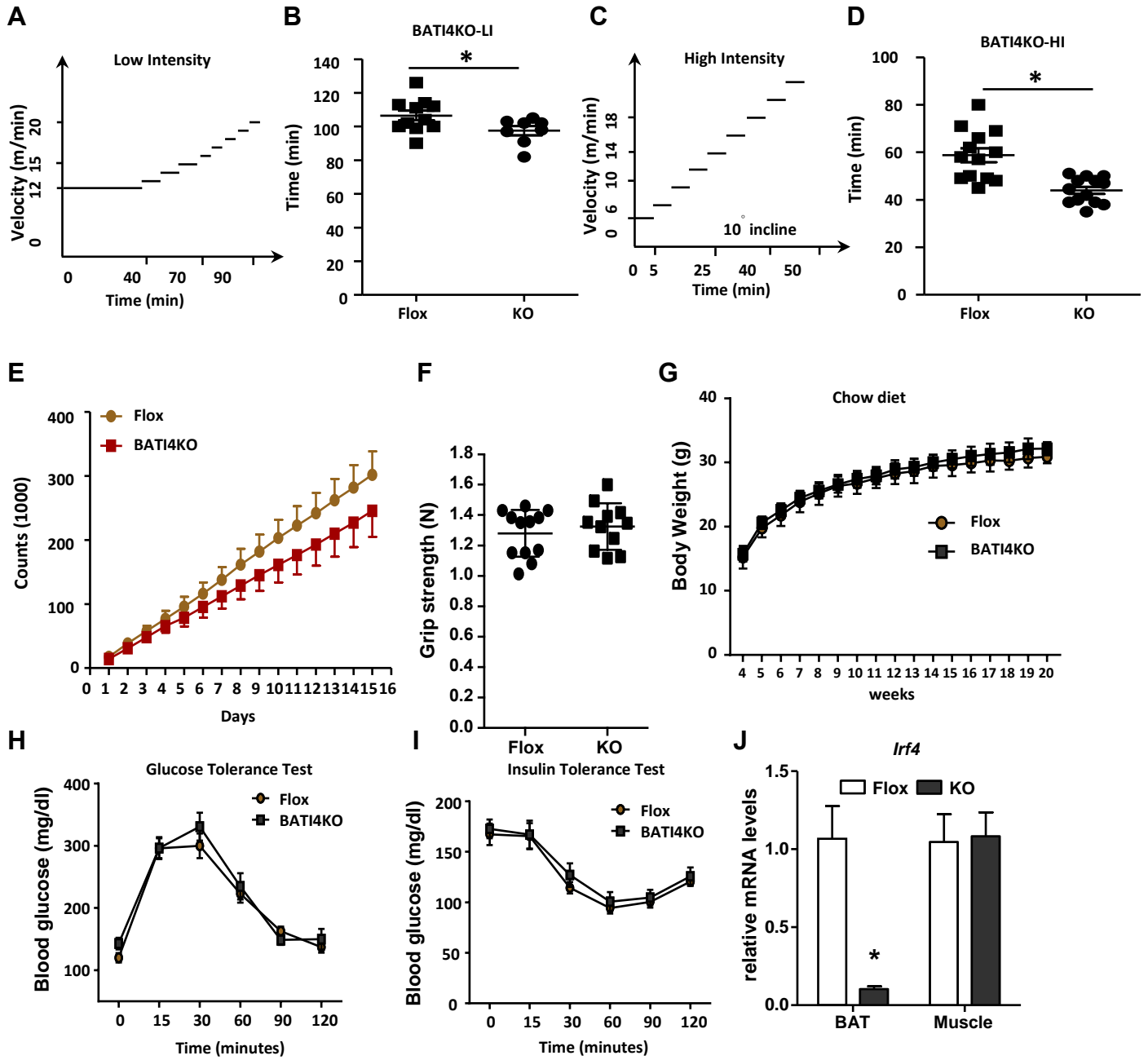


## **Supplemental Information**

### **Brown adipose tissue controls skeletal muscle function via the secretion of myostatin**

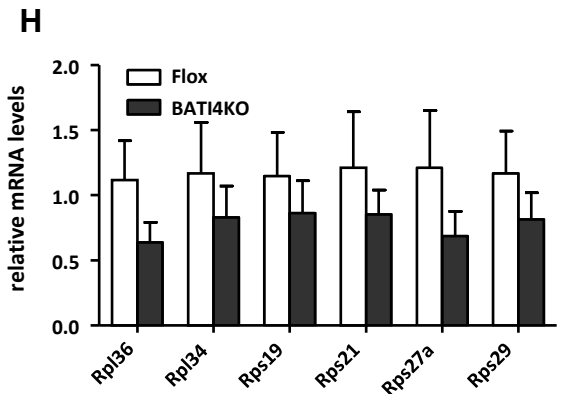
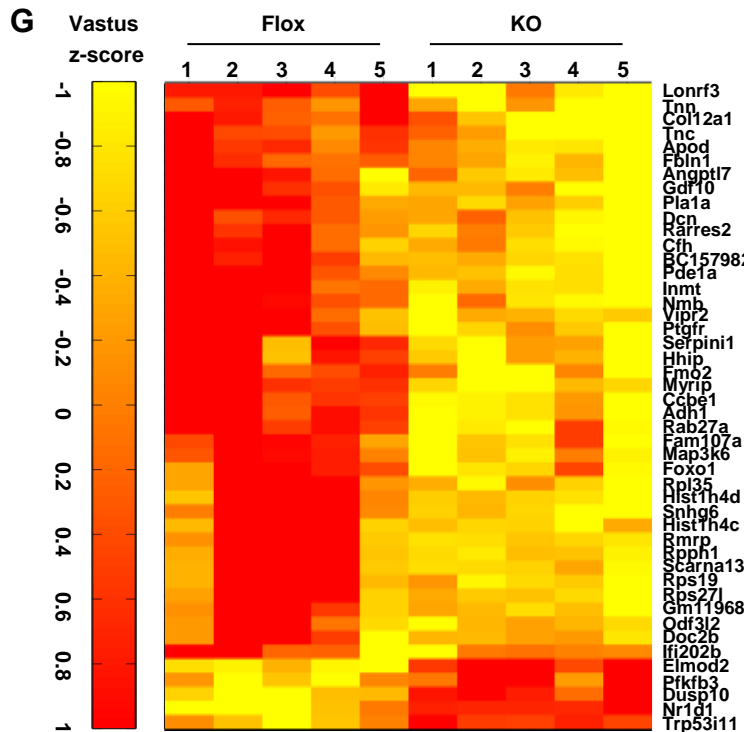
