

Appendix for Evrin et al, "Histone H2A-H2B binding by Pol α in the eukaryotic replisome contributes to the maintenance of repressive chromatin"

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Appendix Figure S1

Tethering of Pol α to the eukaryotic replisome is largely but not completely dependent upon Ctf4 and the CIP-box of Pol1. Control cells (YSS10), *ctf4* Δ (YGDP975), *pol1-4A* cells with mutations in the CIP-box (YCE1073) and *pol1-6A* with mutations in the histone-binding motif (YCE1075) were synchronized in G1-phase at 30°C and then released into S-phase for 20 minutes. S-phase samples used to prepare cell extracts, from which the CMG helicase component Mcm4 was isolated by immunoprecipitation. The associated replisome components were monitored by immunoblotting. The asterisk denotes a non-specific band in the cell extract samples that is recognized by the anti-Ctf4 antibody.

Appendix Figure S2

Telomere length is not affected by mutation of the CIP-box or histone-binding motifs of Pol1. Genomic DNA was extracted from the indicated strains and digested with Xho1, before transfer to nitrocellulose membrane and hybridization with a radiolabelled probe corresponding to the Y' element of yeast telomeres. Telomere length is reduced in *tel1* Δ cells that lack the Tel1 checkpoint kinase, and increased in *pol1-F1463A* cells in which primase is displaced from the carboxyl terminus of Pol1. In contrast, telomere length is not affected by histone-binding mutations in Mcm2 (Mcm2-3A) or Pol1 (Pol1-2A2 and Pol1-6A), or by displacement of Pol1 from the replisome (Pol1-4A).

Appendix Figure S3

Salt sensitivity of the interaction of Pol1NT with chromatin-derived histone complexes. Cells expressing ProteinA-tagged Pol1NT (YCE39) were grown and processed as for the experiments in Figure 3, except that the cell extracts were prepared in the presence of the indicated concentrations of KOAc.

Appendix Figure S4

Pol1NT and FACT can bind simultaneously to the same chromatin-derived histone complexes. **(A)** Repeat of the experiment in Figure 3E. The cells expressed GAL-Pol1NT and also had a second copy of *SPT16* at the *leu2* locus (in addition to the endogenous *SPT16* locus), expressing MYC-tagged versions of either wild type Spt16 (YCE248) or Spt16 with a small deletion at the carboxyl terminus (YCE250) that abolishes interaction of FACT with H3-H4 tetramers. After arresting cells in G2-M phase, cell extracts were prepared and treated with DNase to release chromatin-derived histone complexes, before isolation of Pol1NT by immunoprecipitation. Pol1NT associated in vitro with chromatin-derived histone complexes from both strains, but only co-purified with full-length Spt16-9MYC, rather than Spt16-ΔCT-9MYC (note that endogenous Spt16 was also present in both extracts, but is not shown in the figure). **(B)** MYC-tagged Spt16 was isolated from the same extracts, confirming that full-length Spt16-9MYC was able to co-purify with chromatin-derived histone complexes, but Spt16-9MYC was not. It was not possible to monitor association of MYC-tagged Spt16 with Pol1NT in this case, as the ProteinA tag on Pol1NT associated with the antibodies used to isolate Spt16-9MYC. **(C)** Similar experiment to those described above, except that expression of GAL-Pol1NT was induced in G1-phase cells that were then released into S-phase. Samples were taken 20' after release into S-phase and used to prepare cell extracts. **(D)** Pol1NT

was isolated by immunoprecipitation, after digestion of DNA to release histone complexes from chromatin. As above, Pol1NT co-purified with full-length Spt16-9MYC but not with the histone-binding mutant Spt16-ΔNT-9MYC. Pol1NT purified from both extracts with chromatin-derived histone complexes (probably with FACT containing endogenous wild type Spt16) and with the replisome (via the interaction of Pol1NT with trimeric Ctf4 that also binds directly to the CMG helicase).

Appendix Figure S5

Mutation of conserved residues in Pol1NT block association with chromatin-derived histones from S-phase or G2-M phase cells. **(A)** The amino terminal tails of wild type budding yeast Pol1 (YCE39) or Pol1-6A (YCE226) were induced in cells arrested in G2-M phase (lower panels), or in G1-phase cells before release into S-phase (top panels). **(B)** Samples were taken after induction in G2-M phase cells, or 20' after release into S-phase, and then used to prepare cell extracts, from which Pol1NT was isolated after digestion of DNA to release histone complexes from chromatin. The indicated proteins were monitored by immunoblotting. **(C)** Summary of the ability of Pol1NT with the indicated mutations to interact with Mcm2 and FACT (IPs from extracts not treated with DNase), or with Ctf4 and histones (IPs from extracts treated with DNase – the histone complexes co-purifying with Pol1NT are also bound by Mcm2 and FACT).

