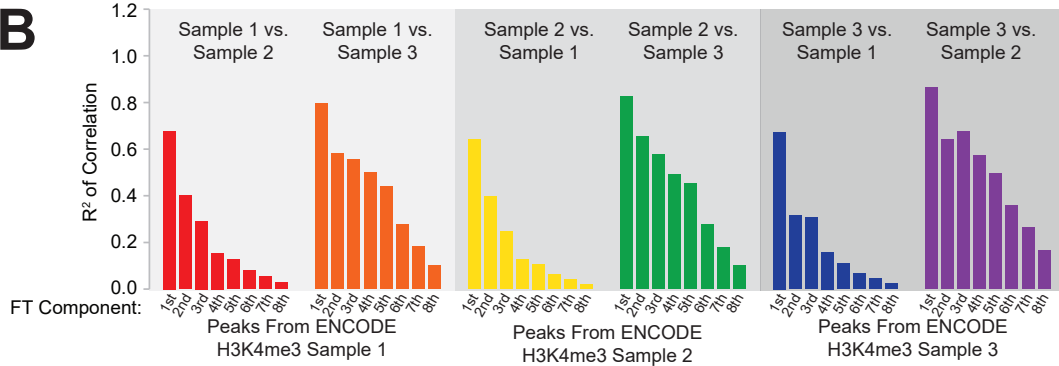


Figure S1

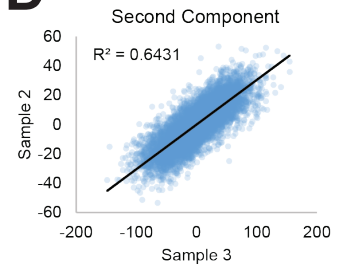
A

	EMD-07-473 ENCSR000AKU	ab8580 ENCSR668LDD	CST-9751S ENCSR000EWA	CST-9751S ENCSR000DWD
Replicated peaks	43,677	21,402	22,884	30,800
Replicated peaks intersecting replicated peaks from all other ENCODE H3K4me3 Datasets in K562	22,455 (51.4%)	17,230 (80.5%)	18,563 (81.1%)	20,248 (65.7%)

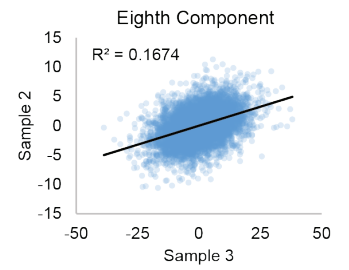
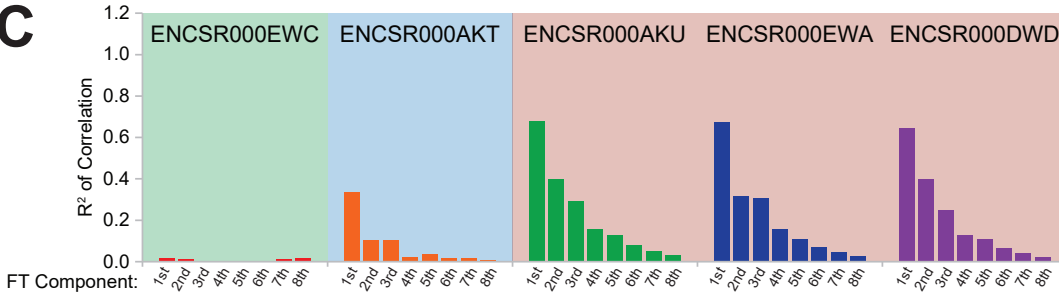
B



