

Construct	Cas13d binary complex	Cas13d ternary complex	Cas13d apo form
Data collection and Image Processing			
Microscope	Arctica Talos	Titan Krios	Arctica Talos
Voltage (kV)	200	300	200
Camera	Gatan K2 Summit	Gatan K2 Summit	Gatan K2 Summit
Defocus range (μm)	0.5-2.0	1.0-3.0	0.8-2.5
Defocus mean \pm std (μm)	1.1 \pm 0.3	2.0 \pm 0.4	1.6 \pm 0.5
Exposure time (s)	12	12	12
Dose rate (e $^-$ /pixel/s)	2.5	3.0	2.5
Total dose (e $^-$ / \AA^2)	56.8	57.3	56.8
Pixel size (\AA)	0.73	0.79	0.73
Exposure rate (ms)	100	200	100
Number of micrographs	1435	2205	1158
Number of particles (processed)	412591	684169	330986
Number of particles (3D classification & refinement)	207849	177910	154889
Number of particles (in final map)	43786	51885	15846
Symmetry	C1	C1	C1
Resolution (global) (\AA)*	3.4	3.3	6.5
Resolution range (homogeneous portion) (\AA)	3.0-5.0	3.0-5.0	6.0-8.0
Map sharpening	Spectral flattening between 8 \AA and 3.4 \AA	Spectral flattening between 8 \AA and 3.3 \AA	Spectral flattening between 10 \AA and 6.5 \AA
Sphericity @ threshold 0.75	0.98	0.89	0.97
Model refinement			
Number of atoms (modeled)	8163	8834	
Protein			
Favored rotamers (%)	97.4	94.6	
Poor rotamers (%)	0.26	0.0	
Ramachandran favored (%)	95.9	91.9	
Ramachandran outliers (%)	0.0	0.0	
C β deviations >0.25 \AA (%)	0.0	0.0	
Cis prolines (%)	0.0	0.0	
Cis prolines (%)	0.0	0.0	
Bad bonds (%)	0.0	0.0	
Bad angles (%)	0.0	0.0	
Nucleic acid			
Probably wrong sugar pockers (%)	0.0	0.0	
Bad backbone conformations (%)	11.8	12.7	
Bad bonds (%)	0.0	0.0	
Bad angles (%)	0.0	0.0	

*Resolution assessment based on frequency-limited refinement using the 0.143 threshold for resolution analysis

Supplementary Table 1. Cryo-EM data collection, image processing, and modeling summary.
Related to Figures 1 and 6