Appendix Figure S6

Functional analysis of the Pol1-2A2 and Pol1-6A alleles. **(A)** Control (YCE482) and *pol1-6A-9MYC* (YCE483) cells were synchronized in G1-phase at 30°C and then released into S-phase. Samples were taken after 20' and used to prepare cell

extracts, from which Pol1-9MYC (or Pol1-6A-9MYC) were isolated by immunoprecipitation. The indicated proteins were monitored by immunoblotting. (B) DNA content was monitored by flow cytometry in the same experiment. (C) Control (YCE542) and *pol1-2A2* (YCE1050) cells were synchronized in G1-phase at 24°C and then released into S-phase for 30', before replisome assembly was monitored as in Figure 6A, via immunoprecipitation of the Sld5 subunit of the CMG helicase. (D) The kinetics of DNA synthesis for the *pol1-2A2* mutant were monitored by flow cytometry, in the same experiment shown above in Figure 1A and Figure 6B (alongside control, *pol1-4A* and *pol1-6A*). Therefore, the control sample in this panel is identical to that shown in Figure 1A and Figure 6B.

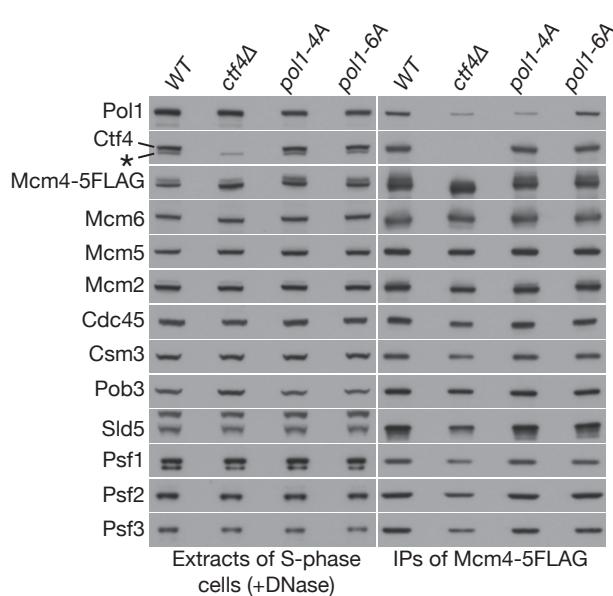
Appendix Figure S7

Cells lacking the Dpb3 and Dpb4 subunits of DNA polymerase ϵ are viable in the absence of the S-phase checkpoint pathway. Diploid cells of the indicated genotype were processed as for Figure 1C-D.

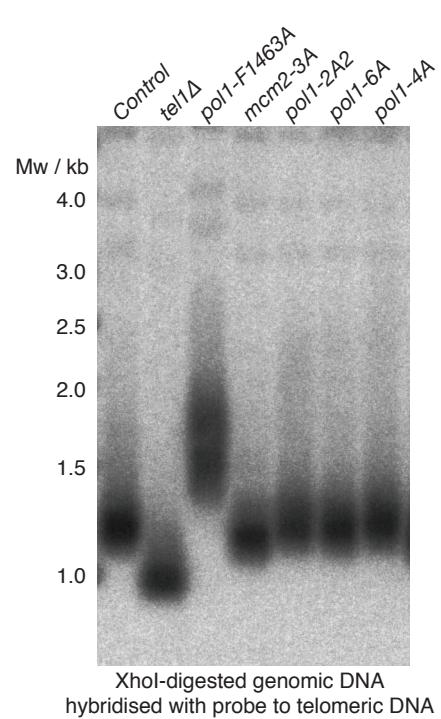
Appendix Figure S8

Amino acid sequences of recombinant tagged forms of HsPOLA1-NTD (1-110) and HsMCM2-NTD (43-160). The sequences of HsPOLA1-NTD and HsMCM2-NTD are in blue, the recognition site for PreScission protease is in orange, the Twin-Strep tag is shown in green and the residues mutated to Alanine in HsPOLA1-NTD are in bold and red.