D



C



E

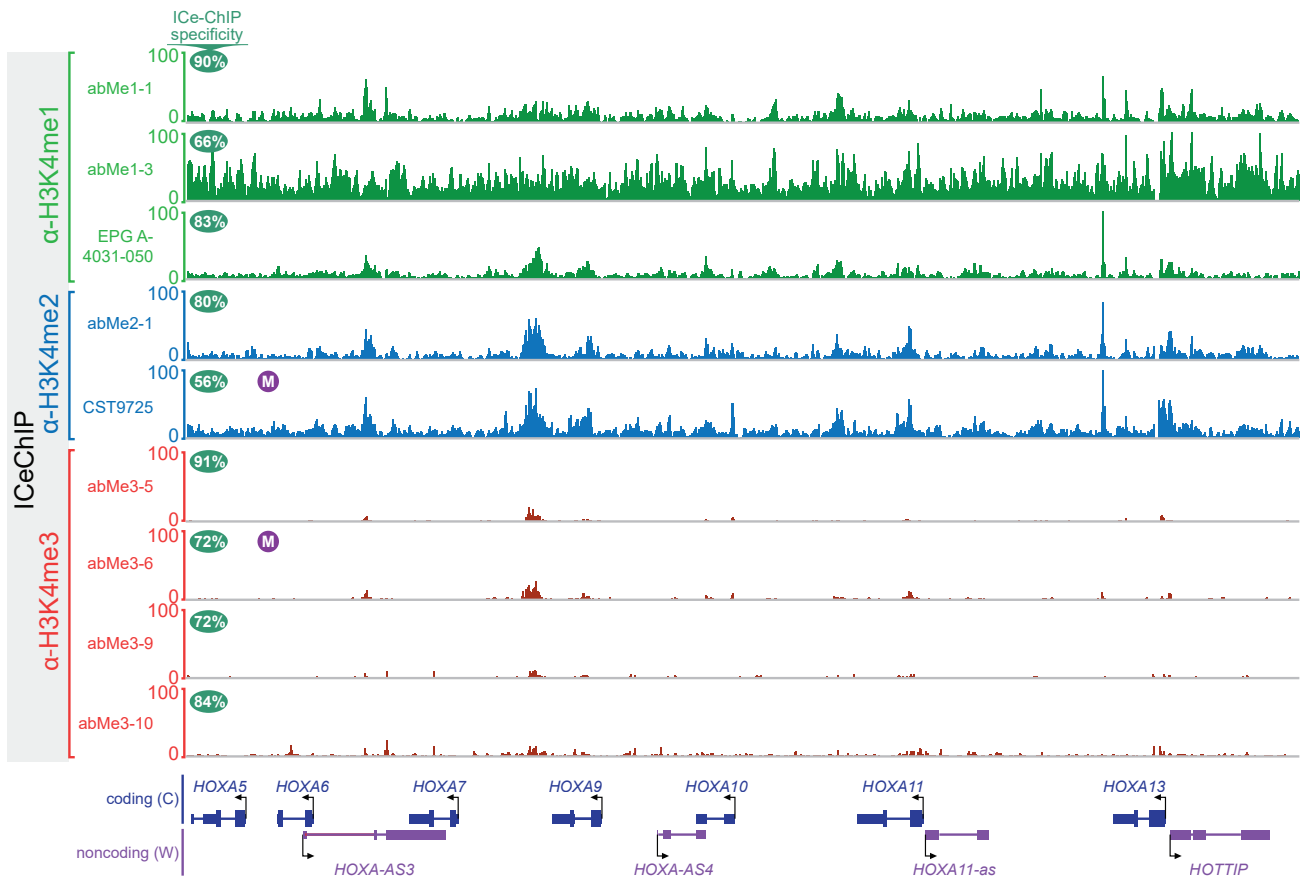


Figure S2

■ H3K4me0
 ■ H3K4me1
 ■ H3K4me2
 ■ H3K4me3
 ■ Signal

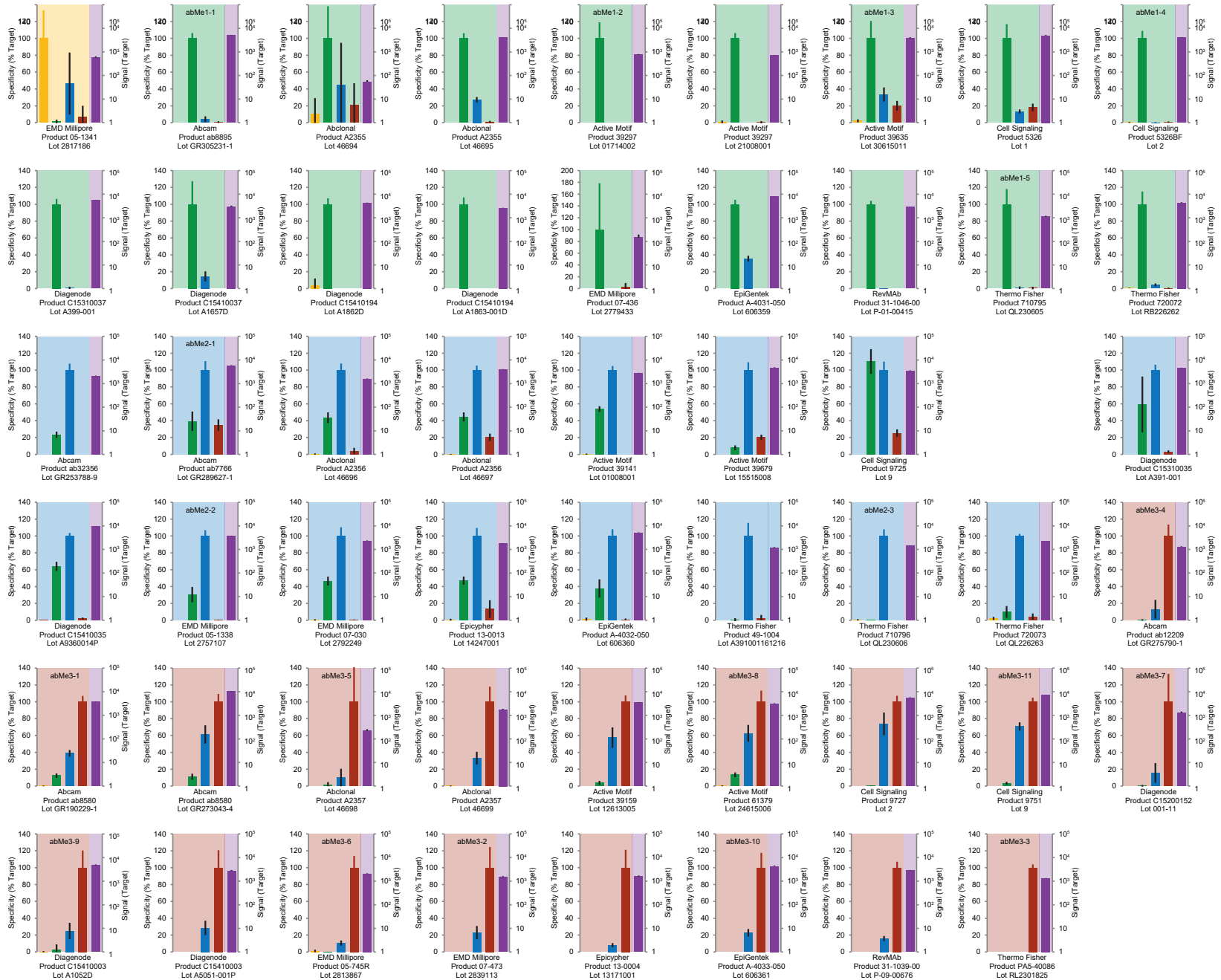


Figure S3

■ H3K4me0
 ■ H3K4me1
 ■ H3K4me2
 ■ H3K4me3
 ■ Enrichment

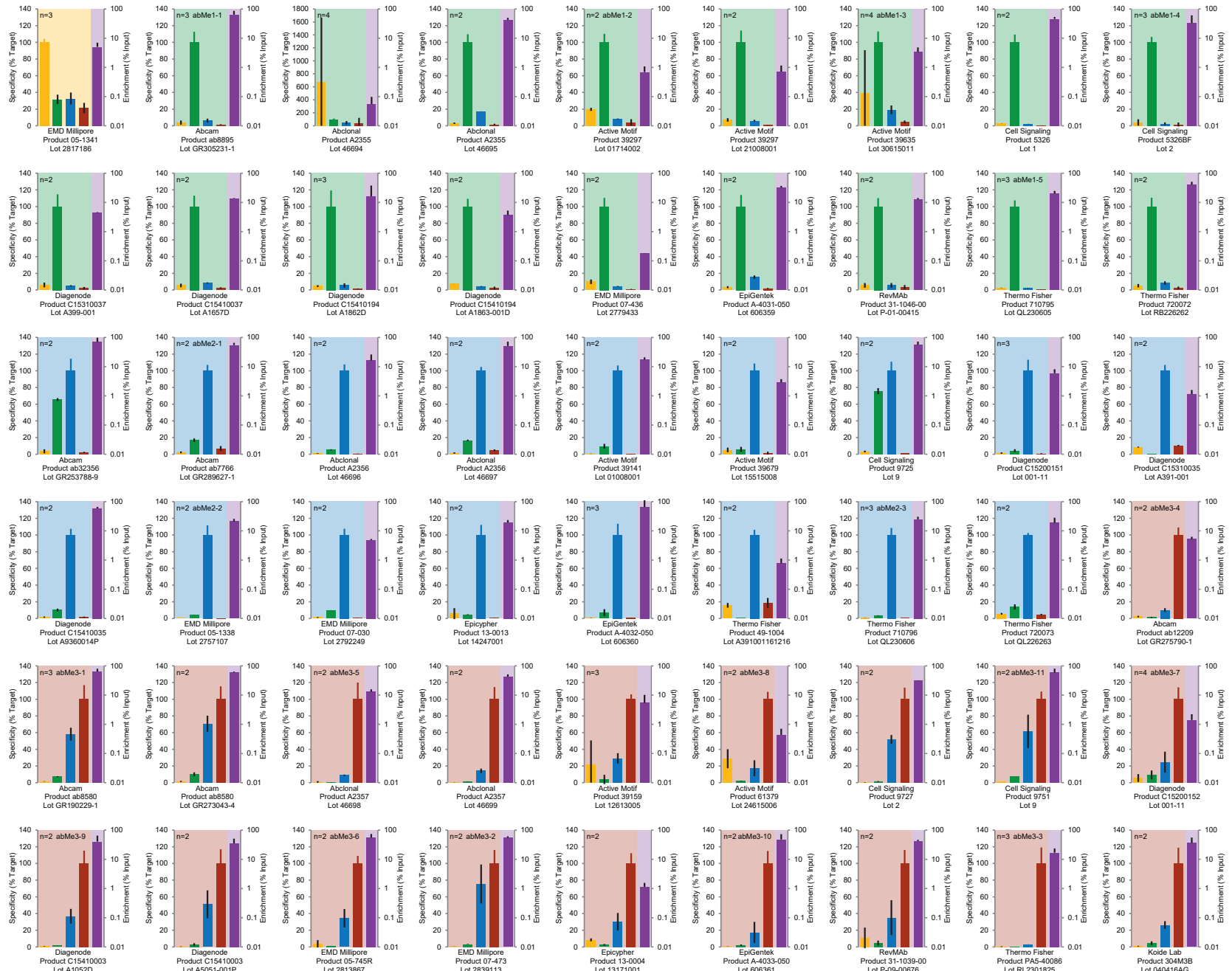
