

Supplemental material

Singhal et al., <https://doi.org/10.1084/jem.20180008>

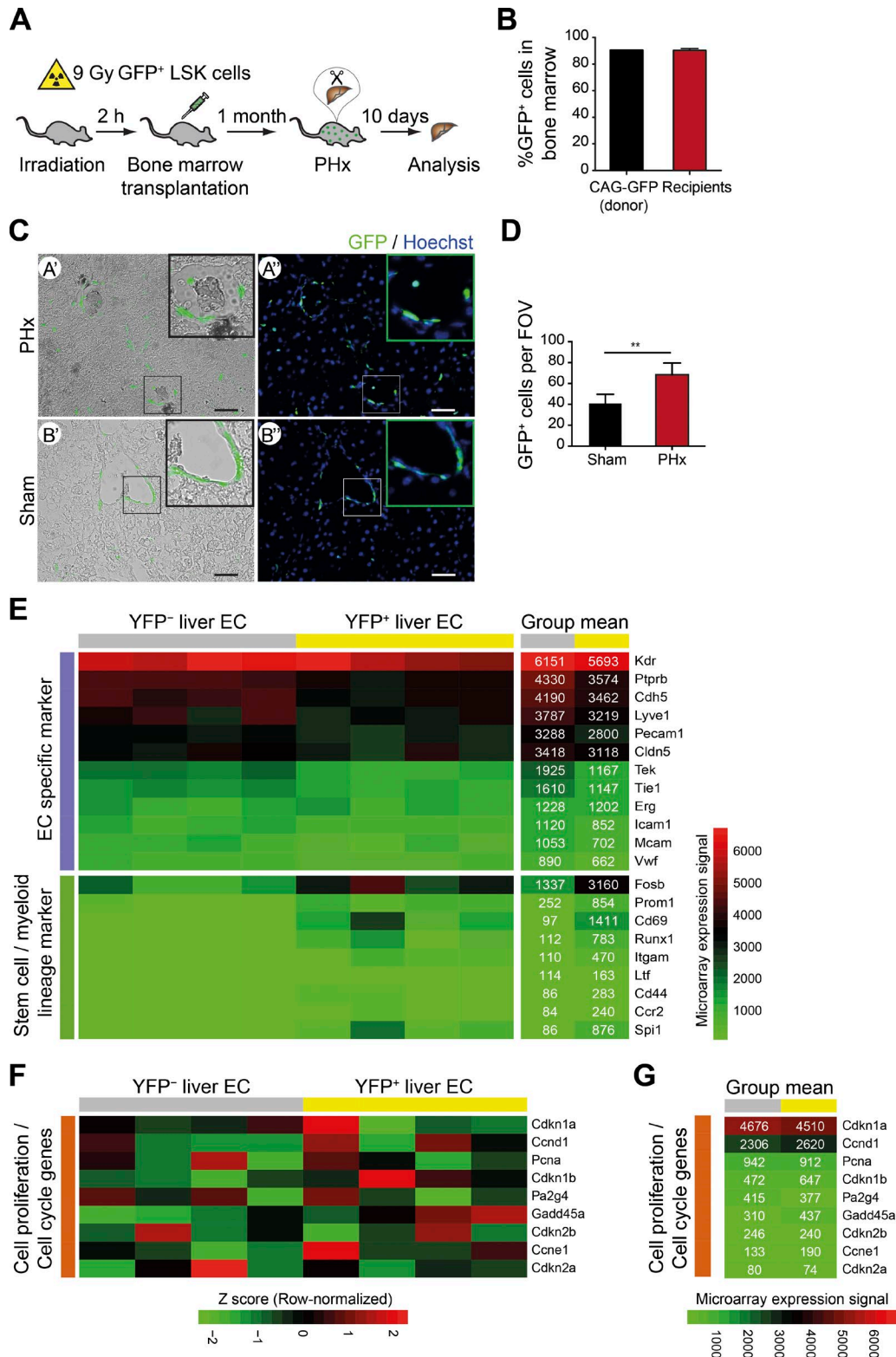


Figure S1. **Irradiation-conditioned bone marrow chimeras illustrate recruitment of BMDMCs in the regenerating liver.** (A) Experimental outline of PHx-induced liver regeneration in irradiation-conditioned GFP⁺ bone marrow-transplanted mice. (B) Whole bone marrow cells were isolated from the CAG-GFP donor mice as well as the recipient mice and were analyzed by FACS to evaluate the donor chimerism (mean \pm SD, $n_{CAG-GFP} = 1$, $n_{Recipient} = 6$ mice). (C) Microscopic analysis of liver sections from sham-operated (lower panels, B' and B'') and PHx mice (upper panels, A' and A'') shows bone marrow-derived GFP⁺ cells integrated into the liver vasculature. Scale bars, 50 μ m. (D) The plot shows the GFP⁺ cell count per field of view (FOV) in the sham-operated and PHx mice (mean \pm SD, $n = 6$ mice). **, $P < 0.01$ (two-tailed Student's *t* test). (E) Microarray-based gene expression analysis of YFP⁻ resident liver ECs and YFP⁺ bone marrow-derived liver ECs was performed. Group mean heat map shows the average of biological replicates ($n = 4$ mice). (F) The heat map represents expression of cell cycle regulatory genes. (G) Group mean heat map shows the average of biological replicates ($n = 4$ mice).