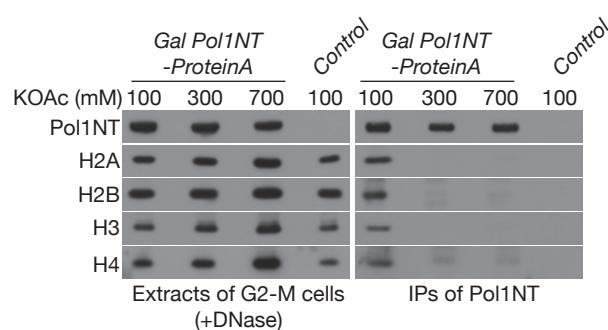
Evrin et al, Appendix Figure S1



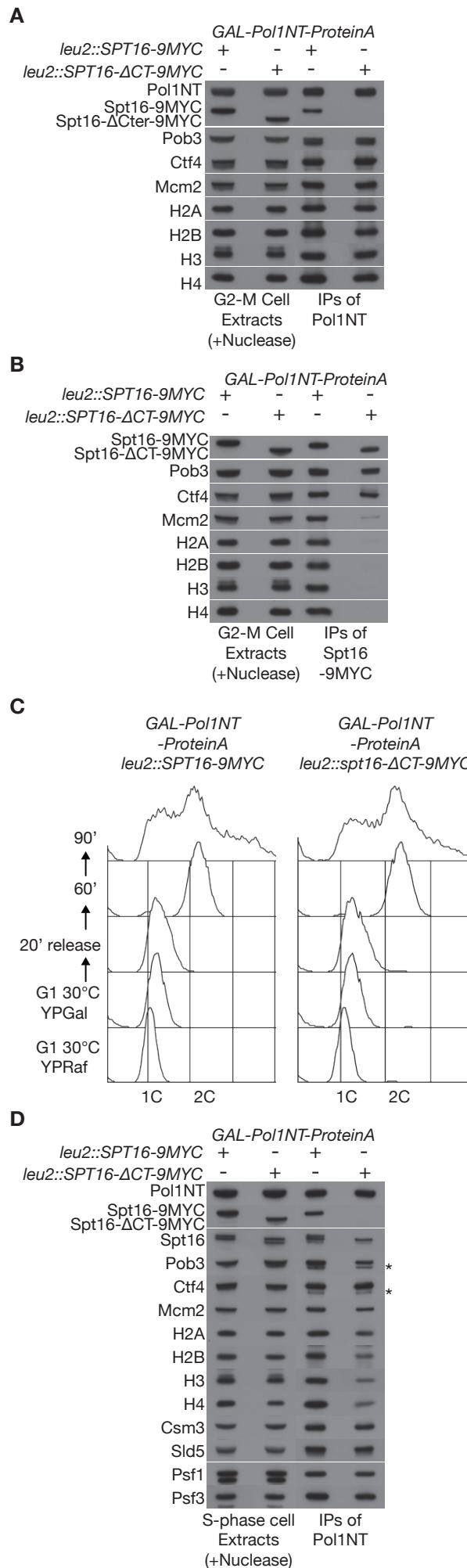
Evrin et al, Appendix Figure S2



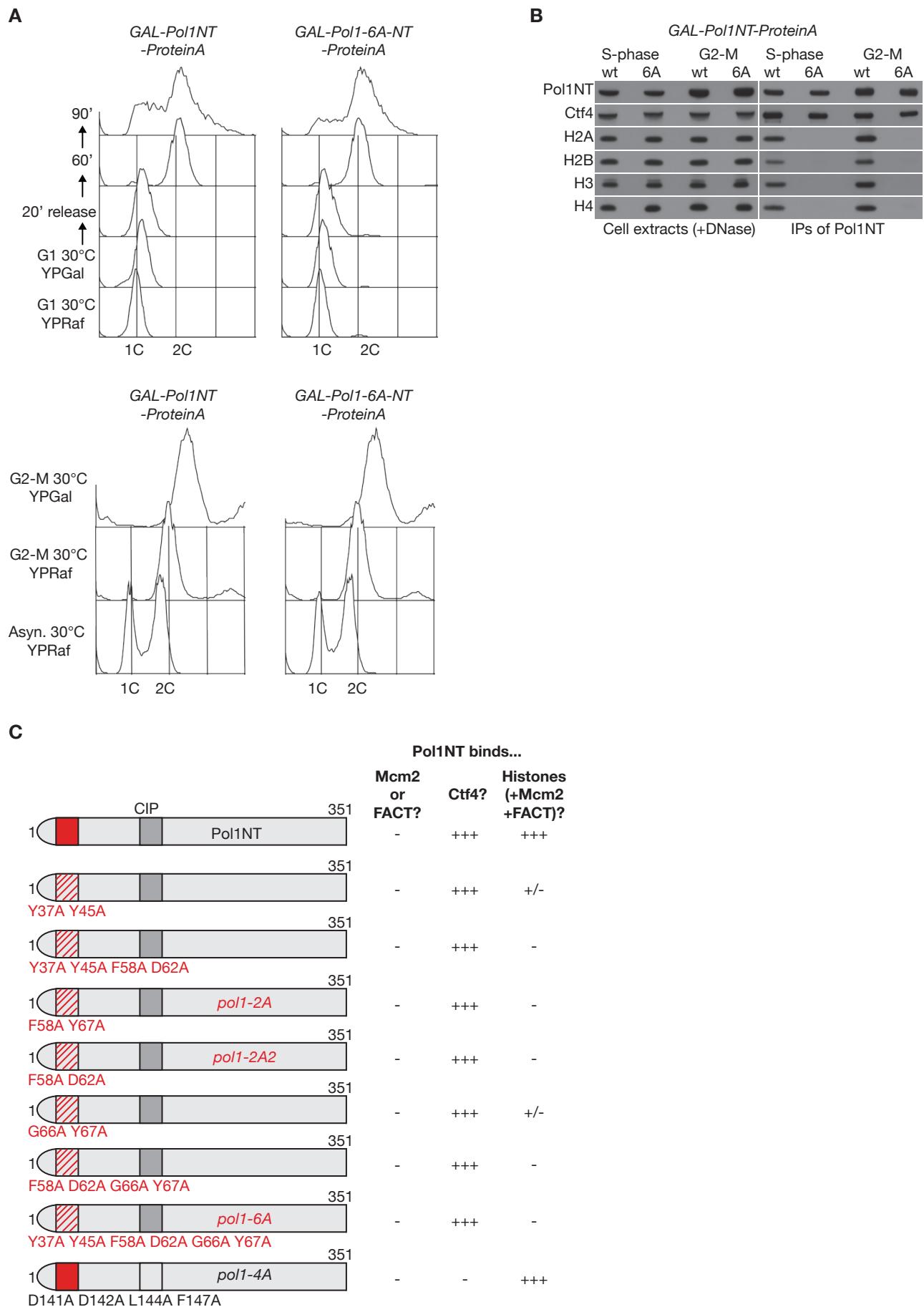
Evrin et al, Appendix Figure S3



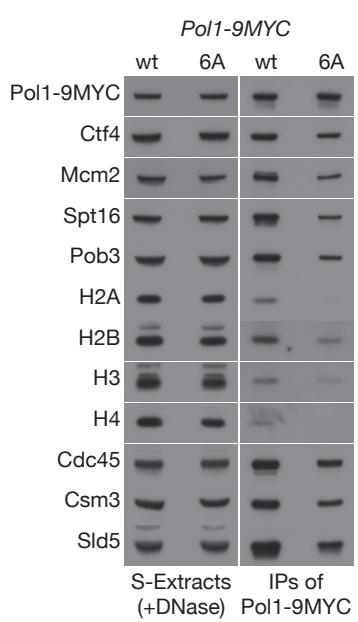