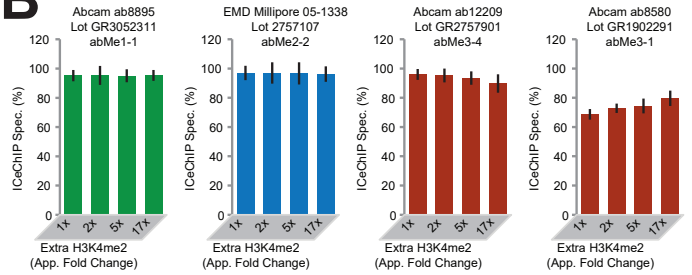


Figure S4

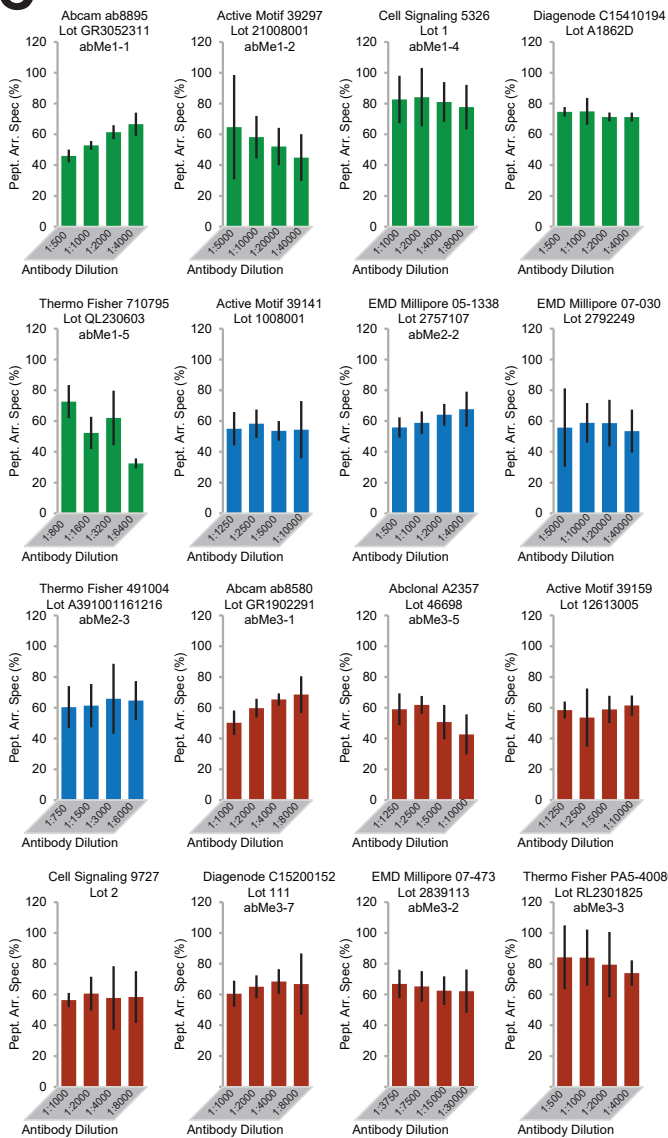
A

	ICeChIP More Specific	Approx. Equally Specific	Pept. Arrays More Specific	Total
α -H3K4me0	0	1	0	1
α -H3K4me1	1	11	5	17
α -H3K4me2	12	2	2	16
α -H3K4me3	1	11	6	18
Total	14	25	13	52