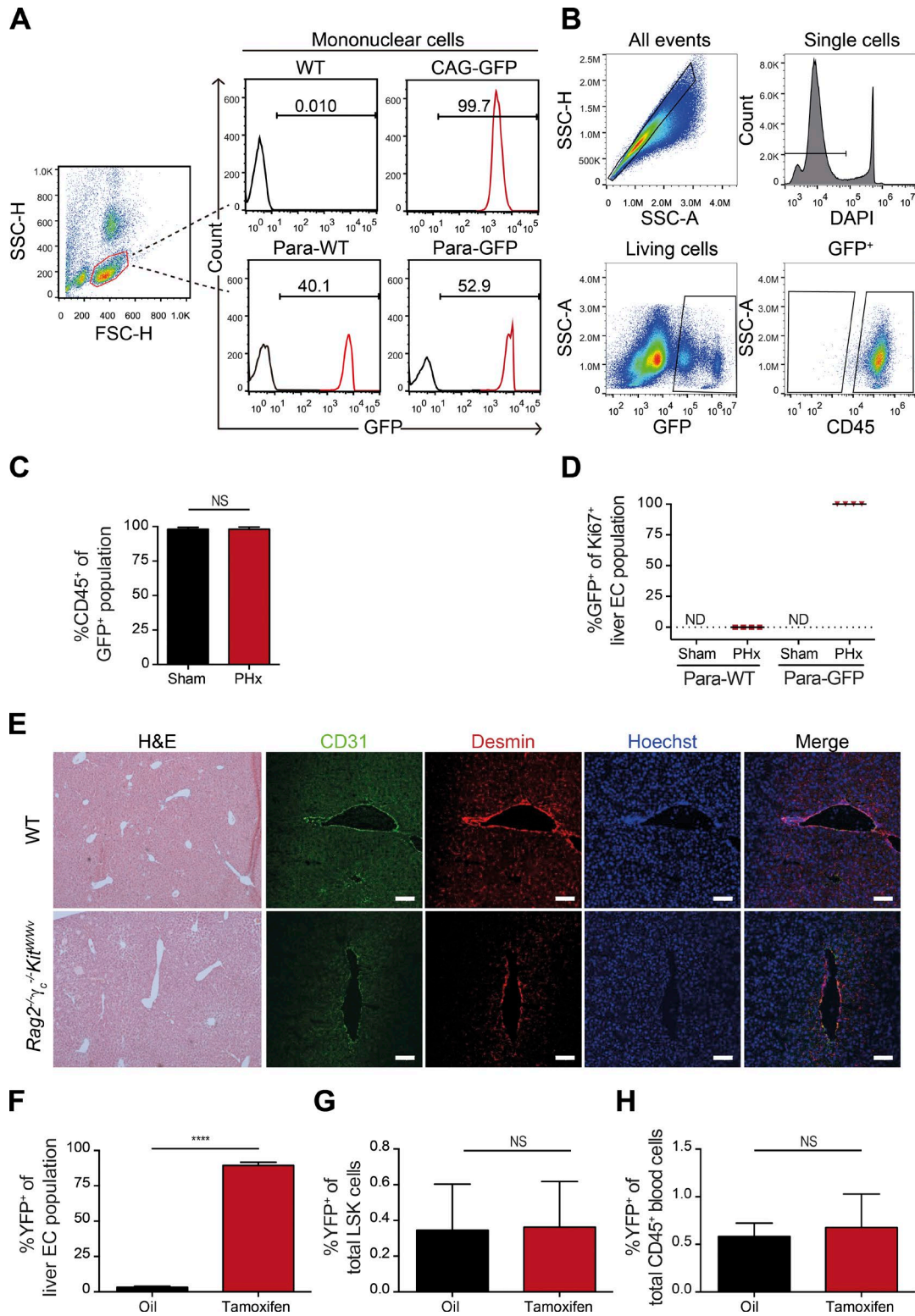


Figure S2. **Nonmyeloablative mouse models to study liver regeneration.** (A) FACS analysis of blood chimerism of parabiosed WT (Para-WT) and GFP (Para-GFP) mice. Blood samples from nonoperated WT and CAG-GFP mice served as negative and positive controls, respectively. (B) Gating strategy for the analysis of the cell identity of GFP⁺ cells in the livers of Para-WT mice. (C) The plot shows the frequency of CD45 positivity among GFP⁺ cells in the liver of Para-WT mice (mean ± SD, n = 3–4 mice). (D) The plot shows the ratio of GFP⁺ cells among Ki67⁺ liver ECs in Para-WT and Para-GFP mice (mean ± SD, n = 3–4 mice). (E) PHx was performed on both WT and *Rag2^{-/-}γ_c^{-/-}Kit^{W/Wv}* mice. Immunohistochemistry and immunofluorescence analysis of the livers of WT and *Rag2^{-/-}γ_c^{-/-}Kit^{W/Wv}* mice after PHx indicate normal liver regeneration in *Rag2^{-/-}γ_c^{-/-}Kit^{W/Wv}* mice. Scale bars, 100 μm. (F–H) *VECad-Cre^{ERT2}xRosa26-YFP^{fl/fl}* mice were transiently administered with either oil or tamoxifen. After a resting period of 1 mo, the frequencies of YFP⁺ cells among liver ECs (F), LSK cells in the bone marrow (G), and circulating immune cells in the peripheral blood (H) were analyzed by FACS (mean ± SD, n = 4 mice). ND, nondetectable; ****, P < 0.0001 (two-tailed Student's *t* test).