Evrin et al, Appendix Figure S4



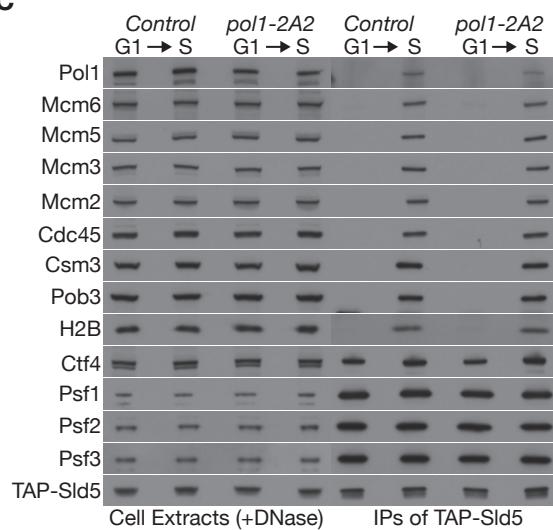
Evrin et al, Appendix Figure S5



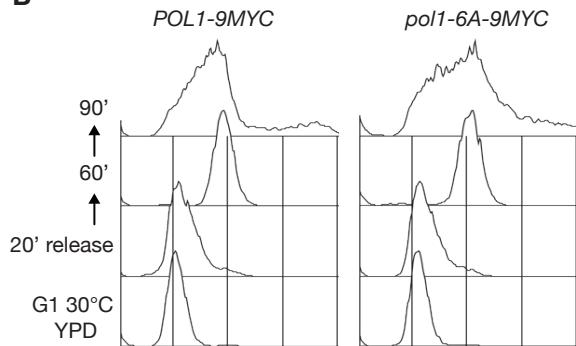
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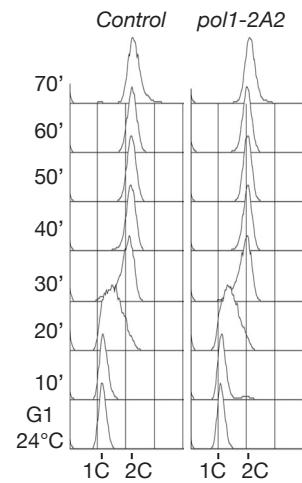
C