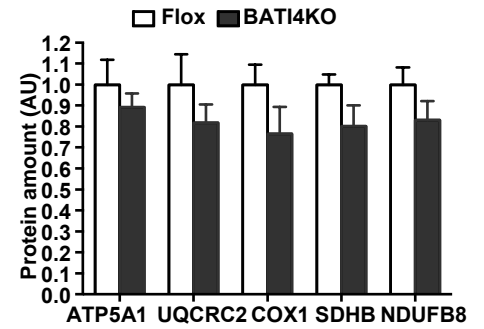
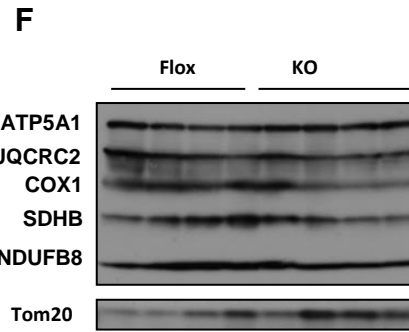
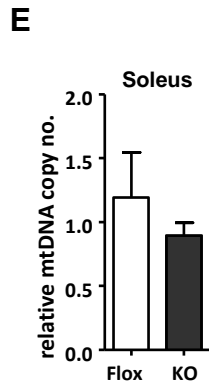
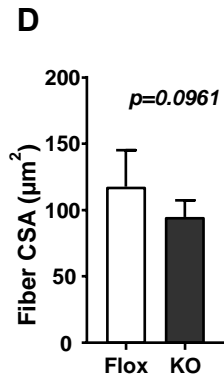
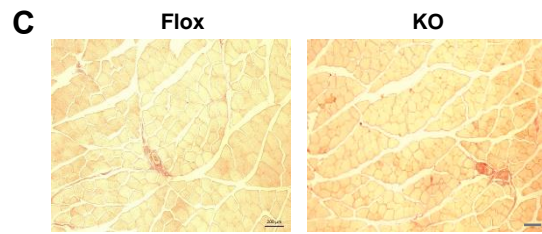
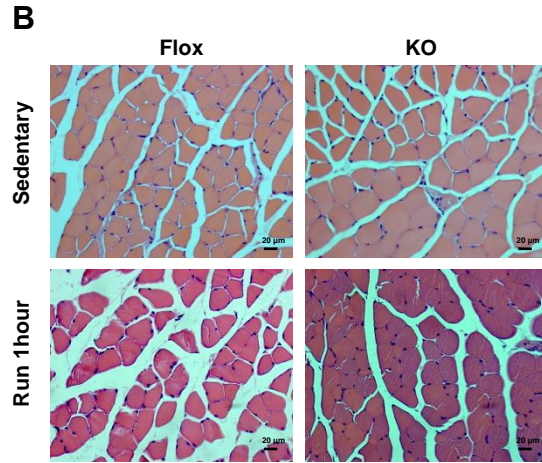
