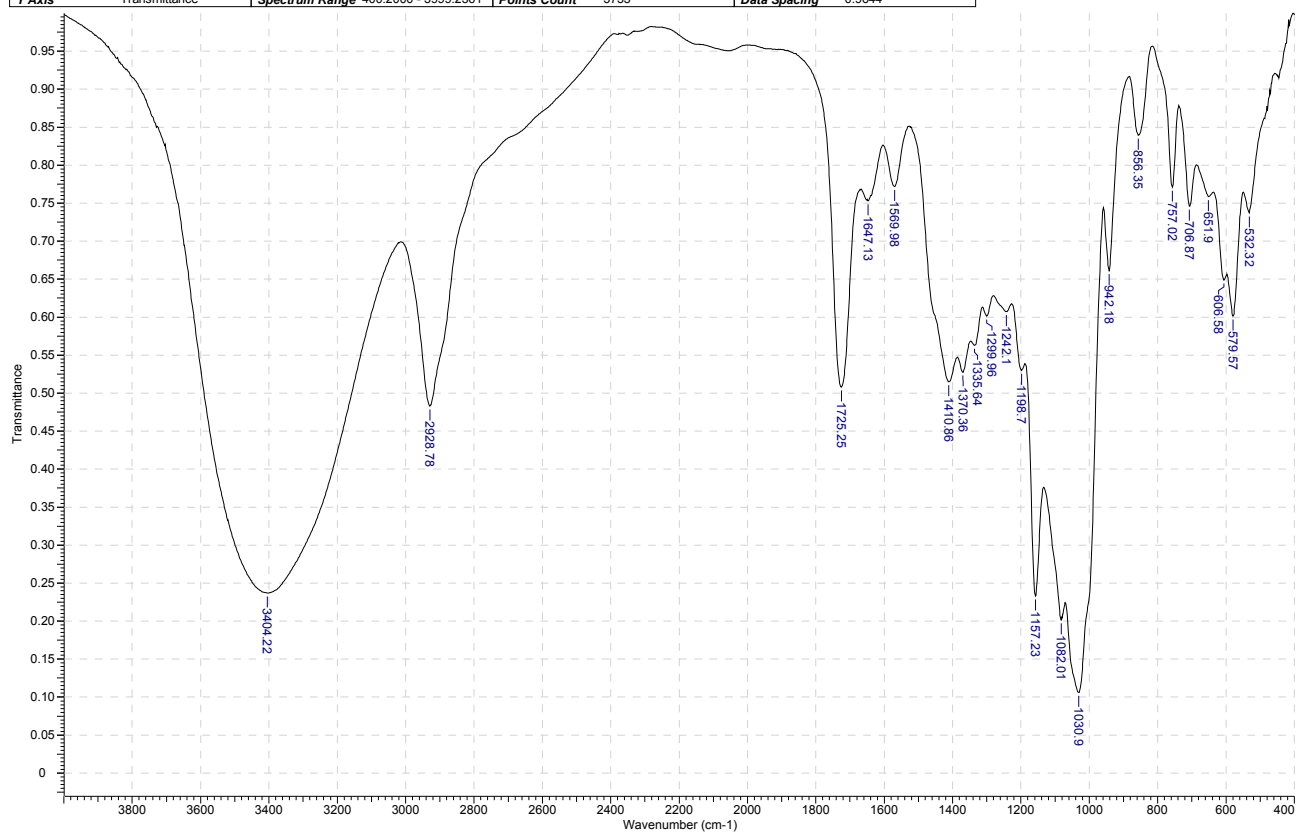


Figure S 24: IR spectrum of carboxylethylated  $\beta$ CD (3) prepared in solution (item 8 in Table 1) as acid, DS  $\approx$  3.0-3.4

<b>Comment</b>	CEBCD_LJ00sol_Na	<b>File Name</b>	E:\DOCUMENTS\1\MOLECULES\ANAL\IR2\S\CEBCD_LJ00SOL_NA.SPC	<b>Date Stamp</b>	27/01/2017 13:01:00
<b>Date</b>	20 Jan 2017 13:01:00	<b>Technique</b>	Infrared	<b>Spectral Region</b>	IR
<b>Spectrum Range</b>	400.2250 - 3999.2360	<b>Points Count</b>	3733	<b>Data Spacing</b>	0.9644
				<b>X Axis</b>	Wavenumber (cm-1)
				<b>Y Axis</b>	Transmittance

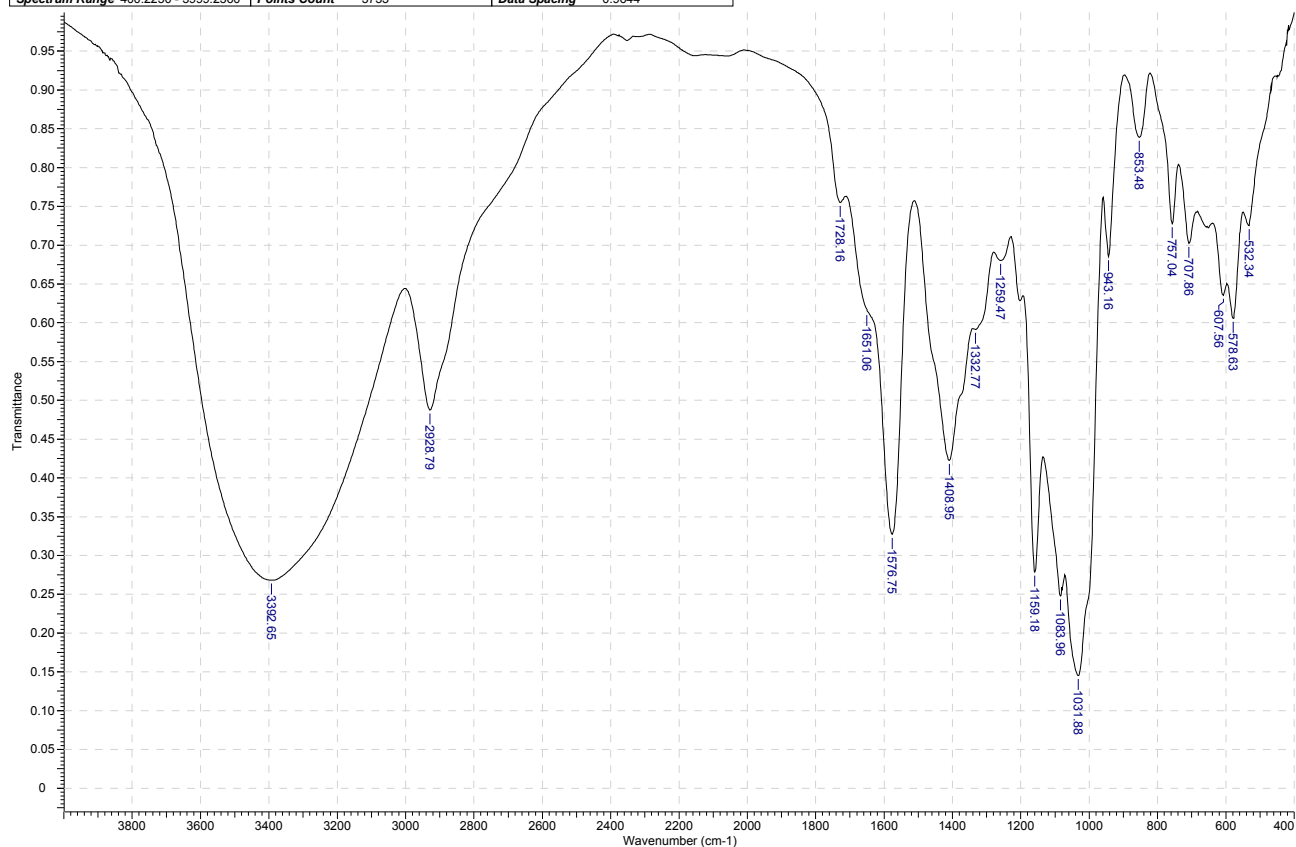


Figure S 25: IR spectrum of carboxylethylated  $\beta$ CD (3) prepared in solution (item 8 in Table 1) as sodium salt, DS  $\approx$  3.0-3.4

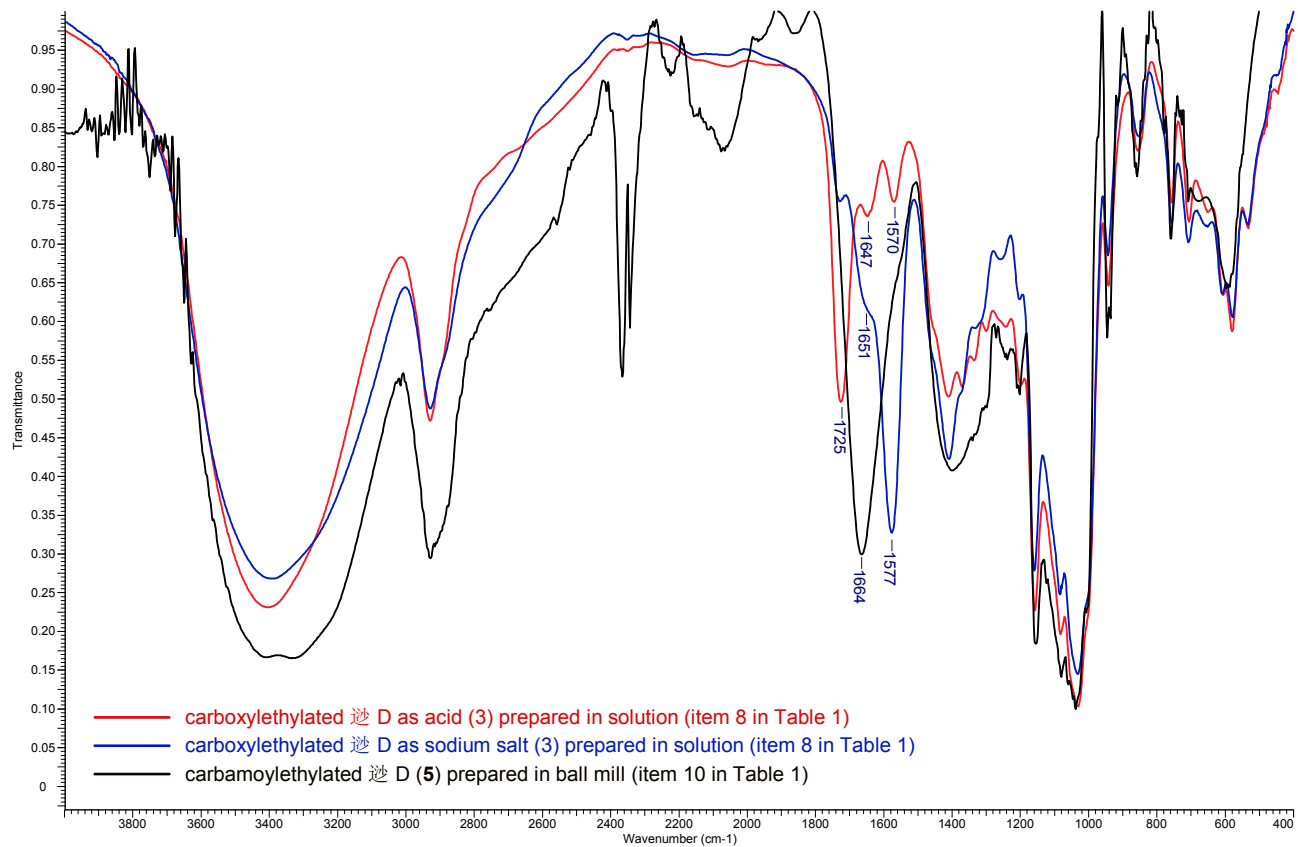


Figure S 26: Overlay of IR spectrum of carboxylethylated  $\beta$ CD acid & salt form (3) prepared in solution (item 8 in Table 1), DS  $\approx$  3.0-3.4 and carbamoylethylated  $\beta$ CD prepared in ball mill (item 10 in table 1)

Acquisition Time (sec)	3.6438	Comment	CAMBCD_LJ01w (30 mg in 0.6ml D2O)_19102016_1H_RG=180		Date	19 Oct 2016 11:40:16	
Date Stamp	19 Oct 2016 11:40:16	File Name	E:\doc\1\molecules\Anal\NMR\CAMBCD_LJ01\7fid		Frequency (MHz)	300.13	
Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Points Count	262144	Pulse Sequence	zg	Receiver Gain	181.00	SW(cyclical) (Hz)	4496.40
Spectrum Offset (Hz)	1096.5278	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.39	Temperature (degree C)	20.860

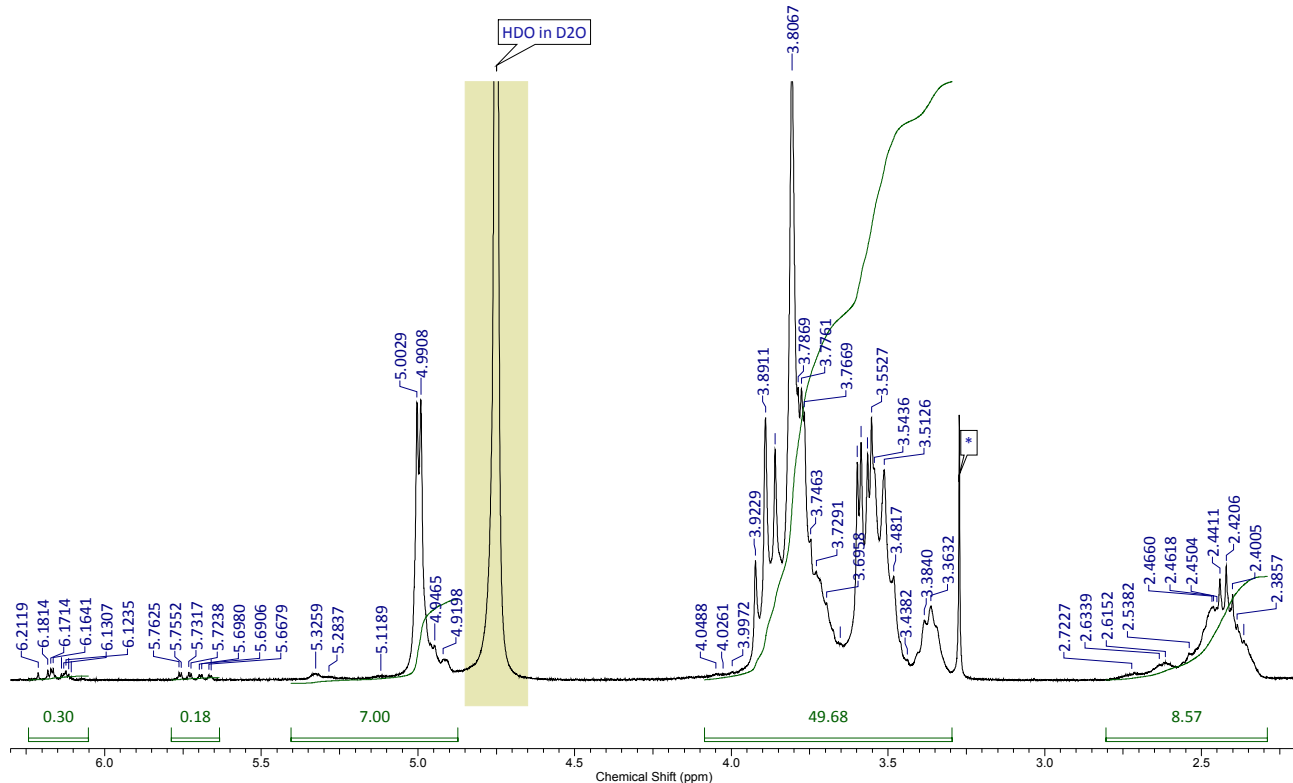


Figure S 27: Proton NMR spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 9 in Table 1),  $DS \approx 3.8-4.3$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059		Date	19 Oct 2016 11:34:02	
File Name	E:\doc\1\molecules\Anal\NMR\CAMBCD_LJ01\8\user	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)		
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)	Owner	psm
Points Count	(256, 256)	Pulse Sequence	hsqcetlap	Solvent	DMSO	Spectrum Type	HSQC-DEPT
Sweep Width (Hz)	(2985.89, 12451.17)	Temperature (degree C)	20.960	Title	CAMBCD_LJ01w (30 mg in 0.6ml D2O)_19102016_HSQC		

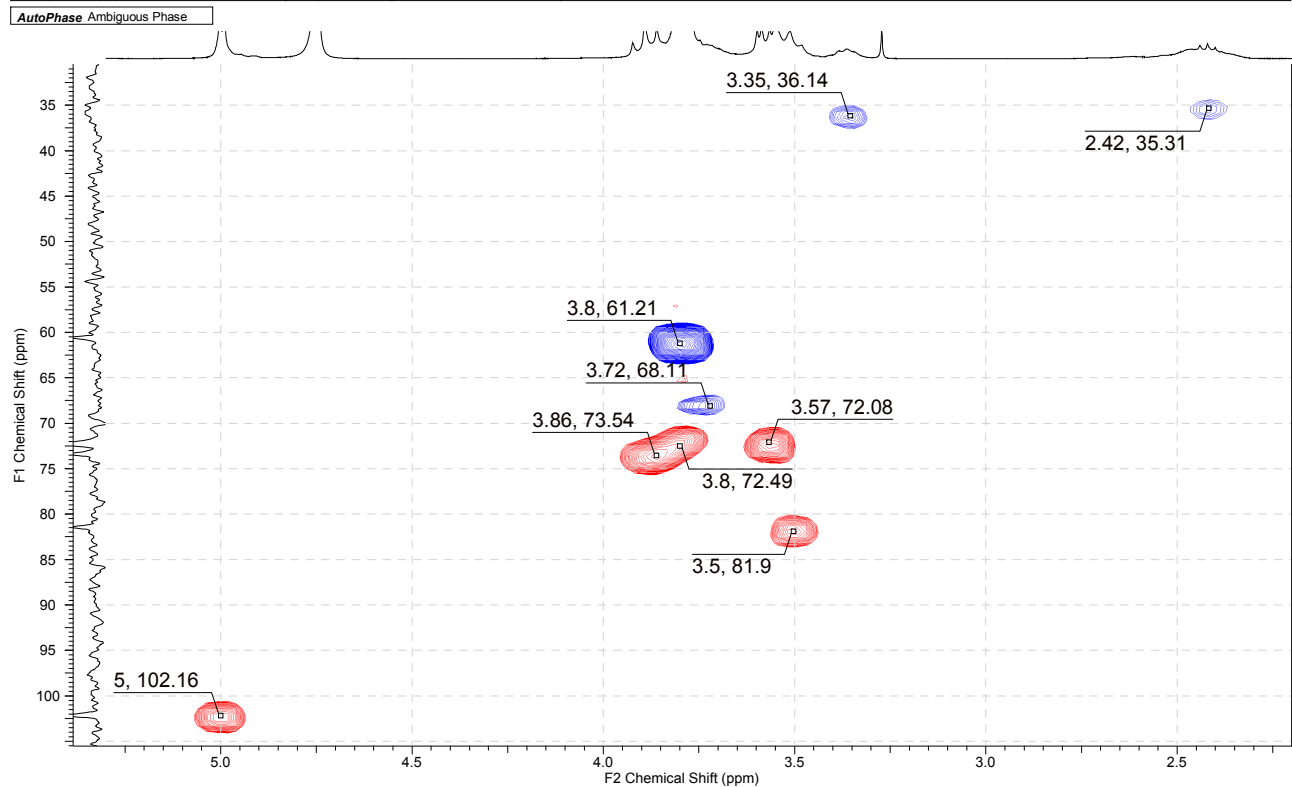


Figure S 28: HSQC-DEPT spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 9 in Table 1),  $DS \approx 3.8-4.3$

Acquisition Time (sec)	1.8088	Comment	CAM1bCD_LJ01 in 700ul D2O	Date	20 Jan 2017 13:01:04		
Date Stamp	20 Jan 2017 13:01:04	File Name	E:\Documents\Anal\CarbonNMR\CAM1bCD_LJ01\1fid				
Frequency (MHz)	75.47	Nucleus	13C	Number of Transients	836	Original Points Count	32768
Owner	psm	Points Count	262144	Pulse Sequence	zgpg.save.txt	Receiver Gain	13004.00
Solvent	CHLOROFORM-d	Spectrum Offset (Hz)	8309.0479	Spectrum Type	STANDARD	SW(cyclical) (Hz)	18115.94
Temperature (degree C)	20.760					Sweep Width (Hz)	18115.87