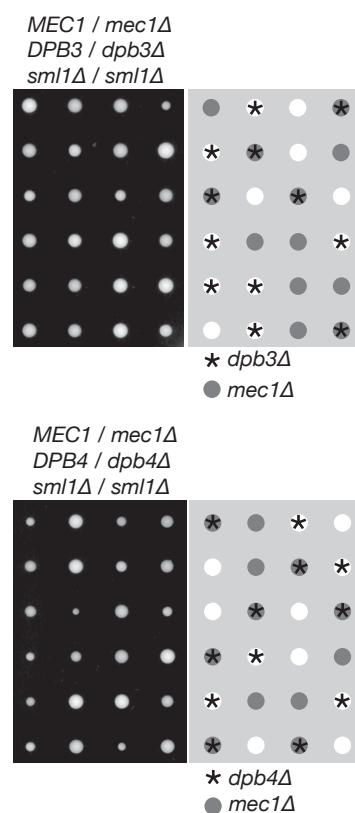
B



D



Evrin et al, Appendix Figure S7



LEVLFQGP = Precision Protease site

SAWSHPQFEKGGGSGGGSGGGSWHPQFEK = TwinStreptag

Sequences of **HsPOLA1** and **HsMCM2** are indicated in **BLUE**

Location of **3A** mutations in **HsPOLA1** are indicated in **bold red**

8His-MBP_Precision protease site_HsPOLA1 (1-110)_TwinStreptag

MNGSHHHHHHHHTSMKIEEGKLVIWINGDKGYNGLAEVGKKFEKDTGIKVT
VEHPDKLEEKFPQVAATGDGPDIIFWAHDRFGGYAQSGLLAEITPDKAFQDK
LYPFTWDARVYNGKLIAYPIAVEALSLIYNKDLLPNPPKTWEEIPALDKEALKAK
GKSALMFNLQEPYFTWPLIAADGGYAFKYENGKYDIKVGVVDNAGAKAGLT
FLVDLIKHNHMNADTDYSIAEAAFNKGGETAMTINGPWAWSNIDTSKVNYGVT
VLPTFKGQPSKPFVGVLSSAGIDAASPNKELAKEFLENYLLTDEGLEAVNKDK
PLGAVALKSYYEEELAKDPRIAATMENAQKGEIMPNIPQMSAFWYAVRTAVIN
AASGRQTVDEALKDAQTNTSSSTGSGTSGST**LEVLFQGP**GS**MAPVHGDDSL**
SDSGSFVSSRARREKSKKGRQEALERLKKAKAGEKYKYEVEDFTGVYEEV
DEEQYSKLVQARQDDDW**IVDDD**GIGY**VEDGREIFDDDLEDDALDADEKG**
SAWSHPQFEKGGGSGGGSGGGSWHPQFEK

8His-MBP_Precision protease site_HsPOLA1-3A (1-110)_TwinStreptag

MNGSHHHHHHHHTSMKIEEGKLVIWINGDKGYNGLAEVGKKFEKDTGIKVT
VEHPDKLEEKFPQVAATGDGPDIIFWAHDRFGGYAQSGLLAEITPDKAFQDK
LYPFTWDARVYNGKLIAYPIAVEALSLIYNKDLLPNPPKTWEEIPALDKEALKAK
GKSALMFNLQEPYFTWPLIAADGGYAFKYENGKYDIKVGVVDNAGAKAGLT
FLVDLIKHNHMNADTDYSIAEAAFNKGGETAMTINGPWAWSNIDTSKVNYGVT
VLPTFKGQPSKPFVGVLSSAGIDAASPNKELAKEFLENYLLTDEGLEAVNKDK
PLGAVALKSYYEEELAKDPRIAATMENAQKGEIMPNIPQMSAFWYAVRTAVIN
AASGRQTVDEALKDAQTNTSSSTGSGTSGST**LEVLFQGP**GS**MAPVHGDDSL**
SDSGSFVSSRARREKSKKGRQEALERLKKAKAGEKYKYEVEDFTGVYEEV
DEEQYSKLVQARQDDAA**IVDDD**GIG**AVEDGREIFDDDLEDDALDADEKG**
SAWSHPQFEKGGGSGGGSGGGSWHPQFEK