B



C



D

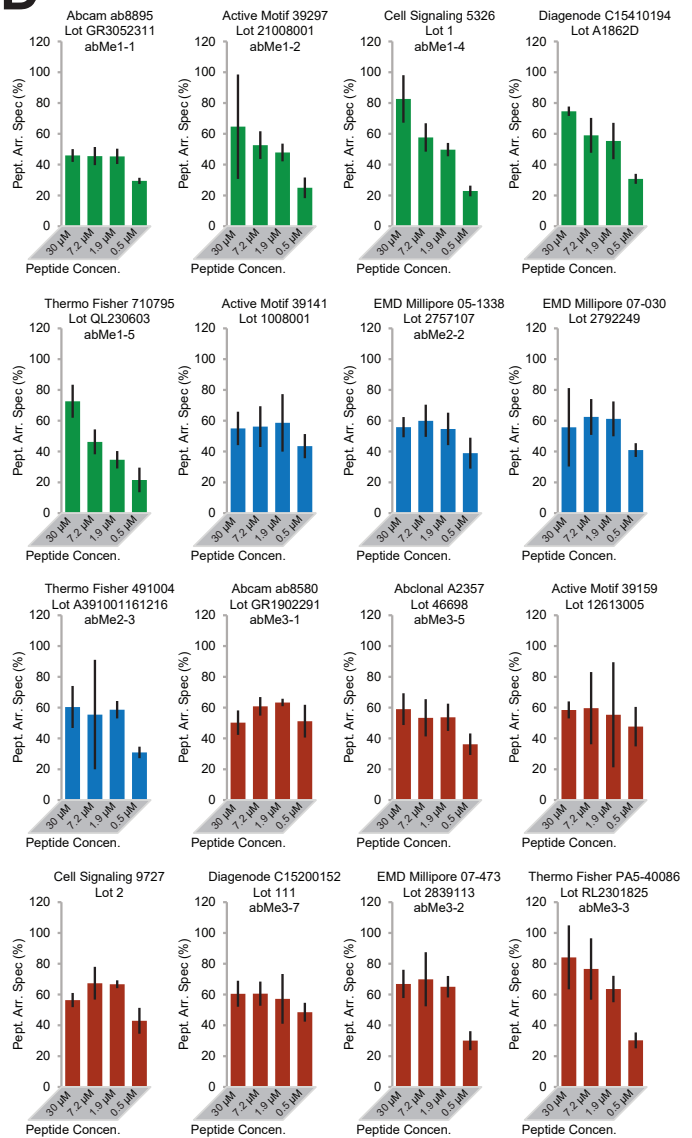


Figure S5

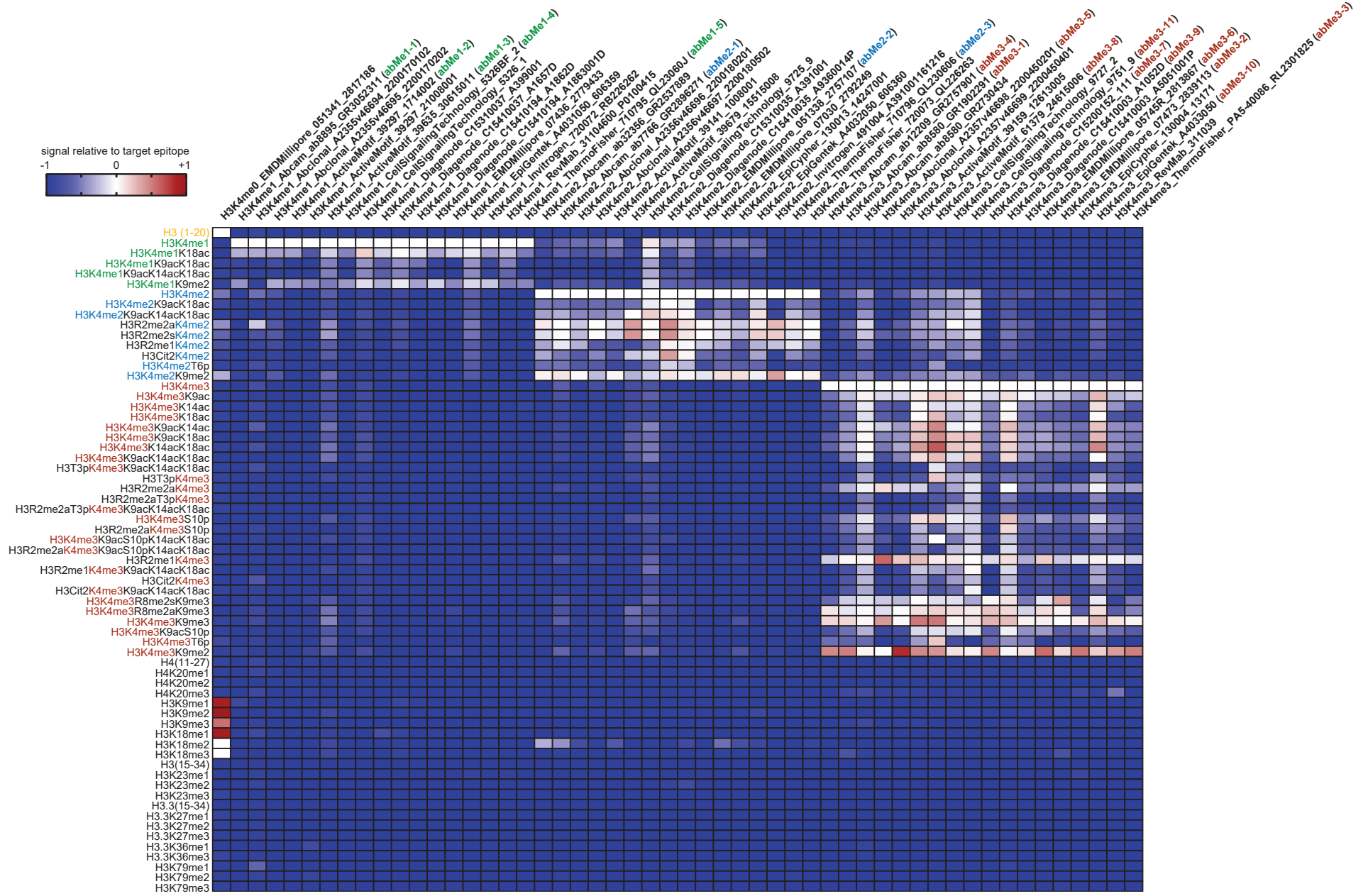