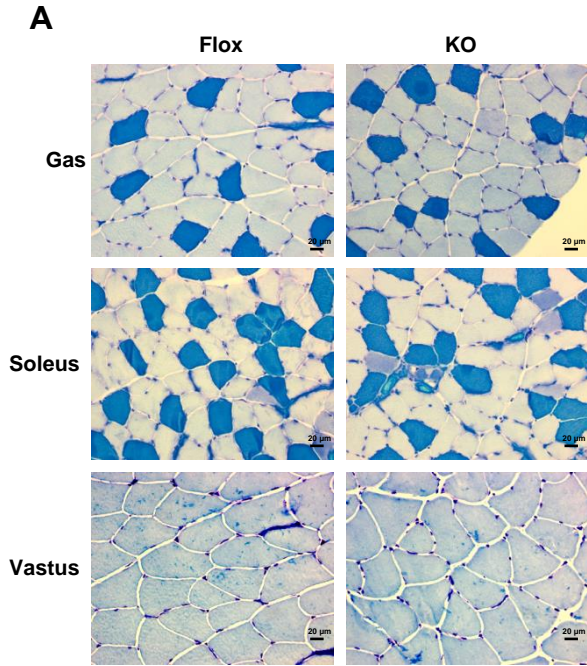
Xingxing Kong, Ting Yao, Peng Zhou, Lawrence Kazak, Danielle Tenen, Anna Lyubetskaya, Brian A. Dawes, Linus Tsai, Barbara B. Kahn, Bruce M. Spiegelman, Tiemin Liu, and Evan D. Rosen