8His-MBP_Precision protease site_HsMCM2 (43-160)_TwinStreptag

MNGLNDIFEAQKIEWHESGSHHHHHHHHTSMKIEEGKLVIWINGDKGYNGL
AEVGKKFEKDTGIKVTVEHPDKLEEKFPQVAATGDGPDIIFWAHDRFGGYAQ
SGLLAEITPDKAFQDKLYPFTWDARVYNGKLIAYPIAVEALSLIYNKDLPNPP
KTWEEIPALDKEALKAKGKSALMFNLQEPYFTWPLIAADGGYAFKYENGKYDI
KVGVVDNAGAKAGLTFLVDLIKHNHMNADTDYSIAEAAFNKGGETAMTINGP
AWSNIDTSKVNYGVTVLPTFKGQPSKPFVGVLSSAGIDAASPNKELAKEFLEN
YLLTDEGLEAVNKDKPLGAVALKSYYEEELAKDPRIAATMENAQKGEIMPNIP
QMSAFWYAVRTAVINAASGRQTVDEALKDAQTNTSSSTGSGTSGST**LEVLFQGP**
GSGRDLPPFEDESEGLLGTEGPLEEEEDGEELIGDGMERDYRAIPELDAYEAE****
GLALDDEDVEELTASQREAERAMRQRDREAGRGLGRMRRGLLYDSDEEDE
ERPARKRRQVERATEDENLYFQGSSAWSHPQFEKGGGSGGGSGGGSWHPQFEK**GSS**

Strain	Genotype
W303-1	<i>MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100 / MATα ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100</i>
W303-1a	<i>MATa ade2-1 ura3-1 his3-11,15 trp1-1 leu2-3,112 can1-100</i>
YSS3	<i>MATa pep4Δ::ADE2</i>
YGDP5	<i>MATa mec1Δ::ADE2 sml1Δ::HIS3</i>
YSS10	<i>MATa MCM4-5FLAG-9HIS (hphNT) pep4Δ::ADE2</i>
YFV18	<i>MATα dna2-4A-9MYC</i>
YFV37	<i>MATa leu2::mURA3-LEU2</i>
YFV38	<i>MATa RDN1-NTS2::mURA3-LEU2</i>
YCE39	<i>MATa ura3-1::GAL-Pol1 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YFV39	<i>MATa RDN1-NTS1::mURA3-LEU2</i>
YCE125	<i>MATa ura3-1::GAL-Pol1 1-200-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE126	<i>MATa ura3-1::GAL-Pol1 1-267-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE127	<i>MATa ura3-1::GAL-Pol1 1-315-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE128	<i>MATa ura3-1::GAL-Pol1 89-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE129	<i>MATa ura3-1::GAL-Pol1 150-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YMP154-1	<i>MATa MCM2-9MYC (K.I.TRP1) pep4Δ::ADE2</i>
YMP177-1	<i>MATa SPT16-9MYC (K.I.TRP1) pep4Δ::ADE2</i>
YBH213	<i>MATa mrc1Δ::K.I.TRP1</i>
YCE217	<i>MATa ura3-1::GAL-Pol1-4A 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE222	<i>MATa ura3-1::GAL-Pol1 G66A Y67A 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE224	<i>MATa ura3-1::GAL-Pol1-F58A D62A G66A Y67A 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE226	<i>MATa ura3-1::GAL-Pol1-6A 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE232	<i>MATa ura3-1::GAL-Pol1-2A2 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE234	<i>MATa ura3-1:: GAL-Pol1-Y37A Y45A F58A D62A 1-351 ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE236	<i>MATa ura3-1::GAL-Pol1-Y37A Y45A 1-351 ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE238	<i>MATa ura3-1::GAL-Pol1 1-351-ProteinA-7His (URA3) mcm2-3A (hphNT) pep4Δ::ADE2</i>
YCE248	<i>MATa ura3-1::GAL-Pol1 1-351-ProteinA-7His (URA3) leu2-3,112::SPT16-9MYC (LEU2) pep4Δ::ADE2</i>
YCE250	<i>MATa ura3-1::GAL-Pol1 1-351-ProteinA-7His (URA3) leu2-3,112::Spt16 1-957-9MYC (LEU2) pep4Δ::ADE2</i>