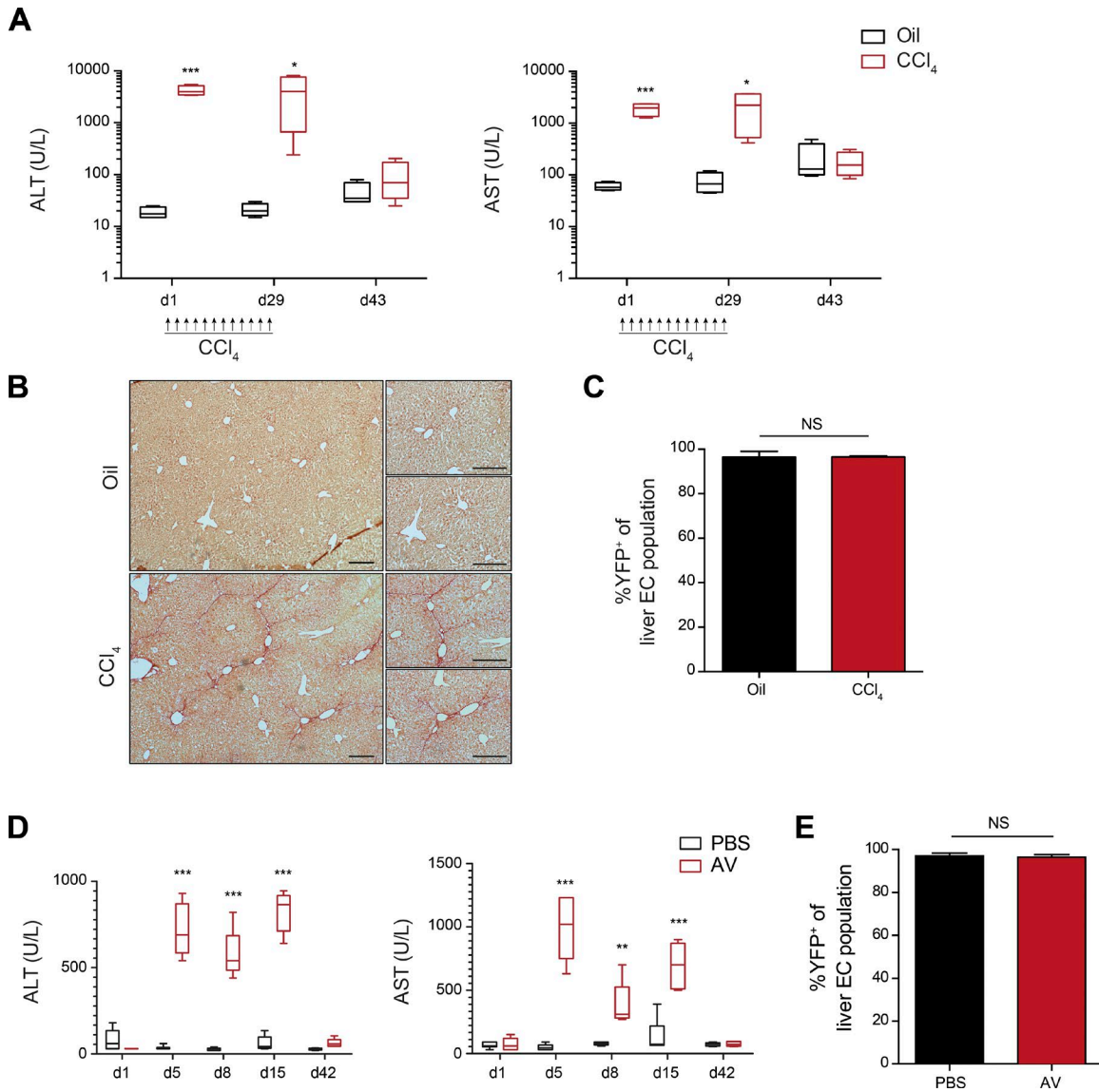
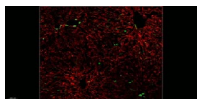


Figure S3. **Liver neovascularization during chronic liver injury.** (A–C) *VECad-Cre^{ERT2}×Rosa26-YFP^{fl/fl}* mice with labeled liver ECs were repeatedly administered with CCl₄ over a period of 4 wk. (A) The plot shows blood plasma ALT/AST levels over the course of the experiment. (B) Sirius red staining was performed on the livers of mice treated with either oil alone or with CCl₄. Scale bars, 200 μm. (C) The frequency of YFP⁺ ECs in the livers of *VECad-Cre^{ERT2}×Rosa26-YFP^{fl/fl}* mice was analyzed by FACS (mean ± SD, n = 4 mice). (D and E) *VECad-Cre^{ERT2}×Rosa26-YFP^{fl/fl}* mice with labeled liver ECs were intravenously injected with 10¹¹ viral particles of empty replication–deficient adenovirus. (D) The plot shows blood plasma ALT/AST levels over the course of the experiment (mean ± SD, n = 6 mice). (E) The frequency of YFP⁺ ECs in the livers of *VECad-Cre^{ERT2}×Rosa26-YFP^{fl/fl}* mice was analyzed by FACS (mean ± SD, n = 6 mice). *, P < 0.05; **, P < 0.01; ***, P < 0.001 (two-tailed Student’s t test).



Video 1. **High-resolution three-dimensional reconstruction of liver vasculature demonstrating bona fide integration of fluorescent-labeled elongated EC into the endothelial layer.** ECs in the video are represented in gray; GFP⁺ cells in green. Surface rendering transparency was set at 50% and recorded at 30 frames per second.