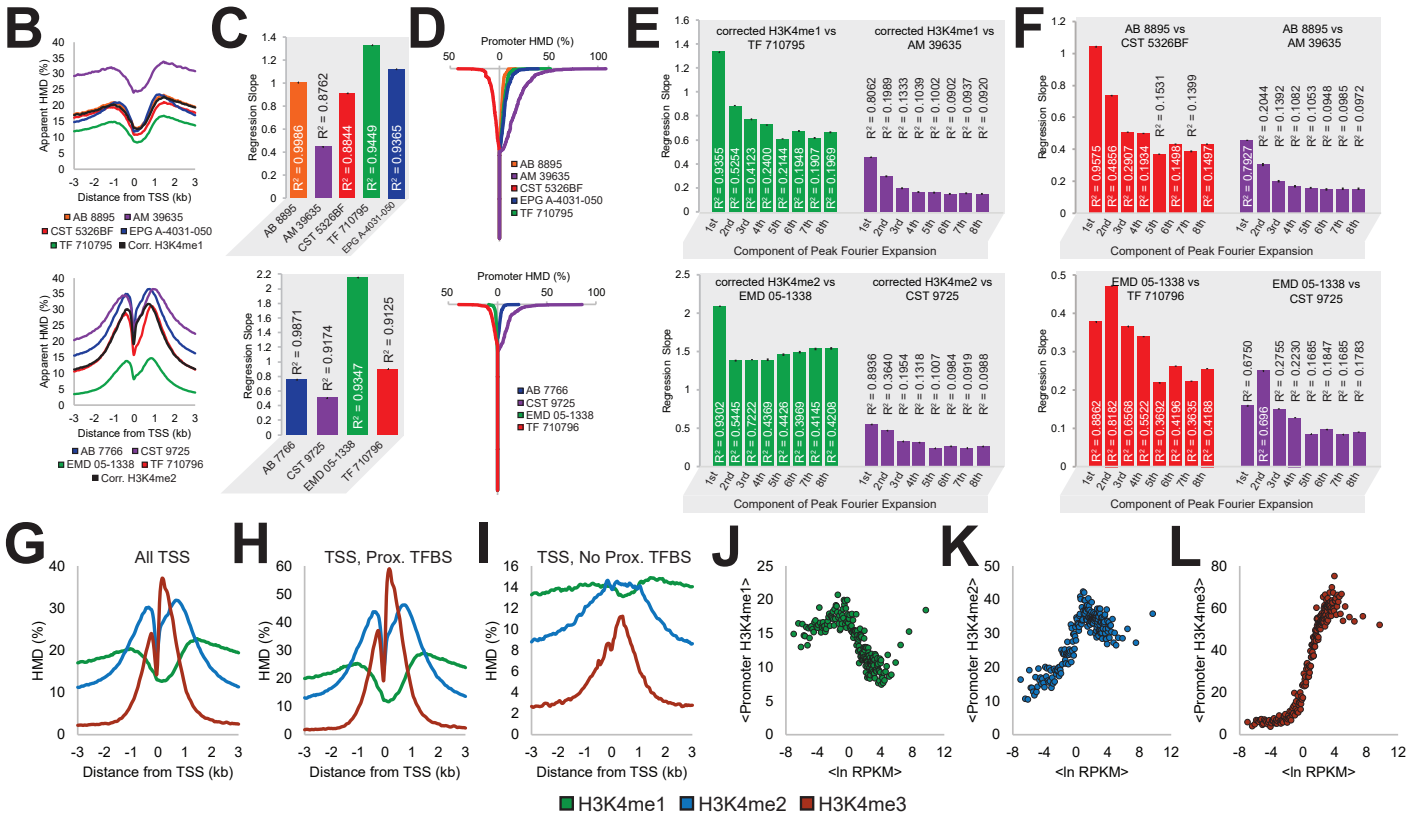
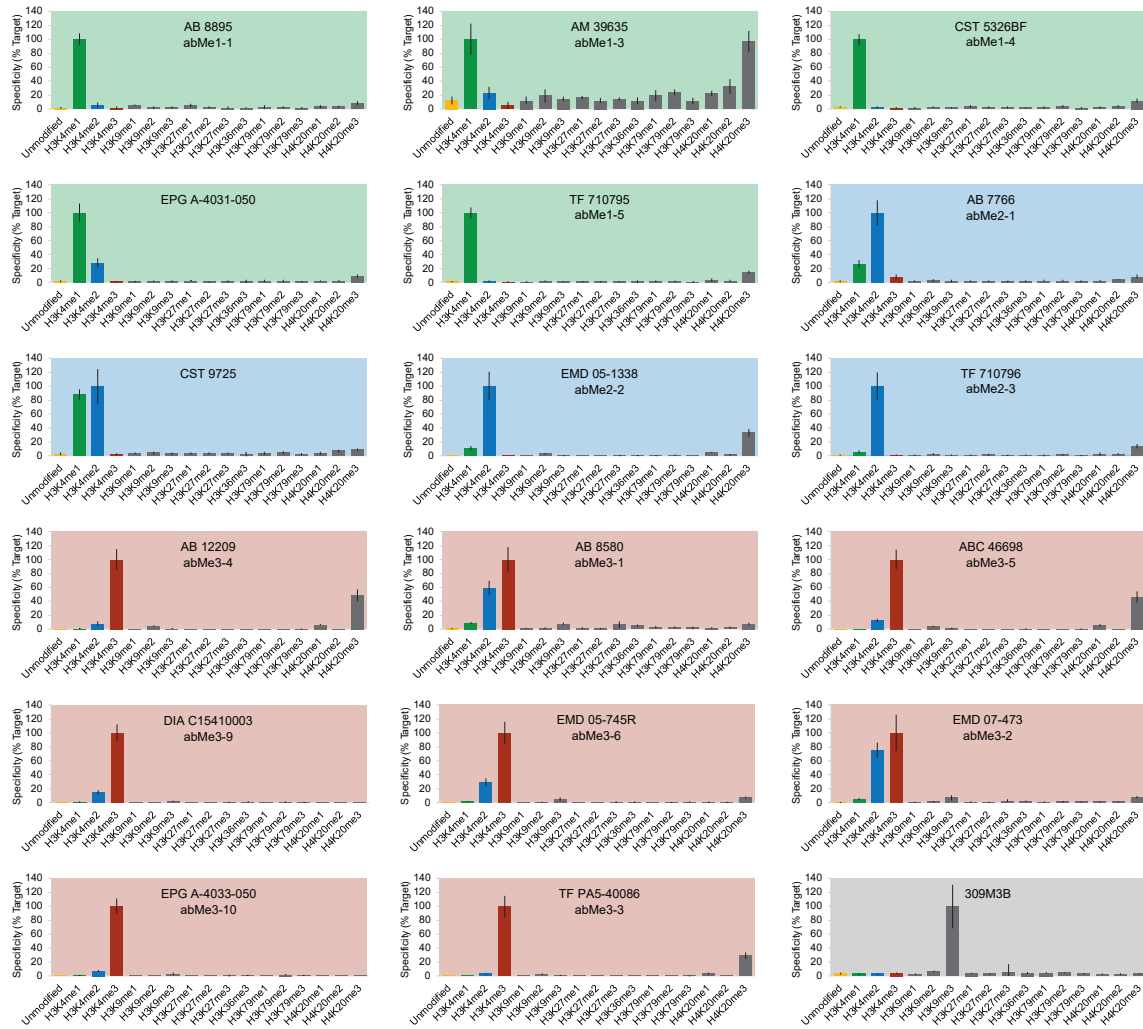


