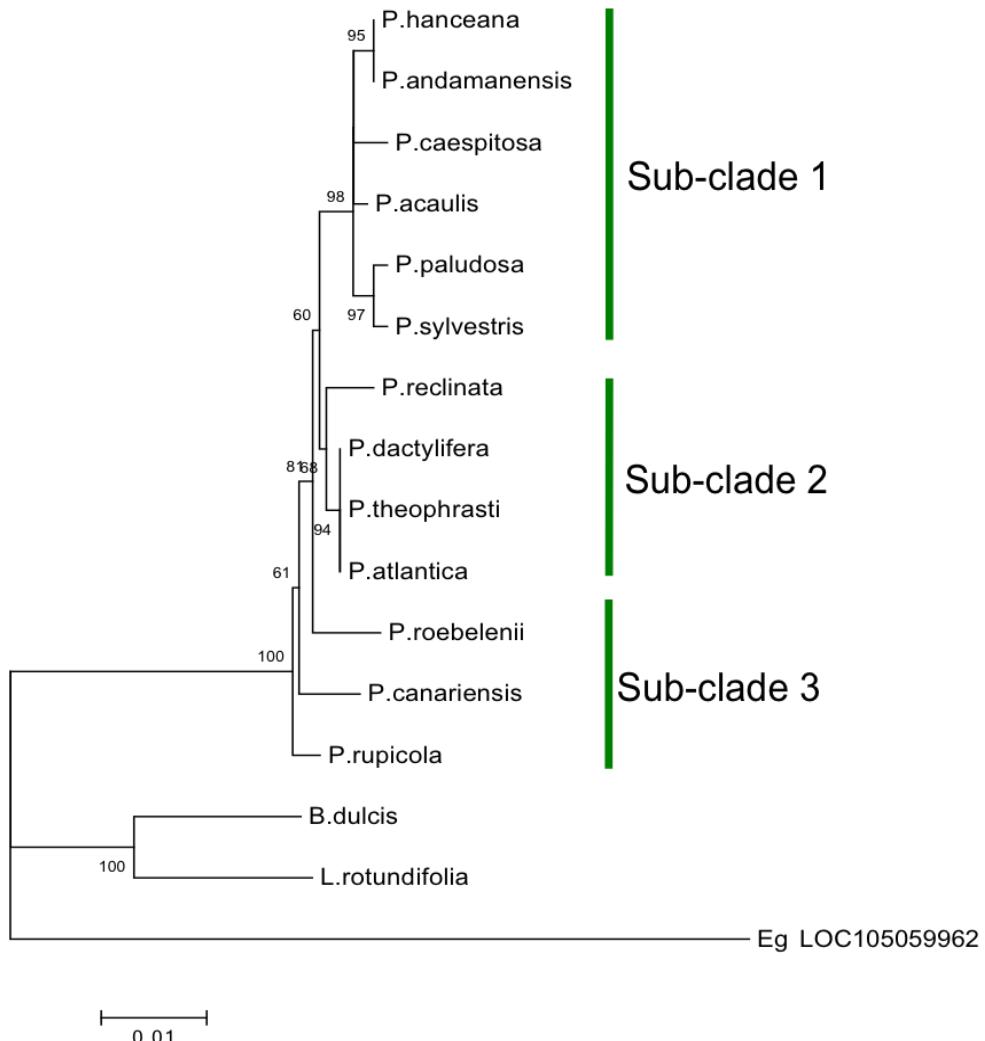