YCE278	<i>MATa pol1-6A-9MYC (hphNT)</i>
YMP287	<i>MATa ura3-1::GAL-Mcm2 1-200-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YMP289	<i>MATa ura3-1::GAL-Mcm4 1-186 -ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE296	<i>MATa mcm2-3A-9MYC (K.I.TRP1) pol1-6A-9MYC (hphNT)</i>
YCE378	<i>MATa ura3-1::GAL-Sp Pol1 1-328-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE405	<i>MATa / MATα VR/VR::ADE2-TEL VIIIL/VIIIL-adh4::URA3-TEL POL1/pol1-6A-9MYC (hphNT)</i>
YCE407	<i>MATa ura3-1::GAL-Hs Pol1 1-339-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YLS409	<i>MATα hmrΔ77-144:SUP4-o HMR::ADE2 (Mol. Cell. Biol. 13, 3919-3928, (1993))</i>
YLS410	<i>MATα hmrΔ77-144:SUP4-o HMR::2EDA (Mol. Cell. Biol. 13, 3919-3928, (1993))</i>
YCE425	<i>MATa ura3-1::GAL-Hs Pol1 1-339-2A2-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE435	<i>MATa ura3-1::GAL-Hs Pol1 1-339-6A-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE449	<i>MATa pol1-4A (URA3)</i>
YCE451	<i>MATa pol1-6A (URA3)</i>
YCE482	<i>MATa POL1-9MYC (HphNT) pep4Δ::ADE2</i>
YCE483	<i>MATa pol1-6A-9MYC (HphNT) pep4Δ::ADE2</i>
YCE485	<i>MATa / MATα VR/VR::ADE2-TEL</i>
YCE488	<i>MATa / MATα VR/VR::ADE2-TEL POL1/pol1-4A (URA3)</i>
YCE486	<i>MATa / MATα VR/VR::ADE2-TEL MCM2/mcm2-3A (hphNT)</i>
YMP502-5	<i>MATa mcm2-3A-9MYC (K.I.TRP1)</i>
YMP531	<i>MATa mcm2-3A (hphNT)</i>
YCE533	<i>MATa / MATα MEC1/mec1Δ::ADE2 sml1Δ::HIS3/sml1Δ::HIS3 POL1 / pol1-4A (URA3)</i>
YCE534	<i>MATa / MATα MEC1/mec1Δ::ADE2 sml1Δ::HIS3/sml1Δ::HIS3 POL1 / pol1 F1463A (hphNT)</i>
YCE535	<i>MATa / MATα MEC1/mec1Δ::ADE2 sml1Δ::HIS3/sml1Δ::HIS3 POL1 / pol1-6A (URA3)</i>
YCE542	<i>MATa TAP-SLD5 (kanMX) POL1 (URA3) pep4Δ::ADE2</i>
YCE544	<i>MATa TAP-SLD5 (kanMX) pol1-4A (URA3) pep4Δ::ADE2</i>
YCE546	<i>MATa TAP-SLD5 (kanMX) pol1-6A (URA3) pep4Δ::ADE2</i>
YCE555	<i>MATa pol1-2A2 (URA3)</i>
YCE597	<i>MATa / MATα MEC1 / mec1Δ::ADE2 sml1Δ::HIS3 / sml1Δ::HIS3 POL1 / pol1-2A (URA3)</i>
YCE609	<i>MATa / MATα VR/VR::ADE2-TEL POL1 / pol1-2A2 (URA3)</i>
YCE611	<i>MATa / MATα VR/VR::ADE2-TEL VIIIL / VIIIL::adh4-URA3-TEL TEL1 / tel1Δ::hphNT</i>
YCE641	<i>MATa leu2::mURA3-LEU2 mcm2-3A (hphNT)</i>
YCE643	<i>MATa leu2::mURA3-LEU2 pol1-2A2-9MYC (hphNT)</i>