**Figure S1: (Related to Figure 1)**

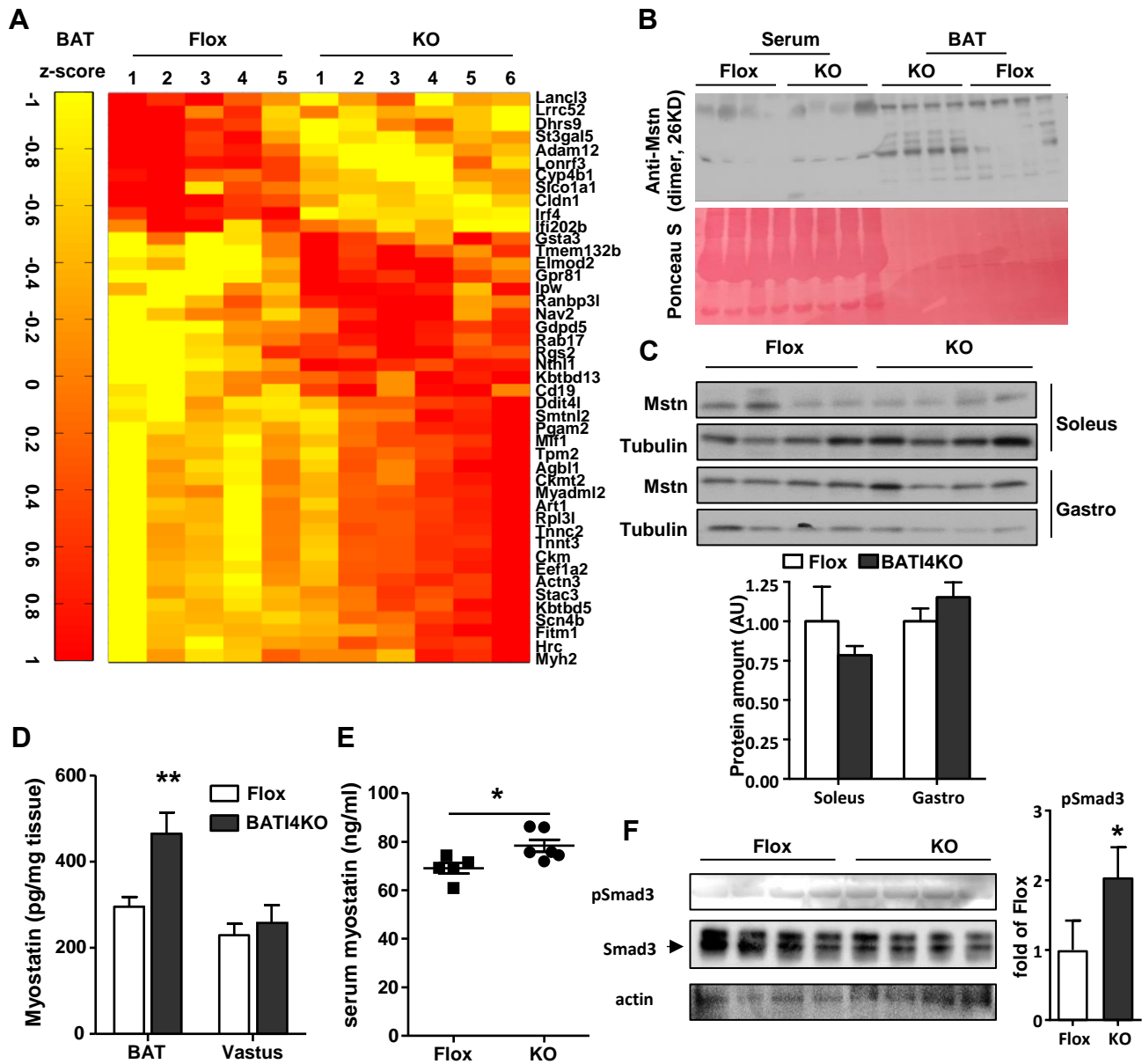
**Reduced exercise capacity in BATI4KO mice.** (A) Schematic of low intensity and (C) high intensity exercise regimens. Time run by BATI4KO and control mice on low intensity (B) and high intensity (D) protocols. (E) Cumulative free wheel running in BATI4KO and control mice (n=8 per group). (F) Grip strength of BATI4KO groups mice. (G) Body weight of BATI4KO and Flox mice on chow diet. (n=9-16/group). (H) Glucose tolerance test in 12-week-old BATI4KO mice on chow diet after intraperitoneal