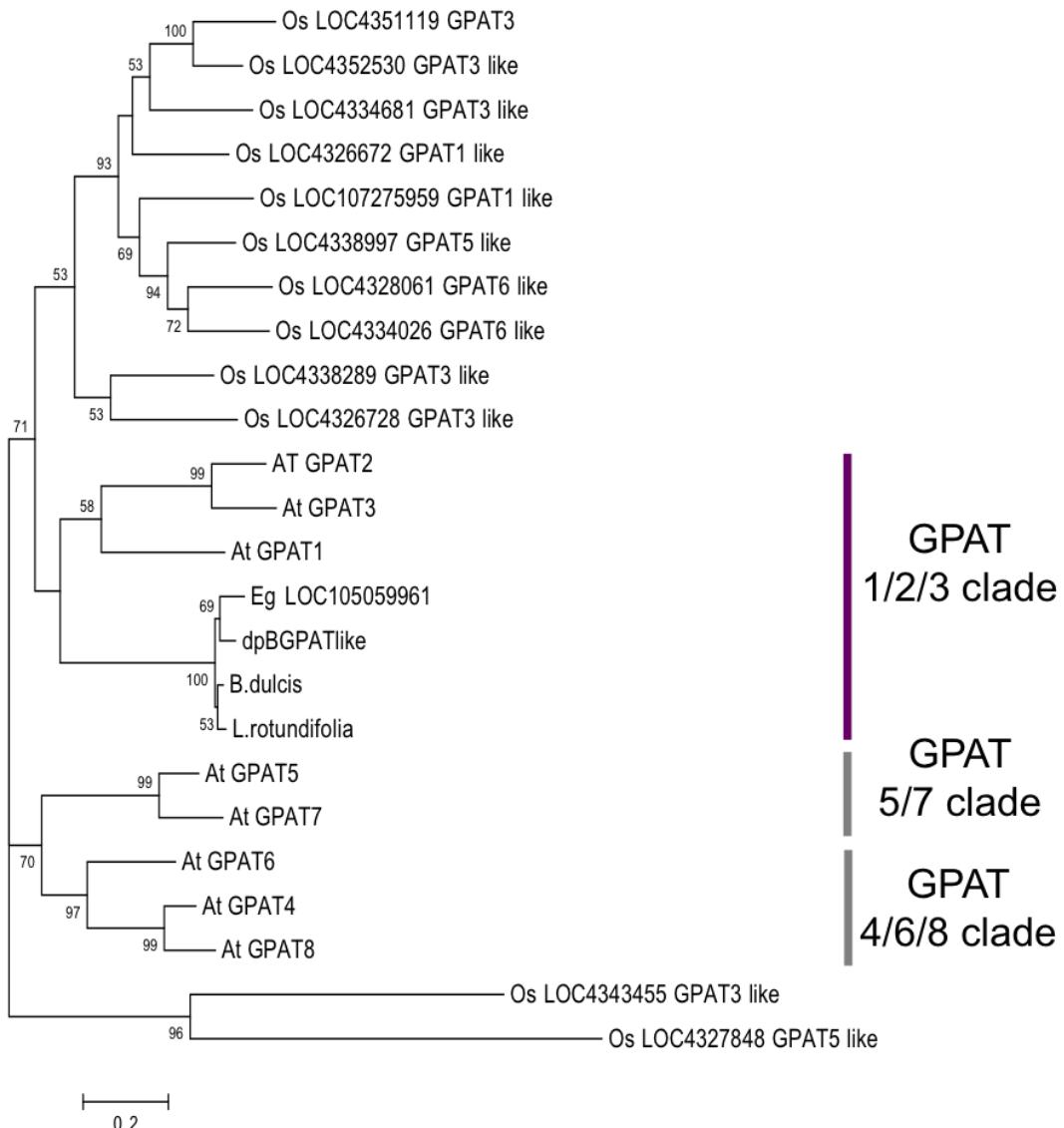