Name	Sequence (5' to 3')	Figure
ternary target	ACGUUUUGAUCUGAAAUUAUUCAGGUCUAUA	1E-G, 2
ssRNA-1 target 1	UACGUACGCCAUCUCAUCCUGCGUGUCUCCUACGAUUUG AUCUGAAAUUAAGAGGUUCGUACAUCACCGACUGCCAU AGAGAGGUUAUCCGCUCACAAUCCACACAACAUACGAGC CGGAAGCAUAAAG	3F
ssRNA-2 target 2	UACGUACGCCAUCUCAUCCUGCGUGUCUCCUACGAUUUG AUCUGAAAUUAUCAGGUUCGUACAUCACCGACUGCCAU AGAGAGGUUAUCCGCUCACAAUCCACACAACAUACGAGC CGGAAGCAUAAAG	3G, S3G
competitor 1 - WT	UCCUUUUGAUCUGAAAUUAUUCAGGUCUCCU	4D-E
competitor 1 - MM1	UCCUAAAACAUCUGAAAUUAUUCAGGUUCUCCU	4D-E
competitor 1 - MM2	UCCUUUUGAAGACAAAUUAUUCAGGUUCUCCU	4D-E
competitor 1 - MM3	UCCUUUUGAUCUGUUUAUUCAGGUUCUCCU	4D-E
competitor 1 - MM4	UCCUUUUGAUCUGAAAUUAAGAGGUUCUCCU	4D-E
competitor 1 - MM5	UCCUUUUGAUCUGAAAUAUUCACCAGUUCUCCU	4D-E
competitor 2 - WT	UCCUGAGUUCAUCCAGUGUAUCCUUCUCCU	4D-E
competitor 2 - MM1	UCCUCUCAUCAUCCAGUGUAUCCUUCUCCU	4D-E
competitor 2 - MM2	UCCUGAGUUGUAGCAGUGUAUCCUUCUCCU	4D-E
competitor 2 - MM3	UCCUGAGUUCAUCGUCAUGUAUCCUUCUCCU	4D-E
competitor 2 - MM4	UCCUGAGUUCAUCCAGUCAUACCUUCUCCU	4D-E
competitor 2 - MM5	UCCUGAGUUCAUCCAGUGUAUCGAAGUUCUCCU	4D-E
competitor 3 - WT	UCCUCAAUGCGUGAGUGUGAAGAUAGUUCU	4D-E
competitor 3 - MM1	UCCUGUUAGCGUGAGUGUGAAGAUAGUUCU	4D-E
competitor 3 - MM2	UCCUCAAUGGCACAGUGUGAAGAUAGUUCU	4D-E
competitor 3 - MM3	UCCUCAAUGCGUGUCACUGAAGAUAGUUCU	4D-E
competitor 3 - MM4	UCCUCAAUGCGUGAGUGACUUGAUAGUUCU	4D-E
competitor 3 - MM5	UCCUCAAUGCGUGAGUGUGAAGUAUCCUUCUCCU	4D-E
competitor 4 - WT	UCCUCCAUGAGAGGUUAUCCGCUCAUCCU	4D-E
competitor 4 - MM1	UCCUGGUAGAGAGGUUAUCCGCUCAUCCU	4D-E
competitor 4 - MM2	UCCUCCAUCUGGUUAUCCGCUCAUCCU	4D-E
competitor 4 - MM3	UCCUCCAUGAGAGACAAUCCGCUCAUCCU	4D-E
competitor 4 - MM4	UCCUCCAUGAGAGGUUAAGGGUCAUCCU	4D-E
competitor 4 - MM5	UCCUCCAUGAGAGGUUAUCCGGAGUUCUCCU	4D-E

DR mut crRNA WT	CACCCGUGCAAAAUGCAGGGUCUAAAACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA G22A	CACCCGUGCAAAAUGCAGGGUCUAAAACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA U23A	CACCCGUGCAAAAUGCAGGGACUAAAACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA A26C	CACCCGUGCAAAAUGCAGGGUCUAAACACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA A27C	CACCCGUGCAAAAUGCAGGGUCUACAACACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA A28C	CACCCGUGCAAAAUGCAGGGUCUAACACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA A29C	CACCCGUGCAAAAUGCAGGGUCUAAACACGACCUCUUAA UUUCAGAUCAAA	3F
DR mut crRNA C30G	CACCCGUGCAAAAUGCAGGGUCUAAAAGGACCUCUUAA UUUCAGAUCAAA	3F
crRNA-1-Es	CACCCGUGCAAAAUGCAGGGUCUAAAACGACCUGAAUA UUUCAGAUCAAA	1, 2, 3G, 4D-E, S3D, S3G
crRNA-1-Ur	CACUGGUGCAAAUUAUCACUAGUCUAAAACGACCUGAAUA UUUCAGAUCAAA	3G
crRNA-1-Rff	UAGUAGUGUGAAUUUACACUACUCUAAAACGACCUGAAUA UUUCAGAUCAAA	3G
crRNA-1-Rfx	AACCCCUCACCAACUGGUUCGGGUUUGAACGACCUGAAUA UUUCAGAUCAAA	3G
crRNA-2	CACCCGUGCAAAAUGCAGGGUCUAAAACGAAGGAUACA CUGGAUGAACUC	4D-E
crRNA-3	CACCCGUGCAAAAUGCAGGGUCUAAAACCUAUCUUCAC ACUCACGCAUUG	4D-E
crRNA-4	CACCCGUGCAAAAUGCAGGGUCUAAAACUGAGCGGAUA ACCUCUCUAUGG	4D-E
pre-crRNA-1	GAACUACACCCGUGCAAAAUGCAGGGUCUAAAACGACC UGAAUAUUUUCAGAUAAAAGCAAUGAACUACACCCGUG CAAAAUGCAGGGUCUAAAACAUUCCUUAUCUAAAAGC AUAUGCUUCUGU	S3A, S3C, S3G-H

Supplementary Table 2. Oligonucleotides used in this study. Related to Figures 1-4, and S3