injection of glucose. (n = 8). (I) Insulin tolerance test in 14-week-old BATI4KO mice after intraperitoneal injection of insulin. (n=8). (J) Q-PCR gene expression analysis of *Irf4* expression in BAT and vastus lateralis of BATI4KO mice. Results are expressed as mean  $\pm$  SEM (n=5-6, \* $p$ <0.05).



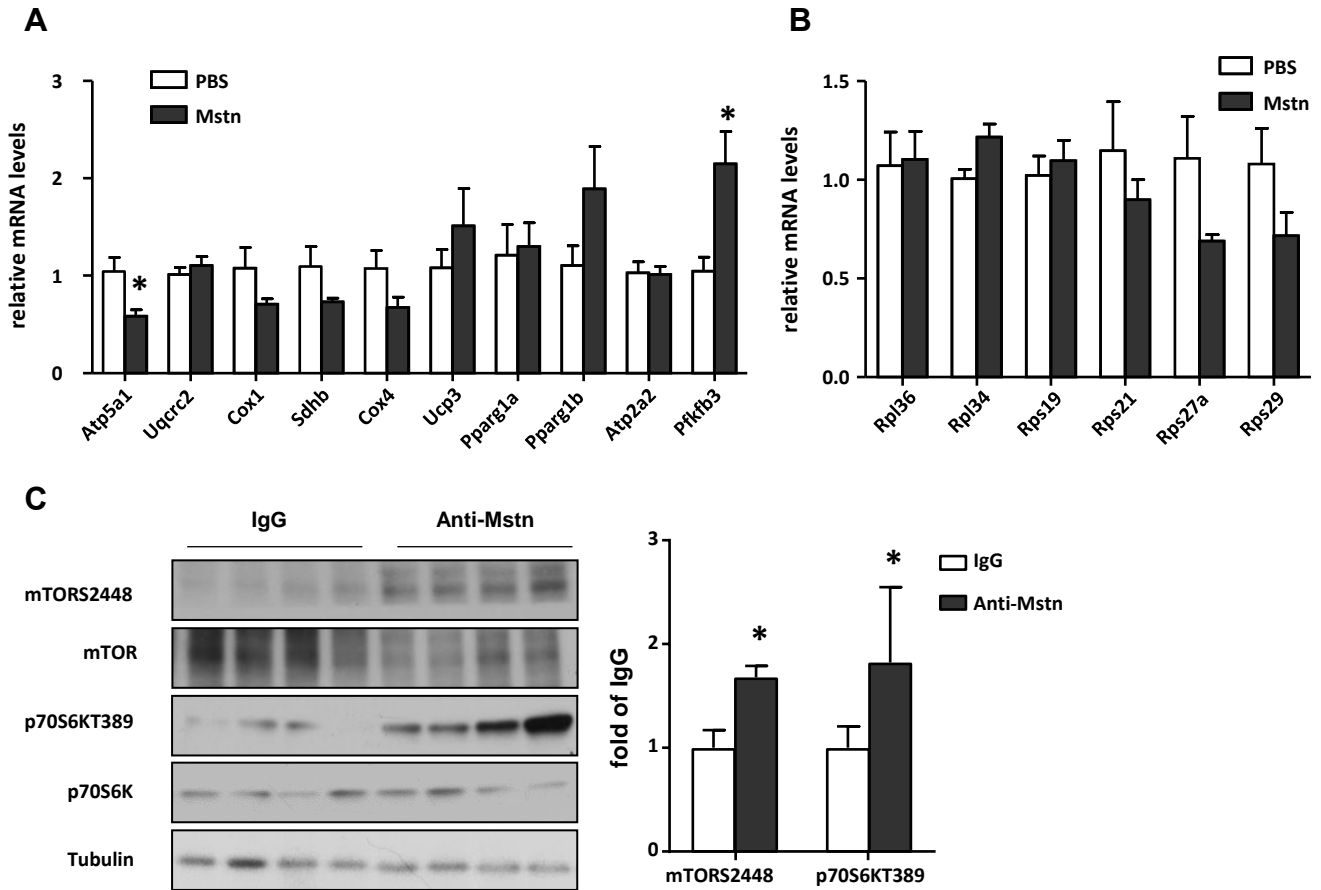
**Figure S2: (Related to Figure 2)**

**BAT14KO mice have altered skeletal muscle.** (A) ATPase staining of muscle sections from BAT14KO and control mice (type I: dark blue; type IIa: white; type IIb: light blue). (B) H&E staining of soleus muscle from BAT14KO and control mice (40X). (C) Sirius Red staining of vastus muscle from BAT14KO and control mice (10X). (D) Muscle fiber cross-sectional area. Values are means  $\pm$  SEM. (E) Mitochondrial DNA number in soleus muscle of BAT14KO and control mice (n=5). (F) Western blot analysis of isolated muscle mitochondria from soleus of BAT14KO and control mice. Protein amount was quantified using Image J. (n=4). (G) Heat map from RNA-seq of vastus lateralis from BAT14KO and control mice,  $|\log_2FC| \geq 1$ , FDR  $\leq 0.05$ . (H) Q-PCR analysis of ribosomal protein gene mRNA expression in the vastus (n=8-10).



**Figure S3: (Related to Figure 3)**

**Direct effect of BAT via secretion of myostatin.** (A) Heat map of differentially expressed genes in BAT of BATI4KO and control mice,  $|\log_2FC| \geq 1$ ,  $FDR \leq 0.05$ . (B, C) Western blot analysis of muscle myostatin levels. Protein amount was quantified using Image J. (n=4). Tissue myostatin levels (D) and serum myostatin levels in BATI4KO group (E) were measured by ELISA kits. (n=5-8, \* $p < 0.05$ ). (F) Western blot analysis of Smad3 phosphorylation in vastus of male BATI4KO vs. control mice. Activity was quantified using Image J (n=4, \* $p < 0.05$  vs. Flox mice).



**Figure S4: (Related to Figure 5)**

**Myostatin mediates the effect of BAT IRF4 loss on exercise capacity.** Q-PCR analysis of mitochondrial gene expression (**A**) and ribosomal protein gene mRNA expression (**B**) in the vastus from myostatin injection groups ( $n=5-6$ ,  $*p < 0.05$ ). (**C**) Western blot analysis of mTOR signaling pathway in vastus lateralis of myostatin antibody injection groups. Activity was quantified using Image J ( $n=4$ ,  $*p < 0.05$  vs. IgG).