**Genus-wide sequencing supports a two-locus model for sex-determination in *Phoenix***

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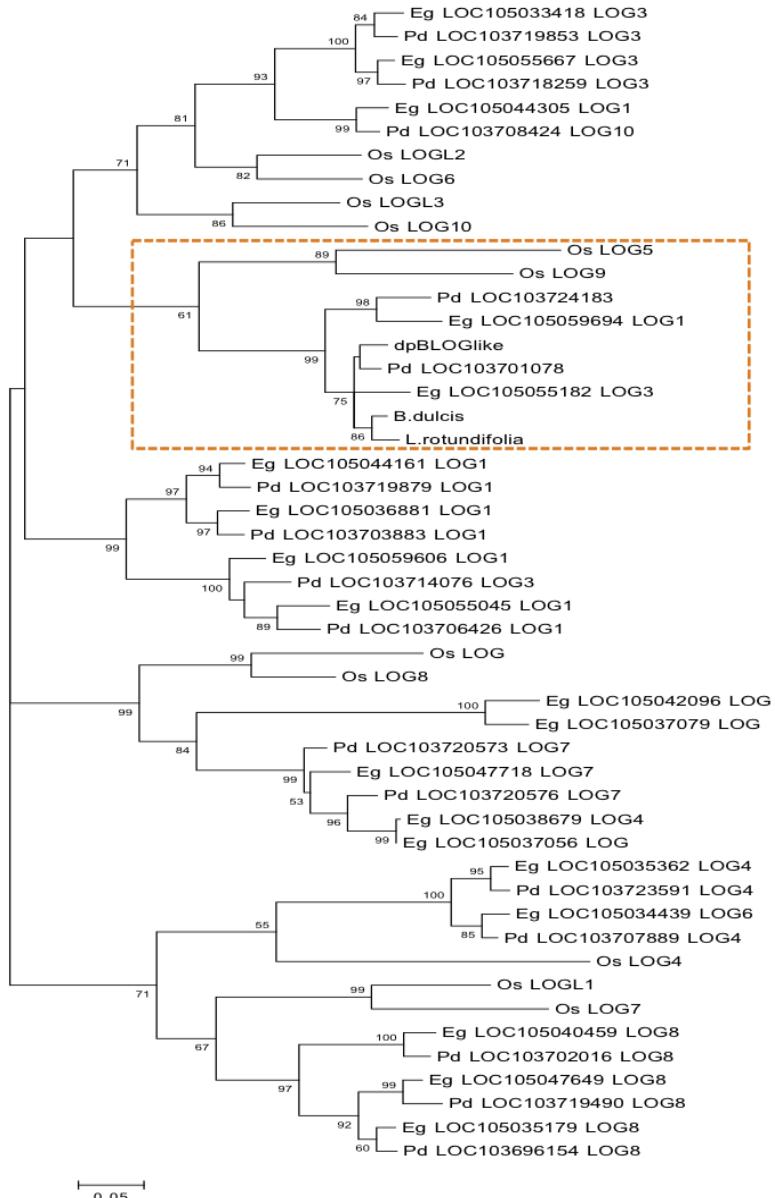


**Supplementary Figure 1. Cyp703 phylogeny.** Maximum likelihood phylogenetic tree based on alignment of k-mer derived *CYP703* cDNA sequences from *Phoenix* and the hermaphrodites *Brahea* and *Livistona*. Branch support values at the nodes (if >50%) were obtained by 1000 bootstrap replications. Branch lengths represent number of substitutions per site. The oil palm *CYP703A* sequence (*Eg LOC105059962*) was utilized to infer *de novo* sequences from other palm species. *Eg*= *Elaeis guineensis*. *B. dulcis*=*Brahea dulcis*. *L. rotundifolia*=*Livistona rotundifolia*