Figure S6

A



■ H3K4me1 ■ H3K4me2 ■ H3K4me3

Figure S7

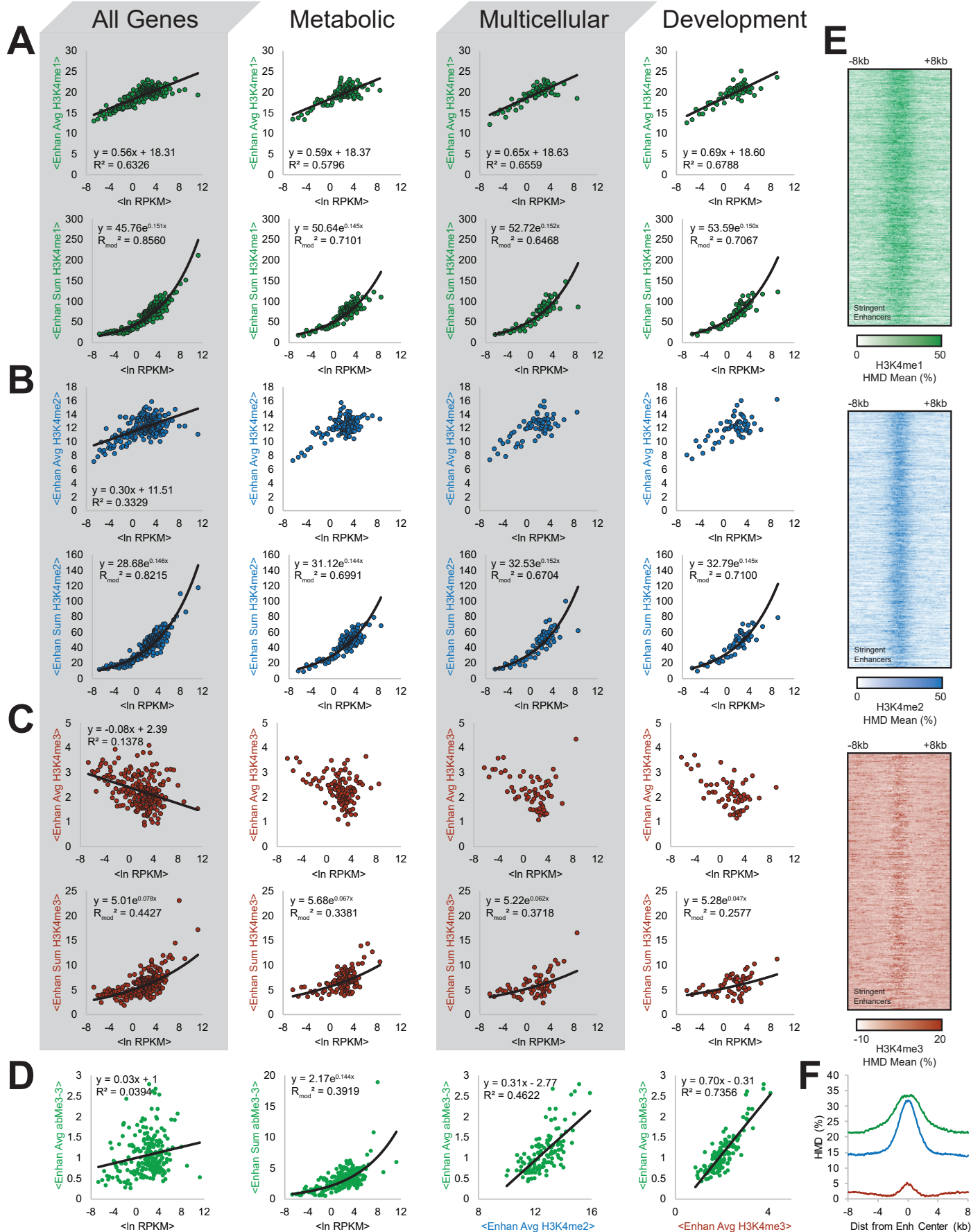


Table S1

Manufacturer	Product	Lot	Antibody Code	IP Methyl. Specificity (% H3K4 Pulldown)	PA Methyl. Specificity (% H3K4 Signal)	Target IP Efficiency (% Input)	Citations	ENCODE Status
EMD Millipore	05-1341	2453179	--	54 ± 5	64 ± 6	5 ± 2	2	--

Manufacturer	Product	Lot	Antibody Code	IP Methyl. Specificity (% H3K4 Pulldown)	PA Methyl. Specificity (% H3K4 Signal)	Target IP Efficiency (% Input)	Citations	ENCODE Status
Abcam	ab8895	GR305231-1	abMe1-1	90 ± 4	96 ± 3	64 ± 22	218	--
Abclonal	A2355	46694	--	37 ± 35	57 ± 19	0.06 ± 0.04	0	--
Abclonal	A2355	46695	--	82 ± 2	78 ± 2	43 ± 7	0	--
Active Motif	39297	01714002	abMe1-2	76 ± 1	100 ± 0	0.7 ± 0.4	11	--
Active Motif	39297	21008001	--	88 ± 2	99 ± 2	0.7 ± 0.4	11	Not Char Stds.
Active Motif	39635	30615011	abMe1-3	66 ± 17	64 ± 4	3 ± 1	1	--
Cell Signaling	5326	1	abMe1-6	94.4 ± 0.1	76 ± 2	46 ± 7	2	--
Cell Signaling	5326BF	2	abMe1-4	93 ± 2	99 ± 1	33 ± 27	2	Char. To Stds.
Diagenode	C15310037	A399-001	--	88 ± 4	99 ± 1	4.4 ± 0.2	0	--
Diagenode	C15410037	A1657D	--	86 ± 3	87 ± 5	14 ± 0.5	2	--
Diagenode	C15410194	A1862D	--	90 ± 3	97 ± 8	16 ± 22	7	Awaiting Char.
Diagenode	C15410194	A1863-001D	--	88 ± 2	100 ± 0	4 ± 2	7	--
EMD Millipore	07-436	DAM1687548	--	87 ± 1	97 ± 6	0.179 ± 0.003	16	--
EpiGentek	A-4031-050	606359	--	83 ± 3	74 ± 2	32 ± 5	0	--
RevMab	31-1046-00	P-01-00415	--	87 ± 6	99.9 ± 0.2	13 ± 1	0	--
Thermo Fisher	710795	QL230603	abMe1-5	94.7 ± 0.5	98 ± 2	20 ± 5	0	--
Thermo Fisher	720072	RB226262	--	86 ± 4	95 ± 2	40 ± 11	0	--