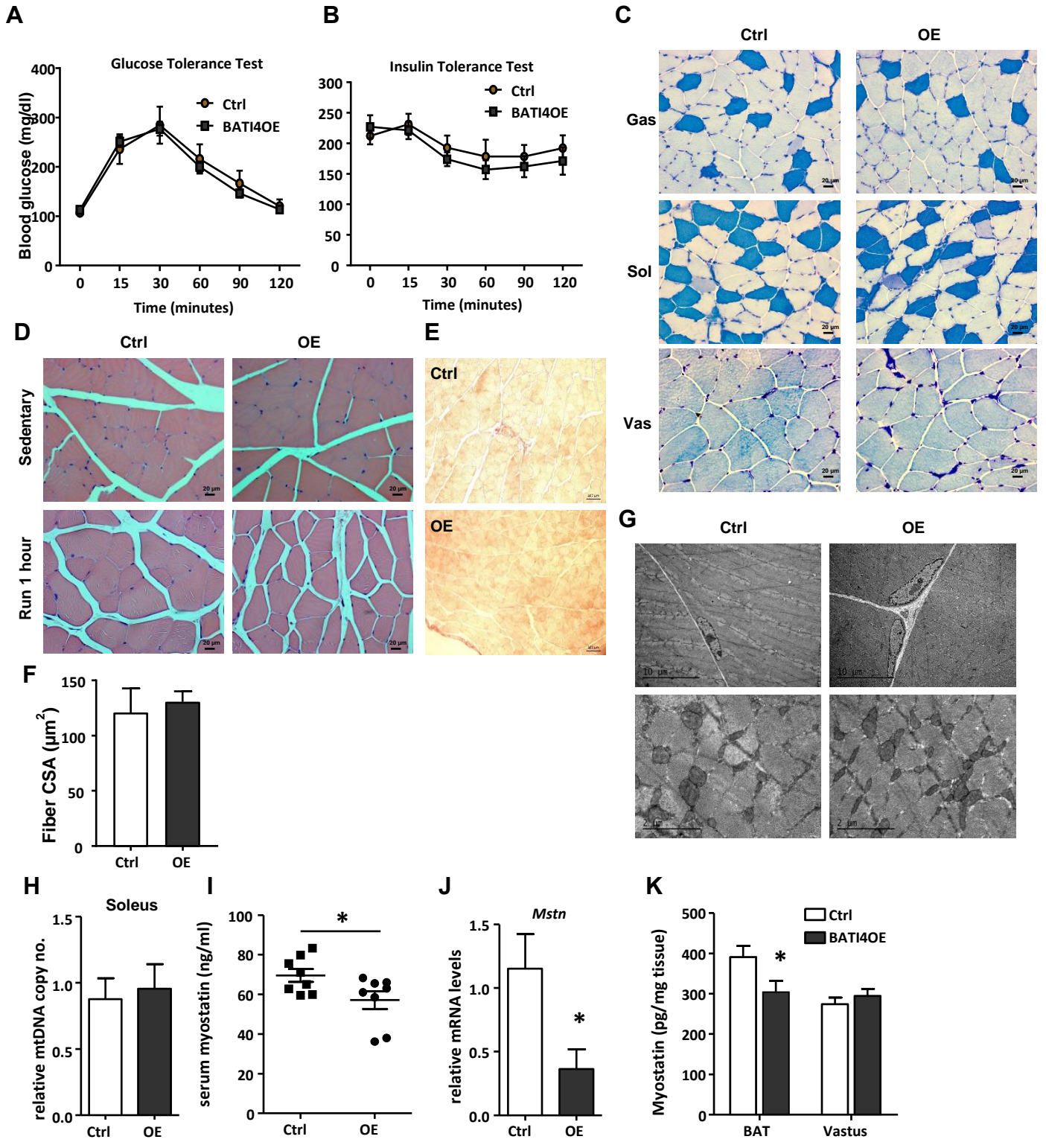