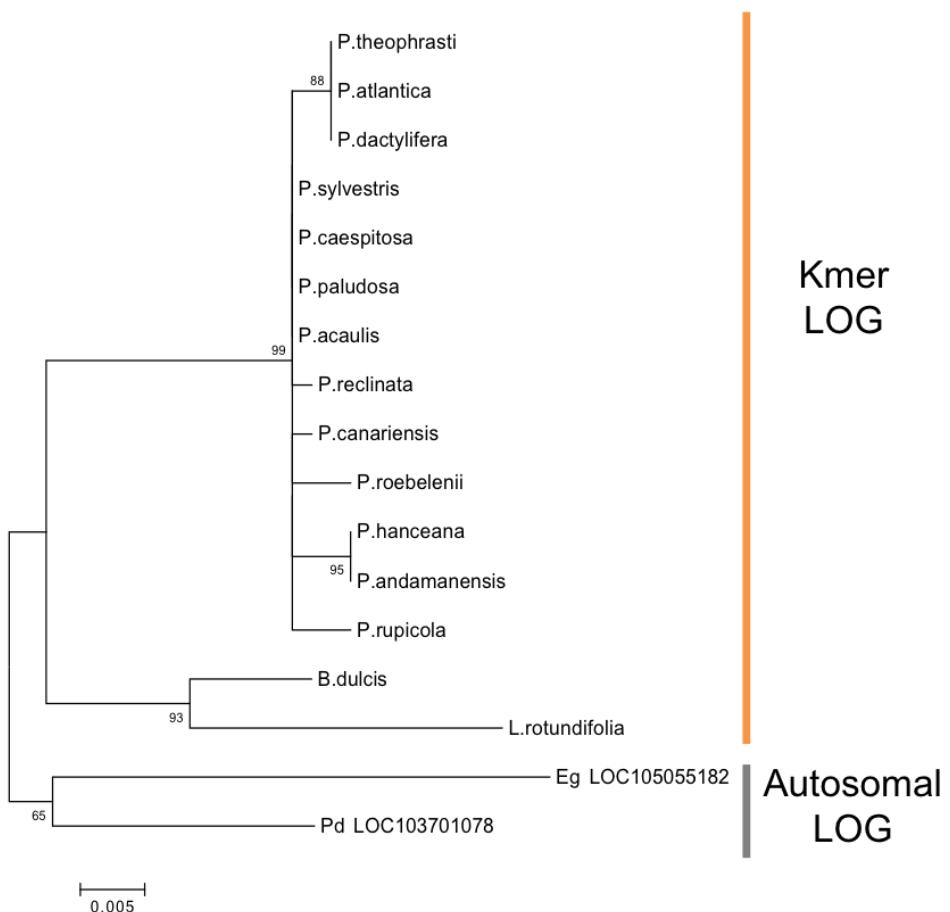


**Supplementary Figure 2. GPAT phylogeny.** Maximum likelihood phylogenetic tree based on alignment of GPAT sequences from *Arabidopsis*, rice and k-mer inferred sequences from *P. dactylifera*, *Brahea* and *Livistona*.

Branch support values indicated at the nodes (if >50%) were obtained by 1000 bootstrap replications. Branch lengths represent number of substitutions per site. The oil palm *GPAT* sequence (Eg\_LOC105059961) was utilized to infer *de novo* GPAT sequences from other palm species. *Arabidopsis* and rice sequences were described by Beisson *et al.*<sup>1</sup> and Men *et al.*<sup>2</sup>. Os= *Oryza sativa*. At= *Arabidopsis thaliana*. Eg= *Elaeis guineensis*. B. dulcis= *Brahea dulcis*. L. rotundifolia = *Livistona rotundifolia*. dpBGPATlike= date palm kmer *GPAT* gene