Manufacturer	Product	Lot	Antibody Code	IP Methyl. Specificity (% H3K4 Pulldown)	PA Methyl. Specificity (% H3K4 Signal)	Target IP Efficiency (% Input)	Citations	ENCODE Status
Abcam	ab32356	GR253788-9	--	58 ± 2	81 ± 2	70 ± 29	35	--
Abcam	ab7766	GR289627-1	abMe2-1	80 ± 4	57 ± 4	55 ± 11	55	--
Abclonal	A2356	46696	--	94 ± 1	68 ± 3	16 ± 10	0	--
Abclonal	A2356	46697	--	81.3 ± 0.7	61 ± 3	51 ± 21	0	--
Active Motif	39141	01008001	--	91 ± 2	65 ± 1	18 ± 3	8	Char. To Stds.
Active Motif	39679	15515008	--	89 ± 1	78 ± 3	3 ± 1	0	--
Cell Signaling	9725	9	--	56 ± 1	42 ± 3	56 ± 16	4	--
Diagenode	C15200151	001-11	--	94 ± 2	----	6 ± 2	0	--
Diagenode	C15310035	A391-001	--	83.8 ± 0.2	62 ± 13	1.1 ± 0.4	0	--
Diagenode	C15410035	A9360014P	--	88 ± 2	60 ± 2	58 ± 8	6	--
EMD Millipore	05-1338	2757107	abMe2-2	95.6 ± 0.1	77 ± 5	21 ± 5	6	--
EMD Millipore	07-030	DAM1479603	--	89.8 ± 0.2	68 ± 2	4.8 ± 0.4	126	--
Epicypther	13-0013	14247001	--	90 ± 5	62 ± 4	19 ± 4	1	--
EpiGentek	A-4032-050	606360	--	92 ± 4	72 ± 6	66 ± 45	0	--
Thermo Fisher	49-1004	A391001161216	--	74 ± 2	97 ± 4	0.8 ± 0.4	2	--
Thermo Fisher	710796	QL230606	abMe2-3	95.3 ± 0.4	99.5 ± 0.8	25 ± 5	0	--
Thermo Fisher	720073	QL226263	--	81 ± 1	86 ± 6	19 ± 9	0	--

Manufacturer	Product	Lot	Antibody Code	IP Methyl. Specificity (% H3K4 Pulldown)	PA Methyl. Specificity (% H3K4 Signal)	Target IP Efficiency (% Input)	Citations	ENCODE Status
Abcam	ab12209	GR275790-1	abMe3-4	88 ± 3	88 ± 9	5 ± 1	6	--
Abcam	ab8580	GR190229-1	abMe3-1	60 ± 3	66 ± 2	63 ± 17	418	Char. To Stds.
Abcam	ab8580	GR273043-4	--	55 ± 4	58 ± 4	59 ± 4	418	--
Abclonal	A2357	46698	abMe3-5	91 ± 2	90 ± 9	13 ± 3	0	--
Abclonal	A2357	46699	--	86 ± 2	75 ± 4	43 ± 8	0	--
Active Motif	39159	12613005	--	66 ± 11	62 ± 5	5 ± 5	80	--
Active Motif	61379	24615006	abMe3-8	67 ± 1	57 ± 3	0.4 ± 0.3	2	--
Cell Signaling	9727	2	--	65 ± 3	58 ± 4	31 ± 0.7	11	Char. To Stds.
Cell Signaling	9751	9	abMe3-11	59 ± 7	57 ± 2	59 ± 22	24	--
Diagenode	C15200152	001-11	abMe3-7	73 ± 12	86 ± 9	1.4 ± 0.8	0	--
Diagenode	C15410003	A1052D	abMe3-9	72 ± 5	78 ± 7	40 ± 24	43	--
Diagenode	C15410003	A5051-001P	--	65 ± 8	78 ± 5	35 ± 16	43	Char. To Stds.
EMD Millipore	05-745R	2813867	abMe3-6	72 ± 8	89 ± 3	55 ± 18	10	--
EMD Millipore	07-473	DAM1623866	abMe3-2	56 ± 7	81 ± 5	54 ± 7	189	Char. To Stds.
Epicypther	13-0004	13171001	--	71 ± 5	93 ± 2	1.1 ± 0.4	1	--
EpiGentek	A-4033-050	606361	abMe3-10	84 ± 10	81 ± 3	48 ± 25	0	--
RevMab	31-1039-00	P-09-00676	--	67 ± 5	86 ± 2	42 ± 5	0	--
Thermo Fisher	PA5-40086	RL2301825	abMe3-3	96.5 ± 0.5	100 ± 0	17 ± 7	0	--
Koide Lab	304M3B	040416AG	--	76 ± 1	----	36 ± 19	0	--

Table S2

Workflow Step	Details and Quality Control
<p>Isolate nuclei ↓ Spike in nucleosomes ↓ Mnase digest ↓ Antibody IP ↓ Purify DNA ↓ qPCR ↓ STOP/GO ↓ NGS</p>	<p>Cells and nuclei should not be cross-linked.</p> <p>Spike-ins contain a pool of on- and off-target barcoded PTMs.</p> <p>Check digestion (>95% mononucleosomes).</p> <p>Start with supplier recommended dilution, ensure bead-antibody complexes are saturated. In rare cases, antibody dilutions may improve specificity.</p> <p>DNA purification with size selection (e.g. Ampure XP beads) can be used to enrich mononucleosome-sized fragments.</p> <p>Phase I: Check the most likely cross-reacting species (e.g. me1, me2, me3 at the same site).</p> <p>Phase II: Check all barcoded nucleosomes in the spike-in panel.</p> <ul style="list-style-type: none">• Does the antibody exhibit specificity for the target?• Is antibody efficiency sufficiently high to minimize noise?• Is the variability between samples within an acceptable range? <p>Double check antibody specificity: All barcodes are captured in the sequencing data.</p> <p>Normalize the data: Express read counts relative to spike-in control.</p>

Figure S1, related to Figure 1. ENCODE ChIP-seq datasets display incongruity between each other and ICeChIP-seq datasets. (A) Concordance of replicated peaks for each H3K4me3 ENCODE dataset with indicated antibody in K562 cells. Top row shows number of called peaks replicated across the two biological replicates for each dataset, and bottom row the number and percentage of such peaks that intersect with peaks common to all four of the ENCODE H3K4me3 datasets. (B) Peak shape and intensity analysis by pairwise correlation between pre-cosinusoidal factors of eight-component discrete Fourier cosine transform of fold changes over control about peaks from ENCODE H3K4me3 Sample 1 (ENCSR000AKU, left), Sample 2 (ENCSR000EWA, centre), or Sample 3 (ENCSR000DWD, right). If only intensity of signal was different (i.e. same data with different scaling, the R^2 would approach 1, and scalar factor reflecting difference would be apparent from the slope. These comparisons (Movie S1) indicate very limited similarity amongst any of two ENCODE data sets, with modest large-scale similarity (early terms) decaying to negligible fine-scale similarity (late terms). (C) R^2 of correlation between pre-cosinusoidal factors of eight-component discrete Fourier cosine transform on antibody-measured peaks of corrected ICeChIP-seq HMD versus antibody-measured fold change over control for H3K4me1, H3K4me2, and H3K4me3 ENCODE ChIP-seq datasets in K562 cells. (D) Example scatterplots showing high correlation (top) and low correlation (bottom) of Fourier Transform components on peaks from ENCODE H3K4me3 Sample 3. (E) Genome browser view at the HOX locus for antibodies not shown in Figure 1A. Green circle represents ICeChIP aggregate specificity, and purple circle with M indicates monoclonality.