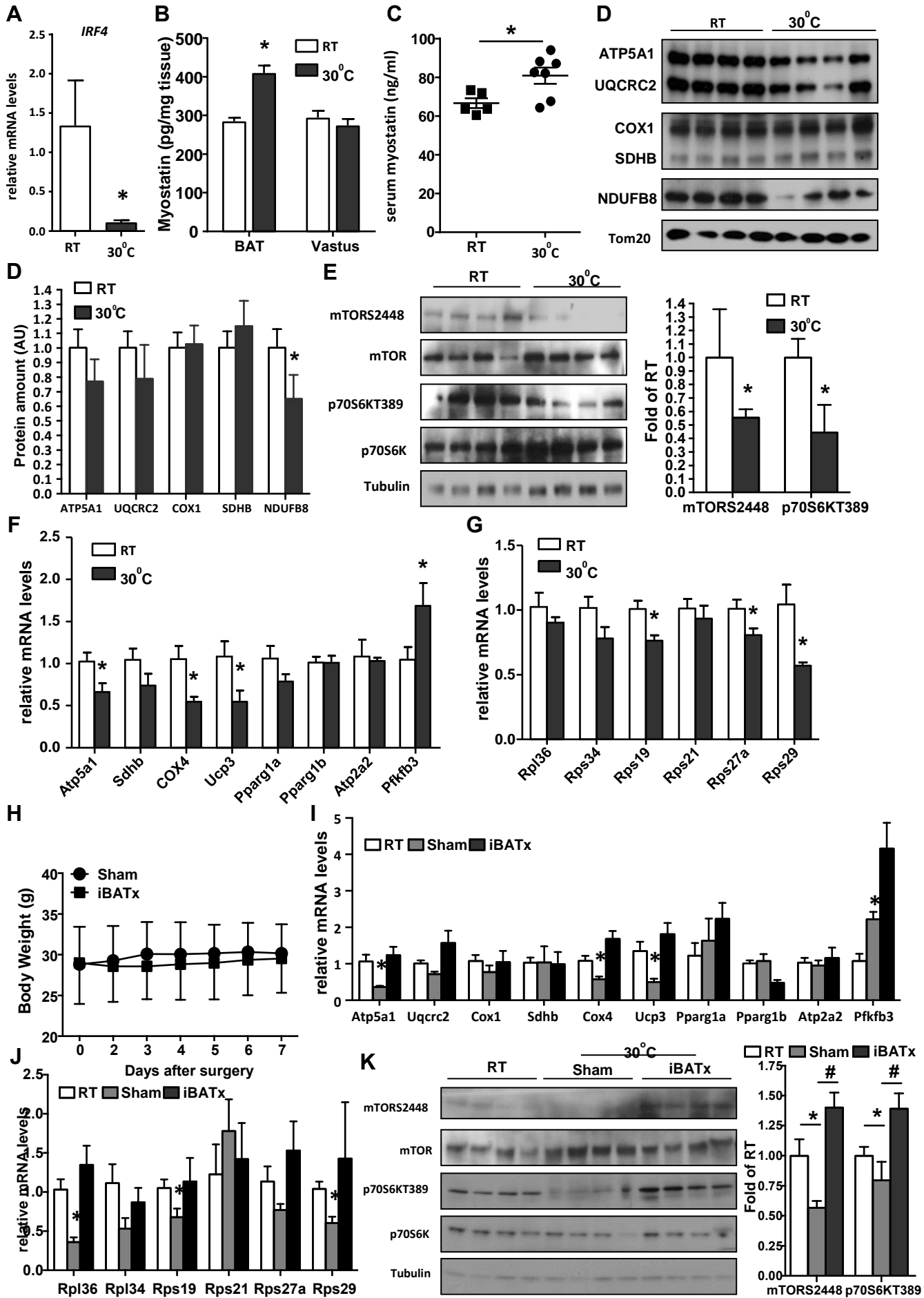