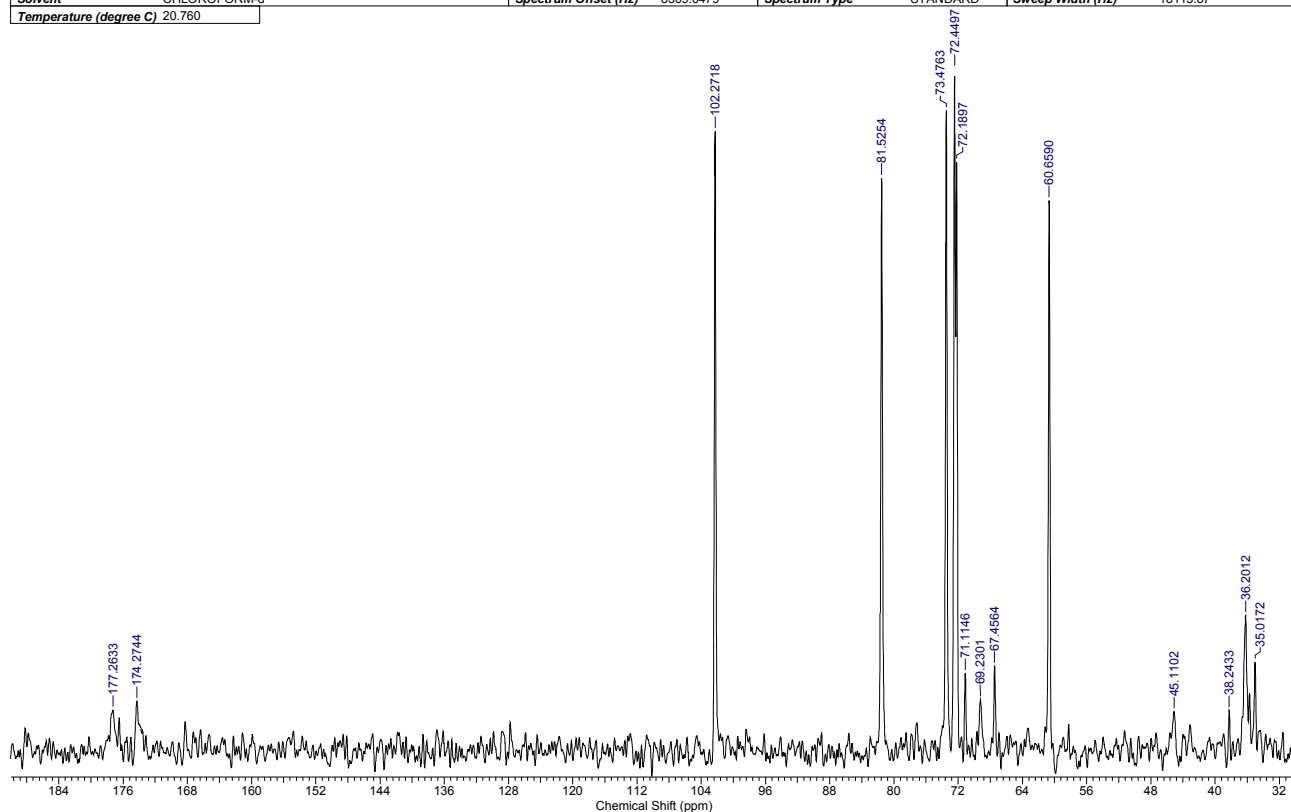


Figure S 29:  $^{13}\text{C}$ -NMR spectrum of carbamoylethylated  $\beta$ CD (**5**) prepared in ball mill (item 9 in Table 1), DS  $\approx$ 3.8-4.3

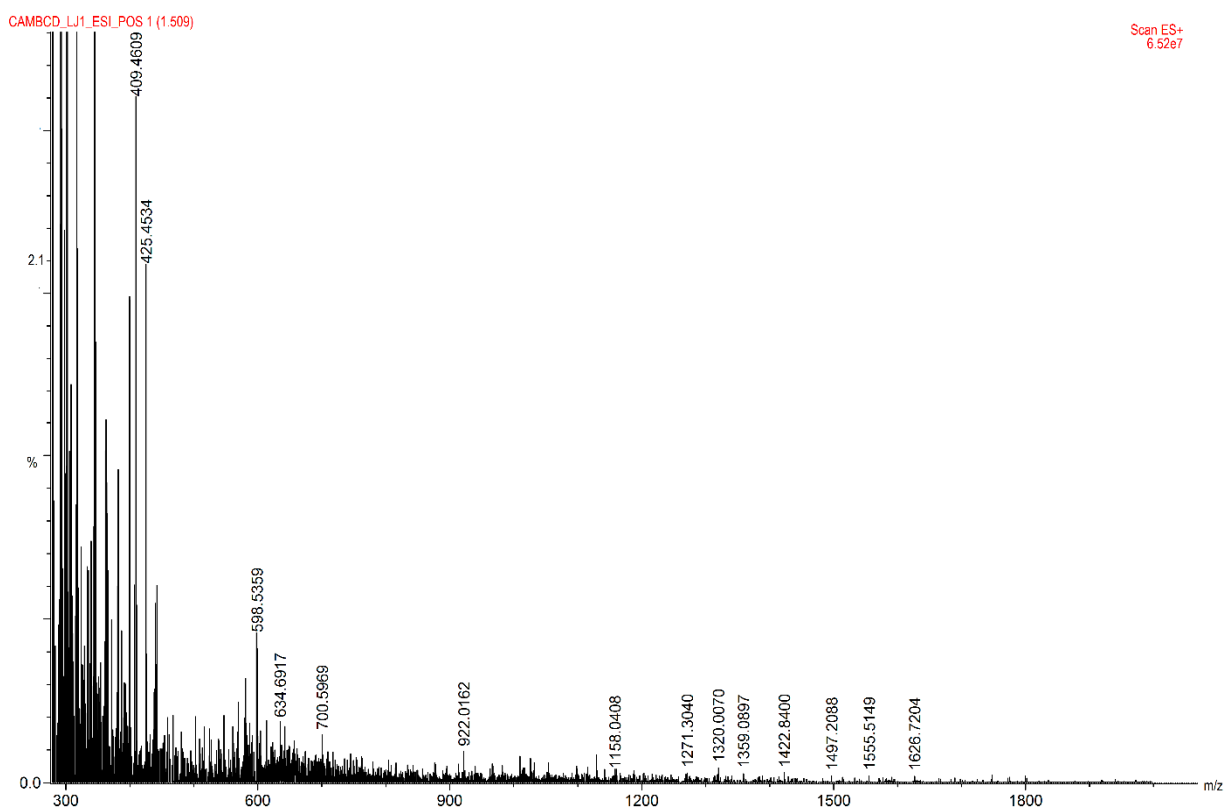
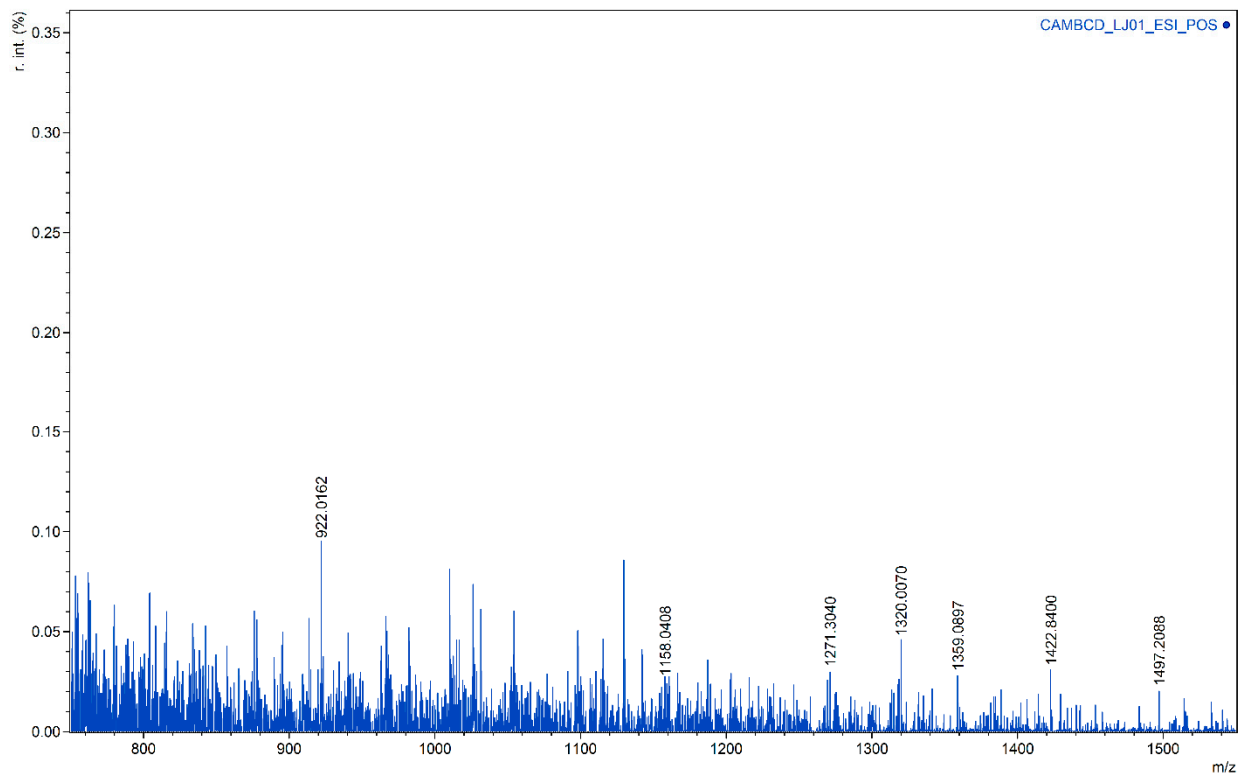


Figure S 30: ESI+ mass spectrum of carbamoylethylated  $\beta$ CD (**5**) prepared in ball mill (item 9 in Table 1), DS  $\approx$ 3.8-4.3



Meas. m/z	Calc. m/z	$\delta$ (Da)	Z	Annotation	Formula
634.6917	635.0289	-0.3372	2	+1 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> +1 HaO	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> C <sub>2</sub> CH <sub>2</sub> COOH) <sub>0</sub> (HCOOH) <sub>0</sub> (H <sub>2</sub> O) <sub>1</sub>
642.6066	643.7032	-1.0966	2	+1 CH <sub>2</sub> CH <sub>2</sub> COONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> C <sub>2</sub> CH <sub>2</sub> COOH) <sub>0</sub> (HCOOH) <sub>0</sub> (H <sub>2</sub> O) <sub>2</sub>
700.5969	700.2153	0.3816	2	+2 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> +1 CH <sub>2</sub> CH <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub> (CH <sub>1</sub> CH <sub>2</sub> CONa) <sub>1</sub>
922.0162	923.7832	-1.7670	2	+5 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> +4 CH <sub>2</sub> HC <sub>2</sub> COONa	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>5</sub> (CH <sub>1</sub> CH <sub>2</sub> CONa) <sub>4</sub>
1158.0408	1157.3590	0.6819	1	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> (bCD)	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na
1271.3040	1271.3907	-0.0867	1	+1 CH <sub>2</sub> CH <sub>2</sub> COOH +1 H <sub>2</sub> O	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>0</sub> (CH <sub>1</sub> C <sub>2</sub> CH <sub>2</sub> COOH) <sub>1</sub> (HCOOH) <sub>0</sub> (H <sub>2</sub> O) <sub>1</sub>

Figure S 31: Peak identification in ESI+ mass spectrum of carbamoylethylated  $\beta$ CD (**5**) prepared in ball mill (item 9 in Table 1), DS  $\approx$ 3.8-4.3

Acquisition Time (sec)	3.6438	Comment	CAMBCD_LJ02_1H_D2O_31102016	Date	31 Oct 2016 11:33:36
Date Stamp	31 Oct 2016 11:33:36	File Name	E:\doc\1\molecules\Anal\NMR\CAMBCD_LJ02\1\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	524288	Pulse Sequence	zg	Receiver Gain	143.70
Spectrum Offset (Hz)	1096.9557	Spectrum Type	STANDARD	SW(cyclical) (Hz)	4496.40
				Temperature (degree C)	19.960
				Owner	root
				Solvent	DEUTERIUM OXIDE

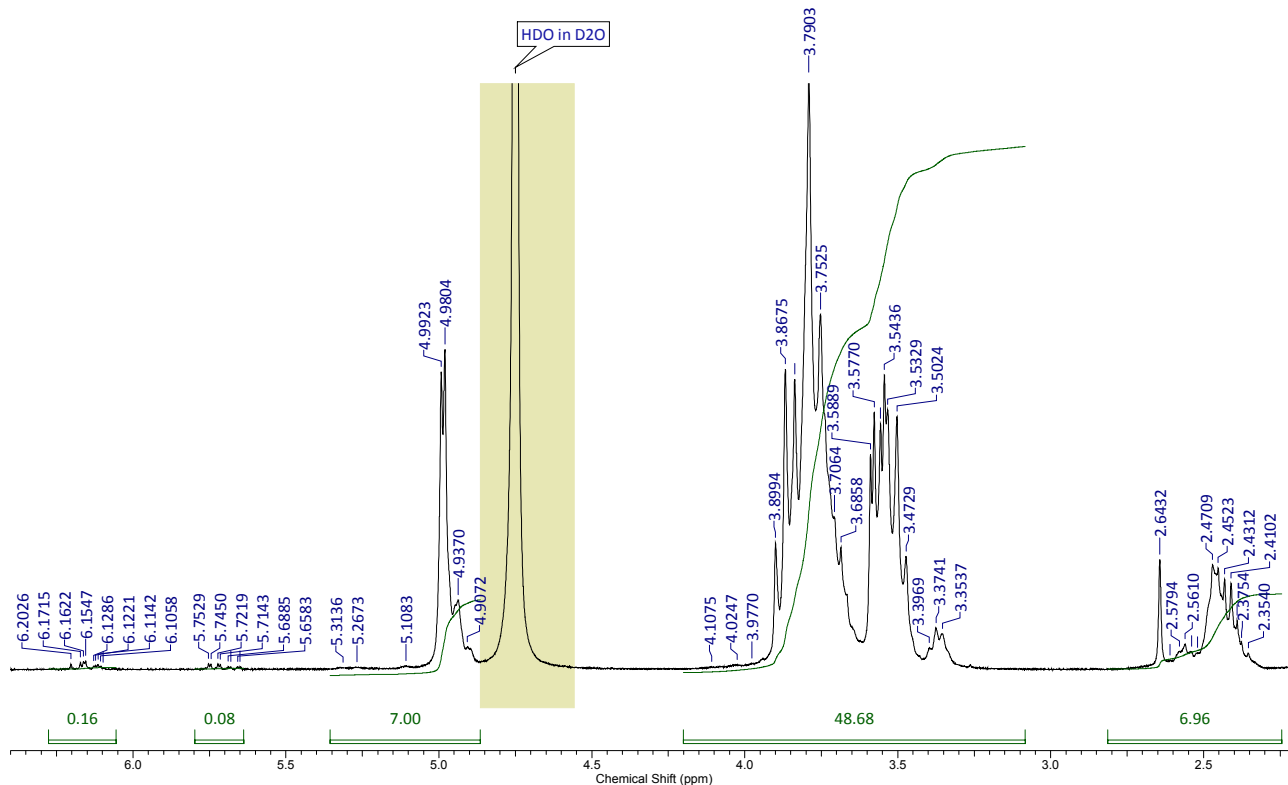


Figure S 32: Proton NMR spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 10 in Table 1), DS  $\approx$ 3.3-3.5

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	31 Oct 2016 12:28:08
File Name	E:\doc\1\molecules\Anal\NMR\CAMBCD_LJ02\2\user	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(256, 1024)	Pulse Sequence	hsgcedetgp	Solvent	DMSO
Sweep Width (Hz)	(2985.89, 12487.79)	Temperature (degree C)	20.060	Spectrum Type	HSQC-DEPT
				Title	CAMBCD_LJ02_HSQC_D2O_31102016

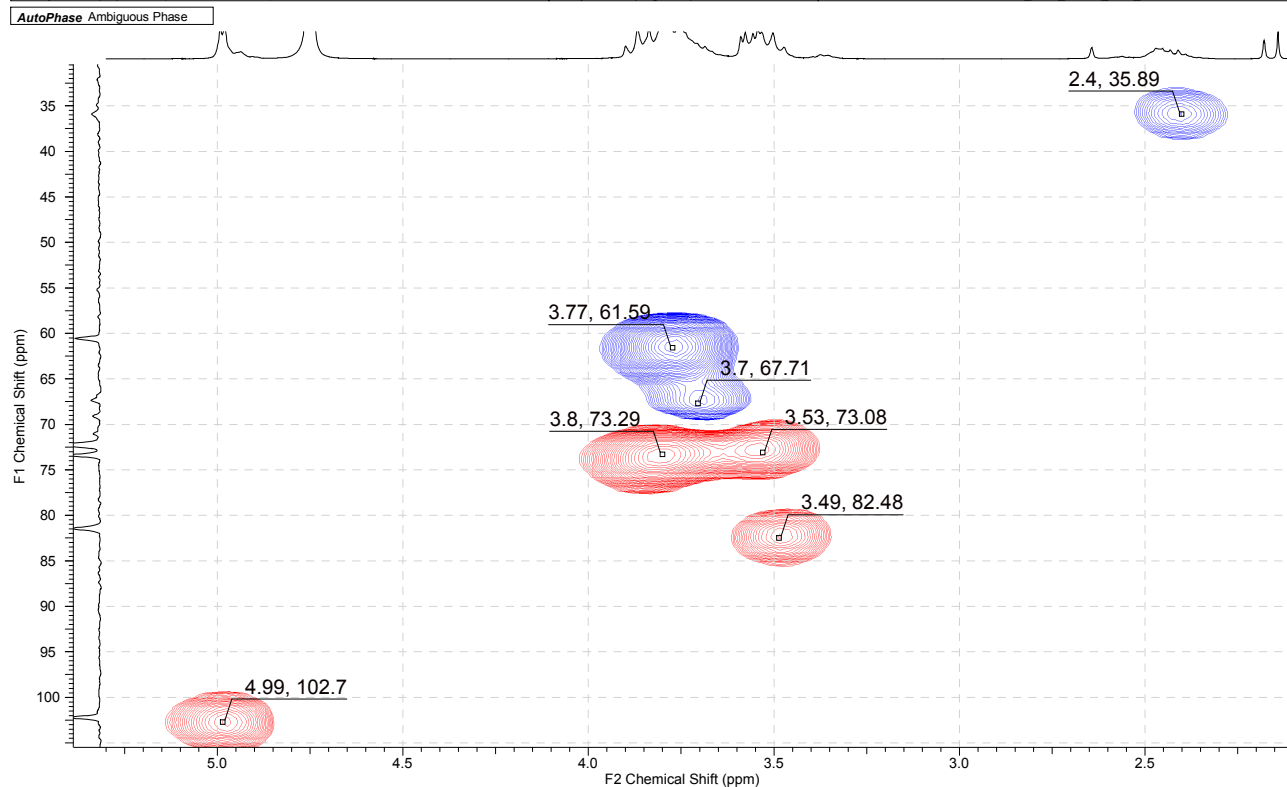


Figure S 33: HSQC-DEPT spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 10 in Table 1), DS  $\approx$ 3.3-3.5

Acquisition Time (sec)	1.8088	Comment	CAMBCD_LJ02_13C_D2O_03112016_2086_notte	Date	03 Nov 2016 13:45:52
Date Stamp	03 Nov 2016 13:45:52	File Name	E:\doc\11\molecules\Anal\NMR\CAMBCD_LJ02\3\fid	Frequency (MHz)	75.47
Nucleus	13C	Number of Transients	1035	Origin	spect
Points Count	262144	Pulse Sequence	zpgpg.save.bt	Original Points Count	32768
Spectrum Offset (Hz)	8301.4463	Receiver Gain	4597.60	SW(cyclical) (Hz)	18115.94
		Spectrum Type	STANDARD	Solvent	DMSO-d6
		Sweep Width (Hz)	18115.87	Temperature (degree C)	21.360