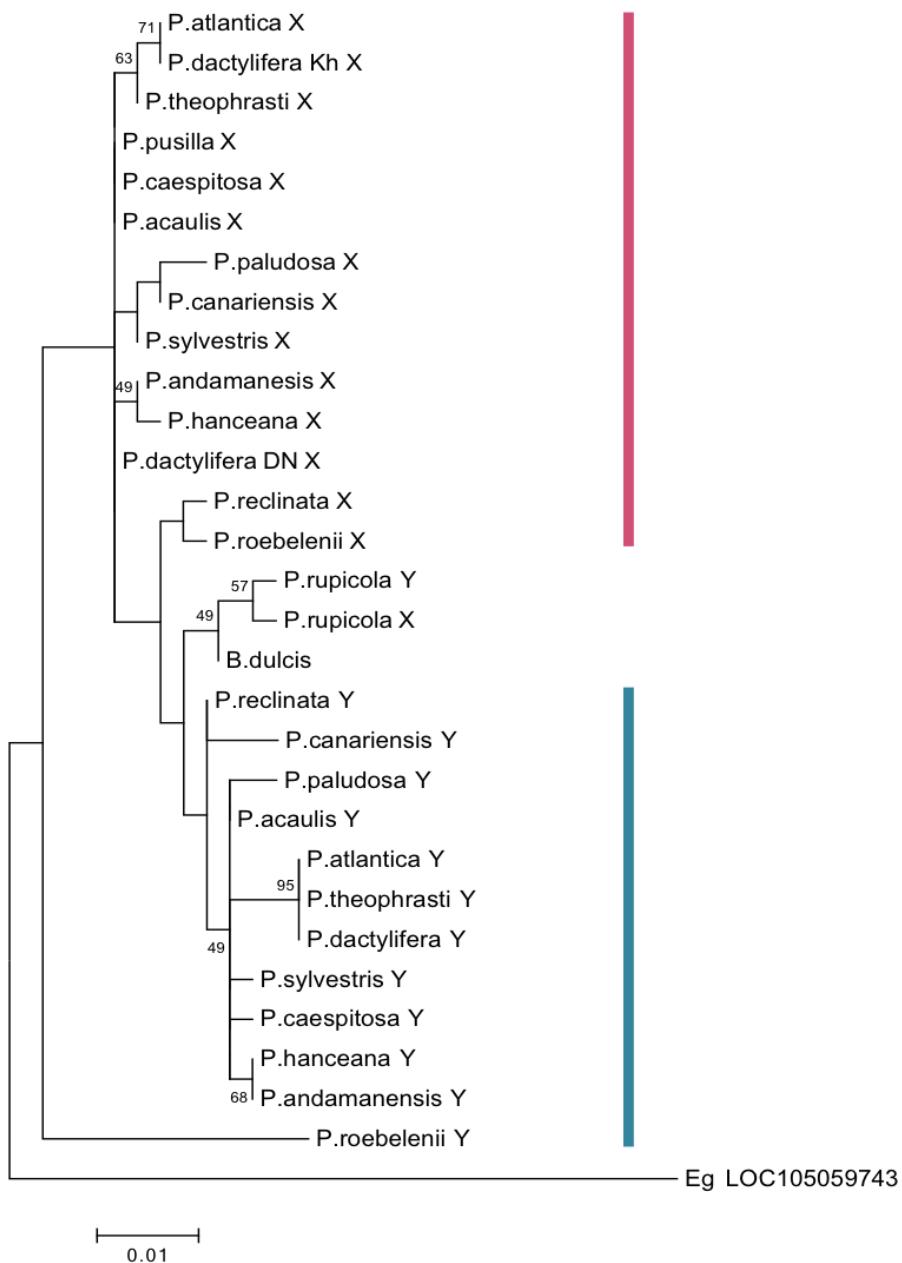


**Supplementary Figure 3. LOG phylogeny.** Maximum likelihood phylogenetic tree based on alignment of all LOG sequences from oil palm, date palm, rice and k-mer inferred sequences from *P. dactylifera*, *Brahea* and *Livistona*. Branch support values indicated at the nodes (if >50%) were obtained by 1000 bootstrap replications. Branch lengths represent number of substitutions per site. The oil palm LOG sequence (Eg\_LOC105055182) was utilized to search the NCBI database for similarities in the entire date palm and oil palm genomes. Os= *Oryza sativa*. Pd= *Phoenix dactylifera*. Eg= *Elaeis guineensis*. B. dulcis= *Brahea dulcis*. L. rotundifolia = *Livistona rotundifolia*. dpBLOGlike= date palm male-specific kmer LOG gene.



**Supplementary Figure 4. Autosomal and male-specific LOG phylogeny.**

Maximum likelihood phylogenetic tree based on alignment of k-mer derived LOG sequences from *Phoenix* and the hermaphrodites *Brahea* and *Livistona*. Branch support values indicated at the nodes were obtained by 1000 bootstrap replications. Branch lengths represent number of substitutions per site. The Oil Palm LOG sequence (Eg\_LOC105055182) was utilized to infer *de novo* sequences from other palm species. The *P. dactylifera* autosomal LOG (Pd\_LOC103701078) was included for reference. Eg= *Elaeis guineensis*. *B. dulcis*= *Brahea dulcis*. *L. rotundifolia* = *Livistona rotundifolia*



**Supplementary Figure 5. Cytidine deaminase phylogeny.** Maximum likelihood phylogenetic tree based on alignment of kmer derived cytidine deaminase cDNA sequences from *Phoenix* and *Brahea*. Branch support values at the nodes (if >40%) were obtained by 1000 bootstrap replications.

Branch lengths represent number of substitutions per site. Cytidine deaminase sequences were inferred *de novo* from sequenced male and female BACs. The oil palm cytidine deaminase gene (Eg\_LOC105059743) was included as an outgroup. Eg= *Elaeis guineensis*. B. dulcis= *Brahea dulcis*. P. dactylifera DN= P. dactylifera Deglet Noor. P. dactylifera Kh= P. dactylifera Khalas. Male alleles for the genus clustered together in general (blue line – Y alleles, red line – X alleles)