Figure S5: (Related to Figure 6)



**Increased exercise capacity following myostatin neutralization or IRF4 overexpression.** (A) Glucose tolerance test in 12-week-old BATI4OE mice on chow diet after intraperitoneal injection of glucose (n = 8). (B) Insulin tolerance test in 14-week-old BATI4OE mice after intraperitoneal injection of insulin (n=8). (C) ATPase staining of muscle sections from BATI4OE and control mice (type I: dark blue; type IIa: white; type IIb: light blue). (D) H&E staining of vastus lateralis from BATI4OE vs. control mice (40X). (E) Sirius Red staining of vastus muscle from BATI4KO and control mice (10X). (F) Muscle fiber cross-sectional area. Values are means  $\pm$  SEM. (G) Electron micrograph of vastus lateralis from BATI4OE and control mice. (H) Mitochondrial DNA number in soleus of BATI4OE and control mice (n=4, 5). Serum myostatin levels (I) and tissue myostatin levels (K) in BATI4OE group were measured by ELISA kits (n=8, \* $p$ <0.05). (J) Q-PCR analysis of *Mstn* mRNA expression in BAT (n=5-6, \* $p$ <0.05) of BATI4OE and control mice.



**Figure S6: (Related to Figure 7)**

**Warming causes myostatin elevation and myopathy in a BAT-dependent manner. (A)** *Irf4* mRNA levels in BAT from mice at RT and 30°C. Tissue myostatin levels **(B)** and serum myostatin levels **(C)** were measured by ELISA kits. (n=5-8, \* $p < 0.05$ ). **(D)** Western blot analysis of isolated mitochondria from vastus of WT mice at RT and 30°C. Protein amount was quantified using Image J (n=4, \* $p < 0.05$  vs. RT mice). **(E)** Western blot analysis of mTOR signaling pathway in vastus of WT mice at RT and 30°C. Activity was quantified using Image J. (n=4, \* $p < 0.05$  vs. RT mice). **(F)** Q-PCR analysis of vastus gene expression (n=5-6, \* $p < 0.05$ ). **(G)** Q-PCR analysis of vastus ribosomal gene expression (n=5-6, \* $p < 0.05$ ). **(H)** Body weight (BW) of iBATx and sham-operated mice after surgery. **(I)** Q-PCR analysis of vastus gene expression before and after iBATx and exposure to 30°C (n=5-6, \* $p < 0.05$ ). **(J)** Q-PCR analysis of vastus ribosomal gene expression before and after iBATx and exposure to 30°C (n=5-6, \* $p < 0.05$ ). **(K)** Western blot analysis of mTOR signaling pathway in vastus from mice before and after iBATx and exposure to 30°C. Activity was quantified using Image J (n=4, \* $p < 0.05$  vs. Flox mice. # $p < 0.05$  vs. iBATx mice).

**Table S3: (Related to STAR Methods) Primers used for qRT-PCR**

Gene	Forward	Reverse
Atp2a2	GAGAACGCTCACACAAAGACC	CAATTCGTTGGAGCCCCAT
Pfkfb3	CCCAGAGCCGGGTACAGAA	GGGGAGTTGGTCAGCTTCG
Irf4	CAGGACTACAATCGTGAGGAGG	GCACATCGTAATCTTGTCTTCCA
Atp5a1	TCTCCATGCCTCTAACACTCG	CCAGGTCAACAGACGTGTGTCAG
Uqcrc2	AAAGTTGCCCGAAGGTAAA	GAGCATAGTTTTCCAGAGAAGCA
COX1	GGATTTGTTCACTGATTCCCATTA	GCATCTGGGTAGTCTGAGTAGCG
SDHB	AATTTGCCATTTACCGATGGGA	AGCATCCAACACCATAGGTCC
COX4	GTTTCAGTTGTACCGCATCCA	TTGTCATAGTCCCACTTGGC
UCP3	TTTCTGCGTCTGGGAGCTT	GGCCCTCTTCAGTTGCTCAT
Pparg1a	GGACATGTGCAGCCAAGACTCT	CACTTCAATCCACCCAGAAAGCT
Pparg1b	GGCAGGTTCAACCCCGA	CTTGCTAACATCACAGAGGATATCTTG
Rpl36	ATGGTCAACGTACCAAAAACCC	TGGGCATACAGGGAATCCTTG
Rpl34	TCGGTGGCCCTATTGAGTG	GCTCGGCTGATACTCGTTTCC
Rps19	GCCTCCAGGCTCTGAAAC	GCTTTGAGGTGGTCTCGACA
Rps21	GTCCATCCAGATGAACGTGG	CCATCAGCCTTAGCCAATCGG
Rps27a	GACCCTTACGGGGAAAACCAT	AGACAAAGTCCGGCCATCTTC
Rps29	GTCTGATCCGCAAATACGGG	AGCCTATGTCTTCGCGTACT
Mstn	AGTGGATCTAAATGAGGGCAGT	GTTTCCAGGCGCAGCTTAC
Actn3	AACAGCAGCGGAAAACCTTCA	GGCTTTATTGACATTGGCGATTT
Asb18	GACTCGCCCCCAGATTACC	TCAGTGCAGATCAGCTTCTGTA
Cacng6	AGAACAGCAGAATCTCGGCTT	CCTGCCACAGTCGCTTGAT
Myod1	CCACTCCGGGACATAGACTTG	AAAAGCGCAGGTCTGGTGAG
Myh8	AACAGAAACGCAATGCTGAGG	TCGCCTGTAATTTGTCCACCA
Tnnt1	CCTGTGGTGCCTCCTTTGATT	TGCGGTCTTTTAGTGCAATGAG
TBP	CCCCTTGTACCCTTCACCAAT	GAAGCTGCGGTACAATTCCAG
36B4	GAGGAATCAGATGAGGATATGGGA	AAGCAGGCTGACTTGGTTGC