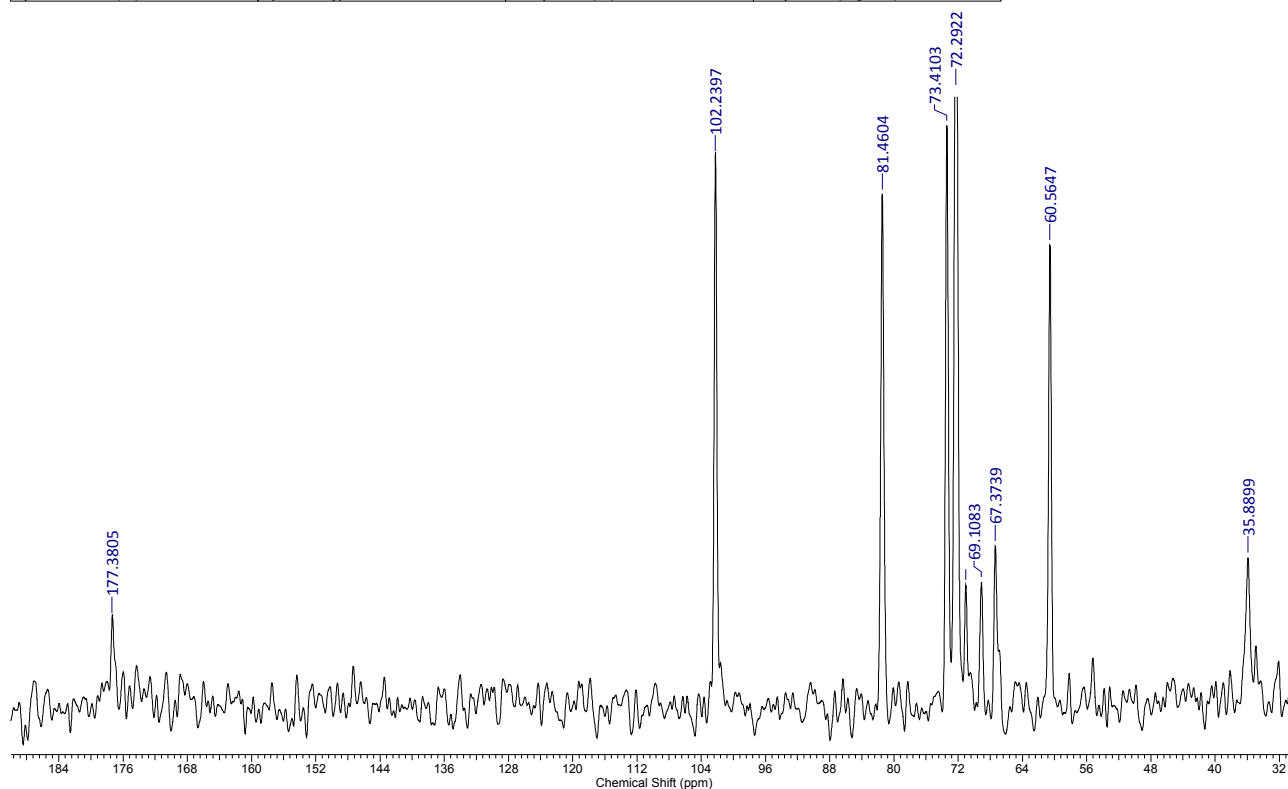


Figure S 34:  $^{13}\text{C}$ -NMR spectrum of carbamoylethylated  $\beta\text{CD}$  (**5**) prepared in ball mill (item 10 in Table 1),  $\text{DS} \approx 3.3\text{-}3.5$

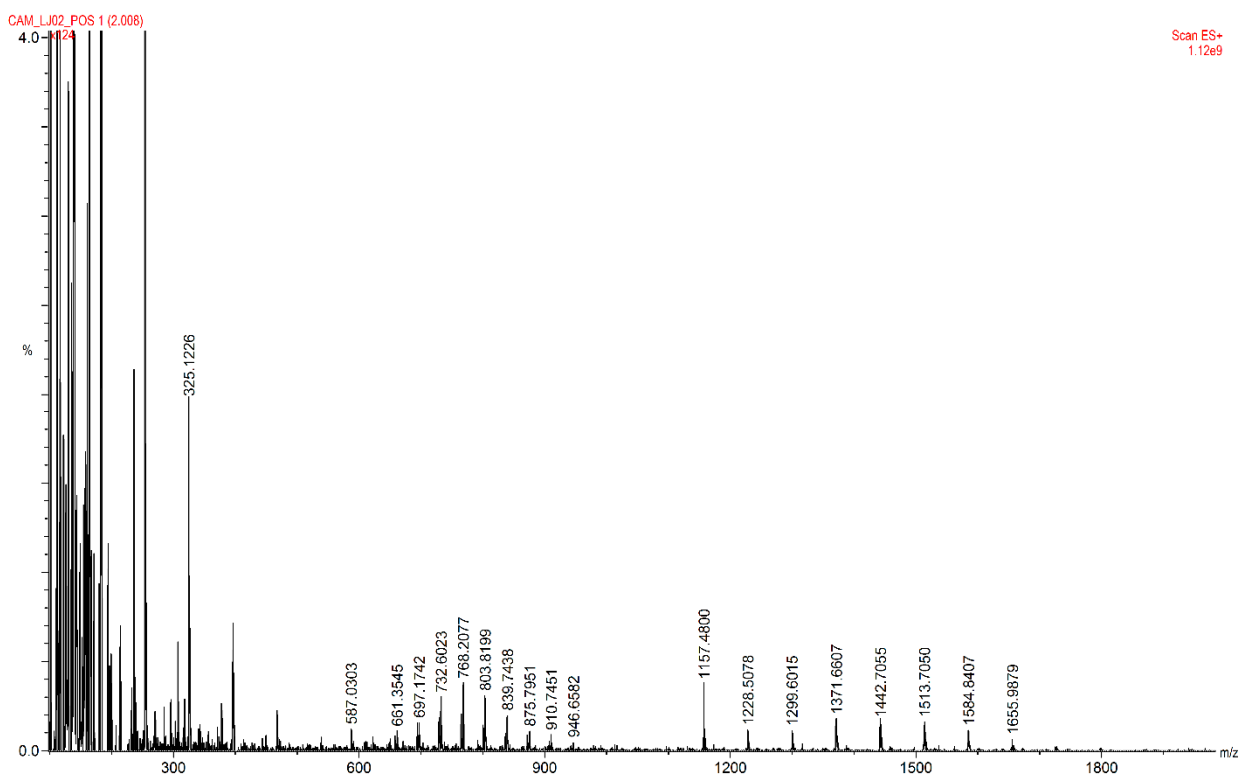
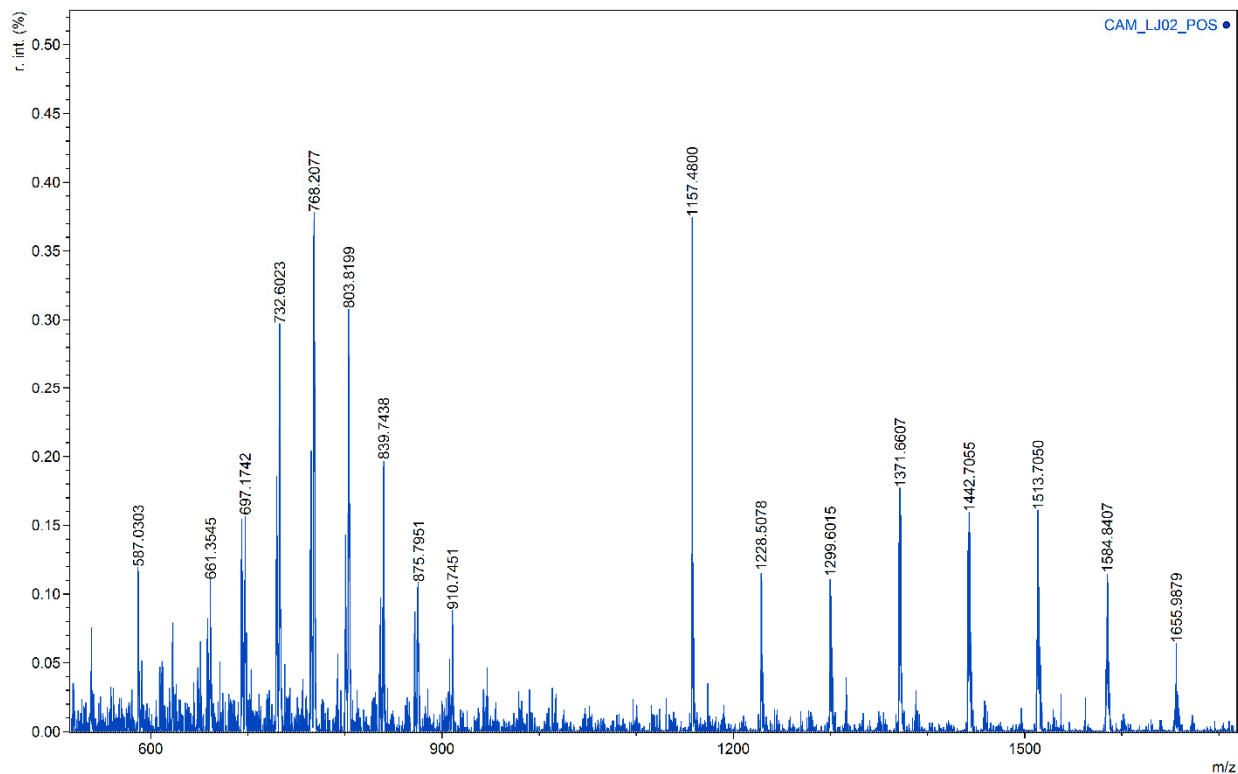


Figure S 35: ESI+ mass spectrum of carbamoylethylated  $\beta\text{CD}$  (**5**) prepared in ball mill (item 10 in Table 1),  $\text{DS} \approx 3.3\text{-}3.5$



Meas. m/z	Calc. m/z	$\delta$ (Da)	Z	Annotation	Formula
661.3545	661.2112	0.1432	2	(+2 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>
697.1742	696.7298	0.4444	2	(+3 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>3</sub>
732.6023	732.2483	0.3539	2	(+4 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>4</sub>
768.2077	767.7669	0.4409	2	(+5 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>5</sub>
803.8199	803.2854	0.5345	2	(+6 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>6</sub>
839.7438	838.8040	0.9398	2	(+7 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>7</sub>
875.7951	875.3185	0.4766	2	(+7 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> +1 CH <sub>2</sub> CH <sub>2</sub> COOH(?)) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>7</sub> /8(CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>1</sub> (?)
910.7451	909.8411	0.9040	2	(+8 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub> +1 CH <sub>2</sub> CH <sub>2</sub> COOH(?)) <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>2</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>8</sub> /9(CH <sub>2</sub> CH <sub>2</sub> COOH) <sub>1</sub> (?)
1157.4800	1157.3590	0.1210	1	C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> (BCD)	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>0</sub>
1228.5078	1228.3961	0.1118	1	+1 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>1</sub>
1299.6015	1299.4332	0.1682	1	+2 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>2</sub>
1371.6607	1370.4703	1.1904	1	+3 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>3</sub>
1442.7055	1441.5074	1.1981	1	+4 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>4</sub>
1513.7050	1512.5446	1.1604	1	+5 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>5</sub>
1584.8407	1583.5817	1.2590	1	+6 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>6</sub>
1655.9879	1654.6188	1.3691	1	+7 CH <sub>2</sub> CH <sub>2</sub> CONH <sub>2</sub>	(C <sub>42</sub> H <sub>70</sub> O <sub>35</sub> Na <sub>1</sub> ) <sub>1</sub> (CH <sub>1</sub> CH <sub>2</sub> CONH <sub>2</sub> ) <sub>7</sub>

Figure S 36: Peak identification in ESI+ mass spectrum of carbamoylethylated  $\beta$ CD (**5**) prepared in ball mill (item 10 in Table 1), DS  $\approx$  3.8-4.3



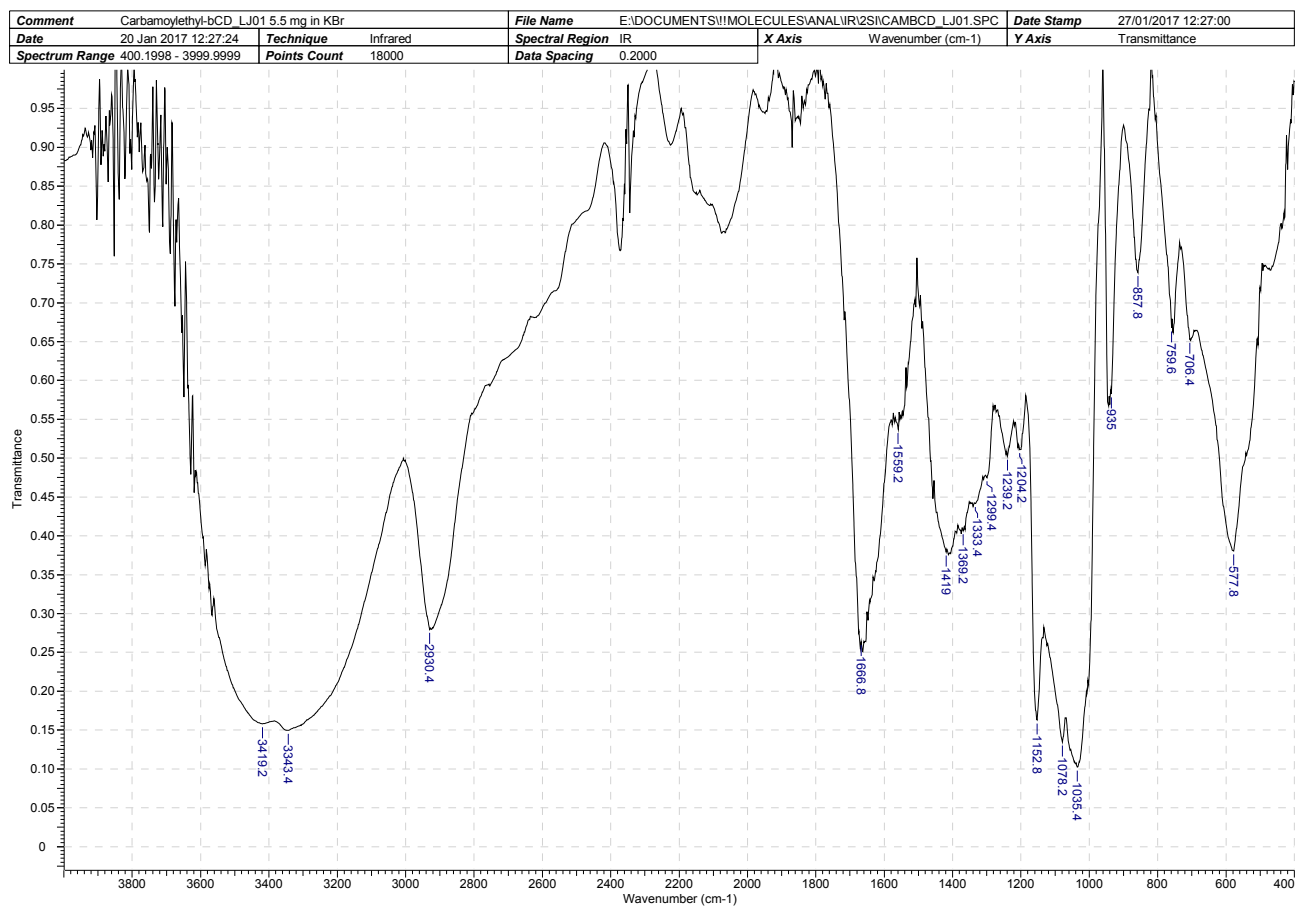


Figure S 37: IR spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 10 in Table 1), DS  $\approx$ 3.3-3.5

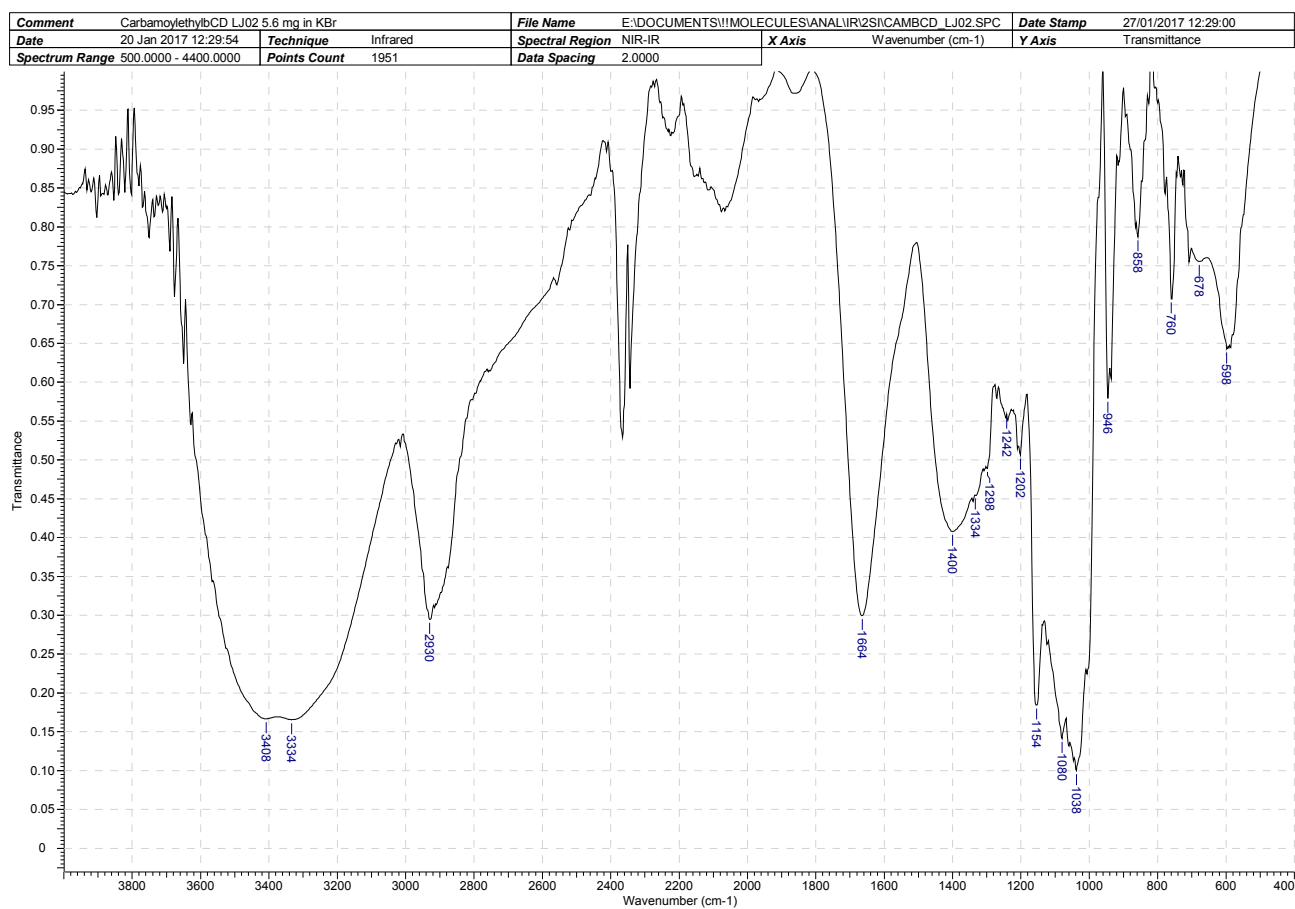


Figure S 38: IR spectrum of carbamoylethylated  $\beta$ CD (5) prepared in ball mill (item 10 in Table 1), DS  $\approx$ 3.3-3.5