**Supplementary Table 1. A table with all samples used for kmer analysis and validation test in this study**

Species	Sex	ID	Collection	Short name	Sequence coverage
<b>Used in kmer analysis</b>					
<i>P. dactylifera</i> ( <i>Deglet Noor</i> )*	F	RIV 2154	USDA, CA	dnPdF	71.3517
<i>P. dactylifera</i> ( <i>Deglet Noor</i> )	M	RIV 2165 PL	USDA, CA	dnPdM	67.531
<i>P. dactylifera</i> ( <i>Khalas</i> )	F	khlsF	Qatar	khlsF2016	69.4486
<i>P. roebilinii</i>	F	RIV 2922 PL	USDA, CA	P03roeF	23.3215
<i>P. roebilinii</i>	M	RIV 2963 PL	USDA, CA	P5roeM	37.2331
<i>P. canariensis</i>	F	RIV 6839 PL	USDA, CA	P08canF	34.7572
<i>P. canariensis</i>	M	RIV 6840 PL	USDA, CA	P09canM	67.0642
<i>P. hanceana</i> ( <i>loureiroi</i> ?)	M	RIV 6823 PL	USDA, CA	P13hanM	28.1128
<i>P. hanceana</i> ( <i>loureiroi</i> ?)	F	RIV 6791 PL	USDA, CA	P14hanF	27.5131
<i>P. paludosa</i>	F	RIV 6817 PL	USDA, CA	P17palF	38.5184
<i>P. paludosa</i>	M	RIV 6819 PL	USDA, CA	P19palM	45.7222
<i>P. reclinata</i>	M	RIV 6807 PL	USDA, CA	P20recM	58.0921
<i>P. reclinata</i>	F	RIV 6811 PL	USDA, CA	P21recF	41.3914
<i>P. sylvestris</i>	F	RIV 2248 PL	USDA, CA	P23sylF	32.3765
<i>P. sylvestris</i>	M	RIV 7395 PL	USDA, CA	P25sylM	27.5639
<i>P. andamanensis</i>	F	N/A	UMH, Spain	Q01andF	36.6741
<i>P. andamanensis</i>	M	N/A	UMH, Spain	Q03andM	41.7865
<i>P. caespitosa</i>	F	N/A	UMH, Spain	Q07caeF	55.0238
<i>P. caespitosa</i>	M	N/A	UMH, Spain	Q09caeM	45.7855
<i>P. acaulis</i>	F	N/A	UMH, Spain	P02acaF	31.0049
<i>P. acaulis</i>	M	N/A	UMH, Spain	Q10acaM	29.0575
<i>P. theophrastii</i>	F	N/A	UMH, Spain	Q17theF	33.7925
<i>P. theophrastii</i>	M	N/A	UMH, Spain	Q19theM	93.2802
<i>P. atlantica</i>	F	N/A	UMH, Spain	Q22atlF	48.0664
<i>P. atlantica</i>	M	N/A	UMH, Spain	Q23atlM	48.9665
<i>P. rupicola</i>	F	N/A	UMH, Spain	P0XrupF	22.906
<i>P. rupicola</i>	M	N/A	UMH, Spain	Q15rupM	36.4488
<i>P. pusilla</i>	F	N/A	UMH, Spain	Q14pusF	30.7199
<i>Brahea dulcis</i>	Herm	N/A	Huntington Gardens, CA	braheaSpp	44.8707
<i>Livistona rotundifolia</i>	Herm	N/A	Huntington Gardens, CA	livistonaSpp	32.4906
<b>Used in validation test</b>					
<i>P. acaulis</i>	F	RIV 3363 PL	USDA, CA	P01acaF	38.33
<i>P. roebilinii</i>	F	RIV 2939 PL	USDA, CA	P04roeF	20.74
<i>P. canariensis</i>	F	RIV 6836 PL	USDA, CA	P06canF	31.86
<i>P. canariensis</i>	M	RIV 6837 PL	USDA, CA	P07canM	26.79
<i>P. reclinata</i>	F	RIV 2975 PL	USDA, CA	P10senF	34.4
<i>P. hanceana</i> ( <i>loureiroi</i> )	F	RIV 6822 PL	USDA, CA	N/A	40.88
<i>P. hanceana</i> ( <i>loureiroi</i> )	M	RIV 6789 PL	USDA, CA	N/A	33.8
<i>P. hanceana</i> ( <i>loureiroi</i> )	F	RIV 6790 PL	USDA, CA	N/A	32.77
<i>P. hanceana</i> ( <i>loureiroi</i> )	M	RIV 6793 PL	USDA, CA	N/A	30.81

<i>P. paludosa</i>	M	RIV 6818 PL	USDA, CA	N/A	31.35
<i>P. reclinata</i>	F	RIV 6812 PL	USDA, CA	N/A	30.73
<i>P. sylvestris</i>	F	RIV 2256 PL	USDA, CA	N/A	43.32
<i>P. sylvestris</i>	M	RIV 2249 PL	USDA, CA	N/A	43.09