YCE645	MAT^a <i>leu2::mURA3-LEU2 pol1-6A-9MYC (hphNT)</i>
YCE647	MAT^a <i>leu2::mURA3-LEU2 pol1-4A-9MYC (hphNT)</i>
YCE649	MAT^a <i>RDN1-NTS2::mURA3-LEU2 mcm2-3A (hphNT)</i>
YCE651	MAT^a <i>RDN1-NTS2::mURA3-LEU2 pol1-2A2-9MYC (hphNT)</i>
YCE653	MAT^a <i>RDN1-NTS2::mURA3-LEU2 pol1-6A-9MYC (hphNT)</i>
YCE655	MAT^a <i>RDN1-NTS2::mURA3-LEU2 pol1-4A-9MYC (hphNT)</i>
YCE657	MAT^a <i>RDN1-NTS1::mURA3-LEU2 mcm2-3A (hphNT)</i>
YCE659	MAT^a <i>RDN1-NTS1::mURA3-LEU2 pol1-2A2-9MYC (hphNT)</i>
YCE661	MAT^a <i>RDN1-NTS1::mURA3-LEU2 pol1-6A-9MYC (hphNT)</i>
YCE663	MAT^a <i>RDN1-NTS1::mURA3-LEU2 pol1-4A-9MYC (hphNT)</i>
YCE665	MAT^a <i>RDN1-NTS1::mURA3-LEU2 mcm2-3A-9MYC (K.I.TRP1) pol1-6A-9MYC (hphNT)</i>
YCE667	MAT^a <i>RDN1-NTS2::mURA3-LEU2 mcm2-3A-9MYC (K.I.TRP1) pol1-6A-9MYC (hphNT)</i>
YCE669	MAT^a <i>leu2::mURA3-LEU2 mcm2-3A-9MYC (K.I.TRP1) pol1-6A-9MYC (hphNT)</i>
YCE674	MAT^a <i>leu2::mURA3-LEU2 top1Δ::kanMX</i>
YCE676	MAT^a <i>RDN1-NTS2::mURA3-LEU2 top1Δ::kanMX</i>
YCE678	MAT^a <i>RDN1-NTS1::mURA3-LEU2 top1Δ::kanMX</i>
YCE694	MAT^a <i>leu2::mURA3-LEU2 dpb4Δ::kanMX</i>
YCE706	MAT^a <i>RDN1-NTS2::mURA3-LEU2 dpb4Δ::kanMX</i>
YCE718	MAT^a <i>RDN1-NTS1::mURA3-LEU2 dpb4Δ::kanMX</i>
YCE728	MAT^a <i>leu2::mURA3-LEU2 dpb3Δ::kanMX</i>
YCE735	MAT^a <i>RDN1-NTS2::mURA3-LEU2 dpb3Δ::kanMX</i>
YCE740	MAT^a <i>RDN1-NTS1::mURA3-LEU2 dpb3Δ::kanMX</i>
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YCE798	MAT^a / MAT^α <i>MEC1/mec1Δ::ADE2 sml1Δ::HIS3 / sml1Δ::HIS3 DPB4 / dpb4Δ::kanMX</i>
YCE799	MAT^a / MAT^α <i>VR/VR::ADE2-TEL TOF2 / tof2-4A</i>
YGDP975	MAT^a <i>MCM4-5FLAG-9HIS (hphNT) ctf4 Δ::kanMX pep4Δ::ADE2</i>
YCE1027	MAT^a <i>ura3-1::GAL-Pol1 1-351-ProteinA-7His (URA3) tel1Δ::hphNT pep4Δ::ADE2</i>
YGDP1045	MAT^a <i>tel1Δ::hphNT</i>
YCE1050	MAT^a <i>TAP-SLD5 (kanMX) pol1-2A2 (URA3) pep4Δ::ADE2</i>
YCE1073	MAT^a <i>MCM4-5FLAG-9HIS (hphNT) pol1-4A (URA3) pep4Δ::ADE2</i>
YCE1075	MAT^a <i>MCM4-5FLAG-9HIS (hphNT) pol1-6A (URA3) pep4Δ::ADE2</i>
YCE1084	MAT^a <i>ura3-1::GAL-Pol1-2A 1-351-ProteinA-7His (URA3) pep4Δ::ADE2</i>
YCE1120	MAT^a / MAT^α <i>HMR / HMR::ADE2</i>

YCE1121	<i>MATa / MATα HMR/ HMR::ADE2 MCM2 / mcm2-3A (HphNT)</i>
YCE1123	<i>MATa / MATα HMR / HMR::ADE2 POL1 / pol1-4A (URA3)</i>
YCE1124	<i>MATa / MATα HMR / HMR::ADE2 POL1 / pol1-6A (URA3)</i>
YCE1125	<i>MATa / MATα HMR / HMR::ADE2 POL1 / pol1-2A2 (URA3)</i>
YCE1126	<i>MATa / MATα HMR / HMR::2EDA</i>
YCE1127	<i>MATa / MATα HMR / HMR::2EDA MCM2 / mcm2-3A (HphNT)</i>
YCE1129	<i>MATa / MATα HMR / HMR::2EDA POL1 / pol1-4A (URA3)</i>
YCE1130	<i>MATa / MATα HMR / HMR::2EDA POL1 / pol1-6A (URA3)</i>
YCE1131	<i>MATa / MATα HMR / HMR::2EDA POL1 / pol1-2A2 (URA3)</i>
YGDP1135	<i>MATa pol1 F1463A (hphNT)</i>

Appendix Table S1

Yeast strains used in this study. All strains are based on the W303 background.