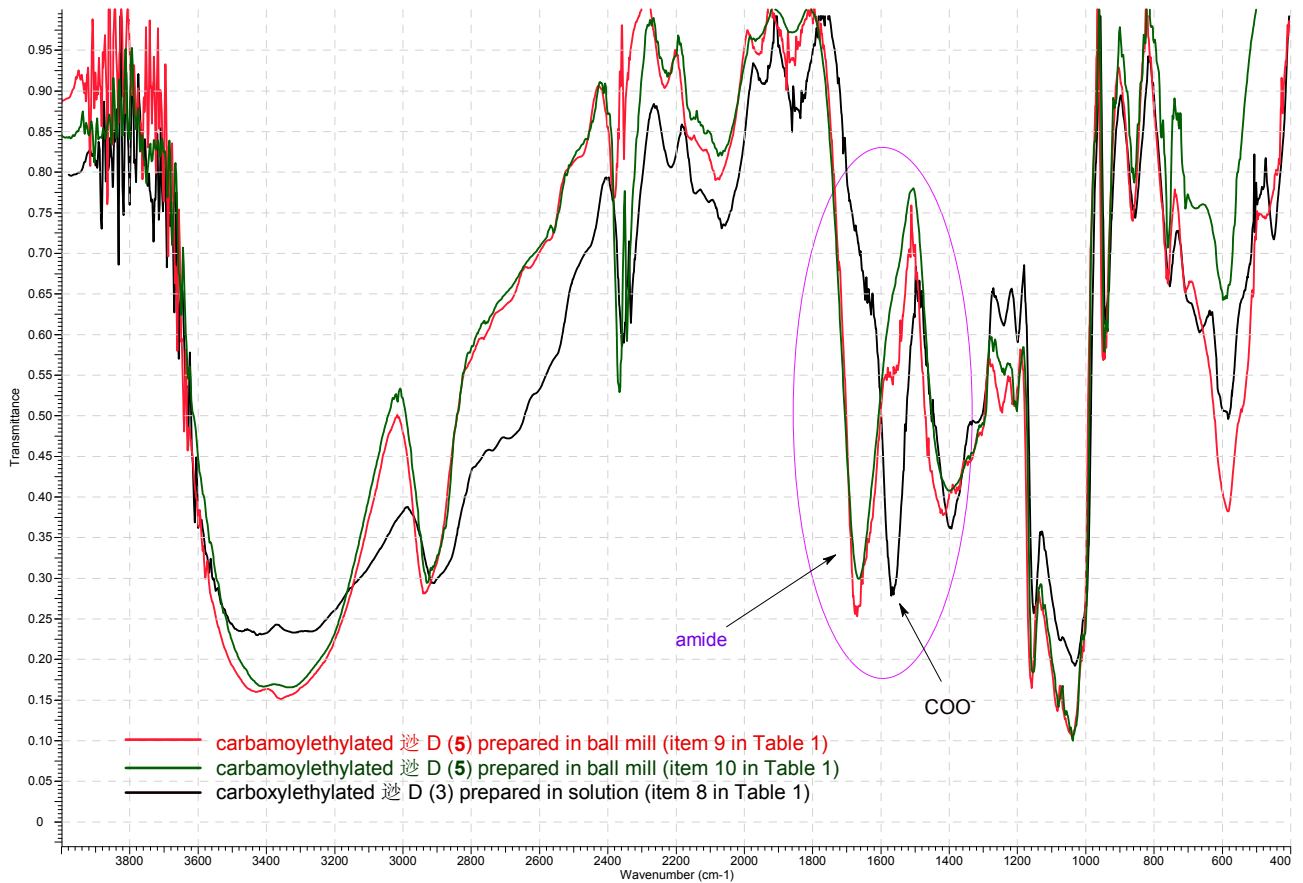


Figure S 39: Overlay of IR spectra of carbamoylethylated βCD (5) prepared in ball mill (item 9&10 in Table 1), DS ≈3.3-3.5 carboxylethylated βCD prepared in solution (item 8 in Table 1)

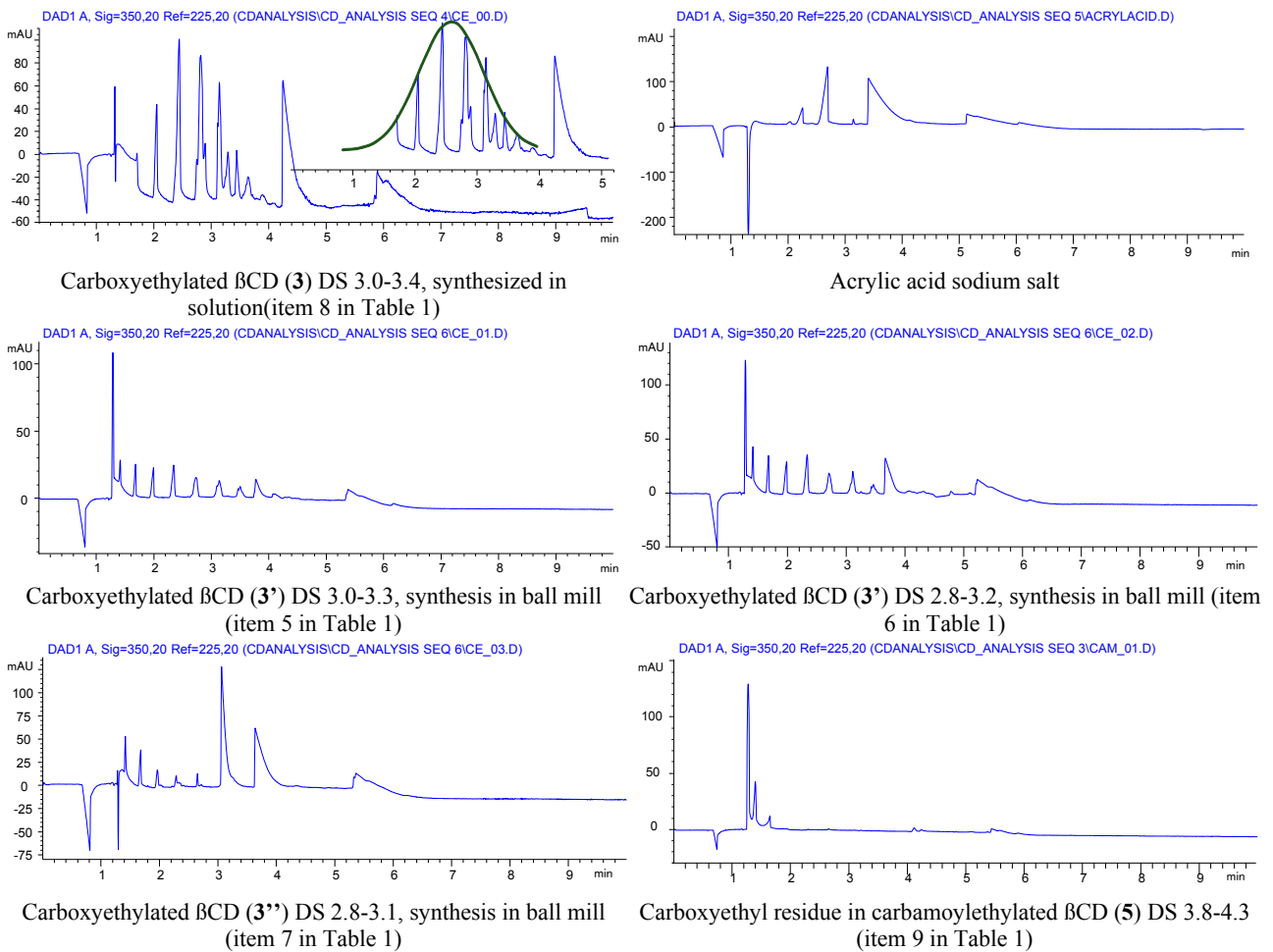


Figure S 40: Capillary electropherogram of carboxylethylated βCDs



Acquisition Time (sec)	4.0000	Comment	samplename: SBbCD_00SOL solvent: d2o temp: 25C probe: sw8099		
Date	Dec 17 2010	Date Stamp	Dec 17 2010	File Name	E:\doc\1\molecules\Anal\NMR\SBbCD_00SOL\s2pul_d2ofid
Frequency (MHz)	399.91	Nucleus	1H	Number of Transients	16
Points Count	65536	Pulse Sequence	s2pul	Receiver Gain	20.00
Spectrum Offset (Hz)	1992.8751	Spectrum Type	STANDARD	Sweep Width (Hz)	10000.00
				Original Points Count	40000
				Solvent	DEUTERIUM OXIDE
				Temperature (degree C)	25.000

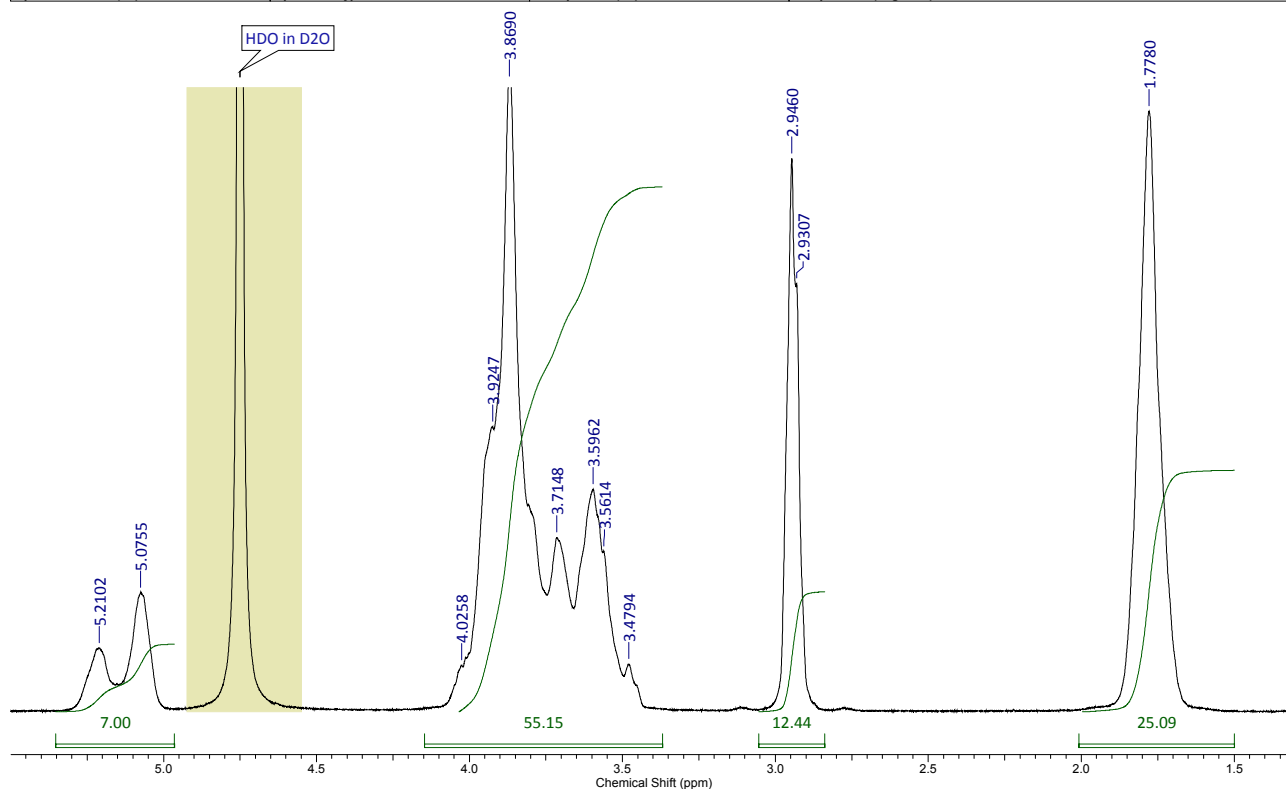


Figure S 41: Proton NMR spectrum of sulfobutylated  $\beta$ CD (4) prepared in solution (item 11 in Table 1),  $DS \approx 6.2-6.5$

Acquisition Time (sec)	(0.1500, 0.0023)	Comment	samplename: SBbCD_00SOL solvent: d2o temp: 25C probe: sw8099		Date	17 Dec 2010 09:21:12	
Date Stamp	Dec 17 2010	File Name	E:\doc\1\molecules\Anal\NMR\SBbCD_00SOL\ghsqc_d2ofid		Frequency (MHz)	(399.91, 100.57)	
Nucleus	(1H, 13C)	Number of Transients	16	Original Points Count	(1500, 64)	Points Count	(8192, 256)
Pulse Sequence	ghsqc	Solvent	d2o	Spectrum Type	HSQC	Sweep Width (Hz)	(9998.78, 28298.12)

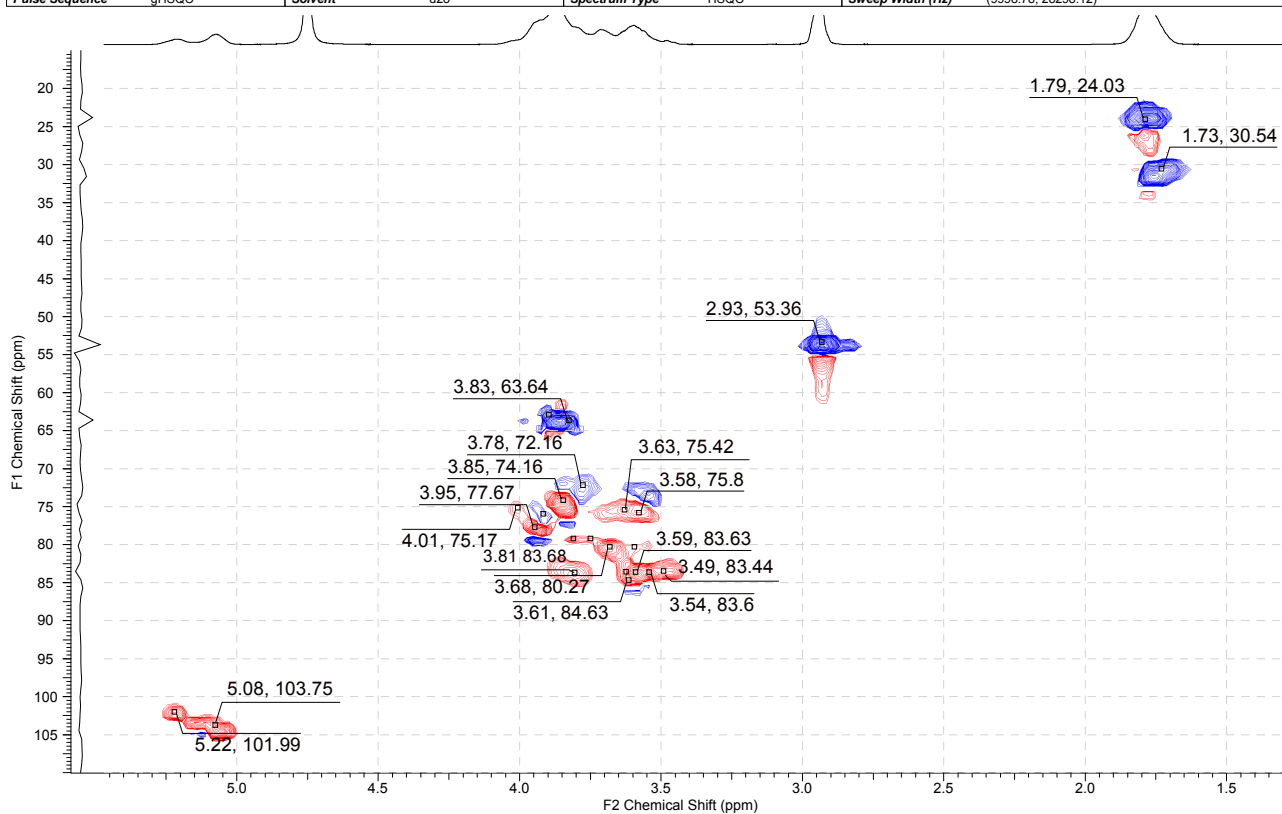


Figure S 42: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD (4) prepared in solution (item 11 in Table 1),  $DS \approx 6.2-6.5$

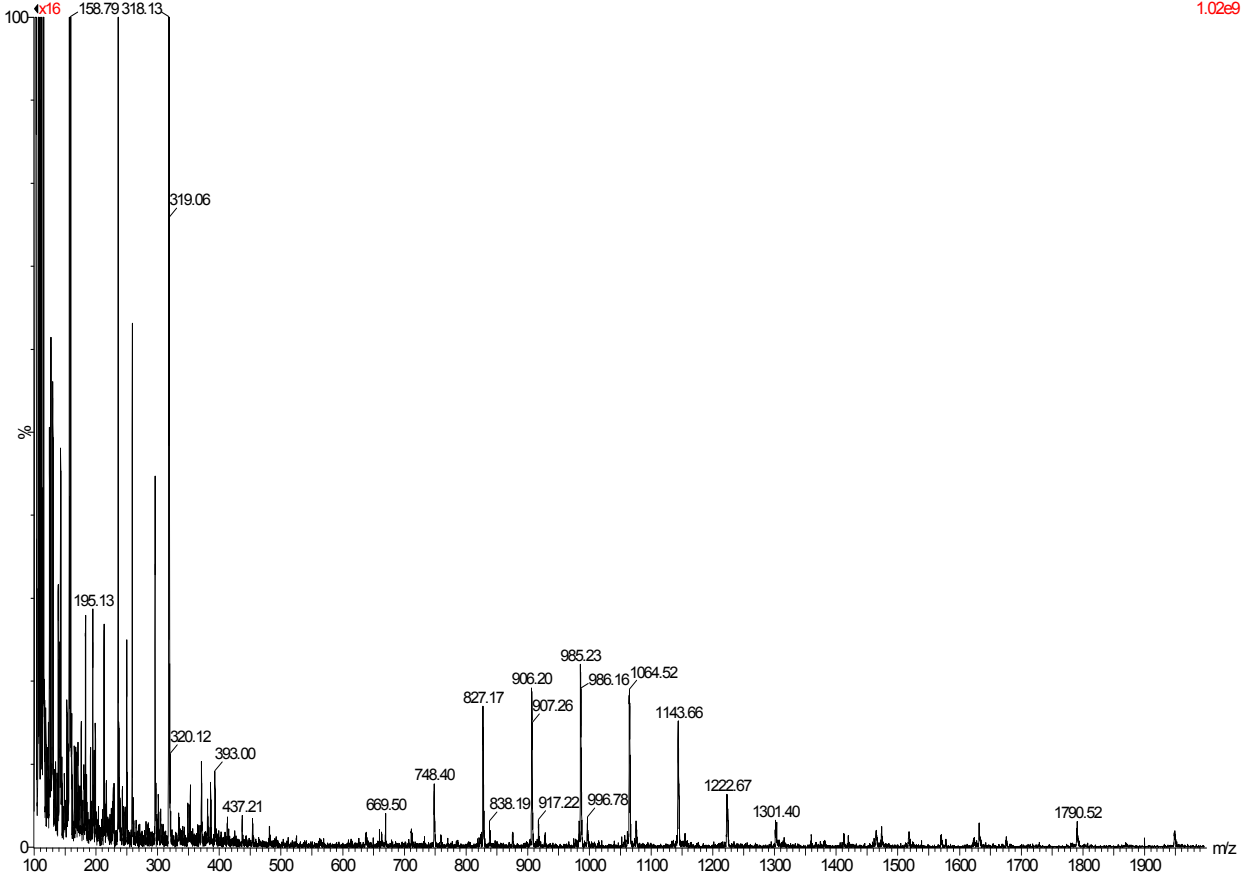
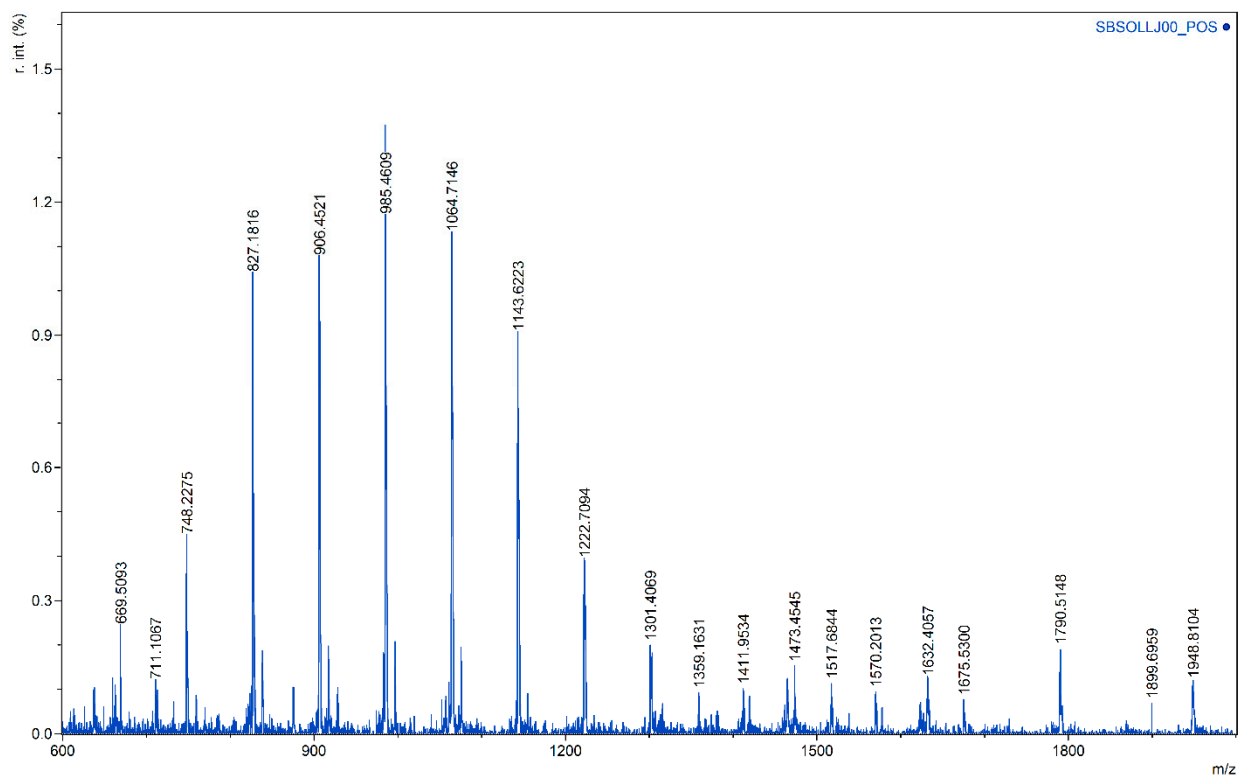