\* *P.* denotes *Phoenix*

**Supplementary Table 2. A table of contigs/scaffolds used in this study**

Name	X, Y, or Autosomal	Length (bp)	NCBI Accession
dpS5Y	Y	567483	MH680965
dpS5X	X	566497	MH680976
dpS5560CH1	Autosomal	1371428	MH680988
dpS5560CH2	Autosomal	1366600	MH680991
dpS5784H1	X	706543	MH680973
dpS5784H2	Y	705158	MH680984
dpS12Y	Y	89036	MH680966
dpS12X	X	89768	MH680977
dpS5828CH1	Autosomal	225201	MH680987
dpS5828CH2	Autosomal	225793	MH680990
dpS5887CH1	Autosomal	30183	MH680989
dpS5887CH2	Autosomal	30198	MH680992
dpS10Y	Y	34743	MH680969
dpS10X	X	34736	MH680980
dpS9X	X	43944	MH680975
dpS9Y	Y	41807	MH680986
dpS6X	X	89164	MH680970
dpS6Y	Y	91697	MH680981
dpS8Y	Y	105672	MH680967
dpS8X	X	104842	MH680978
dpS6284H1	Y	144017	MH680968
dpS6284H2	X	143984	MH680979
dpS11X	X	27585	MH680974
dpS11Y	Y	27658	MH680985
dpS6694H1	Y	147216	MH680971
dpS6694H2	X	147682	MH680982
dpS4X	X	201024	MH680972
dpS4Y	Y	201034	MH680983
dpB13X	X	198875	MH680995
dpB3X	X	282833	MH680993
dpB1X	X	137710	MH680996
dpBLOGautosomal1	Autosomal	101049	MH680997
dpBLOGautosomal2	Autosomal	135777	MH680964
dpB7X	X	349331	MH680994
dpB3Y	Y	289622	MH681003
dpBCYPlike	Y	274394	MH680999
dpB1Y	Y	130775	MH681004
dpBGPATlike	Y	238144	MH681000
dpBLOGlike	Y	111099	MH681001
dpB2Y	Y	781564	MH681002
dpS2X	X	303122	MH680998

**Supplementary Table 3. PCR primers used for identifying male specific regions**

Primer name	Primer sequence	Gene/Contig amplified	Amplicon size(bp)	Genic amplicon
dpBCYP1F	5'- AGTGAGTTCAACTACTACGAT -3'	CYP contig	372	No
dpBCYP1R	5'- GTCATAAACTTGACTAAAGACC -3'	Cyp contig		
dpBCYP2F	5'- CTTCGATTTCTCTTGATCTC -3'	CYP contig	418	No
dpBCYP2R	5'- TCAATCCCTAAACCACTAAC -3'	Cyp contig		
dpBGPAT1F	5'- AGAAAACCTGATATGCTCTTG -3'	GPAT contig	450	Yes
dBpGPAT1R	5'- TGTGATGCACCTGGTAACACT -3'	GPAT contig		
dpBGPAT2F	5'- GGGTCCATTATAAAGTCTTGAA -3'	GPAT contig	550	No
dBpGPAT2R	5'- GCGTGTGATATACATTGTCCTA -3'	GPAT contig		
dpBLOG1F	5'- ATGTAGCTAGACAACCAACA -3'	LOG contig	350	No
dpBLOG1R	5'- CAGCTTATCAGATGACACAATC -3'	LOG contig		
dpBCLOGauto_1F	5'- TGCTTATGTTCTCTTGCTACG -3'	Autosomal log	440	No
dpBCLOGauto_1R	5'- ATCATGGGTGAGAGAGAGTTAC -3'	Autosomal log		
dpBCLOGauto_2F	5'- ATGATGAGAGAGCTATTGTTCA -3'	Autosomal log	365	Yes
dpBCLOGauto21R	5'- AATCATCTGATCAAATGAGAG -3'	Autosomal log		
CYP703_ORF_1F	5'- GAATGGTAAGATCTTGAAGTCC -3'	Cyp Gene	1500	Yes
CYP703_ORF_1R	5'- TAGATGTGGTTCTTGTGTCTGT -3'	Cyp Gene		

## **Supplementary References**

1. Beisson, F., Li, Y., Bonaventure, G., Pollard, M. & Ohlrogge, J. B. The Acyltransferase GPAT5 Is Required for the Synthesis of Suberin in Seed Coat and Root of Arabidopsis. *PLANT CELL ONLINE* **19**, 351–368 (2007).
2. Men, X. *et al.* Glycerol-3-Phosphate Acyltransferase 3 (OsGPAT3) is required for anther development and male fertility in rice. *J. Exp. Bot.* **68**, 513–526 (2017).