

Figure S1, Related to Figure 1. IRTKS is highly expressed in the intestinal crypt domain. (A) Maximum intensity projection of the IRTKS stained mouse intestinal organoid used in Figure 1B. Scale bar, 40 μ m.



Figure S2, Related to Figure 3. Split channel images of IRTKS overexpression and KD/rescue cells. (A-E) SIM projections of Ls174T-W4 cells expressing EGFP-IRTKS constructs (green) and stained with phalloidin (magenta); single channel zooms of the BB are below each image. Scale bars, 5 μ m. (F) Western blot analysis of expression levels of EGFP-IRTKS variants in lysates from Ls174T-W4 cells. Both IRTKS and EGFP antibodies were used because the IRTKS antibody is towards a peptide sequence within the SH3 domain, thus the EGFP-I-BAR alone construct will not be targeted. GAPDH was used as a loading control. (G) Western blot analysis of endogenous IRTKS levels in lysates from WT Ls174T-W4 cells (untransfected), shRNA scramble control, and shRNA IRTKS KD. GAPDH was used as a loading control. (H-I) FRAP curves of shRNA scramble control and shRNA IRTKS KD in Ls174T-W4 cells. (J-N) SIM projections of IRTKS KD Ls174T-W4 cells expressing refractory EGFP-IRTKS constructs in rescue experiments. Zooms of the BB and single IRTKS channel below each image. Scale bars, 5 μ m.



Figure S3, Related to Figure 4. The impact of EPS8 and IRTKS co-expression on the formation of actin-based protrusions. (A) SIM projection of a Ls174T-W4 cell expressing EGFP-EPS8. Dashed box indicates zoom; scale bar, 5 μ m. (B) Quantitation of microvillar length comparing WT Ls174T-W4 cells, EGFP-EPS8 overexpression and EGFP-IRTKS and mCherry-EPS8 dual overexpression, which do not affect microvillar length. 25 cells/condition, 10 microvilli/cell; student's t test (*p<0.0001, ns, not significant) was used to determine the significance. (C) TIRF live-cell imaging of a B16F1 melanoma cell expressing EGFP-EPS8 (green) and mCherry-IRTKS (magenta). Dashed box indicates video frames with time in seconds. Scale bar, 10 μ m. (D-F) *En face* (x-y) and lateral (x-z) SIM projections of HeLa cells expressing either EGFP-IRTKS (D), EGFP-EPS8 (E), or EGFP-IRTKS and mCherry-EPS8 (F) and stained with phalloidin. Scale bars, 10 μ m.



Figure S4, Related to Figures 5 & 6. Images of EPS8 KD/ rescue Ls174T-W4 cells. (A) Pulldown of FLAG-tagged IRTKS SH3 domain co-expressed with EGFP-tagged EPS8 or EGFPtagged EPS8 Δ PR1 reveals binding between EPS8 and the IRTKS SH3 domain. (B) Western blot analysis of endogenous EPS8 levels in lysates from WT Ls174T-W4 cells (untransfected), shRNA scramble control, and shRNA EPS8 KD. GAPDH was used as a loading control. (C) SIM projection of an EPS8 KD Ls174T-W4 cell expressing refractory EGFP-EPS8 (EGFP-EPS8r) construct in rescue experiments; dashed box indicates zoom of the BB. Scale bar, 5 µm. (D) SIM projection of an EPS8 KD Ls174T-W4 cell expressing refractory EGFP-EPS8 Δ PR1 (EGFP-EPS8 Δ PR1r) construct in rescue experiments; dashed box indicates zoom of the BB. Scale bar, 5 µm. (5 µm.



Figure S5, Related to Figure 6. Additional characterization of the IRTKS/EPS8 complex in B16F1 Melanoma Cells. (A-D) TIRF live-cell imaging of B16F1 melanoma cells expressing either EGFP-IRTKS SH3* (green) and mCherry-EPS8 (magenta) (A), EGFP-IRTKS Δ WH2 (green) and mCherry-EPS8 (magenta) (B), or EGFP-EPS8 Δ PR1 (green) and mCherry-IRTKS (magenta) (C), and EGFP-EPS8 Δ PR1 (green) and mCherry-UtrCH (magenta) (D). Dashed boxes indicate zooms. Scale bars, 10 µm. (E) Quantitation of the number of filopodia per µm of cell perimeter in B16F1 melanoma cells from C-E; at least 15 cells/condition. All error bars indicate mean ± SD; all p values calculated using a t test (*p<0.033, **p<0.002, ***p<0.0002, ****p<0.0001).