Figure S 43: ESI+ mass spectrum of sulfobutylated  $\beta$ CD (4) prepared in solution (item 11 in Table 1),  $DS \approx 6.2-6.5$



Meas. m/z	Calc. m/z	$\delta$ (Da)	Z	Annotation	Formula
669.5093	669.1748	0.3345	2	(+1 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)1
711.1067	709.7879	1.3188	3	(+1 C4H8SO3H +5 C4H8SO3Na)/3	(C42H70O35Na3)1(C4H7SO3H)1(C4H7SO3Na)5
748.2275	748.1755	0.0520	2	(+2 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)2
827.1816	827.1761	0.0054	2	(+3 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)3
906.4521	906.1768	0.2752	2	(+4 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)4
985.4609	985.1775	0.2834	2	(+5 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)5
1064.7146	1064.1782	0.5364	2	(+6 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)6
1143.6223	1143.1789	0.4434	2	(+7 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)7
1222.7094	1222.1795	0.5299	2	(+8 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)8
1301.4069	1301.1802	0.2267	2	(+9 C42H70O35Na2)/2	(C42H70O35Na2)1(C4H7SO3H)0(C4H7SO3Na)9
1315.4595	1315.3603	0.0991	1	+1 C4H8SO3Na	(C42H70O35Na1)1(C4H7SO3H)0(C4H7SO3Na)1
1359.1631	1358.1990	0.9642	2	(+2 C4H8SO3H)/2	(C42H70O35Na2)1(C4H7SO3H)2(C4H7SO3Na)8
1473.4545	1473.3617	0.0928	1	+2 C4H8SO3Na	(C42H70O35Na1)1(C4H7SO3H)0(C4H7SO3Na)2
1517.6844	1516.2003	1.4841	2	(+2 C4H8SO3H +10 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)2(C4H7SO3Na)10
1632.4057	1631.3631	1.0427	1	+3 C4H8SO3Na	(C42H70O35Na1)1(C4H7SO3H)0(C4H7SO3Na)3
1675.5300	1674.2017	1.3283	2	(+2 C4H8SO3H +12 C4H8SO3Na)/2	(C42H70O35Na2)1(C4H7SO3H)2(C4H7SO3Na)12
1790.5148	1789.3644	1.1503	1	+4 C4H8SO3Na	(C42H70O35Na1)1(C4H7SO3H)0(C4H7SO3Na)4
1948.8104	1947.3658	1.4446	1	+5 C4H8SO3Na	(C42H70O35Na1)1(C4H7SO3H)0(C4H7SO3Na)5

Figure S 44: Peak identification in ESI+ mass spectrum of sulfobutylated  $\beta$ CD (4) prepared in solution (item 11 in Table 1), DS  $\approx$ 6.2-6.5

Acquisition Time (sec)	3.6438	Comment	SBBCD_LJ01 (20mg in 0.6ml D2O)		Date	30 Sep 2016 15:09:20	
Date Stamp	30 Sep 2016 15:09:20	File Name	D:\Docs\1\Molecules\Anal\SBBCD_LJ01\1fid		Frequency (MHz)	300.13	
Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Points Count	131072	Pulse Sequence	zg	Receiver Gain	90.50	SW(cyclical) (Hz)	4496.40
Spectrum Offset (Hz)	1095.8673	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37	Temperature (degree C)	22.160
						Owner	root
						Solvent	DEUTERIUM OXIDE

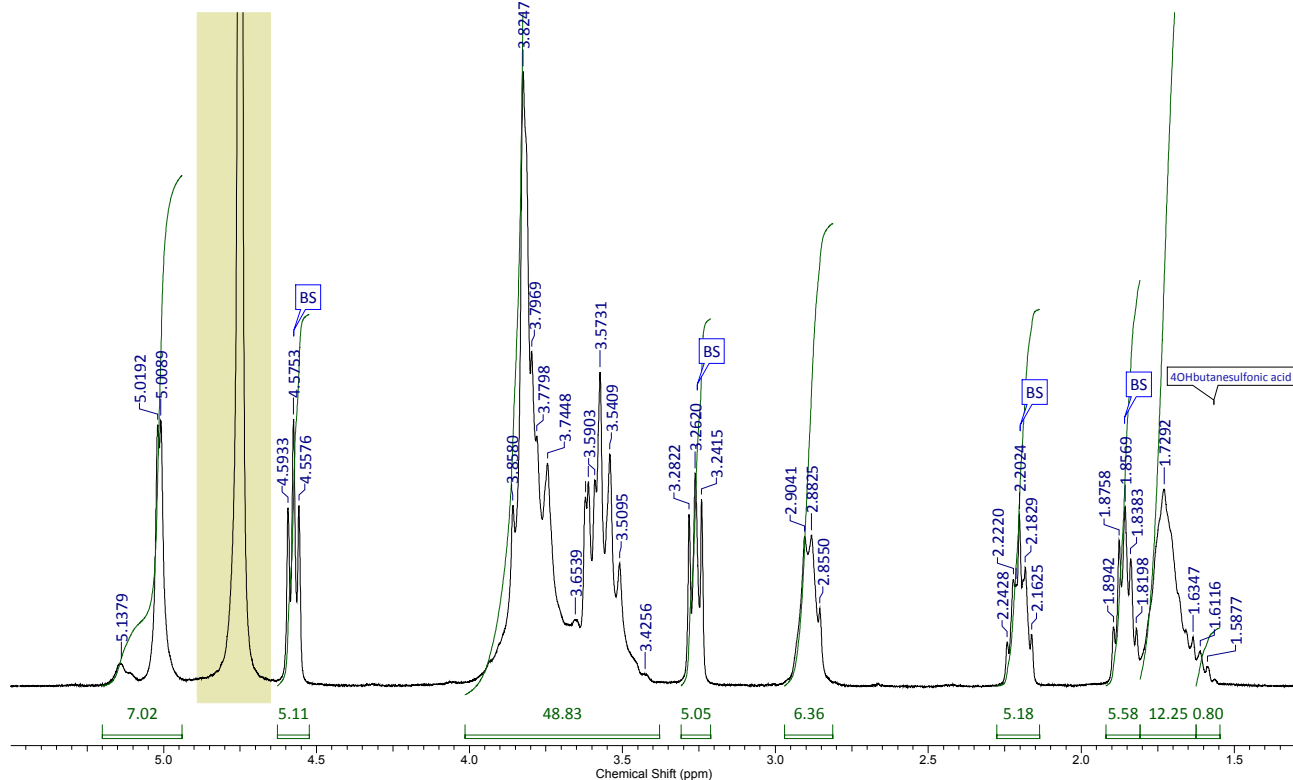


Figure S 45: Proton NMR spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 12 in Table 1),  $DS \approx 3.1-3.3$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059		Date	19 Sep 2016 12:04:24	
File Name	D:\Docs\1\Notebooks\BM Reactions\NMR\SBBCD_LJ01\SBBCD_LJ01\3iser	Frequency (MHz)	(300.13, 75.47)		Original Points Count	(128, 64)	
Nucleus	(1H, 13C)	Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Owner	psm	Points Count	(512, 256)	Pulse Sequence	hsqcdeptp	Solvent	DMSO
Spectrum Type	HSQC-DEPT	Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	22.360		
Title	SBBCD_LJ02_HSQC_D2O_190916_2010_RG=100						

AutoPhase Ambiguous Phase

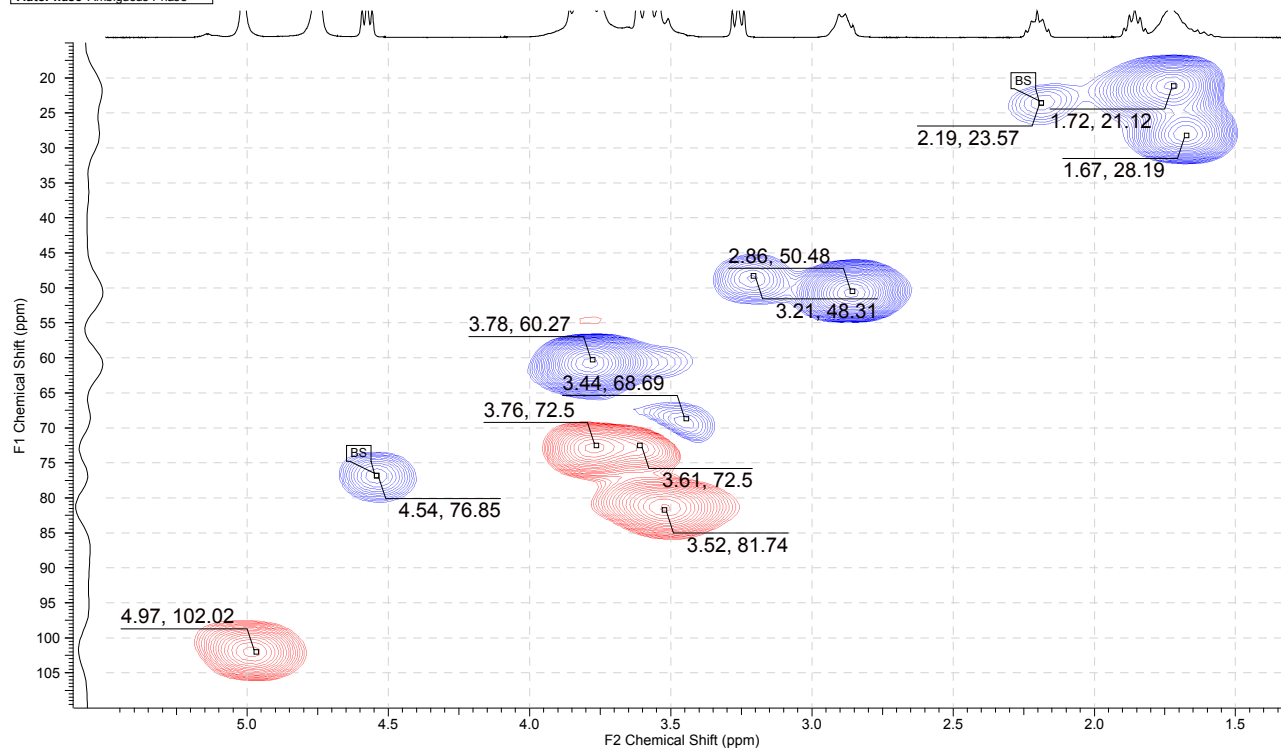


Figure S 46: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 12 in Table 1),  $DS \approx 3.1-3.3$

Acquisition Time (sec)	3.6438	Comment	SBBCD_LJ03 (20mg in 0.6ml D2O)		Date	28 Sep 2016 12:54:56			
Date Stamp	28 Sep 2016 12:54:56	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ03\1.fid						
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Owner	root	Points Count	131072	Pulse Sequence	zg	Receiver Gain	90.50	SW(cyclical) (Hz)	4496.40
Solvent	DMSO-d6	Spectrum Offset (Hz)	1096.1024	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37	Temperature (degree C)	22.160

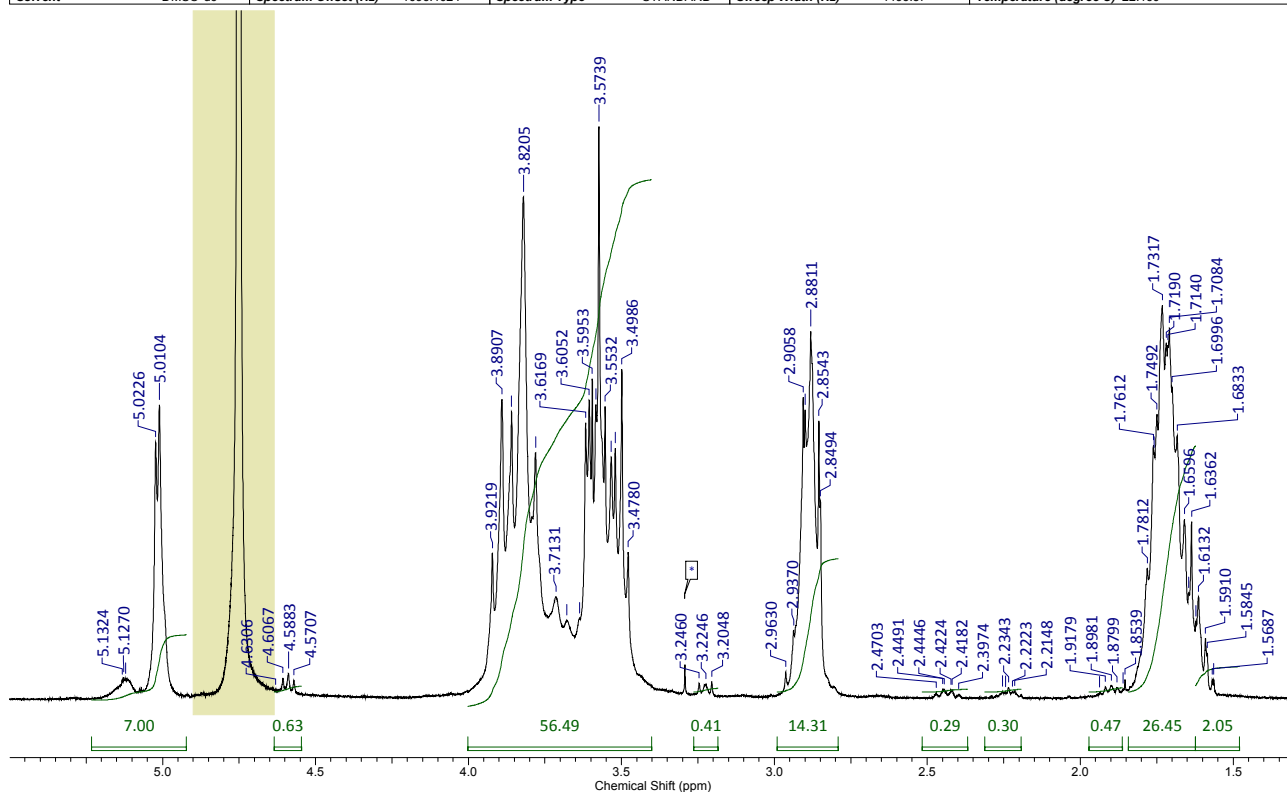


Figure S 47: Proton NMR spectrum of sulfobutylated  $\beta$ CD ( $4'$ ) prepared in ball mill (item 13 in Table 1),  $DS \approx 5.2-5.8$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z6284/0059		Date	28 Sep 2016 13:48:32	
File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ03\2.ser	Frequency (MHz)	(300.13, 75.47)		Nucleus	(1H, 13C)	
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)	Owner	psm
Points Count	(512, 256)	Pulse Sequence	hsgcdetgp	Solvent	DMSO	Spectrum Type	HSQC-DEPT
Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	22.260				

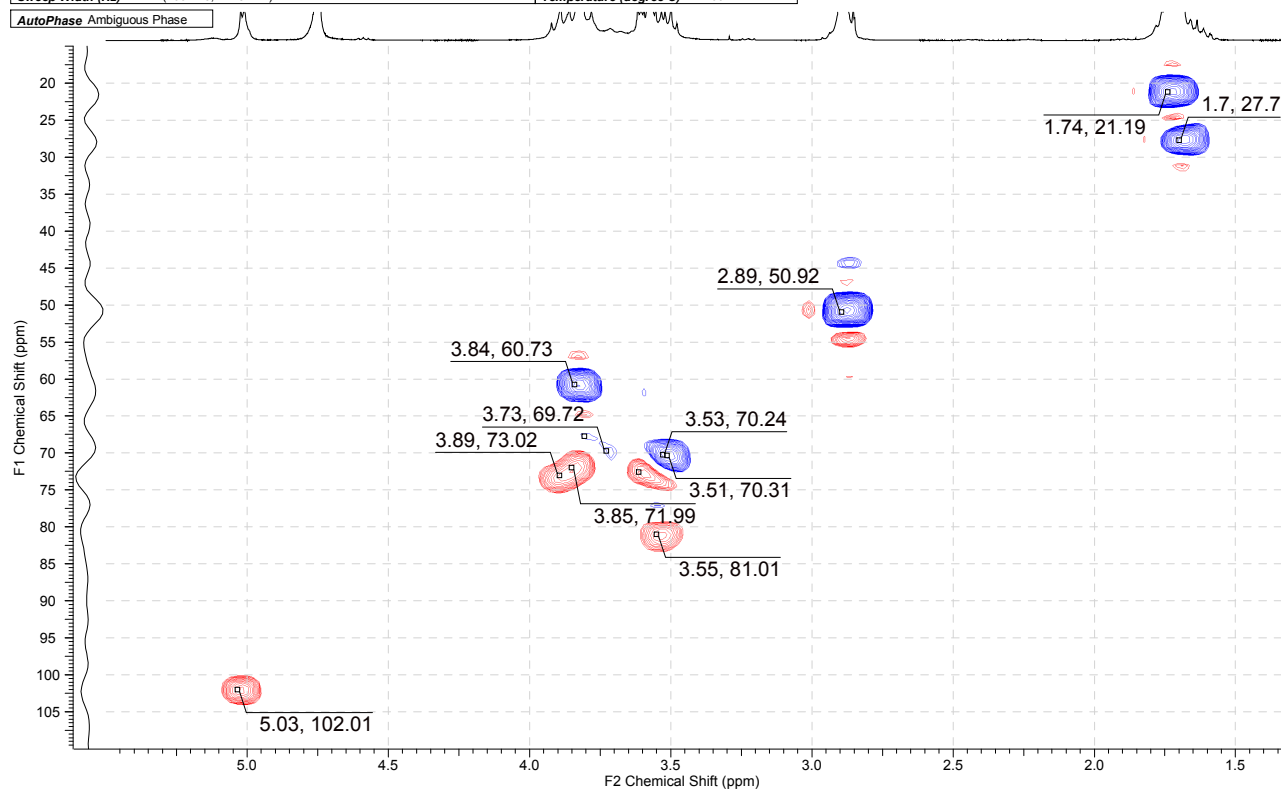


Figure S 48: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD ( $4'$ ) prepared in ball mill (item 13 in Table 1),  $DS \approx 5.2-5.8$