Figure S2, related to Figure 2. Anti-H3K4 methylation antibodies display a broad range of specificities in peptide arrays. The specificity of H3K4 methylform antibody binding on peptide arrays expressed relative to on-target capture. Black error bars represent SD of off-target specificity; colored error bars represent average standard error of on-target signal. Purple bar represents raw fluorescence signal from secondary axis and maps onto secondary axis. Fluorescence measurements for each antibody ($n=6$), independent at the level of spotting, but simultaneously measured against one antibody dilution.

Figure S3, related to Figure 2. Anti-H3K4 methylation antibodies display a broad range of specificities in ICeChIP. The specificity of H3K4 methylform antibody binding in ICeChIP relative to on-target capture. Black error bars represent SD of off-target specificity; colored error bars represent average standard error of on-target signal. Purple bar represents ChIP enrichment and maps onto secondary axis (right). ICeChIP was conducted with 3 μg of mammalian chromatin and 3 μg of each antibody (see Methods). Enrichment of each standard was measured by qPCR (compare to ICeChIP-seq in Fig. S6A); n represents independent ICeChIP experiments averaged for each antibody.

Figure S4, related to Figure 3. ICeChIP and peptide arrays have discrepancies that can be modulated. (A) Agreement of antibody of methyl-form specificity between ICeChIP and peptide arrays, to within 10 percentage points. (B) ICeChIP aggregate specificity for four antibodies when H3K4me2 nucleosomes bearing a different DNA sequence is added in excess of endogenous H3K4me2. One replicate per ICeChIP experiment for a total of four ICeChIPs per antibody. (C, D) Antibody specificity on EpiTitrator peptide arrays with varying amounts of (C) antibody and (D) modified peptide. Approximately half the antibodies tested show marginally altered specificity with increasing dilution, albeit not always in the same direction. Most antibodies show decreased specificity at the most dilute modified peptide concentration, and approximately a third show decreasing specificity with increased modified peptide dilution more broadly. Six fluorescence readings per peptide array experiment, with one independent experiment per antibody dilution.

Figure S5, related to Figure 3. Combinatorial modifications can impact antibody binding in peptide arrays. Heatmap of antibody binding to a wide range of combinatorial and off-target peptides on peptide arrays. Signal is normalized to singly-modified target epitope for a select set of H3K4-proximal combinatorial modifications.

Figure S6, related to Figure 4. Specificity of antibodies is broadly recapitulated in ICeChIP-seq and can impact measured modification profiles. (A) Specificities of antibodies in ICeChIP-seq ex-

periments are identical within experimental error to those measured by qPCR, and a broader range of off-target internal standards are sampled. Error bars represent SD of estimate based on internal variability of ladder members. **(B)** Metagene contours about Refseq TSSs of anti-H3K4me1 (upper panel) and anti-H3K4me2 (lower panel) antibodies and corresponding corrected profiles. **(C)** Correlation between antibody-measured HMD and corrected HMD at antibody called peaks for anti-H3K4me1 and anti-H3K4me2 antibodies. **(D)** Antibody-measured HMD at +1/+2 nucleosomes of genes with no measured HMD in corrected profile for anti-H3K4me1 and anti-H3K4me2 antibodies. **(E)** Correlation and magnitude analysis of pre-cosinusoidal factors for eight term Fourier series comparing corrected HMD versus antibody-measured HMD for anti-H3K4me1 and anti-H3K4me2 antibodies contoured over called peaks in the corrected. **(F)** Similar analysis of pre-cosinusoidal factors of eight-component discrete Fourier cosine transform of measured HMDs by listed antibodies versus high-specificity reference antibodies abMe1-1 (upper panel) or abMe2-2 (lower panel) on peaks from listed antibodies. **(G, H, I)** Metacontours about TSSs of H3K4me1/2/3 HMD for (G) all TSSs (58,951 TSSs), (H) TSSs with a transcription factor binding site (Wang et al. 2014) within 200bp of the TSS (32,531 TSSs), and (I) TSSs without a transcription factor binding site (Wang et al. 2014) within 200bp of the TSS (26,420 TSSs). **(J, K, L)** Average (J) H3K4me1, (K) H3K4me2, and (L) H3K4me3 corrected HMD of +1/+2 nucleosomes versus ln RPKM of genes. Error bars for all correlations represent 99.99% CI for correlation slope.

Figure S7, related to Figure 5. High-quality H3K4 methylation HMD datasets reveal quantitative relationships between enhancer H3K4 methylation and promoter activity. **(A, B, C)** Average (top) and sum (bottom) of (A) H3K4me1, (B) H3K4me2, and (C) H3K4me3 corrected HMD across all enhancers contacting corresponding promoter regions versus ln RPKM for all classes of genes (left), metabolic genes (centre-left), multicellular system process genes (centre-right), and developmental process genes (right). **(D)** From left to right: abMe3-3 measured HMD sum versus ln RPKM, abMe3-3 measured HMD average across enhancers versus ln RPKM, average corrected H3K4me2 enhancer HMD, and average corrected H3K4me3 enhancer HMD. **(E)** Heatmaps of stringently defined enhancer HMD averages for H3K4me1, H3K4me2, and H3K4me3. All heatmaps sorted by ln RPKM of target genes. R_{mod}^2 represents R^2 of linear correlation between actual and predicted/modeled HMD. **(F)** Signal-corrected H3K4me1, H3K4me2, and H3K4me3 modification profiles contoured over stringently-defined enhancers.

Table S1. Summary of Data. Antibody information with ICeChIP and peptide array specificity, IP efficiency, ChIP citations from Google Scholar, and ENCODE validation status (none, awaiting characterization, or characterized to standards). Values represent average \pm SD.

Table S2. Workflow of antibody validation for ICeChIP-seq. Pipeline offers suggested procedure for conducting ICeChIP-seq, including pause points for assessment of antibody quality prior to NGS or biological interpretation.