Acquisition Time (sec)	3.6438	Comment	SBBCD_LJ03 (20mg in 0.6ml D2O)		Date	28 Sep 2016 12:54:56			
Date Stamp	28 Sep 2016 12:54:56	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ03\1.fid						
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64	Origin	spect	Original Points Count	16384
Owner	root	Points Count	131072	Pulse Sequence	zg	Receiver Gain	90.50	SW(cyclical) (Hz)	4496.40
Solvent	DMSO-d6	Spectrum Offset (Hz)	1096.1024	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37	Temperature (degree C)	22.160

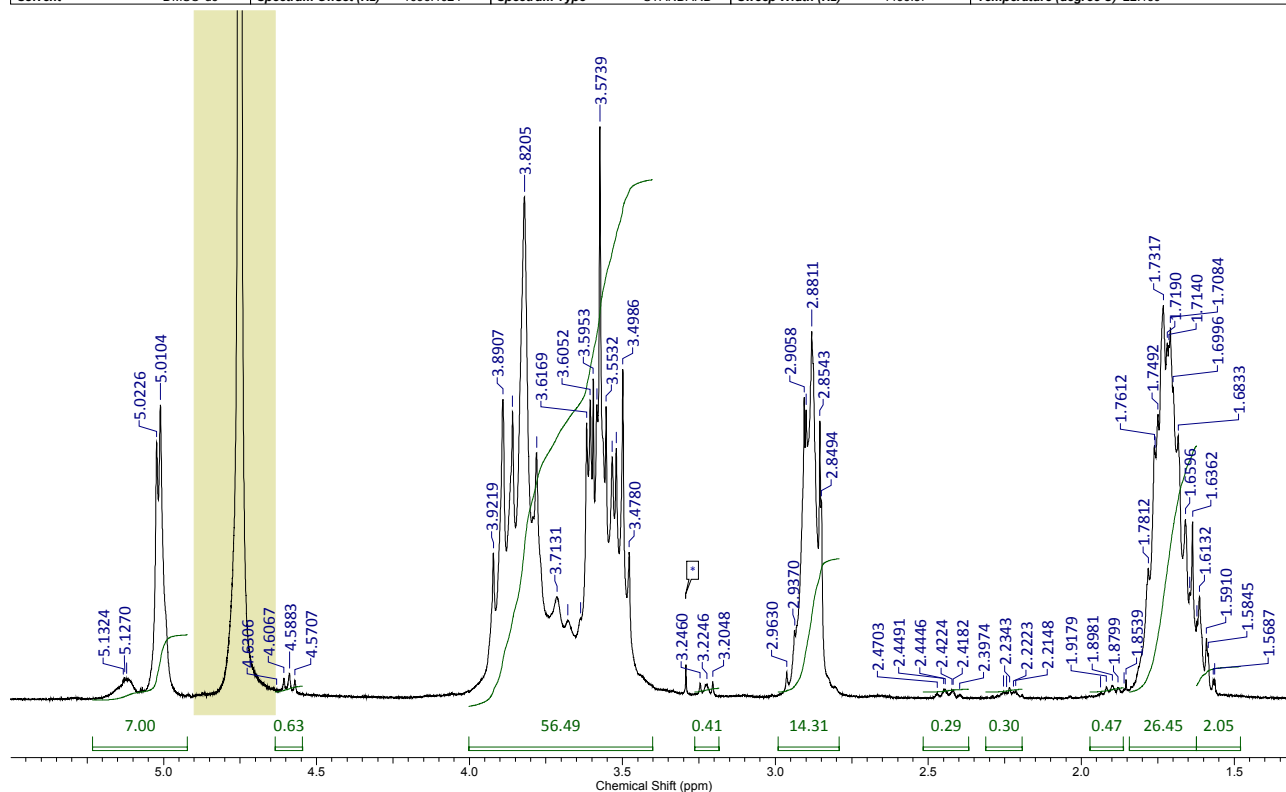


Figure S 49: Proton NMR spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 14 in Table 1), DS  $\approx$  6.2-6.6

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z6284/0059		Date	28 Sep 2016 13:48:32	
File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ03\2.ser	Frequency (MHz)	(300.13, 75.47)		Nucleus	(1H, 13C)	
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)	Owner	psm
Points Count	(512, 256)	Pulse Sequence	hsgcdetgp	Solvent	DMSO	Spectrum Type	HSQC-DEPT
Sweep Width (Hz)	(2991.75, 12451.17)	Temperature (degree C)	22.260				

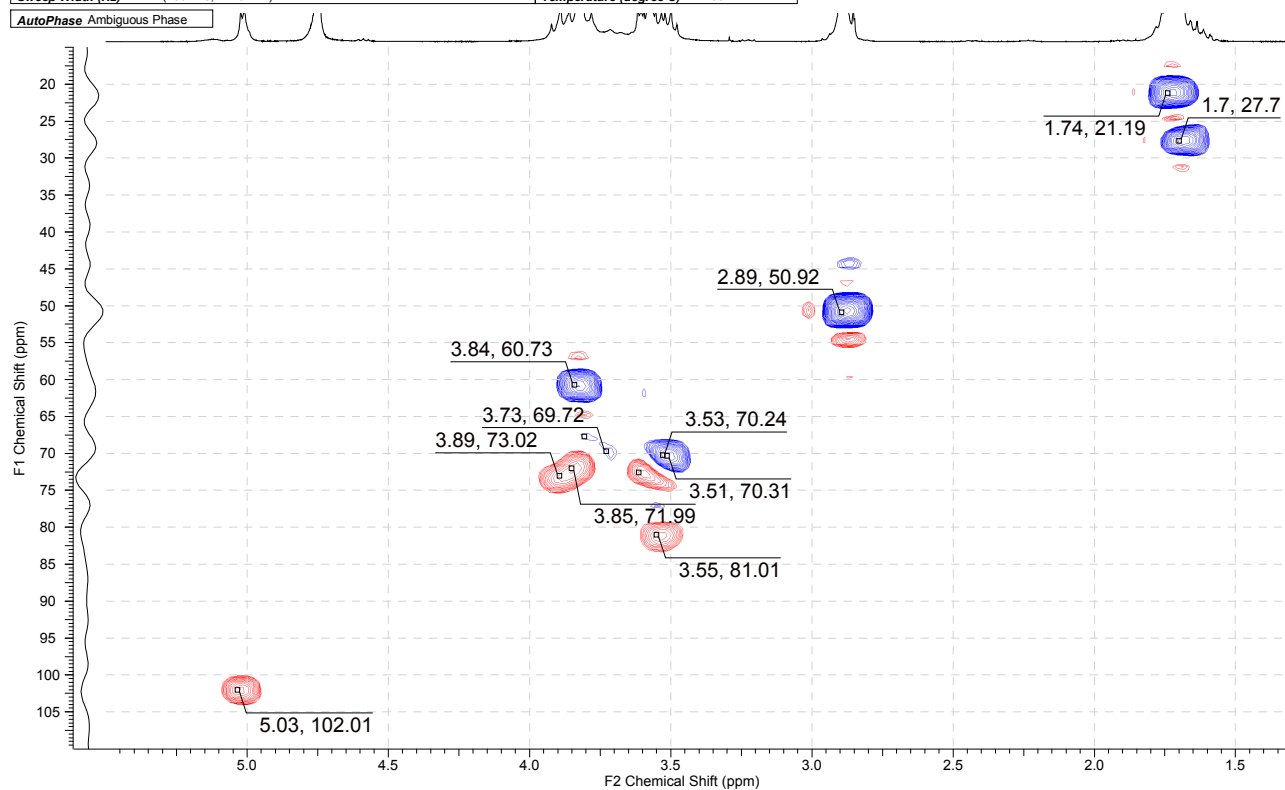


Figure S 50: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 14 in Table 1), DS  $\approx$  6.2-6.6

Acquisition Time (sec)	3.6438	Comment	SBBCD_LJ04 (20mg in 0.6ml D2O)	Date	30 Sep 2016 11:53:04
Date Stamp	30 Sep 2016 11:53:04	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ04\1\fid		
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64
Origin	root	Points Count	131072	Origin	spect
Owner	root	Pulse Sequence	zg	Receiver Gain	90.50
Solvent	DEUTERIUM OXIDE	Spectrum Offset (Hz)	1096.2106	SW(cyclical) (Hz)	4496.40
Temperature (degree C)	22.260			Spectrum Type	STANDARD
				Sweep Width (Hz)	4496.37

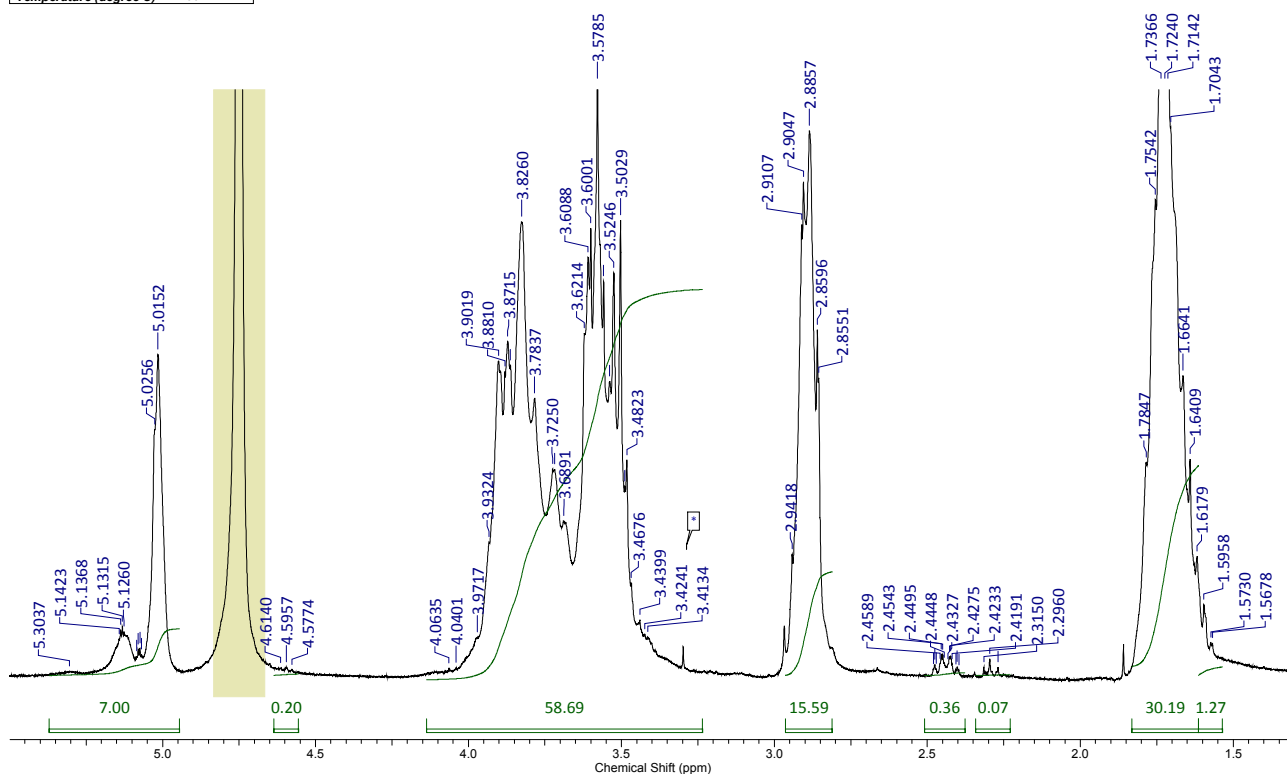


Figure S 51: Proton NMR spectrum of sulfobutylated  $\beta$ CD ( $4'$ ) prepared in ball mill (item 15 in Table 1),  $DS \approx 7.5-7.8$

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	30 Sep 2016 12:46:28
File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD_LJ04\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(512, 256)	Pulse Sequence	hsqcetp	Owner	psm
Sweep Width (Hz)	(2991.75, 12451.17)	Solvent	DMSO	Spectrum Type	HSQC-DEPT
		Temperature (degree C)	22.260		

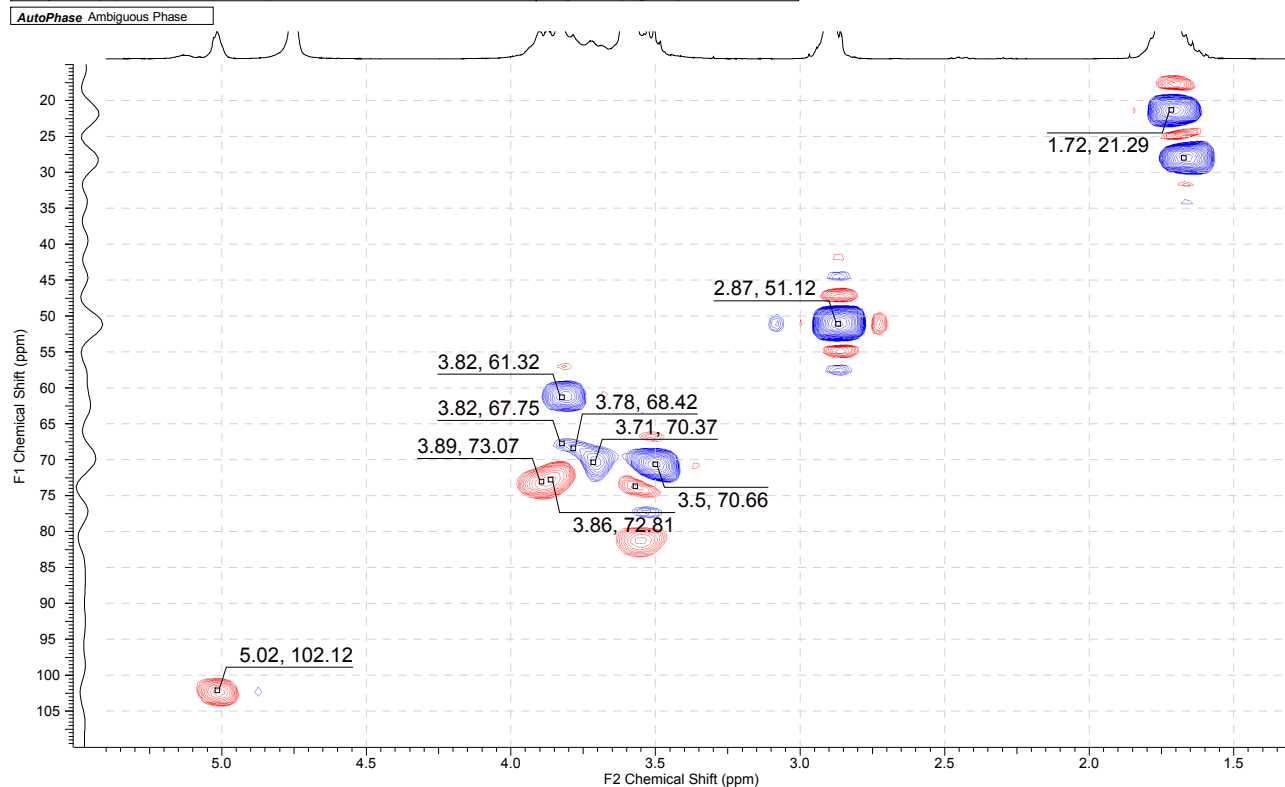


Figure S 52: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD ( $4'$ ) prepared in ball mill (item 15 in Table 1),  $DS \approx 7.5-7.8$

Acquisition Time (sec)	3.6438	Comment	SBBCD-LJ05 1H D2O 031016 2033 RG=90	Date	03 Oct 2016 10:12:48
Date Stamp	03 Oct 2016 10:12:48	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD-LJ05\1fid		
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	58
Owner	root	Points Count	131072	Pulse Sequence	zg
Solvent	CDCI3	Spectrum Offset (Hz)	1096.2448	Spectrum Type	STANDARD
				Receiver Gain	90.50
				Sweep Width (Hz)	4496.37
				Original Points Count	16384
				SW(cyclical) (Hz)	4496.40
				Temperature (degree C)	21.760

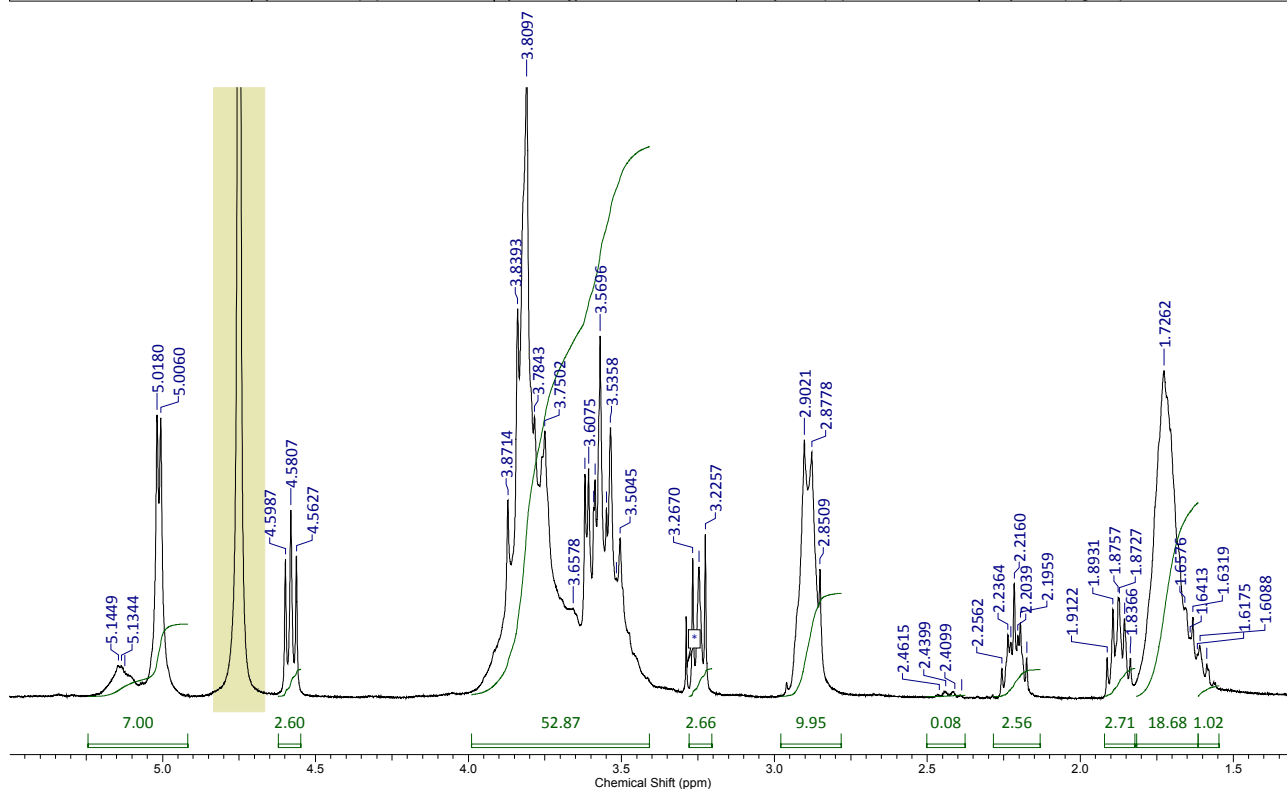


Figure S 53: Proton NMR spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 16 in Table 1), DS  $\approx$  4.4-4.9

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	04 Oct 2016 14:22:56
File Name	E:\doc\1\molecules\Anal\NMR\SBBCD_LJ05\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(1024, 256)	Pulse Sequence	hsqcetdgp	Solvent	DMSO
Sweep Width (Hz)	(2994.67, 12451.17)	Temperature (degree C)	22.560	Title	SBBCD-LJ05 HSQC D2O 031016 2033 RG=90
				Spectrum Type	HSQC-DEPT

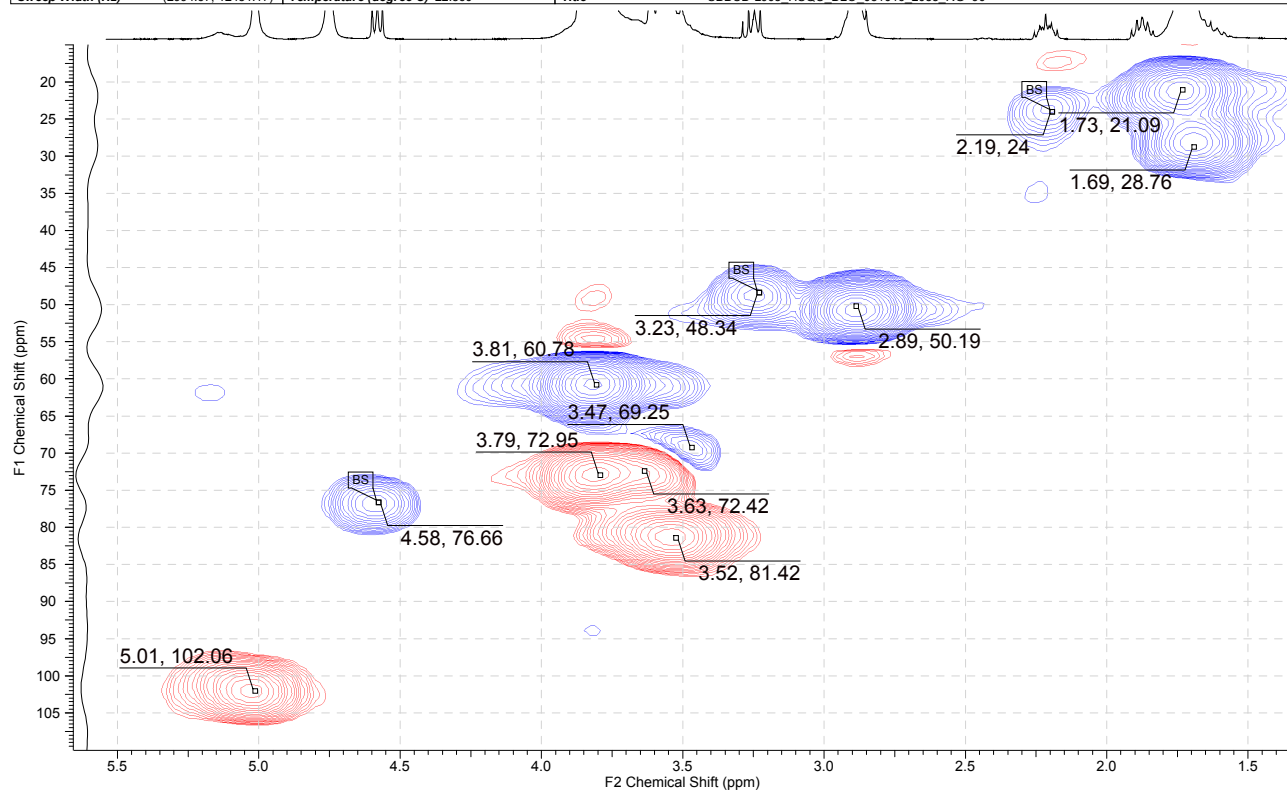


Figure S 54: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 16 in Table 1), DS  $\approx$  4.4-4.9

Acquisition Time (sec)	3.6438	Comment	SBBCD_LJ06_1H_D2O_061016_2045_rg=110	Date	06 Oct 2016 15:30:40
Date Stamp	06 Oct 2016 15:30:40	File Name	D:\Docs\1\Notebooks\BM_Reactions\NMR\SBBCD-LJ06\1.tif		
Frequency (MHz)	300.13	Nucleus	1H	Number of Transients	64
Owner	root	Points Count	131072	Origin	spect
Solvent	CDCI3	Spectrum Offset (Hz)	1096.3082	Receiver Gain	114.00
		Spectrum Type	STANDARD	SW(cyclical) (Hz)	4496.40
				Sweep Width (Hz)	4496.37
				Temperature (degree C)	21.560

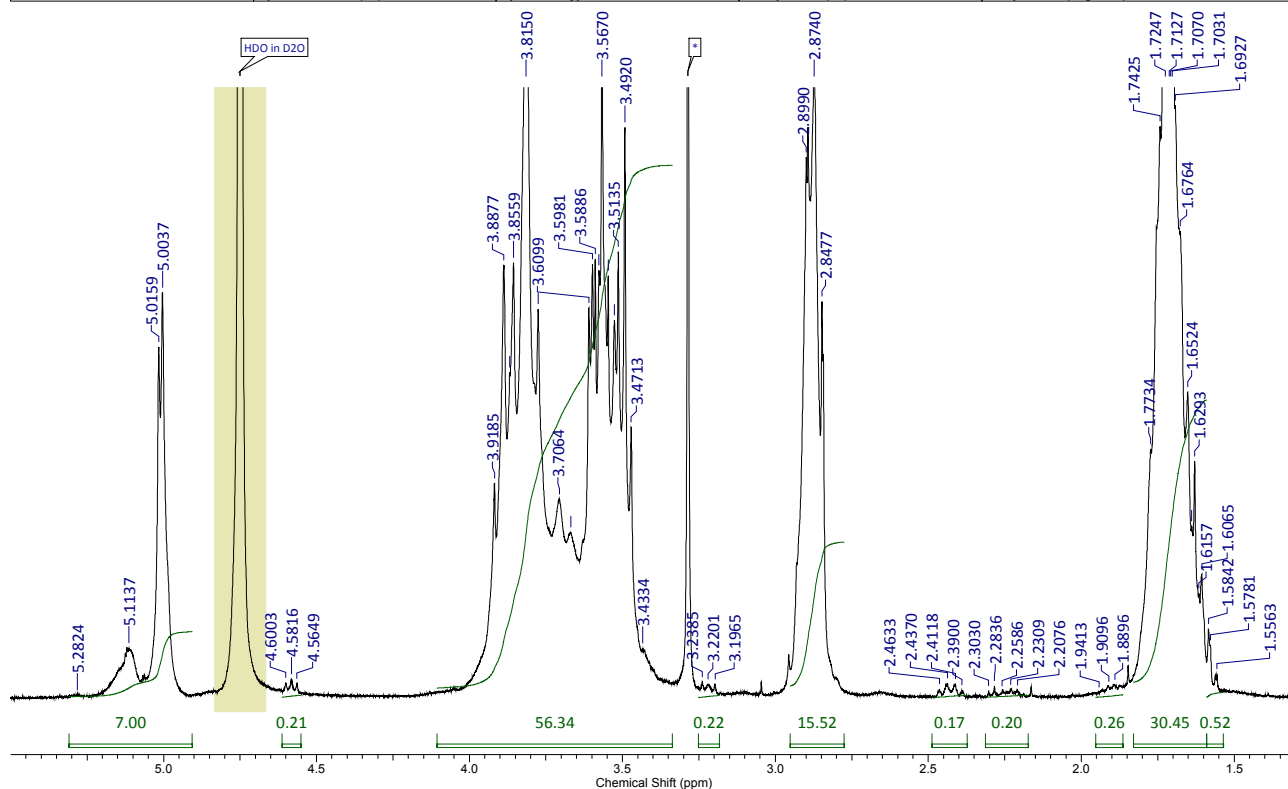


Figure S 55: Proton NMR spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 17 in Table 1), DS  $\approx$  6.9-7.5

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	06 Oct 2016 15:01:30
File Name	E:\docs\1\molecules\Anal\NMR\SBBCD-LJ06\2\ser			Frequency (MHz)	(300.13, 75.47)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(512, 512)	Pulse Sequence	hsqcetdgp	Solvent	DMSO
Sweep Width (Hz)	(2991.75, 12475.59)	Temperature (degree C)	21.560	Title	SBBCD_LJ06_HSQC_D2O_061016_2045_rg=110
				Nucleus	(1H, 13C)
				Owner	psm
				Spectrum Type	HSQC-DEPT

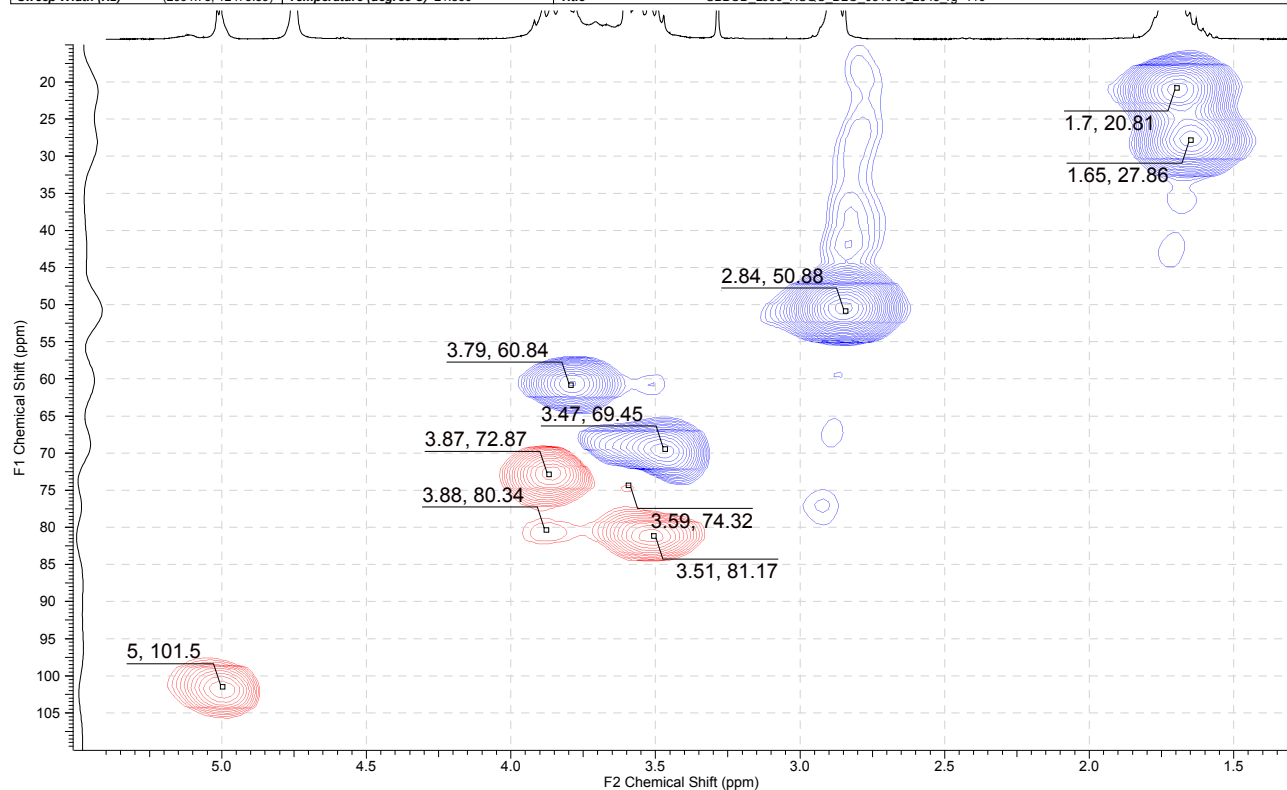


Figure S 56: HSQC-DEPT spectrum of sulfobutylated  $\beta$ CD (4') prepared in ball mill (item 17 in Table 1), DS  $\approx$  6.9-7.5

Acquisition Time (sec)	3.6438	Comment	HOBSANA 1H D2O 120117 2122 RG=180 8 mg/0.8 ml	Date	13 Jan 2017 08:08:48
Date Stamp	13 Jan 2017 08:08:48	File Name	E:\Documents\1\Molecules\Anal\HOBSANA\1\fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	131072	Pulse Sequence	zg	Original Points Count	16384
Spectrum Offset (Hz)	1096.6907	Spectrum Type	STANDARD	Receiver Gain	181.00
				SW(cyclical) (Hz)	4496.40
				Solvent	CHLOROFORM-d
				Sweep Width (Hz)	4496.37
				Temperature (degree C)	20.260

No.	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
1	1.59	23	dd	6.40, 6.30	beta	[1.54 .. 1.65]
2	1.71	14	dddd	8.00, 7.90	gamma	[1.65 .. 1.78]
3	2.86	24	dd	7.80	delta	[2.81 .. 2.92]
4	3.55	24	dd	6.30	alpha	[3.51 .. 3.61]

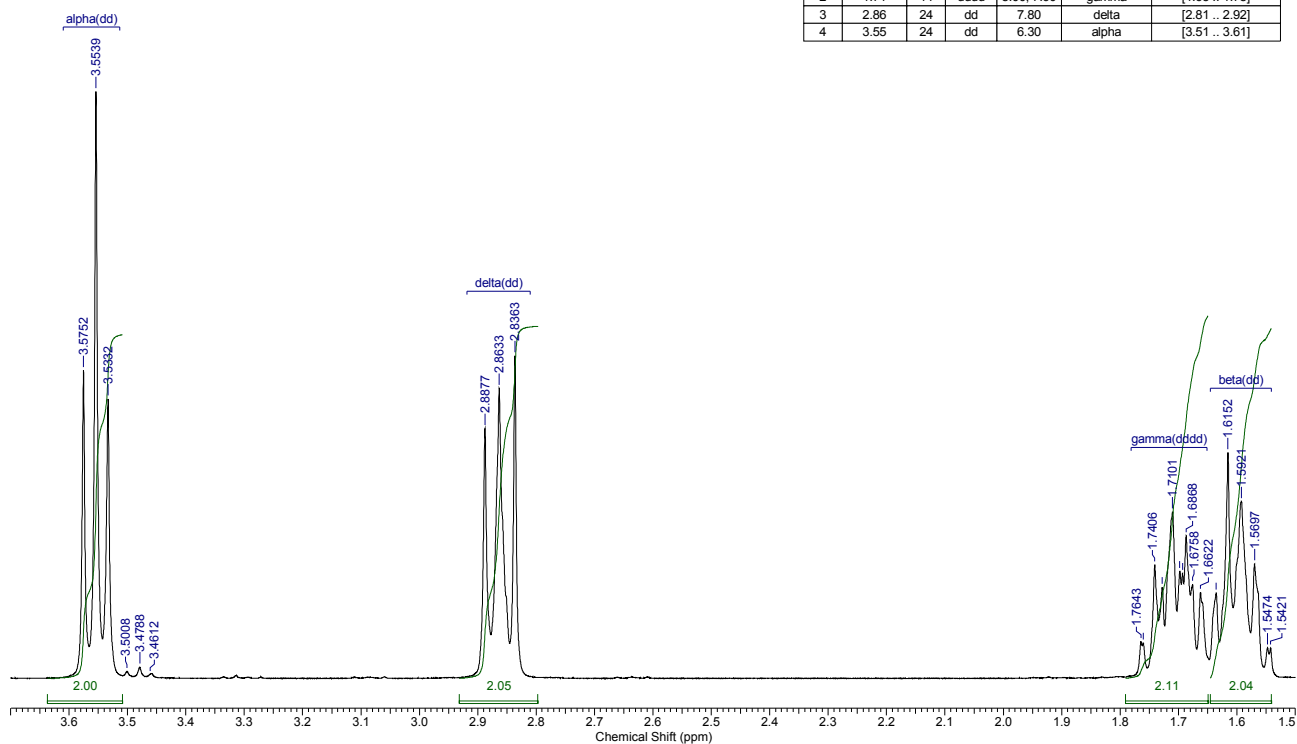


Figure S 57: Proton NMR spectrum of 4-hydroxy-1-butananesulfonic acid sodium salt (HOBSANa)

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	13 Jan 2017 09:02:26
File Name	E:\Documents\1\Molecules\Anal\HOBSANA\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(1024, 512)	Pulse Sequence	hsqcdeptgp	Solvent	DMSO
Sweep Width (Hz)	(2994.67, 12475.59)	Temperature (degree C)	20.360	Spectrum Type	HSQC-DEPT
		Title	HOBSANA HSQC D2O 120117 2122 RG=180 8 mg/0.8 ml		

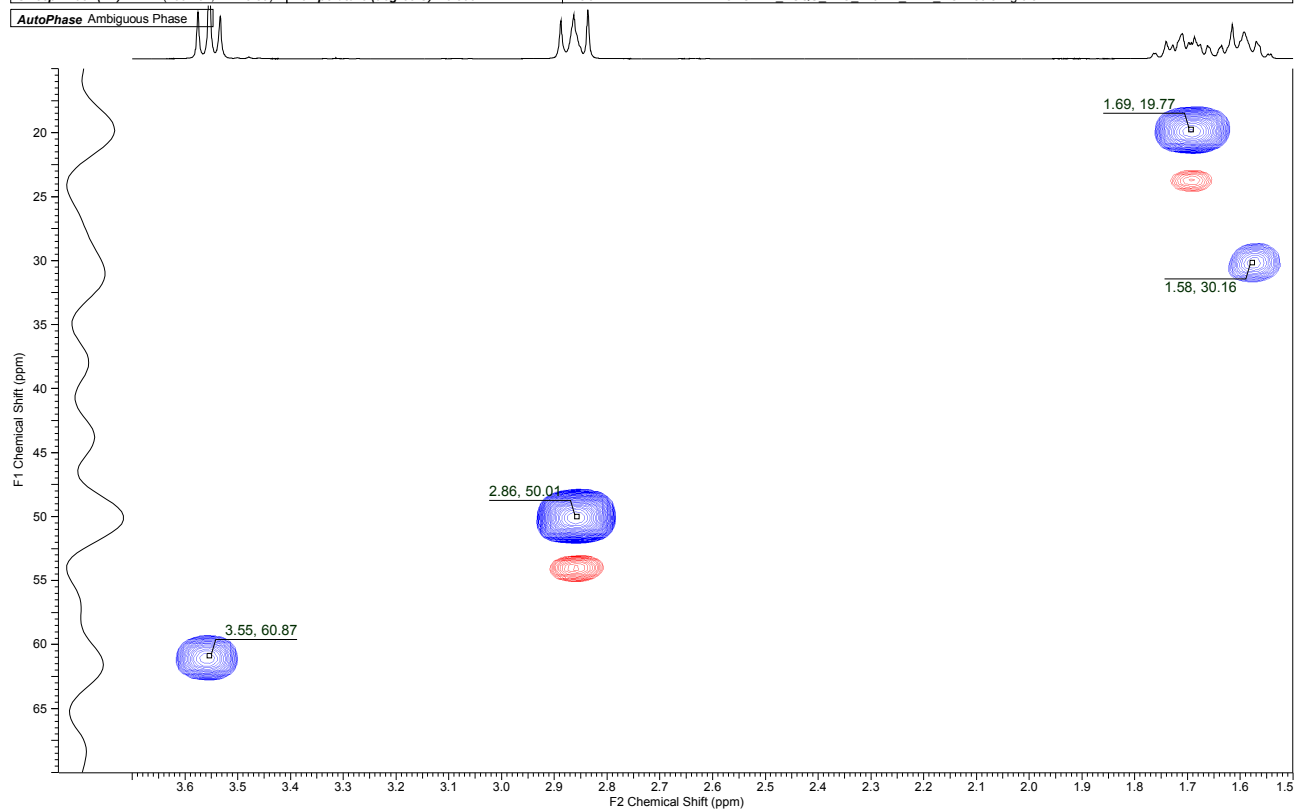


Figure S 58: HSQC-DEPT spectrum of HOBSANa

Acquisition Time (sec)	3.6438	Comment	BIBSANA 1H D2O 130117_2123_RG=180 8 mg/0.8 ml	Date	13 Jan 2017 09:14:56
Date Stamp	13 Jan 2017 09:14:56	File Name	E:\Documents\1\Molecules\Anal\BIBSANA\1fid	Frequency (MHz)	300.13
Nucleus	1H	Number of Transients	64	Origin	spect
Points Count	131072	Pulse Sequence	zg	Receiver Gain	181.00
Spectrum Offset (Hz)	1096.5878	Spectrum Type	STANDARD	Sweep Width (Hz)	4496.37
				Temperature (degree C)	20.460

No.	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
1	1.62	2	m	-	beta	[1.57 .. 1.67]
2	1.71	2	m	-	gamma	[1.67 .. 1.77]
3	2.86	24	dd	7.80	delta	[2.82 .. 2.91]
4	3.48	24	dd	6.30	alpha	[3.43 .. 3.53]

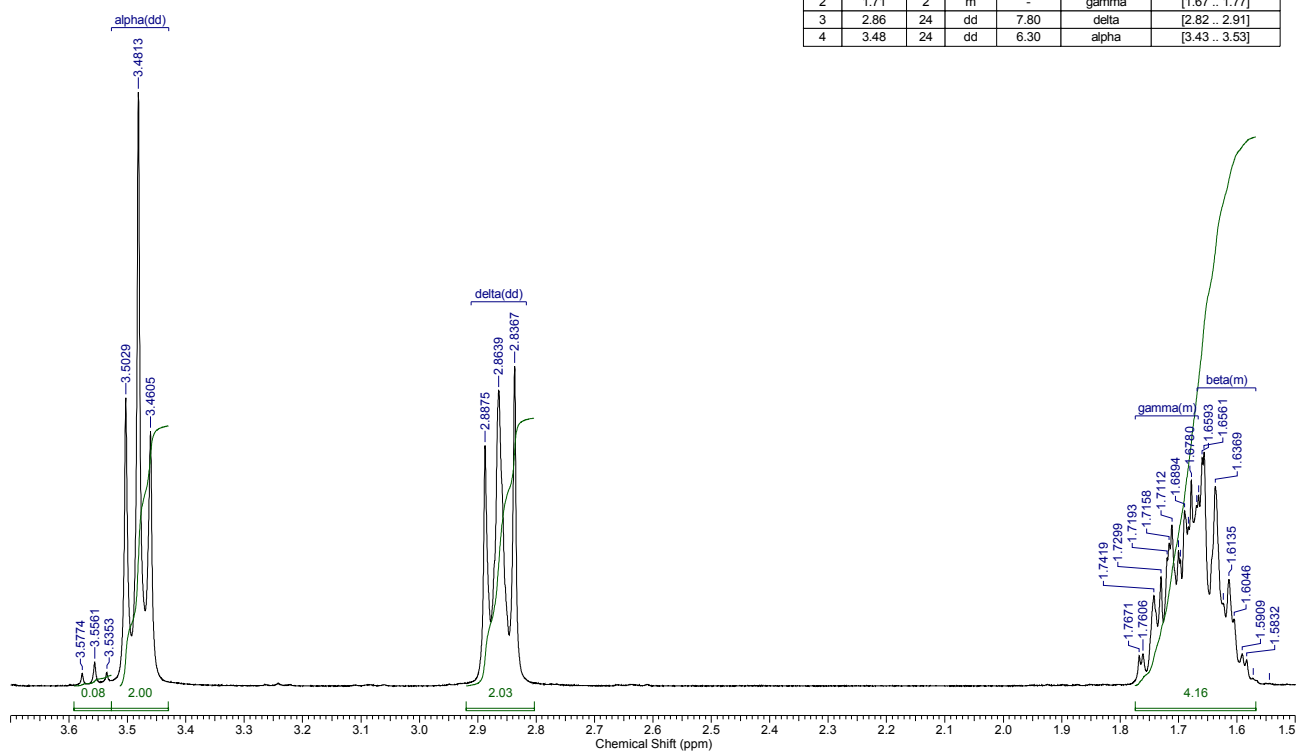


Figure S 59: Proton NMR spectrum of di(1,1'-sulfonatobutyl)ether disodium salt (disodium 4,4'-oxydibutane-1,1'-disulfonate, BIBSANA)

Acquisition Time (sec)	(0.0427, 0.0051)	Comment	5 mm BBO 1H-BB Z-GRD Z8284/0059	Date	13 Jan 2017 10:10:04
File Name	E:\Documents\1\Molecules\Anal\BIBSANA\2\ser	Frequency (MHz)	(300.13, 75.47)	Nucleus	(1H, 13C)
Number of Transients	32	Origin	spect	Original Points Count	(128, 64)
Points Count	(1024, 512)	Pulse Sequence	hsqcadelgp	Solvent	DMSO
Sweep Width (Hz)	(2994.67, 12475.59)	Temperature (degree C)	20.560	Title	BIBSANA HSQC D2O 130117_2123_RG=180 8 mg/0.8 ml

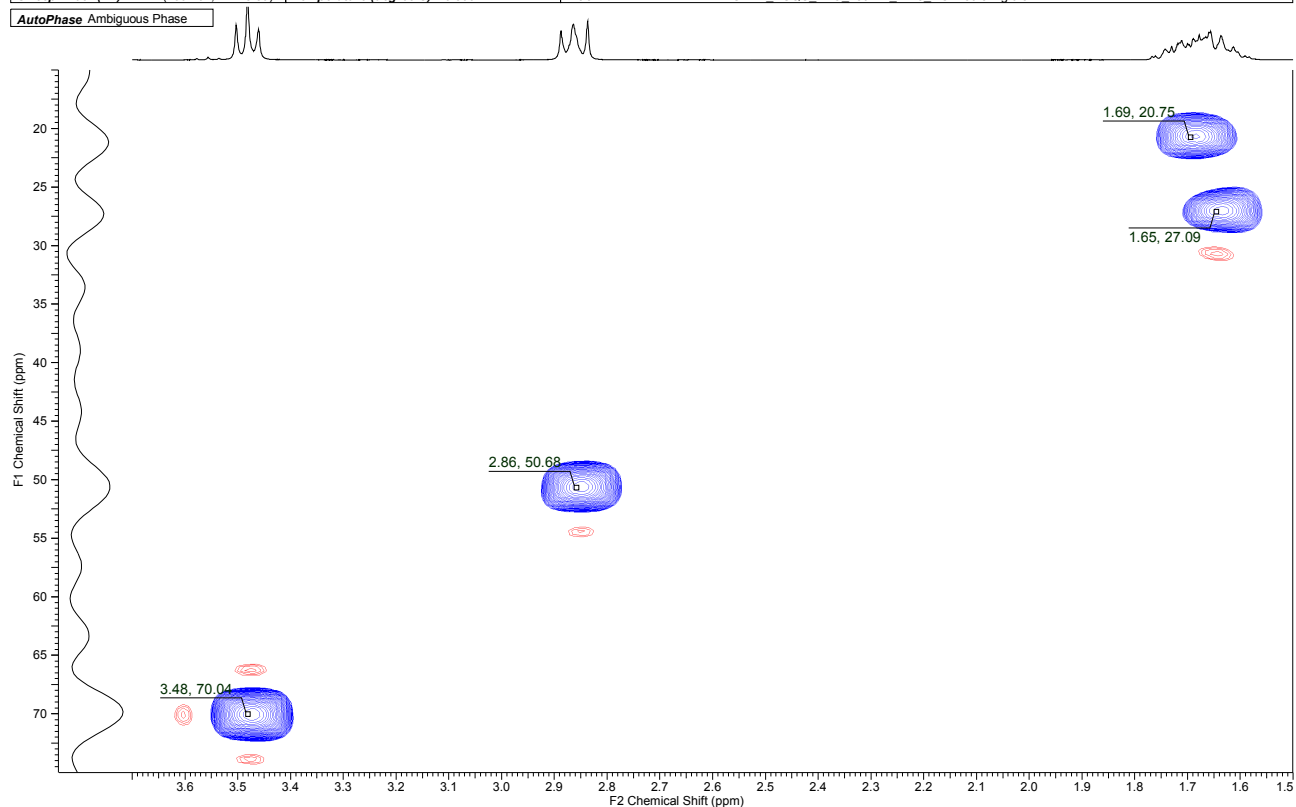


Figure S 60: HSQC-DEPT spectrum of BIBSANA

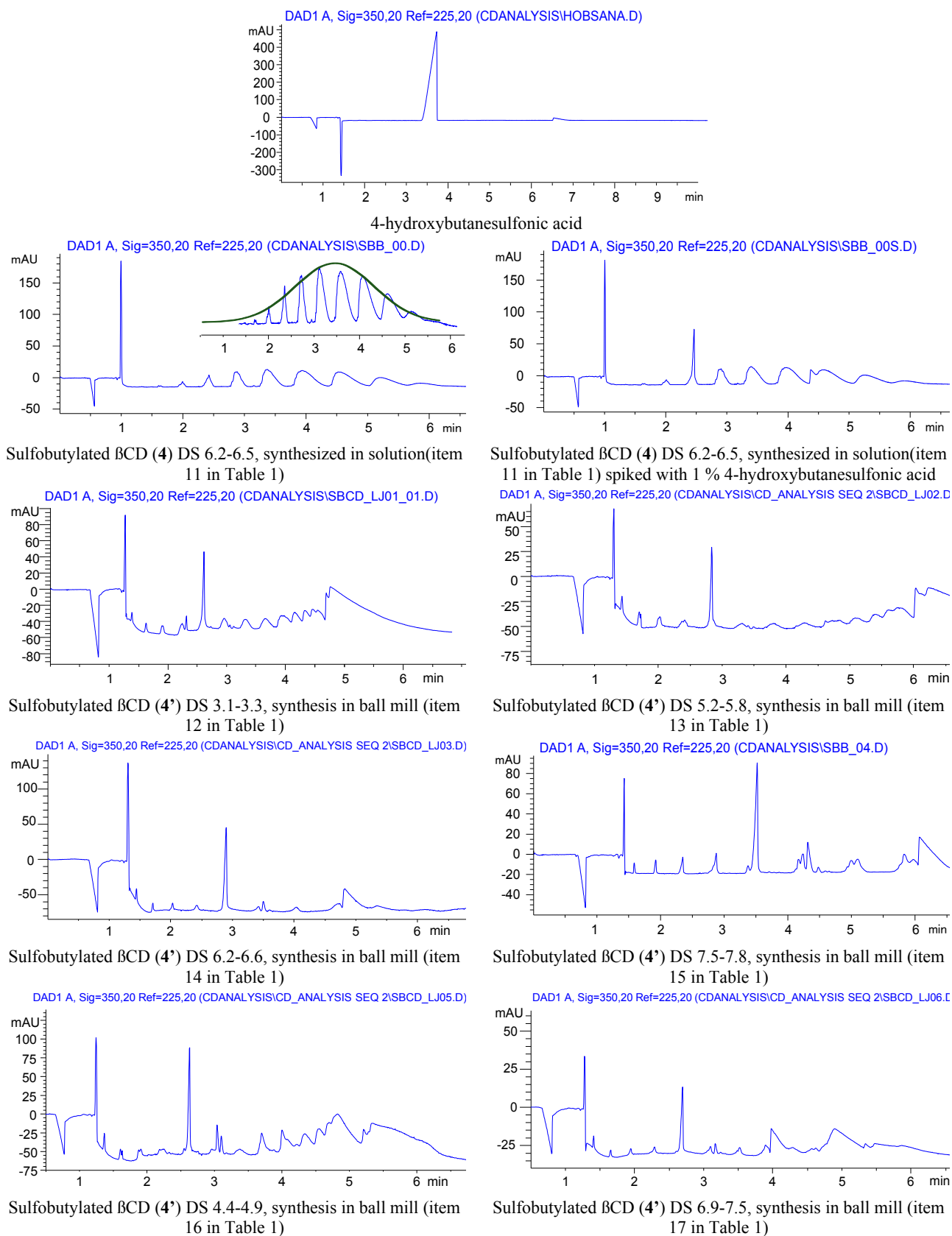
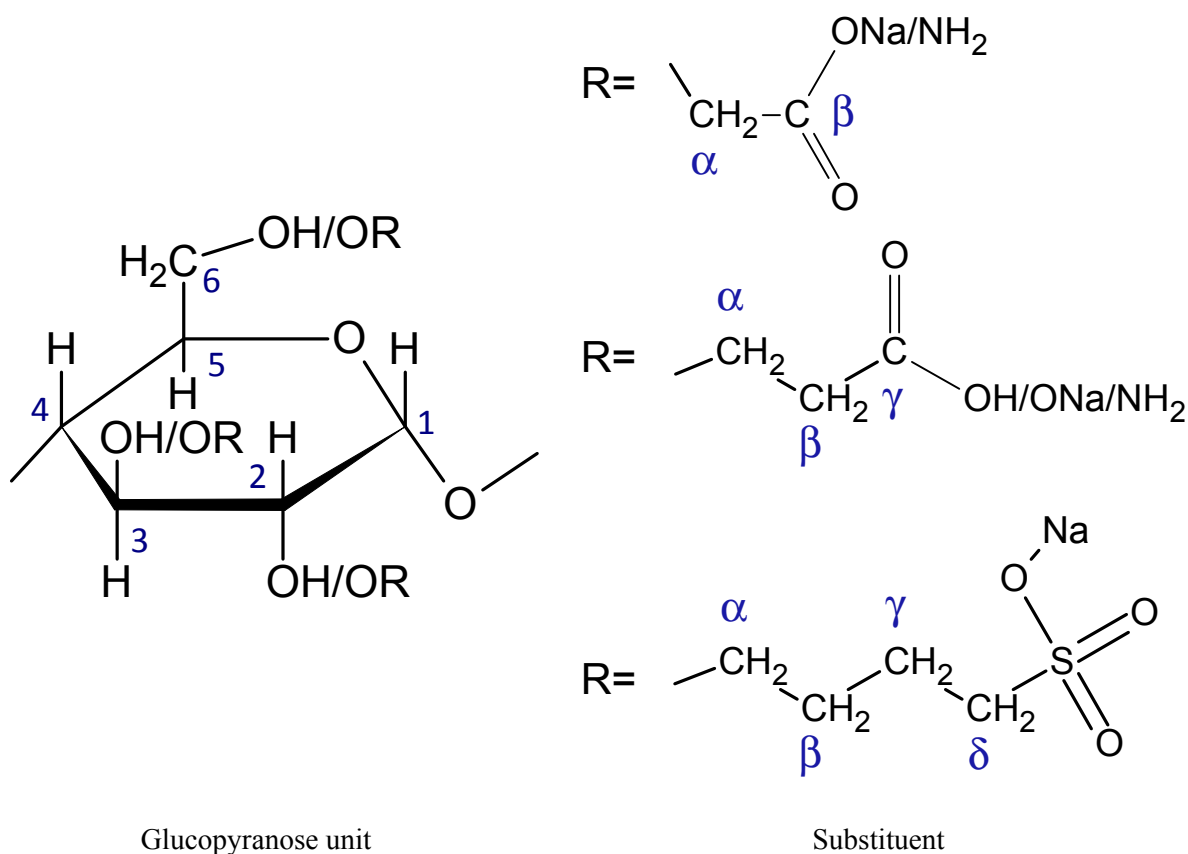


Figure S 61: Capillary electropherogram of sulfobutylated  $\beta$ CDs



Scheme 1

*Scheme S 1: Carbon atom numbering for the NMR assignment (OH: unsubstituted; OR: substituted)*

*Table S 1: Proton assignment of cyclodextrin derivatives prepared in solution (based on <sup>1</sup>H- and HSQC-DEPT experiments)*

	Compound 2 (CMβCD) <sup>1</sup>	Compound 3 (CEβCD) <sup>2</sup>	Compound 5 (CAMβCD) <sup>3</sup>	Compound 4 (SBβCD) <sup>4</sup>	HOBSANa <sup>5</sup>	BIBSANA <sup>6</sup>
C(1)H <sup>7</sup>	5.00, 5.21	4.99	4.97, 4.99	5.02, 5.11	--	--
C(2)H unsubst	3.54-3.61	3.56-3.68	3.41-3.63	3.48-3.61	--	--
C(2)H subst <sup>8</sup>	3.34-3.63	3.43-3.59	3.38-3.60	3.57-3.67	--	--
C(3)H unsubst <sup>8</sup>	3.84-4.04	3.79-3.96	3.63-3.98	3.70-3.90	--	--
C(3)H subst	--	--	--	3.70-3.83	--	--
C(4)H <sup>8</sup>	3.44-3.62	3.43-3.59	3.38-3.60	3.40-3.57	--	--
C(5)H <sup>8</sup>	3.69-3.88	3.73-3.92	3.63-3.98	3.76-3.90	--	--
C(6)H2 unsubst	3.70-3.91	3.70-3.89	3.61-3.93	3.71-3.90	--	--
C(6)H2 subst	--	3.72-3.81(?)	--	3.65-3.73	--	--
alpha to -O	3.86-3.95; 4.03-4.21	3.62-3.74	3.61-3.77	3.43-3.54	3.55	3.48
beta to -O	--	2.35-2.46	2.38-2.51	1.67	1.58	1.65
gamma to -O	--	--	--	1.71	1.69	1.69
delta to -O	--	--	--	2.86	2.86	2.86

<sup>1</sup> CM= Carboxymethyl; <sup>2</sup> CE=Carboxyethyl; <sup>3</sup> CAM=Carbamoyl ethyl; <sup>4</sup> SB=Sulfobutyl; <sup>5</sup> HOBSANa=4-hydroxybutanesulfonic acid sodium salt; <sup>6</sup> BIBSANA: di(1,1'-sulfonatolbutyl)ether disodium salt (disodium 4,4'-oxydibutane-1,1'-disulfonate); <sup>7</sup> main peaks only; <sup>8</sup> shaded assignments may be altered



Table S 2: Carbon assignment of cyclodextrin derivatives prepared in solution (based on <sup>13</sup>C- and HSQC-DEPT experiments)

	Compound 2 (CMβCD) <sup>1</sup>	Compound 3 (CEβCD) <sup>2</sup>	Compound 5 (CAMβCD) <sup>3</sup>	Compound 4 (SBβCD) <sup>4</sup>	HOBSANa <sup>5</sup>	BIBSANA <sup>6</sup>
C1 <sup>7</sup>	100.3, 102.0	102.1	103.1	99.2, 101.1	--	--
C2 unsubst	72.6	72.0	71.9	72.5	--	--
C2 subst <sup>8</sup>	81.1	80.7	81.4	78.3-80.4	--	--
C3 unsubst <sup>7,8</sup>	72.2, 73.8	73.3, 71.1	72.4	70.6	--	--
C3 subst	--	--	--	80.6	--	--
C4 <sup>8</sup>	81.4	81.5	81.4	80.5	--	--
C5 <sup>8</sup>	71.8	72.3	72.7	70.6	--	--
C6 unsubst	61.2	60.4	60.1	60.8	--	--
C6 subst	--	--	--	71.2	--	--
alpha to CD-O <sup>8</sup>	69.7, 70.4	68.6	66.3	70.2	60.9	70.0
beta to CD-O <sup>8</sup>	177.9	38.1	33.4	27.9	30.2	27.1
gamma to CD-O	--	180.7	177.4	21.0	19.8	20.8
			(174.3) <sup>9</sup>			
delta to CD-O	--	--	--	50.2	50.0	50.7

<sup>1</sup> CM= Carboxymethyl; <sup>2</sup> CE=Carboxyethyl; <sup>3</sup> CAM=Carbamoylethyl; <sup>4</sup> SB=Sulfobutyl; <sup>5</sup> HOBSANa=4-hydroxybutanesulfonic acid sodium salt; <sup>6</sup> BIBSANA: di(1,1'-sulfonatobutyl)ether disodium salt (disodium 4,4'-oxydibutane-1,1'-disulfonate); <sup>7</sup> main peaks only; <sup>8</sup> shaded assignments may be altered; <sup>9</sup> protonated carboxyl group (tentative) of the partially hydrolyzed product

Table S 3: Characteristic C=O IR bands of the prepared (carbonyl)ethyl group containing CD derivatives

	Compound 3 (CEβCD)	Compound 5 (CAMβCD) Item 9	Compound 5 (CAMβCD) Item 10
C=O as COO <sup>-</sup>	1570	1559	--
C=O as COOH	1726	--	--
C=O as CONH <sub>2</sub>	--	1667	1664

Table S 4: R<sub>F</sub>-values of the prepared compounds after twice run of the same plate in the same solvent mixture at 7 cm distance in 100 μg

βCD	0.53-0.54				
CMβCD <sup>1</sup>	0.05-0.48	strip with multiple spots	0.45-0.48	mono	
CEβCD <sup>2</sup>	0.15-0.47	strip with multiple spots	0.44-0.47	mono	
CAMβCD <sup>3</sup>	0.54-0.65	strip with multiple spots	0.55-0.57	mono	0.21-0.54 light brown strip no distinctive spots
SBβCD <sup>4</sup>	0.05-0.43	strip with multiple spots	0.40-0.43	mono	

<sup>1</sup> CM= Carboxymethyl; <sup>2</sup> CE=Carboxyethyl; <sup>3</sup> CAM=Carbamoylethyl; <sup>4</sup> SB=Sulfobutyl

## References

- [1] Strohalm, M.; Kavan, D.; Novák, P.; Volný, M. & Havlíček, V., mMass 3: A Cross-Platform Software Environment for Precise Analysis of Mass Spectrometric Data. *Analytical Chemistry*, **2010**, *82*, 4648-4651
- [2] mMass v5.5, An Open Source Mass Spectrometry Tool, [www.mmass.org](http://www.mmass.org)